

1 **“Histological characteristics of HPV-associated and -independent**
2 **squamous cell carcinomas of the vulva: a study of 1594 cases”**

3 Natalia Rakislova¹, Omar Clavero², Laia Alemany^{2,3}, Adela Saco¹, Beatriz
4 Quirós², Belen Lloveras⁴, Maria Alejo⁵, Michael Pawlita⁶, Wim Quint⁷, Marta del
5 Pino⁸, Silvia de Sanjose^{2,3}, Jaume Ordi¹ on behalf of VVAP study group.

6 ¹ Department of Pathology, ISGlobal, Hospital Clínic - Universitat de Barcelona,
7 Barcelona, Spain

8 ² Unit of Infections and Cancer, Cancer Epidemiology Research Program,
9 Catalan Institute of Oncology, IDIBELL, L'Hospitalet de Llobregat, Barcelona,
10 Spain

11 ³ CIBER Epidemiologia y Salud Pública, Barcelona, Spain

12 ⁴ Department of Pathology, Hospital del Mar, Barcelona, Spain

13 ⁵ Department of Pathology, Hospital General d'Hospitalet, L'Hospitalet de
14 Llobregat, Spain

15 ⁶ Division of Molecular Diagnostics of Oncogenic Infections, Research Program
16 Infection, Inflammation and Cancer, German Cancer Research Center (DKFZ),
17 Heidelberg, Germany

18 ⁷ DDL Diagnostic Laboratory, Rijswijk, The Netherlands

19 ⁸ Institute of Gynecology, Obstetrics and Neonatology, Hospital Clínic - Institut
20 d'Investigacions Biomèdiques August Pi I Sunyer (IDIBAPS). Faculty of
21 Medicine-University of Barcelona, Spain

22 Running title:

23 HPV in vulvar cancer

24

25

26

27

28

29 **Corresponding author information:**

30

31 Jaume Ordi MD, PhD.

32 Pathology Department (CDB). Escala 3, Planta 5. Hospital Clinic. 08036 Barcelona.

33 Spain.

34 Tel. +34 93 227 54 50 / 5536

35 Fax. +34 93 227 57 17

36 e-mail: jordi@clinic.cat

37

38

39

40

41

42

43

44

45

46

47

48

49 **ABSTRACT**

50 There are at least two different etio-pathogenic pathways for the development of
51 vulvar squamous cell carcinoma (VSCC): one associated with infection by
52 human papillomavirus (HPV) and another independent of HPV. We aimed to
53 describe the histological characteristics of HPV-associated and HPV-
54 independent tumors and to determine the best strategy to identify HPV in
55 VSCC. A single paraffin block was available for review from a series of 1594
56 VSCCs. In all cases HPV DNA detection was analyzed using the
57 SPF10PCR/DEIA/LiPA25 system and p16 immunohistochemistry (IHC). A tumor
58 was considered as unquestionably HPV-associated if both HPV DNA and p16
59 IHC were positive. A tumor was considered indisputably HPV-independent if
60 both HPV DNA and p16 IHC were negative. Two groups of tumors were
61 classified as non-conclusive: 1) HPV DNA+/p16-; and 2) HPV DNA-/p16+. WHO
62 typing and a thorough histological evaluation were conducted in all cases.

63 441 tumors were HPV DNA+ with 367 cases (23.0%) being HPV DNA+/p16+.
64 These HPV DNA+/p16+ tumors were more frequently basaloid or warty
65 (49.8%), but 36.5% were of the keratinizing type. 1153 tumors were HPV DNA-,
66 with 1060 cases (66.5%) being HPV DNA-/p16-. These HPV DNA-/p16- tumors
67 were mostly keratinizing (81.2%) but were occasionally basaloid or warty
68 (5.2%). The features of HPV DNA-/p16+ cases (n=93) were similar to those of
69 the HPV-associated VSCC, and HPV DNA+/p16- (n=74) cases had a more
70 diverse profile, although they were more similar to HPV-independent tumors.

71 Several histological characteristics were more frequently associated with HPV-
72 related VSCC (koilocytotic-like change, necrosis, moderate to marked
73 pleomorphism, invasive front in nests; $p < 0.001$), however, none of these

Rakislova et al. HPV-positive and –negative vulvar squamous cell carcinomas 4

74 characteristics allowed differentiation between HPV-associated and –
75 independent VSCC. In conclusion, histological criteria do not allow
76 differentiation between HPV-associated and –independent VSCC. p16 alone is
77 a clinically easy strategy to determine HPV status in VSCC.

78

79 INTRODUCTION

80 Vulvar squamous cell carcinoma (VSCC) accounts for less than 5% of the
81 gynecological malignancies but represents more than 90% of the malignant
82 tumors of the vulva ¹. VSCCs can be subdivided into human papilloma virus
83 (HPV)-associated and HPV-independent tumors, with HPV-associated tumors
84 representing one-fifth to one-half of all VSCC ^{2–5}. Increasing evidence indicates
85 that these two different types of VSCC not only have different epidemiological,
86 clinical, pathological and molecular characteristics, but also a different clinical
87 behavior. These differences stress the need of considering HPV-associated
88 and HPV-independent vulvar tumors as two separate entities ^{6–8}.

89 Most pathologists use clinico-pathological criteria to classify vulvar tumors
90 as HPV-associated or –independent. The current WHO classification ⁹
91 recognizes several histological variants of VSCC, including the basaloid, warty,
92 keratinizing and non-keratinizing subtypes. HPV-associated tumors are
93 considered to have a warty or basaloid histology ^{10–13}, and affect younger
94 women ^{14–19}. Contrarily, HPV-independent lesions generally have a keratinizing
95 morphology and occur in elderly women ^{14,20–24}. However, a number of studies
96 indicate that these pathological features have a limited usefulness to determine
97 HPV status ^{14,20–22}. Moreover, a comprehensive study analyzing histological
98 characteristics of HPV-positive and –negative carcinomas has yet to be
99 conducted.

100 Recent studies have shown that HPV DNA alone may be insufficient to
101 identify HPV-associated cancers outside the cervix ^{25–28}. In recent years very
102 sensitive and specific mRNA RT-PCR assays have been developed ²⁸, but

103 these tests are extremely technically complex, and consequently are not
104 adequate for pathology laboratories in routine daily practice.

105 p16 immunohistochemistry (IHC) has shown to be a useful tool to classify
106 VSCC into HPV-associated or –independent and is considered a good
107 surrogate marker of HPV association in the vulva and in other anatomical sites
108 in which HPV-associated and HPV-independent tumors are present^{2,14,16,22,28–}
109 ³². However, many studies evaluating p16 IHC have included a limited number
110 of cases and, consequently, the reliability of p16 positivity as a single marker of
111 HPV–association is not known.

112 Recently, the combination of two markers has been proposed as a reliable
113 tool to provide robust results in terms of HPV status^{2,28}. In this strategy, tumors
114 positive for HPV DNA and p16 would be indisputably HPV-associated, whereas
115 tumors negative for the two biomarkers would be classified as unquestionably
116 HPV-independent. However, it is not known whether the tumors with
117 intermediate features (HPV DNA positive and p16 negative and HPV DNA
118 negative and p16 positive) truly represent HPV-associated or –independent
119 tumors.

120 In this study, we analyzed a large series of 1594 tumors conducting a
121 thorough histological analysis and detection of HPV using HPV DNA, p16 IHC
122 and HPV mRNA. The study had three main goals: 1) to describe the histological
123 characteristics of indisputably HPV-associated (HPV DNA+/p16+) and HPV-
124 independent tumors (HPV DNA-/p16-); 2) to determine the frequency and the
125 features of the tumors with non-conclusive features (HPV DNA-/p16+ and HPV
126 DNA+/p16-); and 3) to determine the best strategy to differentiate HPV-
127 associated and –independent tumors.

128 MATERIAL AND METHODS*129 Study design and materials*

130 We reviewed all the invasive vulvar carcinomas previously analyzed in a
131 retrospective cross-sectional survey coordinated by the Catalan Institute of
132 Oncology (ICO, Barcelona-Spain) in collaboration with DDL Diagnostic
133 Laboratory (Rijswijk, The Netherlands)^{2,33}. In the present analysis we included
134 all the cases fulfilling the following inclusion criteria: 1) invasive squamous
135 carcinoma identified in the block; and 2) adequate material for histological
136 analysis, HPV detection and typing and p16 immunohistochemical stain. The
137 case recruitment protocols have been reported previously. Of the 1709 invasive
138 vulvar tumors included in the study, 47 were excluded because of non-
139 squamous histological types (i.e. basocellular or adenocarcinomas), and 68
140 were excluded because no material was available for p16 staining. Thus, the
141 study included 1594 formalin-fixed paraffin-embedded (FFPE) vulva specimens
142 collected from pathology archives from 38 countries from all continents (Mali,
143 Mozambique, Nigeria, and Senegal in Africa; Argentina, Brazil, Chile, Colombia,
144 Ecuador, Guatemala, Honduras, Mexico, Paraguay, Uruguay, the United States
145 and Venezuela in the Americas; Bangladesh, India, Israel, South Korea, Kuwait,
146 Lebanon, Philippines, Taiwan and Turkey in Asia; Austria, Belarus, Bosnia-
147 Herzegovina, Czech Republic, France, Germany, Greece, Italy, Poland,
148 Portugal, Spain and the United Kingdom in Europe; and Australia and New
149 Zealand in Oceania). Five hundred seventy-five cases (36.1%) were small
150 biopsies from vulvar tumors and 1019 (63.9%) cases came from large surgical
151 excisions or vulvectomy specimens. The study was approved by local and ICO
152 ethics committees.

153 *Histological evaluation*

154 A single FFPE histological block was available from each case for review.

155 Pathology evaluation was blind to HPV results and included a histological
156 subtyping following the WHO 2014 classification⁹. Basaloid tumors are
157 characterized for being composed of small undifferentiated cells with little
158 keratinization and koilocytosis. Warty subtype is defined by its condylomatous
159 features, pleomorphism, striking koilocytic atypia and frequent keratinization.
160 Non-keratinizing carcinoma is composed of sheets or nests of polygonal
161 squamous cells and absence of keratin pearls. The keratinizing type is
162 characterized by differentiated cells with keratin pearls and absence of
163 koilocytosis. When mixed features were identified, the tumor was classified
164 according to the main component, but other secondary components were also
165 recorded.

166 Additionally, the following histological characteristics were reported in all
167 tumors: growth pattern (exophytic or endophytic), percentage of tumor nests
168 showing keratin pearls, percentage of cells showing koilocytotic-like change,
169 individual keratinization, and differentiated features (large, eosinophilic
170 cytoplasm), percentage of tumor showing necrosis, extent of the inflammatory
171 infiltrate adjacent to the tumor, and severity of the pleomorphism (these latter
172 characteristics were semi-quantitatively evaluated as absent, mild, moderate or
173 severe). The invasive front of the tumors was classified as having a nested
174 (large or small nests) and diffuse pattern. Nested invasion consisted of either
175 large geographic nests with frequent central comedo-type necrosis or small
176 nests with frequent central keratin pearls³⁴. A diffuse (or infiltrative) front was
177 defined by irregular cords or single atypical cells in a so-called spray pattern,

178 often with surrounding desmoplastic stroma. Pattern evaluations were
179 performed at low-power (x4 to x10) magnification. Figure 1 shows the main
180 histological features evaluated in all the tumors.

181 *Immunohistochemical staining for p16 and p53*

182 All tumors were stained with p16 monoclonal antibody using the CINtec
183 Histology Kit (clone E6H4; Roche-Mtm-Laboratories, Heidelberg, Germany) ³⁵.
184 Only cases showing positivity in > 25% of tumor cells with strong and diffuse
185 block staining of the basal layer were considered as positive (p16^{INK4a}
186 upregulation) ^{28,36}.

187 p53 immunohistochemistry was performed in the first 192 cases of the
188 whole series of 1594 tumors. p53 was detected with the monoclonal anti-body
189 DO-7 (Dako, Carpinteria, CA, USA). p53 was considered positive when more
190 than 25% of the neoplastic cells showed nuclear staining.

191 *Tissue preparation, nucleic-acid isolation and HPV DNA detection*

192 DNA extraction was performed on whole sections of the formalin-fixed
193 paraffin-embedded (FFPE) tissue from the surgical specimen or from pre-
194 treatment biopsy. The samples were serially sectioned on a microtome. The first
195 and last sections (3 µm) were stained with H&E for histological confirmation of
196 the diagnosis. In-between sections were collected in RNAase-free reaction
197 tubes for DNA isolation (sandwich cutting technique). Sectioning and sample
198 preparation were carried out with the highest measures to avoid cross-
199 contamination. Paraffin blocks lacking tissue were cut in-between the patient
200 samples as controls to ensure the lack of contamination. Processing and
201 pathology diagnosis were done by the reference laboratory.

202 DNA extraction and HPV DNA detection has been previously described³⁷.
203 Briefly, HPV DNA detection was done using SPF10 PCR, DEIA and the LiPA25
204 system (version 1, Laboratory Biomedical Products, Rijswijk, The Netherlands).

205 *HPV mRNA detection*

206 The mRNA extraction and mRNA detection from tissue ribbons were
207 performed as previously described²⁸. For each case, HPV type-specific E6*I
208 mRNA RT-PCR assays were performed for the HPV type(s) previously
209 determined by genotyping and for a cellular ubiquitin C gene as a control for
210 tissue quality. HPV E6*I mRNA assays were developed for the following HPV
211 types: 16, 18, 26, 31, 33, 35, 39, 45, 51, 52, 53, 56, 58, 59, 66, 67, 68, 70, 73,
212 and 82. A second assay was performed to assess the presence of HPV16 E6*I
213 mRNA in all cases, irrespective of the HPV DNA result. Cases with HPV mRNA
214 positive (HPV mRNA+) and/or ubiquitin C mRNA+ signal were considered “RNA
215 valid”. All “RNA invalid” samples [i.e. cases that were HPV mRNA negative
216 (HPV mRNA-) and ubiquitin C mRNA-] were analyzed a second time, and
217 according to the signals obtained, they were classified as “RNA valid” or “RNA
218 invalid”.

219 HPV E6*I mRNA was analyzed in all the HPV DNA positive cases and in a
220 subset of 18 randomly selected HPV DNA cases as a negative control. A total of
221 18 HPV DNA- cases were tested (10 HPV DNA-/p16- and 8 HPV DNA-/p16+
222 cases).

223

224 *Classification of tumors as HPV-associated and HPV-independent. Tumors with*
225 *non-conclusive features*

226 A tumor was considered as unquestionably HPV-associated if both HPV
227 DNA and p16 IHC were positive. A tumor was considered indisputably HPV-
228 independent if both HPV DNA detection and p16IHC were negative. These
229 cases were considered the gold standard for HPV-positivity and –negativity,
230 respectively. Two groups of tumors were classified as non-conclusive for HPV
231 association: 1) tumors testing negative for HPV DNA but with positive staining
232 for p16 (HPV DNA-/p16+); and 2) tumors testing positive for HPV DNA, but with
233 negative result for p16 (HPV DNA+/p16-).

234 *Statistical analysis*

235 The results were reported as means, absolute frequencies and
236 percentages. Chi-square analysis or Fisher's exact test were used for
237 comparisons between nominal variables. Data were analyzed with the program
238 SPSS (version 15.0; SPSS, Inc, Chicago, IL). All statistical tests were two-
239 sided, and a p value of .05 or less was considered statistically significant.

240 **RESULTS**

241 *HPV DNA, p16, p53, and histological typing: HPV-associated, HPV-independent* 242 *tumors and tumors with non-conclusive features*

243 Four hundred forty-one tumors (27.7%) were positive for HPV DNA, and
244 1153 (72.3%) were negative. Table 1 shows the WHO classification of the HPV-
245 positive and -negative tumors. HPV-positive tumors were more frequently
246 basaloid or warty (198 out of 441; 44.9%), while 42.0% were of the keratinizing
247 type. HPV-negative tumors were more frequently keratinizing (905 out of 1153;
248 78.5%), but were occasionally of basaloid or warty type (88 out of 1153; 7.6%).

249 Mixed features, with presence of warty or basaloid along with keratinizing
250 areas were identified in 96/1594 tumors (6.0%). Mixed features were observed
251 in 30/441 (6.8%) HPV-positive tumors (6/160 [3.1%] basaloid, 4/38 [10.5%]
252 warty, 5/56 [8.9%] non-keratinizing, 15/185 [8.1%] keratinizing) and in 66/1153
253 (5.7%) HPV-negative tumors (7/55 [12.7%] basaloid, 9/33 [27.3%] warty, 12/141
254 [8.5%] non-keratinizing, 38/905 [4.2%] keratinizing).

255 Three hundred sixty-seven out of 441 HPV DNA positive tumors (83.2%)
256 were positive for p16 IHC; on the other hand, one thousand sixty out of 1153
257 HPV DNA negative tumors (91.9%) were negative for p16 IHC. The percentage
258 of tumors of basaloid or warty type slightly increased in HPV-associated tumors
259 (49.8%), whereas the percentage of histologically keratinizing tumors slightly
260 increased in HPV-independent tumors (81.2%). Figure 2 shows an example of
261 unquestionably HPV-associated VSCC with keratinizing histology (HPV
262 DNA+/p16+), and two indisputably HPV-independent VSCC (HPV DNA-/p16-)
263 with basaloid and warty characteristics.

264 One hundred sixty-seven tumors (10.5%) showed a non-conclusive profile.
265 Ninety-three tumors were HPV DNA-/p16+; and 74 tumors were HPV
266 DNA+/p16-. Table 2 shows the WHO classification and age distribution of the
267 indisputably HPV-associated tumors (HPV DNA+/p16+), the unquestionably
268 HPV-independent tumors (HPV DNA-/p16- tumors) and the tumors with non-
269 conclusive profiles. The tumors with a HPV DNA-/p16+ profile were similar to
270 the indisputably HPV-associated tumors in terms of WHO typing ($p>0.05$). The
271 HPV DNA+/p16- tumors had indeterminate features, although they were more
272 similar to the unquestionably HPV-independent tumors (slightly higher

273 proportion of warty-basaloid tumors in the HPV DNA+/p16- group compared
274 with HPV DNA-/p16- group; $p < 0.001$).

275 There were marked differences between these two groups in terms of age
276 (61.7 ± 16.7 in the indisputably HPV-associated tumors vs. 71.7 ± 13.4 , in the
277 unquestionably HPV-independent tumors; $p < 0.001$). Interestingly, the mean age
278 of the patients with tumors with a HPV DNA-/p16+ profile was similar to that of
279 the women with indisputably HPV-associated tumors ($p > 0.05$). Contrarily, the
280 HPV DNA+/p16- cases were similar in age distribution to the HPV-independent
281 tumors.

282 From the subset of 192 tumors stained for p53, a positive result was
283 observed in 2 out of 40 (5%) unquestionably HPV-associated tumors (HPV
284 DNA+/p16+ profile) and in 88 out of the 140 (62.9%) unquestionably HPV-
285 independent tumors (HPV DNA-/p16- profile). In the group of HPV DNA-/p16+
286 tumors, p53 was negative in all six tumors tested (0/6; 0%), whereas in the
287 group of HPV DNA+/p16- tumors, 4/6 (66.7%) were positive for p53.

288 *Histological characteristics of unquestionable HPV-associated and –*
289 *independent tumors*

290 Table 3 shows the histological characteristics of the indisputably HPV-
291 associated tumors and the unquestionably HPV-independent tumors, as well as
292 the characteristics of the HPV DNA positive and negative tumors without p16
293 IHC. Several histological characteristics were more frequently associated with
294 HPV-associated tumors (koilocytotic-like change, necrosis, moderate to marked
295 pleomorphism, invasive front in nests; $p < 0.001$), whereas individual
296 keratinization, keratin pearls, highly differentiated cells and a diffuse invasive

297 front were more frequently associated with HPV-independent VSCC ($p < 0.001$).
298 However, none of the histological features allowed reliable differentiation
299 between the two types of VSCC as all of the characteristics were present in a
300 proportion of both types of tumors.

301 *HPV typing and histological classification*

302 Table 4 shows the results of the HPV typing and the histological
303 classification of the tumors according to the typing in the 441 cases positive for
304 HPV DNA and in the group of 367 unquestionable HPV-associated tumors.
305 HPV16 was, by far, the type most frequently identified.

306 p16 IHC was positive in over 90% of cases positive for HPV16 and 45, in
307 over 80% of the cases positive for HPV18 and 33, in over 70% of multiple HPV
308 infections and infections by other high-risk HPV, in 30% of infections by
309 undetermined HPV types and was negative in tumors caused by low-risk HPV.
310 The percentage of tumors with basaloid or warty histology ranged from 70% in
311 HPV33 positive tumors to less than 40% in HPV 18 or HPV45 positive tumors.

312 *HPV mRNA detection*

313 Adequate material for mRNA detection was available in 402 out of the 441
314 (91.1%) HPV DNA positive tumors. HPV mRNA was identified in 350/402
315 tumors (87.1%). Five cases were considered as “RNA invalid”. Type-specific
316 HPV mRNA was not analyzed in the tumors caused by undetermined ($n=19$),
317 low-risk HPV ($n=12$), and in three tumors associated with high-risk types 30, 69,
318 and 102, as the probes were not designed against these viruses. HPV mRNA
319 was positive in 334/352 (94.9%) of the unquestionably HPV-associated tumors

320 (HPV DNA+/p16+) and in only 16/50 (32%) of the HPV DNA+/p16- tumors
321 ($p < 0.001$).

322 In the subset of 18 HPV DNA negative tumors tested for mRNA, eight
323 tumors were HPV DNA-/p16+ and 10 HPV DNA-/p16-. Two out of eight (25%)
324 HPV DNA-/p16+ tumors were positive for HPV mRNA, whereas none of the 10
325 unquestionably HPV-independent tumors (HPV DNA-/p16-) were positive for
326 HPV mRNA.

327 **DISCUSSION**

328 The present study, which includes a large number of VSCCs, confirms
329 previous results obtained by our group^{14,16}, showing that there is significant
330 overlap between the histological types of VSCC and the association with HPV.
331 Indeed, 36.5% of the unquestionable HPV-associated VSCCs were of the
332 conventional keratinizing type, the histological type considered as characteristic
333 of the HPV-independent pathway of VSCC^{3,5,14,16,38–40}. On the other hand, a
334 small, albeit significant, number of indisputably HPV-independent VSCCs
335 showed basaloid or warty features (5.2%), the histological types usually
336 considered as associated with HPV-driven VSCCs^{3,5,14,16,38,41,42}. This high
337 prevalence of HPV infection among keratinizing VSCCs has also been
338 observed by other authors, who have found percentages similar to or higher
339 than those observed in our study^{2,14,20,43–45}. Thus, the present results confirm
340 the limited reliability of the current WHO classification of VSCCs in terms of
341 HPV attribution^{5,14,41}.

342 The poor correlation observed between histological typing and HPV
343 infection may be related, in part, by a certain degree of subjectivity in

344 subclassifying the tumors observed in our study as previously referred by some
345 authors^{14,41}. These difficulties are particularly challenging in the diagnosis of
346 non-keratinizing carcinoma and to a lesser extent, warty carcinomas¹⁴. In
347 addition, the presence of keratinization in basaloid carcinomas can hinder the
348 differential diagnosis with poorly differentiated keratinizing squamous cell
349 carcinomas. Moreover, mixed basaloid-keratinizing squamous carcinomas
350 posing serious diagnostic problems have been described¹⁷.

351 The marked overlap observed in all the histological features evaluated in
352 our study clearly highlights the absence of specific criteria that allow classifying
353 a tumor as HPV-associated or –independent. Indeed, although many of the
354 histological findings evaluated in our study showed significant differences
355 between the two groups, all the characteristics were found in a large proportion
356 of HPV-associated and –independent tumors. A particularly unspecific feature is
357 the presence of koilocytotic-like changes, which were observed in 11.6% of the
358 HPV-independent VSCCs. Moreover, a marked interobserver variability in the
359 separation between pseudokoilocytotic and true koilocytotic changes has been
360 reported^{14,46}. These findings further stress the poor validity of morphology
361 alone to distinguish the two etio-pathogenic types of VSCC^{5,14,20–22,25,41,47}.

362 Our study shows that 10.5% of the tumors had non-conclusive features in
363 terms of HPV attribution assessed by HPV DNA and p16. The most frequent
364 group (HPV DNA-/p16+) was comparable to the unquestionably HPV-
365 associated group in terms of age and histological typing. These findings indicate
366 that these cases may represent false negative results of HPV DNA detection. In
367 these cases the poor tissue quality could have contributed to the false negative
368 HPV DNA result². In contrast, the other non-conclusive group (HPV DNA+/p16-

369) is more heterogeneous. It includes the unusual VSCCs (12 cases) harboring
370 only a low-risk HPV, which have been shown to be negative for p16 IHC ⁴⁸.
371 Some of the cases represented p16 negativity in truly HPV-positive tumors, as
372 shown by the identification of 16 tumors positive not only for HPV DNA but also
373 for HPV mRNA. However, it is likely that some of these cases are false positive
374 HPV DNA cases. This group was more similar to the indisputably HPV-
375 independent group in terms of histological characteristics and age range.
376 Moreover, the infrequent presence of mRNA in these tumors (32.0% vs. 94.9%
377 in the HPV DNA+/p16+ group) after excluding all cases not tested for mRNA
378 HPV, as the test was not designed for low-risk and undetermined HPV types, is
379 also in keeping with this hypothesis. As the whole FFPE block was tested for
380 HPV detection without performing tumor microdissection, we cannot exclude the
381 possibility of contamination from the adjacent embedded tissue not related to
382 the tumor. Thus, our results suggest that HPV DNA detection alone could
383 misclassify up to 10% of cases, whereas despite not being perfect, p16
384 immunohistochemistry could be a more reliable tool, offering even better
385 sensitivity and specificity.

386 p53 IHC was positive in only 5% of the unquestionably HPV-associated
387 tumors and in 66.7% of the unquestionably HPV-independent tumors. These
388 results are in keeping with previous reports showing that p53 abnormalities are
389 infrequent in HPV-associated tumors and present in about two thirds of HPV-
390 independent tumors ^{14,49}. Interestingly, the percentage of p53 positivity in HPV
391 DNA-/p16+ tumors was comparable to the unquestionably HPV-associated
392 neoplasms whereas HPV DNA+/p16- tumors were more similar to the
393 indisputably HPV-independent carcinomas. These results add further evidence

394 indicating that DNA-/p16+ tumors probably represent false negative results of
395 HPV DNA detection, whereas many of the HPV DNA+/p16- tumors are false
396 positive HPV DNA cases.

397 *In situ* hybridization assays that can detect HPV E6/E7 mRNA in FFPE
398 tissue and allow visualization of the transcripts within tumor cells have recently
399 become available. Although the value of these assays needs to be confirmed,
400 they might help in the identification of HR-HPV-associated neoplasia in the
401 vulva, the anus and the head and neck region ⁵⁰.

402 The main strength of the present study is that it includes a very high
403 number of VSCC, and in all the cases, a detailed histological review and
404 thorough molecular analysis including HPV DNA, mRNA and p16 were
405 performed. The correlation of all the data allowed the study to obtain more
406 accurate evidence of HPV involvement in the large series of tumors ²⁸.

407 The study also has some limitations. Only one FFPE block per case was
408 available for review. This may, to some extent, have influenced the histological
409 type distribution and, particularly, may have resulted in a relatively high
410 percentage of non-keratinizing tumors, as keratin pearl formation may have
411 been focal or absent in the area sampled. Additionally, although HPV DNA
412 detection in this study has fulfilled strict quality controls, the different tissue
413 processing techniques performed in a large variety of contributing laboratories
414 could have had an impact on our results. However, the high number of cases
415 analyzed and the strict quality controls used for p16 staining and HPV DNA and
416 RNA detection have probably minimized this effect.

417 Finally, the complete absence of clinical and follow-up data prevents
418 obtaining any conclusions on the prognostic implications of HPV status.
419 Although there are some discordant results on the prognostic significance of
420 HPV in VSCC ^{8,20}, recent evidence indicates a strong association between
421 positive p16 IHC and lower FIGO stage and negative lymph node metastasis ⁷,
422 and that the presence of HPV DNA or positive p16 IHC is an independent
423 prognostic factor ⁶.

424 In conclusion, the present study performed in a large number of cases of
425 vulvar carcinomas confirms that histological criteria do not allow differentiation
426 between HPV-associated and –independent VSCC. Although not perfect, p16 is
427 a good surrogate marker that could even outperform the HPV DNA detection
428 technique.

429

430

431 **REFERENCES**

- 432 1. Kurman R, Ronnett J, Sherman M, Wilkinson E, eds. *Atlas of tumor*
433 *pathology: Tumors of the cervix, vagina, and vulva*, 4th ed. Washington:
434 American Registry of Pathology, 2010; 311-326.
- 435 2. de Sanjosé S, Alemany L, Ordi J, Sara Tous MA, Bigby SM, Joura EA,
436 Maldonado P, Laco J, Bravo IG, Vidal A, Nuria G, Cross P, Wain G V., Karl
437 Ulrich P, Luciano M, Bergeron C, Vaclav M, Adela Rosa S, Félix A,
438 Usubutun A, Seoud M, Hernandez-Suarez G, Nowakowsky AM, Godfrey
439 W, Dalstei V, Hampl M, Kasamatsu ES, Lombardi LE, Tinoco L, Alvarado-
440 Cabrero I, Perrotta M, Bhatla N, Agorastos T, Lynch CF, Goodman MT,
441 Shin H-R, Viarheichyk H, Jach R, M.O.L. Cruz E, Velasco J, Molina C,
442 Bornstein J, Ferrera A, Domingo EJ, Cheng-Yang C, Banjo AF,
443 Castellsagué X, Pawlita M, Lloveras B, Quint WGV, Muñoz N, Bosch FX.
444 Worldwide human papillomavirus genotype attribution in over 2000 cases
445 of intraepithelial and invasive lesions of the vulva. *Eur J Cancer*
446 2013;49:3450–3461.
- 447 3. van der Avoort IA, Shirango H, Hoevenaars BM, Grefte JMM, de Hullu J
448 a, de Wilde PCM, Bulten J, Melchers WJG, Massuger LF a G. Vulvar
449 squamous cell carcinoma is a multifactorial disease following two
450 separate and independent pathways. *Int J Gynecol Pathol* 2006;25:22–
451 29.
- 452 4. van de Nieuwenhof HP, van Kempen LC, de Hullu JA, Bekkers RLM,
453 Bulten J, Melchers WJG, Massuger LFAG. The etiologic role of HPV in
454 vulvar squamous cell carcinoma fine tuned. *Cancer Epidemiol Biomarkers*
455 *Prev* 2009;18:2061–2067.
- 456 5. del Pino M, Rodriguez-Carunchio L, Ordi J. Pathways of vulvar
457 intraepithelial neoplasia and squamous cell carcinoma. *Histopathology*
458 2013;62:161–175.
- 459 6. Wakeham K, Kavanagh K, Cuschieri K, Millan D, Pollock KG, Bell S,
460 Burton K, Reed NS, Graham S V. HPV status and favourable outcome in
461 vulvar squamous cancer. *Int J Cancer* 2017;140: 1134–1146.

- 462 7. Lee LJ, Howitt B, Catalano P, Tanaka C, Murphy R, Cimbak N, DeMaria
463 R, Bu P, Crum C, Horowitz N, Matulonis U, Viswanathan AN. Prognostic
464 importance of human papillomavirus (HPV) and p16 positivity in
465 squamous cell carcinoma of the vulva treated with radiotherapy. *Gynecol*
466 *Oncol* 2016;142: 293–298.
- 467 8. Rakislova N, Saco A, Sierra A, del Pino M, Ordi J. Role of Human
468 Papillomavirus in Vulvar Cancer. *Adv Anat Pathol* 2017;24: 201–214.
- 469 9. Crum CP, Herrington C, McCluggage WG. Tumours of the vulva; epithelial
470 tumors. In: *WHO Classification of Tumours of Female Reproductive*
471 *Organs*; 4th ed. Lyon: International Agency for Research on Cancer
472 (IARC), 2014; 232–241.
- 473 10. Bornstein J, Bogliatto F, Haefner HK, Stockdale CK, Preti M, Bohl TG,
474 Reutter J. The 2015 International Society for the Study of Vulvovaginal
475 Disease (ISSVD) Terminology of Vulvar Squamous Intraepithelial Lesions.
476 *Obstet Gynecol* 2016;127:264–268.
- 477 11. Hoang LN, Park KJ, Soslow RA, Murali R. Squamous precursor lesions of
478 the vulva: current classification and diagnostic challenges. *Pathology*
479 2016;48:291–302.
- 480 12. Bogliatto F, Bohl T, Reutter J, Sideri M, Bornstein J. Last Terminology
481 Applied to the Vulva: : The Challenge of VIN Continues. *J Low Genit Tract*
482 *Dis* 2015;19:47–48.
- 483 13. Darragh TM, Colgan TJ, Thomas Cox J, Heller DS, Henry MR, Luff RD,
484 McCalmont T, Nayar R, Palefsky JM, Stoler MH, Wilkinson EJ, Zaino RJ,
485 Wilbur DC, Members of the LAST Project Work Groups. The Lower
486 Anogenital Squamous Terminology Standardization project for HPV-
487 associated lesions: background and consensus recommendations from
488 the College of American Pathologists and the American Society for
489 Colposcopy and Cervical Pathology. *Int J Gynecol Pathol* 2013;32:76–
490 115.
- 491 14. Santos M, Landolfi S, Olivella A, Lloveras B, Klaustermeier J, Suárez H,
492 Alòs L, Puig-Tintoré LM, Campo E, Ordi J. p16 overexpression identifies
493 HPV-positive vulvar squamous cell carcinomas. *Am J Surg Pathol*

- 494 2006;30:1347–1356.
- 495 15. Toki T, Kurman RJ, Park JS, Kessis T, Daniel RW, Shah K V. Probable
496 nonpapillomavirus etiology of squamous cell carcinoma of the vulva in
497 older women: a clinicopathologic study using in situ hybridization and
498 polymerase chain reaction. *Int J Gynecol Pathol* 1991;10:107–125.
- 499 16. Santos M, Montagut C, Mellado B, Garcia A, Ramon y Cajal S, Cardesa
500 A, Puig-Tintore LM, Ordi J. Immunohistochemical staining for p16 and p53
501 in premalignant and malignant epithelial lesions of the vulva. *Int J*
502 *Gynecol Pathol* 2004;23:206–214.
- 503 17. Kurman RJ, Toki T, Schiffman MH. Basaloid and warty carcinomas of the
504 vulva. Distinctive types of squamous cell carcinoma frequently associated
505 with human papillomaviruses. *Am J Surg Pathol* 1993;17:133–145.
- 506 18. Nogueira MC, Guedes Neto E de P, Rosa MW, Zettler E, Zettler CG.
507 Immunohistochemical expression of p16 and p53 in vulvar intraepithelial
508 neoplasia and squamous cell carcinoma of the vulva. *Pathol Oncol Res*
509 2006;12:153–157.
- 510 19. Preti M, Scurry J, Marchitelli CE, Micheletti L. Vulvar intraepithelial
511 neoplasia. *Best Pract Res Clin Obstet Gynaecol* 2014;28:1051–1062.
- 512 20. Alonso I, Fusté V, Del Pino M, Castillo P, Torné A, Fusté P, Rios J, Pahisa
513 J, Balasch J, Ordi J. Does human papillomavirus infection imply a
514 different prognosis in vulvar squamous cell carcinoma? *Gynecol Oncol*
515 2011;122:509–514.
- 516 21. Ordi J, Alejo M, Fusté V, Lloveras B, Del Pino M, Alonso I, Torné A. HPV-
517 negative vulvar intraepithelial neoplasia (VIN) with basaloid histologic
518 pattern: an unrecognized variant of simplex (differentiated) VIN. *Am J*
519 *Surg Pathol* 2009;33:1659–1665.
- 520 22. Cheng AS, Karnezis AN, Jordan S, Singh N, McAlpine JN, Gilks CB. p16
521 Immunostaining Allows for Accurate Subclassification of Vulvar Squamous
522 Cell Carcinoma Into HPV-Associated and HPV-Independent Cases. *Int J*
523 *Gynecol Pathol* 2016;35:385–393.
- 524 23. Andersen WA, Franquemont DW, Williams J, Taylor PT, Crum CP. Vulvar

- 525 squamous cell carcinoma and papillomaviruses: two separate entities?
526 Am J Obstet Gynecol 1991;165:329–335.
- 527 24. Singh N, Leen SL, Han G, Faruqi A, Kokka F, Rosenthal A, Jiang XR, Kim
528 R, McAlpine JN, Gilks CB. Expanding the morphologic spectrum of
529 differentiated VIN (dVIN) through detailed mapping of cases with p53
530 loss. Am J Surg Pathol 2015;39:52–60.
- 531 25. Alemany L, Cubilla A, Halc G, Kasamatsu E, Quirós B, Masferrer E,
532 Tous S, Lloveras B, Hernández-Suarez G, Lonsdale R, Tinoco L, Alejo M,
533 Alvarado-Cabrero I, Laco J, Guimerà N, Poblet E, Lombardi LE, Bergeron
534 C, Clavero O, Shin H-R, Ferrera A, Felix A, Germar J, Mandys V, Clavel
535 C, Tzardi M, Pons LE, Wain V, Cruz E, Molina C, Mota JD, Jach R,
536 Velasco J, Carrilho C, López-Revilla R, Goodman MT, Quint WG,
537 Castellsagué X, Bravo I, Pawlita M, Muñoz N, Bosch FX, de Sanjosé S.
538 Role of Human Papillomavirus in Penile Carcinomas Worldwide. Eur Urol
539 2016;69:953–961.
- 540 26. Castellsagué X, Alemany L, Quer M, Halc G, Quirós B, Tous S, Clavero
541 O, Alòs L, Biegner T, Szafarowski T, Alejo M, Holzinger D, Cadena E,
542 Claros E, Hall G, Laco J, Poljak M, Benevolo M, Kasamatsu E, Mehanna
543 H, Ndiaye C, Guimerà N, Lloveras B, León X, Ruiz-Cabezas JC,
544 Alvarado-Cabrero I, Kang C-S, Oh J-K, Garcia-Rojo M, Iljazovic E, Ajayi
545 OF, Duarte F, Nessa A, Tinoco L, Duran-Padilla MA, Pirog EC,
546 Viarheichyk H, Morales H, Costes V, Félix A, Germar MJ V, Mena M,
547 Ruacan A, Jain A, Mehrotra R, Goodman MT, Lombardi LE, Ferrera A,
548 Malami S, Albanesi EI, Dabed P, Molina C, López-Revilla R, Mandys V,
549 González ME, Velasco J, Bravo IG, Quint W, Pawlita M, Muñoz N, de
550 Sanjosé S, Xavier Bosch F, ICO International HPV in Head and Neck
551 Cancer Study Group. HPV Involvement in Head and Neck Cancers:
552 Comprehensive Assessment of Biomarkers in 3680 Patients. J Natl
553 Cancer Inst 2016;108:djv403.
- 554 27. Lassen P. The role of Human papillomavirus in head and neck cancer and
555 the impact on radiotherapy outcome. Radiother Oncol 2010;95:371–380.
- 556 28. Halc G, Alemany L, Quiros B, Clavero O, Höfler D, Alejo M, Quint W,

- 557 Pawlita M, Bosch FX, de Sanjose S. Biological relevance of human
558 papillomaviruses in vulvar cancer. *Mod Pathol* 2017;30:549–562.
- 559 29. Hoevenaars BM, van der Avoort IA, de Wilde PCM, Massuger LFAG,
560 Melchers WJG, de Hullu JA, Bulten J. A panel of p16 INK4A , MIB1 and
561 p53 proteins can distinguish between the 2 pathways leading to vulvar
562 squamous cell carcinoma. *Int J Cancer* 2008;123:2767–2773.
- 563 30. Rufforny I, Wilkinson EJ, Liu C, Zhu H, Buteral M, Massoll NA. Human
564 Papillomavirus Infection and p16 INK4a Protein Expression in Vulvar
565 Intraepithelial Neoplasia and Invasive Squamous Cell Carcinoma. *J Low
566 Genit Tract Dis* 2005;9:108–113.
- 567 31. Chung CH, Zhang Q, Kong CS, Harris J, Fertig EJ, Harari PM, Wang D,
568 Redmond KP, Shenouda G, Trotti A, Raben D, Gillison ML, Jordan RC, Le
569 Q-T. p16 protein expression and human papillomavirus status as
570 prognostic biomarkers of nonoropharyngeal head and neck squamous
571 cell carcinoma. *J Clin Oncol* 2014;32:3930–3938.
- 572 32. El-Naggar AK, Westra WH. p16 expression as a surrogate marker for
573 HPV-related oropharyngeal carcinoma: a guide for interpretative
574 relevance and consistency. *Head Neck* 2012;34:459–461.
- 575 33. Serrano B, De Sanjosé S, Tous S, Quiros B, Muñoz N, Bosch X, Alemany
576 L. Human papillomavirus genotype attribution for HPVs 6, 11, 16, 18, 31,
577 33, 45, 52 and 58 in female anogenital lesions. *Eur J Cancer*
578 2015;51:1732–1741.
- 579 34. Jeffus SK, Gehlot A, Holthoff E, Stone R, Spencer H, Kelly T, Post SR,
580 Quick CM. A fibromyxoid stromal response is associated with an
581 infiltrative tumor morphology, perineural invasion, and lymph node
582 metastasis in squamous cell carcinoma of the vulva. *Am J Surg Pathol*
583 2015;39:1226–1233.
- 584 35. Del Pino M, Garcia S, Fusté V, Alonso I, Fusté P, Torné A, Ordi J. Value of
585 p16(INK4a) as a marker of progression/regression in cervical
586 intraepithelial neoplasia grade 1. *Am J Obstet Gynecol* 2009;201:488.e1-
587 7.

- 588 36. Halec G, Holzinger D, Schmitt M, Flechtenmacher C, Dyckhoff G,
589 Lloveras B, Höfler D, Bosch FX, Pawlita M. Biological evidence for a
590 causal role of HPV16 in a small fraction of laryngeal squamous cell
591 carcinoma. *Br J Cancer* 2013;109:172–183.
- 592 37. De Sanjose S, Quint WG, Alemany L, Geraets DT, Klaustermeier JE,
593 Lloveras B, Tous S, Felix A, Bravo LE, Shin H-R, Vallejos CS, de Ruiz PA,
594 Lima MA, Guimera N, Clavero O, Alejo M, Llombart-Bosch A, Cheng-
595 Yang C, Tatti SA, Kasamatsu E, Iljazovic E, Odida M, Prado R, Seoud M,
596 Grce M, Usubutun A, Jain A, Suarez GAH, Lombardi LE, Banjo A,
597 Menéndez C, Domingo EJ, Velasco J, Nessa A, Chichareon SCB, Qiao
598 YL, Lerma E, Garland SM, Sasagawa T, Ferrera A, Hammouda D, Mariani
599 L, Pelayo A, Steiner I, Oliva E, Meijer CJ, Al-Jassar WF, Cruz E, Wright
600 TC, Puras A, Llave CL, Tzardi M, Agorastos T, Garcia-Barriola V, Clavel
601 C, Ordi J, Andújar M, Castellsagué X, Sánchez GI, Nowakowski AM,
602 Bornstein J, Muñoz N, Bosch FX. Human papillomavirus genotype
603 attribution in invasive cervical cancer: a retrospective cross-sectional
604 worldwide study. *Lancet Oncol* 2010;11:1048–1056.
- 605 38. Ueda Y, Enomoto T, Kimura T, Yoshino K, Fujita M, Kimura T. Two distinct
606 pathways to development of squamous cell carcinoma of the vulva. *J Skin
607 Cancer* 2011;951250:7.
- 608 39. Bonvicini F, Venturoli S, Ambretti S, Paterini P, Santini D, Ceccarelli C,
609 Zerbini M, Musiani M. Presence and type of oncogenic human
610 papillomavirus in classic and in differentiated vulvar intraepithelial
611 neoplasia and keratinizing vulvar squamous cell carcinoma. *J Med Virol*
612 2005;77:102–106.
- 613 40. Pinto AP, Miron A, Yassin Y, Monte N, Woo TY, Mehra KK, Medeiros F,
614 Crum CP. Differentiated vulvar intraepithelial neoplasia contains Tp53
615 mutations and is genetically linked to vulvar squamous cell carcinoma.
616 *Mod Pathol* 2010;23:404–412.
- 617 41. Chiesa-Vottero A, Dvoretzky PM, Hart WR. Histopathologic study of thin
618 vulvar squamous cell carcinomas and associated cutaneous lesions: a
619 correlative study of 48 tumors in 44 patients with analysis of adjacent

- 620 vulvar intraepithelial neoplasia types and lichen sclerosus. *Am J Surg*
621 *Pathol* 2006;30:310–318.
- 622 42. Skapa P, Zamecnik J, Hamsikova E, Salakova M, Smahelova J, Jandova
623 K, Robova H, Rob L, Tachezy R. Human papillomavirus (HPV) profiles of
624 vulvar lesions: possible implications for the classification of vulvar
625 squamous cell carcinoma precursors and for the efficacy of prophylactic
626 HPV vaccination. *Am J Surg Pathol* 2007;31:1834–1843.
- 627 43. Siriaunkgul S, Settakorn J, Sukpan K, Srisomboon J, Utaipat U,
628 Lekawanvijit S, Khunamornpong S. HPV detection and genotyping in
629 vulvar squamous cell carcinoma in Northern Thailand. *Asian Pacific J*
630 *Cancer Prev* 2014;15:3773–3778.
- 631 44. Smith JS, Backes DM, Hoots BE, Kurman RJ, Pimenta JM. Human
632 papillomavirus type-distribution in vulvar and vaginal cancers and their
633 associated precursors. *Obstet Gynecol* 2009;113:917–924.
- 634 45. De Vuyst H, Clifford GM, Nascimento MC, Madeleine MM, Franceschi S.
635 Prevalence and type distribution of human papillomavirus in carcinoma
636 and intraepithelial neoplasia of the vulva, vagina and anus: a meta-
637 analysis. *Int J Cancer* 2009;124:1626–1636.
- 638 46. McCluggage WG, Walsh MY, Thornton CM, Hamilton PW, Date A,
639 Caughley LM, Bharucha H. Inter- and intra-observer variation in the
640 histopathological reporting of cervical squamous intraepithelial lesions
641 using a modified Bethesda grading system. *Br J Obstet Gynaecol*
642 1998;105:206–210.
- 643 47. Dong F, Kojiro S, Borger DR, Growdon WB, Oliva E. Squamous Cell
644 Carcinoma of the Vulva: A Subclassification of 97 Cases by
645 Clinicopathologic, Immunohistochemical, and Molecular Features (p16,
646 p53, and EGFR). *Am J Surg Pathol* 2015;39:1045–1053.
- 647 48. Guimerà N, Lloveras B, Lindeman J, Alemany L, van de Sandt M, Alejo M,
648 Hernandez-Suarez G, Bravo IG, Molijn A, Jenkins D, Cubilla A, Muñoz N,
649 de Sanjose S, Bosch FX, Quint W. The Occasional Role of Low-risk
650 Human Papillomaviruses 6, 11, 42, 44, and 70 in Anogenital Carcinoma
651 Defined by Laser Capture Microdissection/PCR Methodology. *Am J Surg*

- 652 Pathol 2013;37:1299–1310.
- 653 49. Trietsch MD, Spaans VM, ter Haar NT, Osse EM, Peters AAW,
654 Gaarenstroom KN, Fleuren GJ. CDKN2A(p16) and HRAS are frequently
655 mutated in vulvar squamous cell carcinoma. *Gynecol Oncol* 2014;135:
656 149–155.
- 657 50. Mills AM, Dirks DC, Poulter MD, Mills SE, Stoler MH. HR-HPV E6/E7
658 mRNA In Situ Hybridization: Validation Against PCR, DNA In Situ
659 Hybridization, and p16 Immunohistochemistry in 102 Samples of Cervical,
660 Vulvar, Anal, and Head and Neck Neoplasia. *Am J Surg Pathol* 2017;41:
661 607–615.
- 662

663 **LEGEND OF FIGURES**

664 **FIGURE 1.** Histological characteristics evaluated in the study: **A)** koilocytotic-like
665 change; **B)** necrosis; **C)** moderate to marked pleomorphism; **D)** individual
666 keratinization, keratin pearls, and highly differentiated cells; **E)** invasive front in
667 nests; **F)** diffuse invasive front. Hematoxylin and eosin staining.

668 **FIGURE 2. A)** Keratinizing VSCC unquestionably HPV-associated tumor (HPV
669 DNA+/p16+); **B)** indisputably HPV-independent VSCC (HPV DNA-/p16-) of
670 basaloid type; **C)** indisputably HPV-independent VSCC (HPV DNA-/p16-) of
671 warty type. Hematoxylin and eosin staining (A, B, C) and p16
672 immunohistochemistry (A', B', C').

673

674

Table 1. WHO classification of the human papillomavirus (HPV)-associated and -independent tumors, defined by HPV DNA detection.

	HPV DNA+	HPV DNA-	p-value
	(n=441)	(n=1153)	
	n (%)	n (%)	
Keratinizing	185 (42.0)	905 (78.5)	0.000
Non-keratinizing	56 (12.7)	141 (12.2)	0.786
Basaloid	160 (36.3)	55 (4.8)	0.000
Warty	38 (8.6)	33 (2.9)	0.000
Verrucous	1 (0.2)	15 (1.3)	0.087
Spindle cell	1 (0.2)	4 (0.3)	0.999

The statistical significant p-values are highlighted in bold (corrected p-value for multiple comparison; $0.05/6=0.008$).

Table 2. WHO classification of the human papillomavirus (HPV)-associated and –independent tumors, defined by HPV DNA and p16 detection.

	Gold standard groups		Non-conclusive groups		p-value ¹
	HPV DNA+ p16+ (n=367) n (%)	HPV DNA- p16- (n=1060) n (%)	HPV DNA- p16+ (n=93) n (%)	HPV DNA + p16- (n=74) n (%)	
Keratinizing	134 (36.5)	861 (81.2)	44 (47.3)	51 (68.9)	0.000
Non-keratinizing	49 (13.4)	124 (11.7)	17 (18.3)	7 (9.5)	0.403
Basaloid	152 (41.4)	27 (2.5)	28 (30.1)	8 (10.8)	0.000
Warty	31 (8.4)	29 (2.7)	4 (4.3)	7 (9.5)	0.000
Verrucous	0 (0.0)	15 (1.4)	0 (0.0)	1 (1.4)	0.016
Spindle cell	1 (0.3)	4 (0.4)	0 (0.0)	0 (0.0)	1.000
Age range [min-max]	[22-97]	[18-104]	[24-94]	[33-92]	
Mean age (SD)	61.7 (16.7)	71.7 (13.4)	64.8 (15.0)	68.8 (14.4)	0.000

1) p-value: comparison of the gold standard for positivity and negativity (HPV DNA+ and p16+ vs HPV DNA- and p16-)

SD: Standard deviation.

The statistical significant p-values are highlighted in bold (corrected p-value for multiple comparisons; $0.05/6=0.008$).

Table 3. Histological characteristics of the human papillomavirus (HPV)-associated and –independent tumors.

	HPV DNA+ (n=441) n (%)	HPV DNA- (n=1153) n (%)	HPV DNA+ and p16+ (n=367) n (%)	HPV DNA- and p16- (n=1060) n (%)	p-value ¹
Pattern of growth					
Exophytic	106 (24.0)	221 (19.2)	83 (22.6)	206 (19.4)	0.191
Endophytic	335 (76.0)	932 (80.8)	284 (77.4)	854 (80.6)	0.191
Koilocytotic-like change	154 (34.9)	145 (12.6)	121 (33.0)	123 (11.6)	0.000
Inflammatory infiltrate	283 (64.2)	720 (62.4)	234 (63.8)	658 (62.1)	0.566
Individual keratinization	223 (50.6)	832 (72.2)	170 (46.3)	780 (73.6)	0.000
Keratin pearls	307 (69.6)	1009 (87.5)	246 (67.0)	946 (89.2)	0.000
Differentiated cells ≥10%	339 (76.9)	1071 (92.9)	271 (73.8)	999 (94.2)	0.000
Necrosis	279 (63.3)	610 (52.9)	231 (62.9)	547 (51.6)	0.000
Pleomorphism moderate/marked	371 (84.1)	858 (74.4)	313 (85.3)	775 (73.1)	0.000
Invasive front					
Large or small nests	260 (59.0)	484 (42.0)	224 (61.0)	438 (41.3)	0.000
Diffuse	181 (41.0)	669 (58.0)	143 (39.0)	622 (58.7)	0.000

1) p-value: comparison of gold standards for positivity and negativity, that is HPV DNA+ AND p16+ vs HPV DNA- AND p16-. The statistical significant p-values are highlighted in bold (corrected p-value for multiple comparisons; 0.05/11=0.005).

Table 4. Histological World Health Organization (WHO) classification according to human papillomavirus (HPV) genotyping.

	HPV DNA positive (n=441)			HPV DNA positive and p16 positive (n=367)		
	WHO histological classification			WHO histological classification		
	Basaloid/warty	Keratinizing/non-keratinizing/verrucous/spindle cell		Basaloid/warty	Keratinizing/non-keratinizing/verrucous/spindle cell	
	n	n (%)	n (%)	n	n (%)	n (%)
HPV infection						
Single HPV16	296	139 (47.0)	157 (53.0)	269	133 (49.4)	136 (50.6)
Single HPV33	23	15 (65.2)	8 (34.8)	20	14 (70.0)	6 (30.0)
Single HPV18	16	5 (31.3)	11 (68.7)	13	5 (38.5)	8 (61.5)
Single HPV45	13	5 (38.5)	8 (61.5)	13	5 (38.5)	8 (61.5)
Single high risk infections by HPV other than 16, 18, 33 or 45^a	38	17(44.7)	21(55.3)	28	13(46.4)	15(53.6)
Multiple infections including at least one high risk HPV type^b	24	11(45.8)	13 (54.2)	18	10 (55.5)	8(44.4)
Low-risk HPV infections^c	12	2(16.7)	10(83.3)	0	-	-
Undetermined	19	4 (21.1)	15 (78.9)	6	4 (66.7)	2 (33.3)

^a Includes infections by HPV 26 (1 case), 30 (1 case), 31 (4 cases), 35 (1 case), 39 (4 cases), 51 (2 cases), 52 (9 cases), 56 (6 cases), 58 (4 cases), 68 (2 cases), 69 (1 case), 70 (1 case), 73 (1 case), and 102 (1 case)

^b Includes a) double infections by HPV6 and 16 (n=4); HPV16 and 18 (n=2); HPV16 and 31 (n=1); HPV16 and 33 (n=1); HPV16 and 51 (n=1); HPV18 and 11 (n=1); HPV18 and 44 (n=2); HPV18 and 74 (n=1); HPV31 and 33 (n=1); HPV31 and 42 (n=1); HPV33 and 56 (n=1); HPV35 and 66 (n=1); HPV44 and 45 (n=1); HPV44 and 58 (n=1); HPV44 and 66 (n=1); b) triple infections HPV31, 33 and 58 (n=1); HPV51, 68 and 73 (n=1); and c) quadruple infections HPV31, 33, 44 and 45 (n=1); and HPV51, 53, 54 and 58 (n=1)

^c Includes single infections by HPV 6 (n=2), 11 (n=1), 44 (n=3), 53 (n=1), 61 (n=2), 74 (n=2) and a double infection 42 and 7 (n=1).

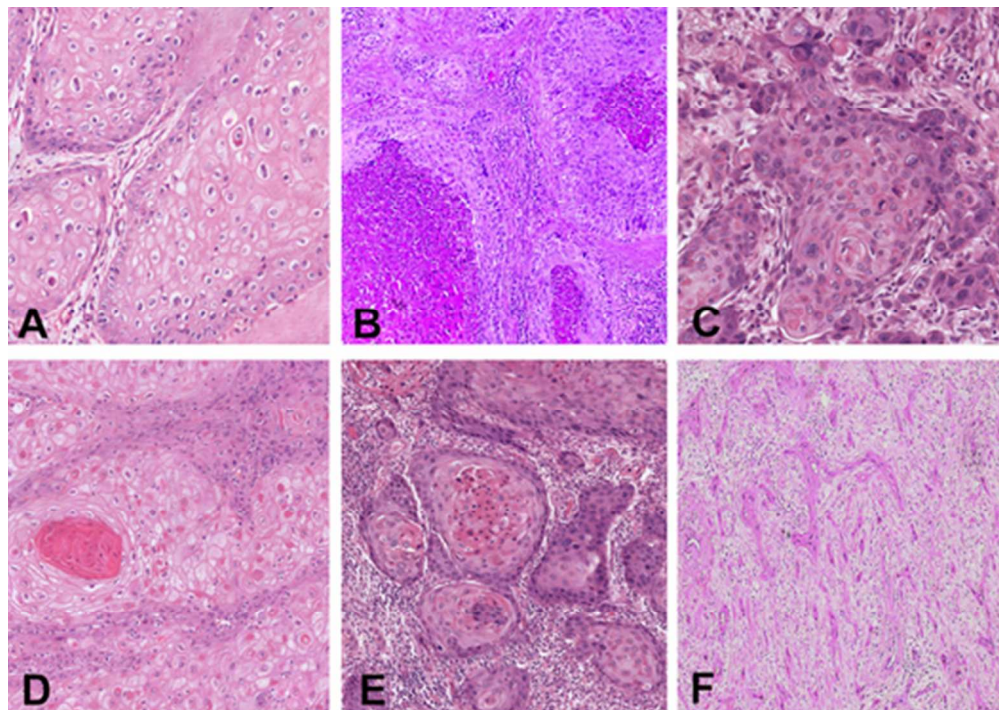


FIGURE 1. Histological characteristics evaluated in the study: A) koilocytotic-like change; B) necrosis; C) moderate to marked pleomorphism; D) individual keratinization, keratin pearls, and highly differentiated cells; E) invasive front in nests; F) diffuse invasive front. Hematoxylin and eosin staining.

22x15mm (600 x 600 DPI)

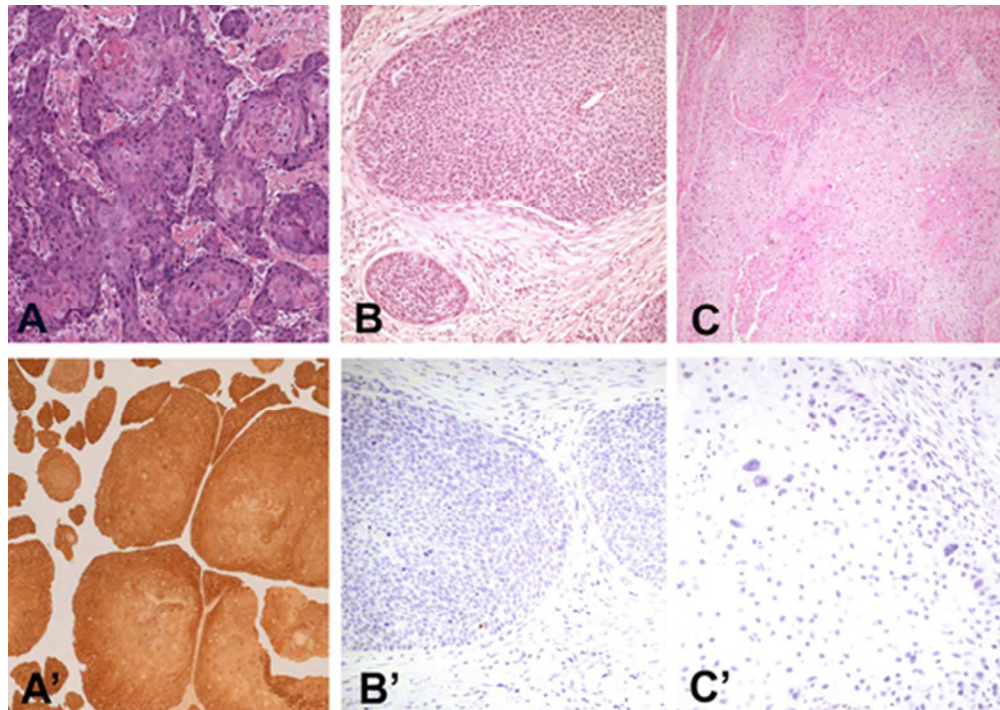


FIGURE 2. A) Keratinizing VSCC unquestionably HPV-associated tumor (HPV DNA+/p16+); B) indisputably HPV-independent VSCC (HPV DNA-/p16-) of basaloid type; C) indisputably HPV-independent VSCC (HPV DNA-/p16-) of warty type. Hematoxylin and eosin staining (A, B, C) and p16 immunohistochemistry (A', B', C').

22x15mm (600 x 600 DPI)