A novel orally available inhibitor of focal adhesion signaling increases survival in a xenograft model of diffuse large B-cell lymphoma with central nervous system involvement

Rosa Bosch,^{1,2} María José Moreno,^{1,2,3} Rebeca Dieguez-Gonzalez,¹ María Virtudes Céspedes,^{1,3} Alberto Gallardo,⁴ Manuel Trias,⁵ Albert Grañena,⁶ Jorge Sierra,⁷ Isolda Casanova,^{1,3} and Ramon Mangues^{1,3}

¹Grup d'Oncogènesi i Antitumorals, Institut d'Investigacions Biomèdiques Sant Pau, Barcelona; ²Department of Biochemistry and Molecular Biology, Universitat de Barcelona; ³CIBER en Bioingeniería, Biomateriales y Nanomecidicina (CIBER-BBN), Barcelona; ⁴Department of Pathology, Clínica Girona, Girona; ⁵Department of Surgery, Hospital de la Sant Creu i Sant Pau, Barcelona; ⁶Department of Hematology, Hospital Universitari de Bellvitge, Barcelona; and ⁷Department of Hematology, Hospital de la Santa Creu i Sant Pau, Barcelona, Spain

ABSTRACT

Central nervous system dissemination is a relatively uncommon but almost always fatal complication in diffuse large B-cell lymphoma patients. Optimal therapy for central nervous involvement in this malignancy has not been established. In this paper, we aimed to evaluate the therapeutic effect of E7123, a celecoxib derivative that inhibits focal adhesion signaling, in a novel xenograft model of diffuse large B-cell lymphoma with central nervous system involvement. Cells obtained after disaggregation of HT subcutaneous tumors (HT-SC cells) were intravenously injected in NOD/SCID mice. These mice received oral vehicle or 75 mg/kg of E7123 daily until they were euthanized for weight loss or signs of sickness. The antitumor effect of E7123 was validated in an independent experiment using a bioluminescent mouse model. Intravenously injected HT-SC cells showed higher take rate and higher central nervous system tropism (associated with increased expression of β 1-integrin and p130Cas proteins) than HT cells. The oral administration of E7123 significantly increased survival time in 2 independent experiments using mice injected with unmodified or bioluminescent HT-SC cells. We have developed a new xenograft model of diffuse large B-cell lymphoma with central nervous system involvement that can be used in the pre-clinical evaluation of new drugs for this malignancy. E7123 is a new, well-tolerated and orally available therapeutic agent that merits further investigation since it may improve current management of diffuse large B-cell lymphoma patients with central nervous system involvement.

۲

Introduction

Central nervous system (CNS) dissemination has been reported in 2-10% of diffuse large B-cell lymphoma (DLBCL) patients.^{1,2} Although relatively uncommon, it is fatal in over 90% of patients.^{3,4} Most CNS metastases occur in the setting of DLBCL relapse and their treatment is mostly palliative.⁵ The factors most consistently associated with risk of CNS recurrence are elevated serum lactate dehydrogenase and involvement of more than one extranodal site.⁶⁹ CNS-directed prophylactic intrathecal therapy is often used to reduce the risk of CNS recurrence in these patients but its benefit has been questioned in recent years.^{1,10,11} The addition of rituximab to standard cyclophosphamide, doxorubicin, vincristin and prednisone (CHOP), has improved the outcome of DLBCL patients,¹²⁻¹⁶ but the protective effect of rituximab on CNS recurrence is controversial.^{2,17-21} Thus, the optimal treatment for CNS dissemination in DLBCL has not yet been established and new therapies are needed in order to prevent or treat this fatal condition. However, the lack of appropriate animal models that reproduce CNS involvement in DLBCL hinders the development of new drugs.

Brain metastasis is a multistep process of tumor cell attach-

ment to microvessel endothelial cells, extravasation into the brain, interaction with the local microenvironment and proliferation. This process involves precise regulation of cell-cell adhesion and cell-extracellular matrix (ECM) adhesion.^{22,28} The cell-ECM adhesion is maintained by the focal adhesion complexes, composed of transmembrane receptors known as integrins, structural proteins (i.e. vinculin, talin, paxilin) and signaling proteins (i.e. FAK, p130Cas, HEF1, Pyk2).²⁴ A critical step in the CNS metastasis process is tumor cell adhesion to the vascular basement membrane of the brain which is mediated by integrin signaling.^{25,26} Thus, agents that efficiently block tumor cell adhesion mediated by focal adhesion signaling in DLBCL may prevent CNS progression.

We recently found that E7123, a novel non-COX-2 celecoxib derivative capable of inhibiting focal adhesion signaling, had *in vitro* antitumor effect against DLBCL cell lines and reduced tumor volume in a NOD/SCID subcutaneous model of DLBCL.²⁷ Our group has also shown that subcutaneous conditioning of cells prior to intravenous injection of DLBCL cell lines increased tumor take rate and cell dissemination capacity as compared to direct intravenous injection in NOD/SCID mice.²⁸ In this paper, we aimed to evaluate the therapeutic effect of E7123 in a novel xenograft mouse model

©2013 Ferrata Storti Foundation. This is an open-access paper. doi:10.3324/haematol.2012.071811 The online version of this article has a Supplementary Appendix. RB and MJM as well as IC and RM contributed equally to this manuscript.

Manuscript received on June 7, 2012. Manuscript accepted on May 15, 2013. Correspondence: rmangues@santpau.cat of DLBCL with CNS involvement. The animal model was generated by performing a subcutaneous passage of HT cells, which had previously shown CNS tropism after direct intravenous injection, before their intravenous injection in NOD/SCID mice.

Design and Methods

Cell line and compounds

HT human DLBCL cell line (DMSZ Cell Line Bank) was cultured in RPMI 1640 supplemented with 10% fetal bovine serum, 1% glutamine, 100 units/mL penicillin/streptomycin (Life Technologies) and incubated at 37°C in a humidified atmosphere containing 5% CO₂. HT-SC cells or bioluminescent HT-Luc-SC cells were obtained from HT or HT-Luc subcutaneous tumors, respectively (*see below*). E7123 was synthesized by the HSCSP Pharmacy Department and Laboratories Esteve S.A.

Lentiviral infection of HT cells

A plasmid containing the Luciferase gene (Luc) was constructed by subcloning and replacing the GFP1 gene of the plasmid pSIN-DUAL-GFP1-GFP2, kindly provided by Dr Mary Collins (College Medical School, Cleveland, OH, USA). Luciferase cDNA was obtained from the plasmid pPK-CMV-F3 (C-Luc) (Promokine, Heidelberg, Germany). Lentiviral particles containing the pSIN-DUAL-Luciferase-GFP2 vector, were used to infect HT cells as described previously by our group.²⁷ Infected cells, termed HT-Luc cells, were used for *in vivo* experiments.

Animal experiments

Animal procedures were approved by the Hospital Sant Pau Animal Ethics Committee according to established guidelines. NOD/SCID mice were subcutaneously inoculated with HT or HT-Luc cells. Cells obtained from subcutaneous tumors, HT-SC or HT-Luc-SC, were intravenously injected in mice to evaluate the aggressiveness of the subcutaneous conditioning and to evaluate E7123 antitumor effect. Mice were orally administered with E7123 (75 mg/kg) or vehicle (PEG:FBS) until they were euthanized (*Online Supplementary Figures S1 and S2*). End points were a 15% loss in body weight or the appearance of motor problems or evident signs of sickness. *In vivo* bioluminescence imaging (BLI) was performed as previously described.²⁹ Dorsal BLI pictures were taken once a week (ORCA-II Deep Cooling BTW Imaging System, Hamamatsu Photonics, Hamamatsu City, Japan) and quantified as described in the *Online Supplementary Appendix*.

Immunohistochemistry

Paraffin-embedded tissue sections of HT-SC or HT infiltrated brains were used to detect CD20, CD10, Ki67 (DAKO, Carpinteria, CA, USA), p130Cas (Neomarkers, Fremont, CA, USA) and β 1-integrin (Upstate, Billerica, MA, USA) proteins. Immunohistochemical reactions were performed in a DAKO Autostainer Link48 following the manufacturer's instructions. All slides were viewed with an Olympus BX51 microscope. Images were acquired using an Olympus DP72 digital camera and processed with the Olympus Cell^D Imaging 3.3 software (Olympus Corporation, Tokyo, Japan).

Western blot analysis

HT-SC cell lysates were used to evaluate protein expression. Western blots were performed as previously described.³⁰ The following primary anti-human antibodies diluted in TBS-T, containing 0.1% BSA, were used: mouse anti-p130Cas, anti-AKT, anti-FAK, anti-FAK Tyr397, anti-Pyk2, anti-Lyn, anti-Mcl1, anti-integrinβ1 (BD Pharmingen, San Jose, CA, USA), rabbit anti-AKT

Statistical analysis

Differences in survival between control HT or HT-SC groups and between HT-SC or HT-Luc-SC mice receiving vehicle or drug were calculated using Kaplan-Meier curves and the log rank test. Fisher's exact test was used to calculate differences between categorical variables. *P*<0.05 was considered statistically significant. Further details are described in the *Online Supplementary Appendix*.

Results

Subcutaneous passage of the HT cell line enhanced engraftment

Previous studies revealed that direct intravenous injection of DLBCL cells in NOD/SCID mice showed inefficient take rate and took a long time to generate lymphoma. However, when we performed a subcutaneous passage of the same DLBCL cell lines before their intravenous injection in mice their aggressiveness was dramatically increased, producing a disseminated lymphoma in all mice in a short period of time.²⁸ In a preliminary experiment, we observed that direct intravenous injection of HT cells produced lymphoma in 2 of 10 mice at 66 and 74 days after injection, and that both mice showed CNS infiltration. In order to increase the take rate and the aggressiveness of these cells, we performed a subcutaneous passage of HT cells before their intravenous injection, and compared their take rate and the animal survival with control mice directly injected with HT cells, without previous subcutaneous passage (see Design and Methods). Cells obtained after disaggregation of HT subcutaneous tumors were named HT-SC cells. In the intravenously injected HT-SC group, one mouse had to be excluded from the experiment as a tumor developed in the injection point. The remaining mice in this group (n=7) were euthanized between Days 40 and 51 post-injection, when they reached a 15% weight loss. In contrast, only 2 of 10 HT control mice were euthanized at Days 78 and 89 after injection as a consequence of weight loss. The remaining mice in the HT control group were sacrificed at Day 100 without any sign of disease and histopathological analysis revealed no tumor development. There was a statistically significant difference in survival times between groups (P < 0.01) as shown by Kaplan-Meier analysis (Figure 1A). Interestingly, mice from the experimental HT-SC group showed a visible cranial inflammation at approximately Day 30 after injection.

Intravenous injection of HT-SC cells generated DLBCL in the CNS

Most mice showed motor difficulties (generally circular motion) before being sacrificed. At necropsy, we observed that mice from the experimental group showed an evident cranial inflammation in comparison with mice from the HT control group (Figure 1B). When analyzed microscopically, we observed that 100% of HT-SC mice showed brain infiltration, especially in leptomeninges (Figure 2A-H), whereas only one of 2 infiltrated HT control mice showed this involvement (Table 1). Infiltrated brains of the HT-SC mice were positive for CD20 (B-cell antigen) (Figure 2I) and CD10 (germinal center marker) (Figure 2J) immunostaining, confirming that HT-SC cells maintained the germinal center DLBCL phenotype found in the original HT cells. Immunostaining with the proliferation marker Ki67 (Figure 2K) showed a proliferation index of 70-80% in all lymph nodes, a common proliferation rate in DLBCL pathology

HT-SC cells showed higher p130Cas and β 1-integrin expression than HT cells

Based on previous evidence showing that in vivo passages were associated with changes in adhesion properties,^{28,31-33} and that the expression of some adhesion molecules could be associated with CNS-tropism,^{23,25} we studied the expression of β 1-integrin, p130Cas, FAK-P, Pyk2, Lyn and Src adhesion proteins in HT cell lysates before (control HT) and after (HT-SC) the subcutaneous passage. We also assessed other proteins associated with the focal adhesion pathway, such as Akt-P, Akt and Mcl-1.27 We detected a marked increase in p130Cas and β 1-integrin expression levels and a slight increase in Src and Mcl-1 expression levels in HT-SC cells as compared to HT cells. However, there was no alteration in FAK, FAK-P, Akt, Akt-P, Pyk2 and Lyn proteins during the subcutaneous conditioning of cells (Figure 3A). Immunohistochemistry of β 1integrin and p130Cas was performed in HT cells before the subcutaneous passage (HT control), in the subcutaneous tumors, in the cells obtained after tumor disaggregation (HT-SC), and in brain tissue sections (Figure 3B). The intensity of the β 1-integrin and p130Cas immunostaining was clearly higher in subcutaneous tumors and in HT-SC cells than in the control HT cells. We also observed that this high intensity was retained in HT-SC cells at the brain metastases generated after their intravenous injection in mice. β 1-integrin and p130Cas staining was always localized in the cytoplasm and membrane in HT cells, subcutaneous tumors, HT-SC cells and infiltrated brains.

E7123 increased survival time in the DLBCL animal model with CNS involvement

Ten days after intravenous injection of HT-SC cells, mice were randomly divided into two groups and treated orally with 75 mg/kg E7123 or vehicle. Mice were treated

every day until they were sacrificed for weight loss or signs of sickness. We used PEG-400:FBS 3:1 as a vehicle because it showed no toxicity in healthy mice and permitted the total solubilization of E7123 without changing its antitumor activity *in vitro*.²⁷ All mice receiving vehicle were sacrificed between Days 39 and 48 post-injection. However, mice receiving E7123 were sacrificed between 43 and 85 days after injection. There was a statistically significant difference in survival time between both groups (P=0.01) as assessed by the log rank test (Table 2). Table 2 also shows the microscopic infiltration of brain, lymph nodes or bone marrow of all mice.

E7123 increased survival time in a validation experiment using a bioluminescent mouse model of DLBCL with CNS infiltration

In order to validate the antitumor effect of E7123 in the new xenograft model of DLBCL disseminated to the CNS, we performed the same procedure using bioluminescent subcutaneously-conditioned HT cells (HT-Luc-SC).

HT-Luc-SC cells were intravenously injected in 20 NOD/SCID mice that were randomly divided into two groups receiving vehicle or 75 mg/kg E7123. The first CNS bioluminescent signal was observed in 3 vehicle-treated and 2 E7123-treated mice on Day 17 post-injection. CNS bioluminescence was observed in all mice between Days 17 and 57 after cell injection. Figure 4A shows the time course BLI signal in CNS of representative vehicle and E7123-treated mice. We could observe that the BLI signal in the CNS of the vehicle-treated mouse increased faster than in the E7123-treated mouse. BLI intensity quantification curves for both groups are represented in Figure 4B. Thus, DLBCL cell growth in CNS was delayed in E7123treated mice. However, E7123 did not block lymphoma cell invasion of the CNS since there were no significant differences in the time of initial detection of CNS BLI signal between groups.

E7123 also increased mice survival time in this model since vehicle-treated mice were killed between 36 and 58 days post-injection, whereas E7123-treated mice were euthanized between 48 and 91 days post-injection (Figure 4C). Two mice from each group were excluded from the

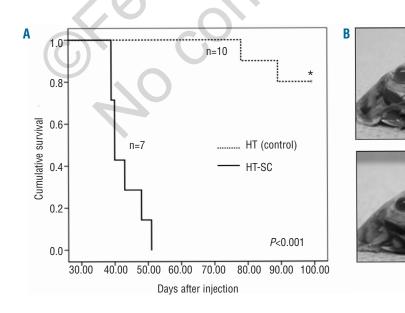


Figure 1. Survival in HT-SC mice is significantly lower than in control HT mice. (A) Kaplan-Meier analysis shows a significant reduction in survival time in HT-SC (n=7) as compared to control HT mice (n=10). *P* value for the log rank test was <0.01. (B) Macroscopic evidence for central nervous system involvement of DLBCL in NOD/SCID mice injected with HT-SC cells via the tail vein. *The remaining mice (n=8) were euthanized 95 days after injection without any sign of disease. experiment because they did not develop lymphoma. Details of the histopathological features and clinical evolution of HT-Luc-SC injected mice are shown in Table 3.

Therefore, we confirmed that E7123 increased mouse survival time in a bioluminescent model of DLBCL mice with CNS infiltration. In addition, as observed in the luminescent images, E7123 slowed down the growth of lymphoma cells within the CNS.

Discussion

In this study, we showed that the oral administration of a novel focal adhesion inhibitor, E7123, increased survival of mice with DLBCL involving the CNS. The subcutaneous passage of HT cells before their intravenous injection in NOD/SCID mice allowed us to generate a xenograft model of DLBCL with CNS involvement that can be used for evaluation of drugs for this malignancy. We observed that, during the subcutaneous passage, HT cells acquired an increased expression level of β 1-integrin and p130Cas proteins. This increased expression was associated with enhanced aggressiveness and CNS tropism of lymphoma cells.

The outcome of DLBCL patients with CNS occurrence is extremely poor and they do not appear to benefit from current prophylactic protocols.^{2,9-11} Thus, there is a need to develop new treatments for this fatal condition. However, the development of new therapies against this malignancy is hindered by the lack of appropriate animal models to evaluate them. Some authors reported xenograft models of primary CNS lymphoma (PCNSL) performed after injection of the aggressive Burkitt's lymphoma cells directly into the brain,³⁴⁻³⁷ whereas others generated syngeneic models of CNS lymphoma after intraocular or intracerebral injection of murine lymphoma cells in immunocompetent hosts.^{38,39} An important advantage of the latter models in respect to xenograft models of CNS lymphoma using immunodeficient mice is that syngeneic models

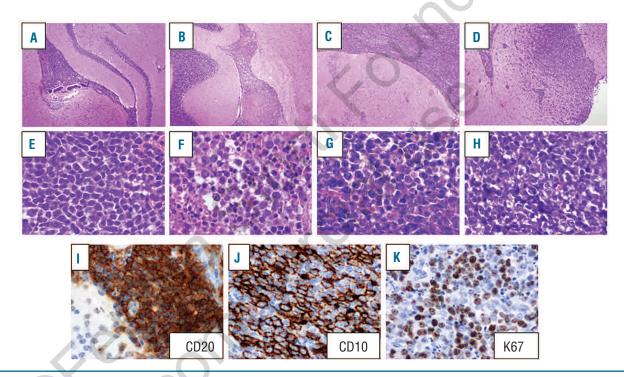


Figure 2. Histological and immunohistochemical phenotype of the DLBCL model with CNS involvement. H&E staining of representative brains in HT-SC mice. Lymphoma cells infiltrated the leptomeninges in most mice (A-D), reaching the parenchyma in isolated cases (D). Original magnification x400 and corresponding magnification x400 (E-H). Immunohistochemical analyses of CD20 (I), CD10 (J) and Ki67 (K) in representative sections of HT-SC infiltrated brains. Original magnification x400. Slides were viewed with an Olympus BX51 microscope. Images were acquired using an Olympus DP72 digital camera and processed with the Olympus Cell[®] Imaging 3.3 software.

| Table 1. Histopathological features and clinical ev | volution of NOD/SCID mice after intravenous | injection of HT or HT-SC cells. |
|---|---|---------------------------------|
| | | |

| Clinical features | Control HT | HT-SC | Р |
|---|----------------------------------|------------------------------------|-------------------------|
| N. of positive mice, n. (%) | 2/10 (20) | 7/7 (100) | 0.002 |
| Survival time (days) ¹ | 83.5 ± 7.8 | 42.9 ± 4.8 | <0.001 |
| N. of positive mice with CNS infiltration, n. (%) Meningeal infiltration, n. (%) Parenchymal infiltration, n. (%) | 1/2 (50) 1/2 (50) 1/2 (50) | 7/7 (100) 7/7 (100) 4/7 (57) | 0.222 0.222 1.000 |
| N. of positive mice with LN infiltration, n. (%) | 0/2 (0) | 1/7 (14) | 1.000 |
| N. of positive mice with BM infiltration, n. (%) | 1/2 (50) | 2/7 (29) | 1.000 |

Positive mice are those mice that developed lymphoma. P values were calculated using the log rank test for survival time and Fisher's exact test for categorical variables. CNS: central nervous system; LN: lymph node; BM: bone marrow. Values represent the mean ± SD.

allow the study of tumor-induced immunity.^{40,41} However, they are technically difficult to perform and they do not use human DLBCL cells. Here, we generated a rapid xenograft model of DLBCL with CNS involvement achieved after a simple intravenous injection of HT-SC cells in NOD/SCID mice. Cells maintained the typical morphology of DLBCL cells and retained the germinal center DLBCL marker expression of the original HT cells. Our model also showed a 100% of meningeal infiltration, the most typical infiltrating pattern found in DLBCL patients with CNS involvement.⁵ Moreover, the new biolumines-

cent model of DLBCL with CNS infiltration complements currently available murine models since it allows the study of the mechanisms of entrance, migration and growth of DLBCL cells in the CNS and their inhibition by drugs.

HT-SC cells used to generate this xenograft model showed higher levels of β 1-integrin and p130Cas expression than HT cells. It has recently been reported that β 1-integrin interaction with brain vessels is sufficient to promote proliferation and metastasis within the brain in carcinomas and lymphomas.^{25,26,42} Moreover, p130Cas overexpression is associated with metastases in different cancer

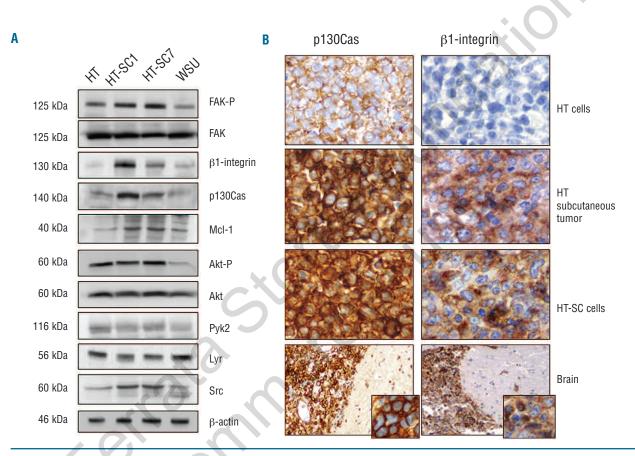


Figure 3. Expression of p130Cas and β 1-integrin in HT and HT-SC cells. (A) Immunoblotting of FAK (Tyr 397), FAK, β 1-integrin, p130Cas, Pyk2, Lyn, AKT (Thr 308) and AKT in protein extracts of control HT or HT-SC cells one day (HT-SC1) or seven days (HT-SC7) after disaggregation of subcutaneous tumors. WSU cell line lysate was used as positive control. HT-SC protein extracts showed higher expression levels of β 1-integrin and p130Cas than HT control cells. Equal loading was checked by immunobotting with β -actin. (B) Immunostaining analysis of p130Cas and β 1-integrin in control HT cells, HT subcutaneous tumor, HT-SC cells (original magnification x400) and HT-SC infiltrated brain sections (original magnification x100, inset x400). Slides were viewed with an Olympus BX51 microscope. Images were acquired using an Olympus DP72 digital camera and processed with the Olympus Cell^o Imaging 3.3 software.

Table 2. Histopathological features and clinical evolution of HT-SC injected NOD/SCID mice after administration of vehicle or E7123.

| Clinical features | Vehicle | E7123 | Р |
|---|------------------------------------|------------------------------------|-----------|
| N. of positive mice, n. (%) | 7/7 (100) | 9/9 (100) | _ |
| Survival time (days) ¹ | 42.9 ± 4.8 | 61.1 ± 15.4 | 0.001 |
| N. of positive mice with CNS infiltration, n. (%) Meningeal infiltration, n. (%) Parenchymal infiltration, n. (%) | 7/7 (100) 7/7 (100) 4/7 (57) | 9/9 (100) 9/9 (100) 7/9 (78) | 0.596 |
| N. of positive mice with LN infiltration, n. (%) | 1/7 (14) | 2/9 (22) | 1.000 |
| N. of positive mice with BM infiltration, n. (%) | 2/7 (29) | 2/9 (22) | 1.000 |

Positive mice are those that developed lymphoma. P values were calculated using the log rank test for survival time and Fisher's exact test for categorical variables. CNS: central nervous system; LN: lymph node; BM: bone marrow. 'Values represent the mean ± SD.

types.^{43,44} β 1-integrin and p130Cas are components of the same signaling pathway which is associated with tumor invasion.⁴⁵ Thus, the high β 1-integrin and p130Cas expression levels observed in HT-SC cells could partly explain the higher aggressiveness and capacity to migrate and grow into the CNS of DLBCL lymphoma cells as compared to HT cells. However, further functional studies are needed to determine whether β 1-integrin and/or p130Cas are responsible for HT-SC cells tropism to the CNS and/or growth into the brain. In a previous study, the non-COX2 inhibitor celecoxib derivative E7123 showed an *in vitro* antitumor

effect in different DLBCL cell lines by deregulation of focal adhesion proteins, which were expressed in the majority of human DLBCL patient samples. The oral administration of E7123 was well tolerated and reduced tumor growth in a subcutaneous xenograft of DLBCL.²⁷ In this paper, we showed that the focal adhesion inhibitor E7123 increases mice survival time in 2 independent experiments using a new xenograft model of DLBCL with CNS involvement. BLI images confirmed that E7123 did not block the CNS invasion, but delayed the growth of the DLBCL cells once in the brain, suggesting that E7123 may be capable of

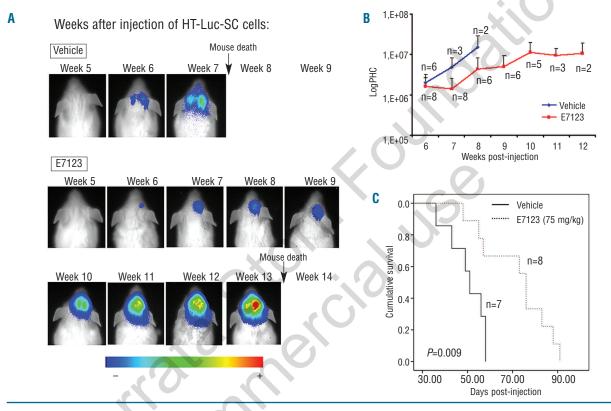


Figure 4. E7123 antitumor effect in an independent experiment using a bioluminescent model of DLBCL with CNS involvement. (A) Representative BLI images showing lymphoma HT-Luc-Sc cell growth evolution in the CNS in vehicle (PEG:FBS) or treated groups (75 mg/kg E7123). Vehicle-treated and E7123-treated mice were sacrificed on Days 56 and 92 post-injection of cells, respectively. Arbitrary color bars illustrate relative light intensity levels from firefly luciferase, ranging from low (blue) to high (red). Corresponding grayscale photographs and bioluminescent images were superimposed for each animal. (B) BLI signal quantification curves in the CNS for vehicle and E7123-treated mice. At week 6 after tumor cell injection, one vehicle-treated mouse could not be evaluated for BLI because it showed signs of sickness that precluded the use of anesthesia. BLI signal was quantified and expressed as Photon Counts (PHC). Curves represent the kinetics of BLI intensity mean for each group. Error bars represent the standard error. (C) Survival time in HT-Luc-SC mice treated with E7123 (75 mg/kg) was significantly higher than in control mice (PEG:FBS). *P* value for the log rank test was <0.01. Kaplan-Meier analysis showed the significant increase in survival time in treated mice (n=8) as compared to control mice (n=7).

Table 3. Histopathological features and clinical evolution of HT-Luc-SC injected NOD/SCID mice after administration of vehicle or E7123.

| Clinical features | Vehicle | E7123 | Р |
|---|----------------|------------|-------|
| N. of positive mice, n. (%) | 8/10 (80) | 8/10 (80) | - |
| N. of positive mice with CNS infiltration, n. (%) | 7/8 (87.5) | 8/8 (100) | 1.000 |
| Meningeal infiltration, n. (%) | 7/7 (100) | 8/8 (100) | - |
| Parenchymal infiltration, n. (%) | 5/7 (71.4) | 7/8 (87.5) | 0.596 |
| N. of positive mice with LN infiltration, n. (%) | 0/8 (0) | 2/8 (25) | 0.471 |
| N. of positive mice with BM infiltration, n. (%) | 1/8 (12.5) | 2/8 (25) | 1.000 |
| Survival time (days)1 of CNS positive mice | 50.1 ± 8.3 | 74±14.8 | 0.009 |

Positive mice are those that developed lymphoma. p values were calculated using the log rank test for survival time and Fisher's exact test for categorical variables. CNS: central nervous system; LN: lymph node; BM: bone marrow. Values represent the mean \pm SD.

crossing the blood-brain barrier. Most drugs included in the standard therapy for DLBCL, CHOP-R, do not reach the CNS^{11,20} and, consequently, do not prevent CNS relapse. Interestingly, E7123 parent compound, celecoxib, crossed the blood-brain barrier⁴⁶ and prolonged mice survival time in a xenograft model of PCNSL.³⁶ The clinical use of celecoxib, however, is associated with risk of cardiovascular events associated with COX-2 inhibition.⁴⁷ E7123, in contrast, does not inhibit COX-2. Thus, E7123 is likely to be effective for the treatment of CNS relapse in DLBCL without showing cardiovascular toxicity. Considering the unproven efficacy of current drugs in CNS relapse treatment, and the physical discomfort associated with intrathecal administration, orally available E7123 could be an alternative treatment in combination with current therapy to treat CNS involvement in DLBCL in the future.

In conclusion, E7123 is a new, well tolerated and orally bioavailable therapeutic agent that increases survival time in a novel xenograft model of DLBCL with CNS involvement. We believe E7123 merits further investigation, as findings to date suggest it may improve current management and Quality of Life of DLBCL patients with this fatal condition.

Acknowledgments

The authors would like to thank Montse Gómez and Iris Rodríguez for their technical support, and Carolyn Newey for editing the English. We also want to thank Josep Nomdedeu for his critical reading of the manuscript.

Funding

This work was supported by Instituto de Salud Carlos III FI08/00007 to RB, FI10/00758 to MJM, Sara Borrell CD09/00014 to RD, FIS PI08/0672 to JS, FIS PI11/00872 to JS, FIS PI06/1294 to RM, MiCINN PIB2010BZ-00563 to RM, CIBER-BBN CBV6/01/1031 to RM, AGAUR 2009-SGR-1437 to RM, Red de BioBancos RD09/0076/00081 to RM, Fundación de Investigación Médica Mútua Madrileña FMMA 04/305-475 to RM, FMMA2006-168 to JS, Fundació La Caixa BM05-258-0 to RM, Fundació La Marató TV3 100830 to JS; Fundació d'Investigació Sant Pau and Fundació José Carreras.

Authorship and Disclosures

Information on authorship, contributions, and financial & other disclosures was provided by the authors and is available with the online version of this article at www.haematologica.org.

References

- Korfel A. Prevention of central nervous system relapses in diffuse large B-cell lymphoma: which patients and how? Curr Opin Oncol. 2011;23(5):436-40.
- Tai WM, Chung J, Tang PL, Koo YX, Hou X, Tay KW, et al. Central nervous system (CNS) relapse in diffuse large B cell lymphoma (DLBCL): pre- and post-rituximab. Ann Hematol. 2011;90(7):809-18.
- 3. van Besien K, Ha CS, Murphy S, McLaughlin P, Rodriguez A, Amin K, et al. Risk factors, treatment, and outcome of central nervous system recurrence in adults with intermediate-grade and immunoblastic lymphoma. Blood. 1998;91(4):1178-84.
- Hollender A, Kvaloy S, Lote K, Nome O, Holte H. Prognostic factors in 140 adult patients with non-Hodgkin's lymphoma with systemic central nervous system (CNS) involvement. A single centre analysis. Eur J Cancer. 2000;36(14):1762-8.
- Gleissner B, Chamberlain M. Treatment of CNS dissemination in systemic lymphoma. J Neurooncol. 2007;84(1):107-17.
- Herrlinger U, Glantz M, Schlegel U, Gisselbrecht C, Cavalli E. Should intra-cerebrospinal fluid prophylaxis be part of initial therapy for patients with non-Hodgkin lymphoma: what we know, and how we can find out more. Semin Oncol. 2009;36(4 Suppl 2):S25-34.
- Boehme V, Zeynalova S, Kloess M, Loeffler M, Kaiser U, Pfreundschuh M, et al. Incidence and risk factors of central nervous system recurrence in aggressive lymphomaa survey of 1693 patients treated in protocols of the German High-Grade Non-Hodgkin's Lymphoma Study Group (DSHNHL). Ann Oncol. 2007;18(1):149-57.
- Shimazu Y, Notohara K, Ueda Y. Diffuse large B-cell lymphoma with central nervous system relapse: prognosis and risk factors according to retrospective analysis from a single-center experience. Int J Hematol.

2009;89(5):577-83.

- 9. Kridel R, Dietrich PY. Prevention of CNS relapse in diffuse large B-cell lymphoma. Lancet Oncol. 2011;12(13):1258-66.
- McMillan A. Central nervous system-directed preventative therapy in adults with lymphoma. Br J Haematol. 2005;131(1):13-21.
 Siegal T, Goldschmidt N. CNS prophylaxis
- Siegal I, Goldschmidt N. CNS prophylaxis in diffuse large B-cell lymphoma: if, when, how and for whom? Blood Rev. 2012; 26(3):97-106.
- Pfreundschuh M, Trumper L, Osterborg A, Pettengell R, Trneny M, Imrie K, et al. CHOP-like chemotherapy plus rituximab versus CHOP-like chemotherapy alone in young patients with good-prognosis diffuse large-B-cell lymphoma: a randomised controlled trial by the MabThera International Trial (MInT) Group. Lancet Oncol. 2006;7 (5):379-91.
- Coiffier B, Lepage E, Briere J, Herbrecht R, Tilly H, Bouabdallah R, et al. CHOP chemotherapy plus rituximab compared with CHOP alone in elderly patients with diffuse large-B-cell lymphoma. N Engl J Med. 2002;346(4):235-42.
- 14. Feugier P, Van Hoof A, Sebban C, Solal-Celigny P, Bouabdallah R, Ferme C, et al. Long-term results of the R-CHOP study in the treatment of elderly patients with diffuse large B-cell lymphoma: a study by the Groupe d'Etude des Lymphomes de l'Adulte. J Clin Oncol. 2005;23(18):4117-26.
- 15. Habermann TM, Weller EA, Morrison VA, Gascoyne RD, Cassileth PA, Cohn JB, et al. Rituximab-CHOP versus CHOP alone or with maintenance rituximab in older patients with diffuse large B-cell lymphoma. J Clin Oncol. 2006;24(19):3121-7.
- Sehn LH, Donaldson J, Chhanabhai M, Fitzgerald C, Gill K, Klasa R, et al. Introduction of combined CHOP plus rituximab therapy dramatically improved outcome of diffuse large B-cell lymphoma in British Columbia. J Clin Oncol. 2005;23(22): 5027-33.
- 17. Boehme V, Schmitz N, Zeynalova S, Loeffler

M, Pfreundschuh M. CNS events in elderly patients with aggressive lymphoma treated with modern chemotherapy (CHOP-14) with or without rituximab: an analysis of patients treated in the RICOVER-60 trial of the German High-Grade Non-Hodgkin Lymphoma Study Group (DSHNHL). Blood. 2009;113(17):3896-902.

- Feugier P, Virion JM, Tilly H, Haioun C, Marit G, Macro M, et al. Incidence and risk factors for central nervous system occurrence in elderly patients with diffuse large-Bcell lymphoma: influence of rituximab. Ann Oncol. 2004;15(1):129-33.
- Villa D, Connors JM, Shenkier TN, Gascoyne RD, Sehn LH, Savage KJ. Incidence and risk factors for central nervous system relapse in patients with diffuse large B-cell lymphoma: the impact of the addition of rituximab to CHOP chemotherapy. Ann Oncol. 2010;21(5):1046-52.
 Yamamoto W, Tomita N, Watanabe R,
- Yamamoto W, Tomita N, Watanabe R, Hattori Y, Nakajima Y, Hyo R, et al. Central nervous system involvement in diffuse large B-cell lymphoma. Eur J Haematol. 2010;85 (1):6-10.
- Kridel R, Dietrich PY. Prevention of CNS relapse in diffuse large B-cell lymphoma. Lancet Oncol. 2011;12(13):1258-66.
- Zheng Y, Lu Z. Paradoxical roles of FAK in tumor cell migration and metastasis. Cell Cycle. 2009;8(21):3474-9.
- Preusser M, Capper D, Ilhan-Mutlu A, Berghoff AS, Birner P, Bartsch R, et al. Brain metastases: pathobiology and emerging targeted therapies. Acta Neuropathol. 2012;123 (2):205-22.
- Hynes RO. Integrins: versatility, modulation, and signaling in cell adhesion. Cell. 1992;69(1):11-25.
- del Zoppo GJ, Milner R. Integrin-matrix interactions in the cerebral microvasculature. Arterioscler Thromb Vasc Biol. 2006; 26(9):1966-75.
- Carbonell WS, Ansorge O, Sibson N, Muschel R. The vascular basement membrane as "soil" in brain metastasis. PLoS

One. 2009;4(6):e5857.

- Bosch R, Dieguez-Gonzalez R, Cespedes MV, Parreno M, Pavon MA, Granena A, et al. A novel inhibitor of focal adhesion signaling induces caspase-independent cell death in diffuse large B-cell lymphoma. Blood. 2011;118(16):4411-20.
- Bosch R, Moreno MJ, Dieguez-Gonzalez R, Cespedes MV, Gallardo A, Nomdedeu J, et al. Subcutaneous passage increases cell aggressiveness in a xenograft model of diffuse large B cell lymphoma. Clin. Exp. Metastasis. 2012;29(4):339-47.
- Vilalta M, Degano IR, Bago J, Aguilar E, Gambhir SS, Rubio N, et al. Human adipose tissue-derived mesenchymal stromal cells as vehicles for tumor bystander effect: a model based on bioluminescence imaging. Gene Ther. 2009;16(4):547-57.
- Casanova I, Bosch R, Lasa A, Parreno M, Cespedes MV, Brunet S, et al. A celecoxib derivative inhibits focal adhesion signaling and induces caspase-8-dependent apoptosis in human acute myeloid leukemia cells. Int J Cancer. 2008;123(1):217-26.
- Fidler IJ. Selection of successive tumour lines for metastasis. Nat New Biol. 1973;242 (118):148-9.
- 32. Andreassen K, Mortensen B, Winberg JO, Huseby NE. Increased resistance towards oxidative stress accompanies enhancement of metastatic potential obtained by repeated in vivo passage of colon carcinoma cells in syngeneic rats. Clin Exp Metastasis. 2002;19(7):623-9.
- 33. Yasoshima T, Denno R, Kawaguchi S, Sato N, Okada Y, Ura H, et al. Establishment and characterization of human gastric carcinoma lines with high metastatic potential in the

Kellonn

liver: changes in integrin expression associated with the ability to metastasize in the liver of nude mice. Jpn J Cancer Res. 1996;87 (2):153-60.

- 34. Kim JA, Kim SJ, Do IG, Jin J, Nam DH, Ko YH, et al. Hypoxia-associated protein expression in primary central nervous system diffuse large B-cell lymphoma: does it predict prognosis² Leuk Lymphoma. 2011;52(2):205-13.
- Kadoch C, Dinca EB, Voicu R, Chen L, Nguyen D, Parikh S, et al. Pathologic correlates of primary central nervous system lymphoma defined in an orthotopic xenograft model. Clin Cancer Res. 2009;15(6):1989-97.
- Wang W, Kardosh A, Su YS, Schonthal AH, Chen TC. Efficacy of celecoxib in the treatment of CNS lymphomas: an in vivo model. Neurosurg Focus. 2006;21(5):E14.
- Muldoon LL, Lewin SJ, Dosa E, Kraemer DF, Pagel MA, Doolittle ND, et al. Imaging and therapy with rituximab anti-CD20 immunotherapy in an animal model of central nervous system lymphoma. Clin Cancer Res. 2011;17(8):2207-15.
- 38. Mineo JF, Scheffer A, Karkoutly C, Nouvel L, Kerdraon O, Trauet J, et al. Using human CD20-transfected murine lymphomatous B cells to evaluate the efficacy of intravitreal and intracerebral rituximab injections in mice. Invest Ophthalmol Vis Sci. 2008; 49(11):4738-45.
- Assaf N, Hasson T, Hoch-Marchaim H, Pe'er J, Gnessin H, Deckert-Schluter M, et al. An experimental model for infiltration of malignant lymphoma to the eye and brain. Virchows Arch. 1997;431(6):459-67.
- 40. Galand C, Donnou S, Molina TJ, Fridman WH, Fisson S, Sautes-Fridman C. Influence

of Tumor Location on the Composition of Immune Infiltrate and Its Impact on Patient Survival. Lessons from DCBCL and Animal Models. Front Immunol. 2012;3:98.

- Donnou S, Galand C, Touitou V, Sautes-Fridman C, Fabry Z, Fisson S. Murine models of B-cell lymphomas: promising tools for designing cancer therapies. Adv Hematol. 2012;98(3):701-4.
- Yoshimasu T, Sakurai T, Oura S, Hirai I, Tanino H, Kokawa Y, et al. Increased expression of integrin alpha3beta1 in highly brain metastatic subclone of a human non-small cell lung cancer cell line. Cancer Sci. 2004;95(2):142-8.
- Kim M, Gans JD, Nogueira C, Wang A, Paik JH, Feng B, et al. Comparative oncogenomics identifies NEDD9 as a melanoma metastasis gene. Cell. 2006;125(7):1269-81.
- 44. Fromont G, Vallancien G, Validire P, Levillain P, Cussenot O. BCAR1 expression in prostate cancer: association with 16q23 LOH status, tumor progression and EGRP/KAI1 staining. Prostate. 2007;67(3): 268-73.
- 45. Sansing HA, Sarkeshik A, Yates JR, Patel V, Gutkind JS, Yamada KM, et al. Integrin alphabeta1, alphavbeta, alpha6beta effectors p130Cas, Src and talin regulate carcinoma invasion and chemoresistance. Biochem Biophys Res Commun. 2011;406(2):171-6.
- Dembo G, Park SB, Kharasch ED. Central nervous system concentrations of cyclooxygenase-2 inhibitors in humans. Anesthesiology. 2005;102(2):409-15.
 Chi L, Li L, Karan R, Kuman R, Kuma
- Chakraborti AK, Garg SK, Kumar R, Motiwala HF, Jadhavar PS. Progress in COX-2 inhibitors: a journey so far. Curr Med Chem. 2010;17(15):1563-93.