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Escape from adamantane: scaffold optimization of novel P2X7 antagonists featuring complex polycycles

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ABSTRACT

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Scaffold replacement

The adamantane scaffold, despite being widely used in medicinal chemistry, is not devoid of problems. In the recent years we have developed new polycyclic scaffolds as surrogates of the adamantane group with encouraging results in multiple targets. As an adamantane scaffold is a common structural feature in several P2X7 receptor antagonists, herein we report the synthesis and pharmacological evaluation of multiple replacement options of adamantane that maintain a good activity profile. Molecular modeling studies support the binding of the compounds to a site close to the central pore, rather than to the ATP-binding site and shed light on the structural requirements for novel P2X7 antagonists.

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The P2X7 receptor is an ionotropic ligand-gated purinergic receptor which is activated by extracellular ATP. ¹ Upon the application of physiological concentrations of this messenger, the P2X7 receptor acts as a cation selective ion channel, allowing the passage of Na⁺, K⁺ and Ca²⁺ through the plasma membrane, activating several signal transduction pathways for the correct function of the organism.² However, when this receptor is exposed to high concentrations of ATP for a long period of time, in the high micromolar range, it induces the formation of a non-selective large pore³ permeable to molecules up to 900 Da, which ultimately triggers cell apoptosis.⁴

This ion channel is present in virtually all tissues of the body, being especially abundant in cells of hematopoietic lineage. Due to the widespread presence of the P2X7 receptor in the mammalian organism, its malfunction has been linked to a vast number of pathological conditions, including rheumatoid arthritis (RA), osteoporosis, Parkinson's disease, multiple sclerosis and cancer, among other relevant inflammatory, immune or musculoskeletal disorders. Taking into account that modulation of the P2X7 offers an ample possibility of therapeutic intervention, it is no surprise that in recent years several academic and non-academic groups have directed their research efforts to the development of potential P2X7 antagonists.

Resulting from this endeavour several structurally diverse small molecule inhibitors have come to light, placing the most promising ones into clinical trials (Figure 1). Unfortunately, all the candidates to date have failed in this process, resulting in a void in the P2X7 therapeutic pipeline.

Figure 1. Structures of unsuccessful P2X7 clinical candidates. Two additional drugs (AFC-5128 and EVT-401, of undisclosed structures, also failed in clinical trials).

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One of the most promising chemical classes of P2X7 antagonists is found in the adamantyl derivatives. Indeed, this class of compounds had its most auspicious example in the AZD9056 (Figure 1), which successfully progressed to phase II clinical trials; regrettably it failed to show significant efficacy in several inflammatory diseases (osteoarthritis, RA, COPD and Chrohn's disease). Unexpectedly after this failure, the adamantane core was not only not abandoned, but regained interest as a scaffold for P2X7 antagonists. Proof of this are the several patents filled by AstraZeneca, Lundbeck, Renovis and Abbot. 10 Even more curious is the case of the academic groups that also resisted abandoning this scaffold, eagerly pursuing the preparation of adamantyl derivatives as P2X7 antagonists. Worth mentioning are the compounds developed by Lee et al., 11 such as **2**, and the C-2 disubstituted adamantanes developed by Battilocchio et al., such as **3**. Despite the aforementioned combination of industry and academic efforts, modifying other moieties of the adamantyl derivatives has not met the expectation to overcome the drawbacks found in the past and send a compound to clinic. Surprisingly, little to no effort has been dedicated to exploring the possibilities of the adamantane moiety replacement. Among the few examples reported so far, we find only weakly active compounds such as norbornane 4,11 and cubane 5 (Figure 2).¹³

1
$$IC_{50} = 11.5 \text{ nM}$$
2 $IC_{50} = 13 \text{ nM}$
3 $IC_{50} = 7.7 \text{ } \mu\text{M}$
4 $IC_{50} = 2.8 \text{ } \mu\text{M}$
5 $IC_{50} = 0.4 \text{ } \mu\text{M}$

Figure 2. Selected adamantane derivatives (1-3) with activity as P2X7 receptor antagonists and examples of adamantane replacements (4-5).

Taking into account the vast experience of our group in successfully replacing the adamantane with other tailor made polycyclic moieties and the capability of the aforementioned compounds to undergo structural modification we decided to explore this understudied path. ¹⁴

For this purpose, we envisaged four types of suitable scaffolds that may act as adamantane replacement for their assessment in terms of P2X7 receptor activity. The aliphatic polycyclic scaffolds were judiciously chosen taking into account several factors, first of all the maintenance of the desirable drug-like properties shown by the adamantane compounds. Secondly, the fact that they already proved as suitable adamantane replacements in our hands, yielding to potent compounds. Finally, considering that when this work was carried out no crystal structures of the P2X7 were available, 15 we aimed to explore the chemical space in the drug-binding site. To this aim, starting from the known nanomolar inhibitor 1, the bulkier substitution possibilities were embodied by I, its saturated analogue II, which display a similar shape index as adamantyl compounds (see below), and III, which exhibited a greater length alternative with restricted free rotation of the phenylhydrazine moiety. On the other hand, taking into account that smaller analogues of adamantane such as the norbornane and the cubane scaffolds exemplified by 4 and 5 led to poor antagonists, just one smaller scaffold (**IV**) was envisaged (Figure 3).

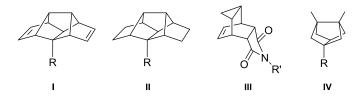


Figure 3. Adamantane replacement scaffolds. R = CONHNHAr, $\mathbb{R}^2 = NHAr$.

Diene **6**, envisaged as a suitable precursor of **I** and **II**, was accessed through a concise 7-step synthetic route previously developed by our group. ¹⁶ Upon treatment of **6** with thionyl chloride, the resulting acid chloride **7** was obtained and readily reacted with the desired hydrazine, to yield hydrazides **8-11** in good yields. The saturated analogues **12-14** were successfully prepared through a double bond reduction with hydrazide monohydrate under oxygen atmosphere. ¹⁷ The compounds with the general structure **III** were easily accessed through the reaction of 1,3,5-cyclohepatriene with maleic anhydride, which furnished the polycyclic core in good yield. ¹⁸ Upon reaction of **15** with the corresponding phenylhydrazine, the compounds **16 - 18** were dehydrated under Dean-Stark conditions to yield the cyclized adducts **19 - 21** (Scheme 1).

As the pharmacology trends for the already tested series **I** and **II** placed the 2-Cl substitution among the preferred one (see below), for the scaffold **IV**, only compound **24** was prepared. Hence, the hydrazide **24** was accessed from the known bisnoradamantane monoacid precursor and, paralleling the procedure for the preparation of derivatives **I**, was treated with thionyl chloride and readily reacted with 2-chlorophenylhydrazine (Scheme 1).

The structure of all new compounds was confirmed by accurate mass measurement, IR, ¹H and ¹³C NMR spectroscopy.

Scheme 1. Reagents and conditions: (a) $SOCl_2$, Δ ; 2 h, quantitative yield. (b) substituted phenylhydrazine hydrochloride, triethylamine, anh. THF, overnight, 92-99% yield. (c) hydrazine monohydrate, O_2 , ethanol, rt, 24 h, >95% yield. (d) substituted phenylhydrazine, ethanol, rt, overnight. (e) toluene, Dean-Stark, Δ , 47-81% overall yield for d and e. R = see Table 1 for details. See supporting information for details.

All new compounds were evaluated for their antagonistic effects on 2'(3')-O-(4-benzoylbenzoyl)-ATP (BzATP)-induced ethidium bromide uptake in human embryonic kidney 293 (HEK 293) cells stably expressing the human P2X7 receptor, using compound KN62 as a positive control (Table 1).²⁰

In order to rationalize the results, the binding mode of all the compounds was analysed by docking in the extracellular domain of the homology models of the receptor hP2X7 and rP2X7, using AutoDock version 4.2.²¹ In agreement with the very recent crystal structures of five P2X7 antagonists in complex with a mammalian P2X7 receptor, ¹⁵ our calculations supported the ability of the surrogate adamantane scaffolds to work as allosteric inhibitors. They fill a hydrophobic cavity close to the central pore of hP2X7, localized deeper in the receptor than the ATP binding site of hP2X7 (Figure 4A-B). The calculations also provided some clues about the observed structure-activity relationship (SAR).

Regarding the aromatic moiety substitution, we took advantage of the SAR previously developed by Abbott around the adamantane derivative 1 that revealed electron-withdrawing substitutents in the ortho position of the phenyl ring as the preferred group. In fact, within our series, the ortho position clearly showed to be the preferred one for the activity (compare 9 vs 10 and 13 vs 14, table 1). Interestingly, the introduction of a substituent in the para position was not only detrimental for the activity of the monosubstituted derivative (e.g., 10), but was able to abolish the benefitial effect of the ortho group (e.g., 11). This observation is explained by the steric clash of the paratrifluoromethyl group with the residue Phe95 of the receptor that has been previously shown to play a key role by forming π -stacking interactions with distinct inhibitors (Figure 4C).

Within this series, the most potent compound, $8 (IC_{50} = 96 \text{ nM})$, bearing a chlorine atom in the ortho position, was nearly 9 times less potent than its adamantane counterpart, $1 (IC_{50} = 11.5 \text{ nM})$. Notwithstanding, 8 was much more potent than previously studied replacements for adamantane (e.g., 4 and 5). Moving from dienes I to alkanes II led to higher IC_{50} values (compare 8 vs 12 and 9 vs 13), so full saturation of the structure is detrimental for the activity. This is rationalized as the dihedral angle formed by the amide and the aromatic ring, suffers a slight tilt in the saturated compound, compared to its more active unsaturated analogue (Figure 4D).

As the hydrogen atom of the CONH group did not seem to play an essential role in the binding, we drew our attention to the conformationally restricted series III. Neither the unsubstituted phenyl derivative, 19, nor the ortho substituted analogues 20 and 21, displayed significant activity, a fact that indicates that a degree of conformational freedom in the aromatic moiety is required when designing P2X7 inhibitors.

Fully unexpectedly, taking into account the weak inhibitory activity displayed by known smaller replacement of adamantane, such as **4** and **5** (see above), compound **24** was revealed as the most potent analogue synthesized in this work, displaying an activity in the low nanomolar range ($IC_{50} = 18 \text{ nM}$) which pairs the one displayed by its adamantane analogue **1** ($IC_{50} = 11.5 \text{ nM}$).

Strikingly, upon docking in the receptor, hydrazide **24** showed to adopt a completey different orientation (Figure 4E-F) than the one shown by its adamantane analogue **1** and **8**, which parallels the adamantane compound in the binding site. Moreover, upon docking in the rP2X7 receptor, no unfavourable interactions were observed for **24**, suggesting good antagonist potency. In view of the fact that several classes of inhibitors display significantly reduced the binding affinity in rP2X7 receptor, complicating

further in vivo studies, our docking results further highlight the potential interest of bisnoradamantane derivatives as surrogates of adamantane.

Table 1. P2X7 receptor antagonist activity of novel compounds.

Compound	R	IC ₅₀ (nM)
8	2-Cl	96 ± 8
9	2-CF ₃	132 ± 20
10	4-CF ₃	<20% b
11	2-Cl, 4-CF ₃	1557 ± 500
12	2-Cl	933 ± 156
13	2-CF ₃	366 ± 37
14	4-CF ₃	<20% ^b
19	Ħ	<20% ^b
20	2-Cl	<20% ^b
21	2-F	<20% ^b
24	2-Cl	18 ± 2
KN-62	2-Cl	96 ± 11

^aExperiments were assessed in the ethidium accumulation assay using hP2X7R-expressing HEK293 cells. The IC₅₀ values were obtained from concentration-response curves. Data are expressed as the means SD of at least 3-6 experiments. ^bPercent inhibition at concentration 10 μM.

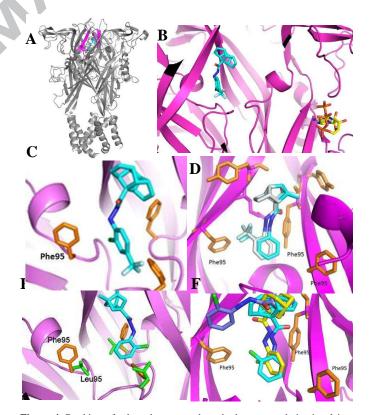


Figure 4. Docking of selected compounds and adamantane derivative 1 into the human P2X7 receptor. A. Compounds binding site in the whole extracellular domain. B. Compounds binding site in relation with the ATP binding site. C. Steric clash of the trifluoromethyl group of 10. D. Superposition of 8 (light blue) and 12 (grey) in the binding site. E. Bisnoradamantane derivative 24 in the receptor binding pocket. Position 95 being Leu in the rP2X7 and Phe in the hP2X7. F. Superposition of 1 (yellow), 8 (light blue) and 24 (purple) in their binding site.

To further explore the suitability as an adamantane replacement we predicted their physicochemical properties using DataWarrior. Of note, **24** and its isomer **1** displayed very similar physicochemical properties (Table 2). One of the main concerns when moving adamantane derivatives further into drug discovery programs is its easy metabolization by oxidation at the bridgehead positions. Taking into account that scaffold **IV** features two of the bridgehead positions substituted its metabolization by oxidation might be more difficult.

Table 2. Relevant physicochemical properties for a representative example of each scaffold predicted by DataWarrior. ^a

each scaling predicted by Balan values.										
Compound (scaffold)	M. W.	cLogP	H-	H-	PSA	SI^b	MF^{c}	sp^3		
	(g/mol)		A	D	(\mathring{A}^2)			atoms		
1	304.82	4.32	3	2	41.13	0.52	0.53	10		
8 (I)	324.81	3.57	3	2	41.13	0.48	0.74	8		
12 (II)	328.84	4.12	3	3	41.13	0.48	0.74	12		
20 (III)	314.77	2.64	4	1	49.41	0.50	0.64	7		
24 (IV)	304.82	4.29	3	2	41.13	0.52	0.73	10		

^aAll the series were predicted not to be mutagenic or tumorogenic agents. ^bShape index. ^cMolecular flexibility.

In conclusion, eleven new compounds have been synthesized from four available polycyclic compounds as P2X7 antagonists. Several of these novel compounds displayed reasonable activity with 24, being the most potent with low nanomolar potency. To the best of our knowledge this is the first time, within the P2X7 antagonist context, that the adamantane scaffold has been successfully replaced by a different polycyclic structure. On top of that, 24 is predicted to have physicochemical properties that parallel those of adamantane, reinforcing its suitability as a scaffold to develop drug-like compounds. Taken together, we can conclude that the smaller bisnoradamantane scaffold IV revealed as a very promising surrogate of adamantane regarding P2X7 inhibition, in contrast to a hit-to-lead study reported by AstraZeneca that mentioned that no replacements for adamantane were found.²⁵

Research towards the development of novel polycyclic scaffolds to function as adamantane replacement and the exploration of further analogues of **24** are in progress to reach derivatives with optimum P2X7 activity and enhanced ADME properties.

Acknowledgments

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References and notes

- 1. Burnstock, G. Pharmacol. Rev. 1972, 24, 509-581.
- (a) Burnstock, G. Pharmacol Rev. 2006, 58, 58–86. (b) Coddou, C.; Yan, Z.; Obsil, T.; Huidobro-Toro, J.P; Stojilkovic, S. S. Pharmacol. Rev. 2011, 63, 641–683.
- Yan, Z; Li, S; Liang, Z; Tomic, M.; Stojilkovic, S. S. J. Gen. Physiol. 2008, 132, 563–573.
- Khadra, A.; Tomic, Melanija; Y., Zonghe; Z., H.; Sherman, A.; Stojilkovic, S. S. Biophys. J. 2013, 104, 2612–2621.
- Parvathenani, L. K.; Tertyshnikova, S.; Greco, C. R.; Roberts, S. B.; Robertson, B.; Posmantur, R. J. Biol. Chem. 2003, 278, 13309–13317.

- (a) De Marchi, E.; Orioli, E.; Dal Ben, D.; Adinolfi, E. Adv. Protein Chem. Struct. Biol. 2016, 104, 39–79. (b) Tewari, M; Seth, P. Ageing Res. Rev. 2015, 24, 328–342. (c) Skaper, S. D.; Debetto, P.; Giusti, P. FASEB J. 2010, 24, 337–345. (d) North, R. A.; Jarvis, M. F. Molec. Pharmacol. 2003, 83, 759–69. (e) Roger, S.; Pelegrin, P. Expert Opin. Investig. Drugs. 2011, 20, 875–80.
- (a) Arulkumaran, N.; Unwin, R. J.; Tam, F. W. Expert Opin. Investig. Drugs. 2011, 20, 897–915. (b) Di Virgilio, F.; Ceruti, S.; Bramanti, P.; Abbracchio, M. P. Trends Neurosci. 2009, 32, 79–87
- (a) Friedle, S. A.; Curet, M. A.; Watters, J. J. Recent Pat. CNS Drug Discov.2010, 5, 35–45. (b) Bartlett, R.; Stokes, L.; Sluyter, R. Pharmacol. Rev. 2014, 66, 638–75.
- (a) Keystone, E.C.; Wang, M.M.; Layton, M.; Hollis, S.; McInnes, I. B. Ann. Rheum. Dis. 2012. 71, 1630–1635. (b) Eser, A.; Colombel, J. F.; Rutgeerts, P.; Vermeire, S.; Vogelsang, H.; Braddock, M.; Persson, T.; Reinisch, W. Inflamm. Bowel Dis. 2015, 21, 2247-2253. (c) https://openinnovation.astrazeneca.com/azd9056.html, accessed on Dec. 12th, 2016.
- (a) Gunosewoyo, H.; Kassiou, M. Expert Opin. Ther. Pat. 2010, 20, 625–646. (b) Park, J.-H.; Kim, Y.-C. Expert Opin. Ther. Pat. doi: 10.1080/13543776.2017.1246538
- Lee, W.-G.; Lee, S.-D.; Cho, J.-H.; Jung, Y.; Kim, J.-H.; Hien, T. T.; Kang, K.-W.; Ko, H.; Kim, Y.-C. J. Med. Chem. 2012, 55, 3687–3698.
- Battilocchio, C.; Guetzoyan, L.; Cervetto, C.; Di Cesare Mannelli, L.; Frattaroli, D.; Baxendale, I. R.; Maura, G.; Rossi, A.; Sautebin, L.; Biava, M.; Ghelardini, C.; Marcoli, M.; Ley, S. V. ACS Med. Chem. Lett. 2013, 4, 704–709.
- (a) Gunosewoyo, H.; Guo, J. L.; Bennett, M. R.; Coster, M. J.; Kassiou, M. Bioorg. Med. Chem. Lett. 2008, 18, 3720–3723. (b) Wilkinson, S. M.; Gunosewoyo, H.; Barron, M. L.; Boucher, A.; McDonnell, M.; Turner, P.; Morrison, D. E.; Bennett, M. R.; McGregor, I. S.; Rendina, L. M.; Kassiou, M. ACS Chem. Neurosci. 2014, 5, 335–339.
- (a) Camps, P.; Duque, M. D.; Vázquez, S.; Naesens, L.; De Clercq, E.; Sureda, F. X.; López-Querol, M.; Camins, A.; Pallàs, M.; Prathalingam, S. R.; Kelly, J. M.; Romero, V.; Ivorra, D.; Cortés, D. Bioorg. Med. Chem. 2008, 16, 9925-9936. (b) Duque, M. D.; Camps, P.; Profire, L.; Montaner, S.; Vázquez, S.; Sureda, F. X.; Mallol, J.; López-Querol, M.; Naesens, L.; De Clercq, E.; Prathalingam, S. R.; Kelly, J. M. Bioorg. Med. Chem. 2009, 17, 3198-3206. (c) Duque, M. D.; Camps, P.; Torres, E.; Valverde, E.; Sureda, F. X.; López-Querol, M.; Camins, A.; Prathalingam, S. R.; Kelly, J. M.; Vázquez, S. Bioorg. Med. Chem. 2010, 18, 46-57. (d) Duque, M. D.; Ma, C.; Torres, E.; Wang, J.; Naesens, L.; Juárez- Jiménez, J.; Camps, P.; Luque, F. J.; DeGrado, W. F.; Lamb, R. A.; Pinto, L. H.; Vázquez, S. J. Med. Chem. 2011, 54, 2646-2657. (e) Torres, E.; Duque, M. D.; López-Querol, M.; Taylor, M. C.; Naesens, L.; Ma, C.; Pinto, L. H.; Sureda, F. X.; Kelly, J. M.; Vázquez, S. Bioorg. Med. Chem. 2012, 20, 942-948. (f) Rey-Carrizo, M.; Torres, E.; Ma, C.; Barniol-Xicota, M.; Wang, J.; Wu, Y.; Naesens, L.; DeGrado, W. F.; Lamb, R. A.; Pinto, L. H.; Vázquez, S. J. Med. Chem. 2013, 56, 9265-9274. (g) Torres, E.; Leiva, R.; Gazzarrini, S.; Rey-Carrizo, M.; Frigolé-Vivas, M.; Moroni, A.; Naesens, L.; Vázquez S. ACS Med. Chem. Lett. 2014, 5, 831-836. (h) Rey-Carrizo, M.; Barniol-Xicota, M.; Ma, C.; Frigolé-Vivas, M.; Torres, E.; Naesens, L.; Llabrés, S.; Juárez-Jiménez, J.; Luque, F. J.; DeGrado, W. F.; Lamb, R. A.; Pinto, L. H.; Vázquez, S. J. Med. Chem. 2014, 57, 5738-5747. (i) Valverde, E.; Sureda, F. X.; Vázquez, S. Bioorg. Med. Chem. 2014, 22, 2678-2683. (j) Rey-Carrizo, M.; Gazzarrini, S.; Llabrés, S.; Frigolé-Vivas, M.; Juárez-Jiménez, J.; Font-Bardia, M.; Naesens, L.; Moroni, A.; Luque, F. J.; Vázquez, S. Eur. J. Med. Chem. 2015, 96, 318-329. (k) Valverde, E.; Seira, C.; McBride, A.; Binnie, M.; Luque, F. J.; Webster, S. P.; Bidon-Chanal, A.; Vázquez, S. Bioorg. Med. Chem. 2015, 23, 7607–7617.
- During the writing of this manuscript, the crystal structures of a mammalian P2X7 receptor complexed with five structurallyunrelated antagonists was reported, see: Karasawa, A.; Kawate, T. eLIFE 2016, 5, 322153.
- Camps, P., Pujol, X., Rossi, R. A., Vázquez, S. Synthesis. 1999, 854–58.
- 17. Menges, N.; Balci, M. Synlett. 2014, 25, 671-676.
- Abou-Gharbia, M.; Patel, U. R.; Webb, M. B.; Moyer, J. A.;
 Andree, T. H.; Muth, E. A. J. Med. Chem. 1988, 31, 1382–1392.
- (a) Camps, P.; Iglesias, C.; Rodríguez, M. J.; Grancha, M. D.; Gregori, M. E.; Lozano, R.; Miranda, M. A.; Figueredo, M.;

- Linares, A. A. Chem. Ber. **1988**, 121, 647–654. (b) Camps, P.; Font-Bardia, M.; Pérez, F.; Solans, X.; Vázquez, S. Angew. Chem. Int. Ed. Engl. **1995**, 34, 912–914. (c) Camps, P.; Lukach, A. E.; Rossi, R. A. J. Org. Chem. **2001**, 66, 5366–5373.
- P2X7 receptor antagonist activity. Dye uptake using the ethidium ion assay. HEK293 cells stably expressing P2X7 receptor were maintained in Dulbecco's modified Eagle's medium (DMEM) supplemented with 10% fetal bovine serum (FBS), 2 mM Lglutamine, and antibiotics (50 U/mL penicillin and 50 mg/mL streptomycin) in a humidified 5% CO2 atmosphere at 37 °C. We used Lipofectamine as a transfection reagent with a pcDNA3.1 vector-based plasmid harbouring hP2X7 receptor (Invitrogen). After diluting to 2.5 x 10⁶ cells/mL, buffer composed of 10 mM HEPES, 5mM N-methyl-D-glutamine, 5.6 mM KCl, 10 mM Dglucose, and 0.5 mM CaCl₂ (pH 7.4) supplemented with either 280 mM sucrose or 140 mM NaCl, and an 80 μL aliquot was added to each well of 96-well culture plates. The test com- pounds and 2'(3')-O-(4-benzoylbenzoyl)-ATP (BzATP) were then added, and the cells were incubated for 2 h in a humidified 5% CO2 atmosphere at 37°C. After incubation, a Bio-Tek FL600 fluorescence plate reader was used to measure the absorbance at an excitation wavelength of 530 nm and an emission wavelength of 590 nm. The inhibition (percent) of ethidium ion uptake was expressed as a relative value of the maximum accumulation when stimulated with BzATP only. To calculate the IC50 values, we calculated a series of dose-response data using nonlinear regression analysis (i.e., per-centage accumulation of ethidium bromide vs compound concentration).
- 21. Homology modelling. Structural models of the P2X7 receptor were produced based on the zebrafish P2X4 receptor crystal structure in the closed and ATP-bound states (Protein Data Bank code 4DW0 and 4DW1, respectively). Modeller version 9.12 was used to generate one hundred versions of each model. The five with the lowest energy were further analysed with MolProbity and those with the greatest percentage of residues in allowed regions of the Ramachandran plot were selected for use in this study. The ModLoop server was used to model the nonconserved loop region between the β2 and β3 strands. Molecular docking simulations. Molecular docking was carried out using AUTODOCK version 4.2. The target cavity file for each docking simulation consisted of the P2X7 receptor extracellular domain. The auxiliary program AutoGrid was used to generate the affinity grid files
- Nelson, D. W.; Sarris, K.; Kalvin, D. M.; Namovic, M. T.; Grayson, G.; Donnelly-Roberts, D. L.; Harris, R.; Honore, P.; Jarvis, M. F.; Faltynek, C. R.; Carroll, W. A. J. Med. Chem. 2008, 51, 3030–3034.
- Caseley, E. A.; Muench, S. P.; Baldwin, S. A.; Simmons, K.; Fishwick, C. W.; Jiang, L.-H. Bioorg. Med. Chem. Lett. 2015, 25, 3164–2167.
- 24. DataWarrior version 4.4.4.
- Baxter, A.; Bent, J.; Bowers, K.; Braddock, M.; Brough, S.;
 Fagura, M.; Lawson, M.; McInally, T.; Mortimore, M.; Robertson, M.; Weaver, R.; Webborn, P. Bioorg. Med. Chem. Lett. 2003, 13, 4047–4050.
- 26. Hattori, M.; Gouaux, E. Nature 2012, 485, 207-212.
- Webb, B.; Sali, A. Curr. Protoc. Bioinformatics 2006, unit 5.6. DOI: 10.1002/cphi.3
- Davis, I. W.; Leaver-Fay, A.; Chen, V. B.; Block, J. N.; Kapral, G. J.; Wang, X.; Murray, L. W.; Arendall, W. B.; Snoeyink, J.; Richardson, J. S. Nucleic Acids Res. 2007, 35(suppl 2), W375-W383.
- 29. Fieser, A.; Do, R. K. G.; Šali, A. Protein Sci. 2000, 9, 1753-1773.
- Morris, G.M.; Huey, R.; Lindstrom, W.; Sanner, M.F.; Belew, R.K.; Goodsell, D.S.; Olson, A.J. J. Comput. Chem. 2009, 30, 2785–2791.

Supplementary Material

Experimental procedures and characterization details for the new compounds.

Graphical Abstract

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Escape from adamantane: scaffold optimization of novel P2X7 antagonists featuring complex polycycles

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Marta Barniol-Xicota, Seung-Hwa Kwak, So-Deok Lee, Emily Caseley, Elena Valverde, Lin-Hua Jiang, Yong-Chul Kim* and Santiago Vázquez*

