# **Bacterial cellulose for increasing barrier**

# 2 properties of paper products

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#### 19 ABSTRACT

20 Bacterial cellulose was combined with wood cellulose papers in order to obtain 21 biomaterials with increased barrier properties. For this purpose, different parameters 22 were assessed: two producing bacterial strains (Komagataeibacter xylinus and 23 *Gluconacetobacter sucrofermentans*), two paper supports to hold bacterial cellulose 24 (filter paper and eucalyptus paper), two kinds of combined biomaterials (composite and 25 bilayer) and two drying temperatures (90°C and room temperature). Papers with increased barrier properties (100° of water contact angle, 1220s of water drop test and 26 air permeability  $<1\mu m (Pa \cdot s)^{-1}$ ) were obtained by the addition of bacterial cellulose to 27 each paper support. However, due to the lower initial barrier properties of filter paper, 28 29 higher improvements were produced with this paper. In addition, bacterial cellulose 30 provided smoother surfaces with higher gloss without a detrimental effect on physical 31 properties. Higher resistance to water absorption was obtained with K. xylinus possibly 32 explained by its longer size fibers than G. sucrofermentans, as analysed by SEM. 33 Smoothness and gloss were specially increased in the bilayer biomaterial although 34 resistance to air and water were further improved in the composite. In this biomaterial 35 drying at high temperature had a detrimental effect. SEM analysis of the products 36 obtained showed the intimate contact among fibers of bacterial cellulose and wood 37 paper. Results obtained show the contribution of bacterial cellulose to improve the 38 properties of paper and its potential for the design of new added value paper products 39 from biomass. 40 41 42 43 44 45 46 Keywords: bacterial cellulose, barrier properties, hydrophobicity, air permeability, 47 *water resistance, cellulose paper* 

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## 49 **INTRODUCTION**

50 Cellulose is the most abundant polymer of the Earth as a main component of 51 plant biomass. Due to its availability, it has been traditionally used as a raw material for 52 the production of a great diversity of industrial products including, paper, cardboard, 53 textiles, food additives and pharmaceutical products, among others. The renewed 54 interest for biomass valorization has fostered the research for the transformation and 55 modification of plant residues into increased value products as biofuels and 56 biomaterials, such as nanocellulose (Tuck et al. 2012; Beltramino et al. 2015, 2016). 57 One of the main problems found is the intimate association of cellulose with lignin and 58 hemicelluloses in plant biomass, in lignocellulose (Gilbert 2010). Deconstruction of 59 plant cell wall requires the development of technology to improve the separation and 60 upgrading of its lignocellulosic components in valuable new products (Gilbert 2010; 61 Quintana et al. 2013, 2015). Besides plants, some microorganisms can also produce 62 cellulose. Bacterial cellulose shows identical molecular composition to plant cellulose, 63 but it shows a major advantage: it is not associated to lignin and hemicellulose, it is a 64 high purity polymer.

65 Comparison of plant and bacterial cellulose show several properties that are favorably increased in bacterial cellulose, among which degree of polymerization and 66 67 crystallinity, that are remarkably high (Klemm et al. 2005). An important property of 68 bacterial cellulose is biocompatibility, that together with its elevated mechanical 69 strength has prompted its use in medical applications such as scaffold for tissue and skin 70 regeneration, artificial blood vessels, and as thickening food additive (Lin et al. 2013). 71 These applications are correlated to its high water holding capacity determined by a 72 structure of well separated cellulose nanofibrils, what makes bacterial cellulose a highly 73 porous material that can show up to more than 90% water content. However, this water 74 holding property is notably diminished after air drying, probably as a consequence of 75 the hydrogen-bond formation among cellulose fibrils (Klemm et al. 2005; Hagiwara et 76 al. 2010).

Mechanical properties of bacterial cellulose makes it an excellent candidate for
the restoration of damaged paper documents where its surface lining does not affect
document readability (Santos et al. 2015, 2016a, 2017). Application of bacterial

cellulose as a reinforcing agent of pulps in papermaking has also been studied showing
variable results depending on the pulp source (Yamanaka et al. 1989; Pommet et al.

2008; Gao et al. 2010; Tang et al. 2013; Xiang et al. 2017b), while its application for the

electronic and magnetic papers (Chawla et al. 2009; Shah et al. 2013; Lim et al. 2016).

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83 production of nanocomposites can give a diversity of high added value products such as

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85 On the other hand, barrier properties in papers (impermeability to air, water, 86 water vapor, oxygen, fats, microorganisms, etc.), that are especially important in the 87 food packaging sector, are currently provided by plastic films produced from 88 petrochemical products. However, due to the increase in social awareness regarding the 89 harmful environmental impact and the unsustainable life cycle of these materials, 90 research is focusing on the creation of new biomaterials from renewable resources, 91 which besides having these advanced barrier properties, may even become 92 biodegradable (Cusola et al. 2014). Bacterial cellulose, because of its specific 93 properties, can fulfil these requirements (Klemm et al. 2011; Osong et al. 2016). In fact, 94 previous works have demonstrated bacterial cellulose can decrease wettability and 95 permeability of paper (Gao et al. 2010; Santos et al. 2017; Xiang et al. 2017b).

96 The main purpose of this work was to combine bacterial cellulose with wood 97 cellulose in order to increase barrier properties of paper without a detrimental effect on 98 mechanical properties. Different aspects were taken into account, such as the microbial 99 strain, the paper type and the way of joining bacterial cellulose and paper. For this 100 purpose, bacterial cellulose produced by two different microbial strains was firstly 101 characterized. Then, the bacterial cellulose was combined with two wood paper types to 102 obtain two kind of biomaterials: composites or bilayers. In the composites, bacterial 103 cellulose was directly synthesized by the growth of the producing bacteria on the 104 surface of filter or eucalyptus paper sheets. In the bilayers, bacterial cellulose films were 105 previously synthesized and then coated over the surface of the paper sheets. The 106 properties of the resulting paper products were analyzed in terms of their mechanical 107 strength, optical and barrier properties, and SEM morphology.

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## **108 MATERIALS AND METHODS**

#### 109 Bacterial strains

110 Strains Komagataeibacter xylinus (DMS-2004) and Gluconacetobacter 111 sucrofermentans (CECT 7291) were obtained from the DSMZ German Collection of 112 Microorganisms and Cell Cultures and from the Spanish Type Culture Collection, 113 respectively. They were grown in Hestrin–Schramm (HS) solid medium (Hestrin 1954) in agar plates for maintenance. Suspensions of bacterial cells were obtained by gentle 114 115 shaking and inoculated in flasks containing 100 mL of HS liquid medium which were 116 incubated under static conditions for 4-7 days. Following, the surface bacterial films 117 produced were cut in small pieces (1x1 cm) in sterile conditions and shaken in HS 118 liquid medium at 700 rpm for 30 min to detach cells from the cellulose films. The 119 suspensions obtained were filtered through a gauze to remove film portions, centrifuged 120 at 4000 rpm for 10 min and, after discarding supernatants, pellets were resuspended in Ringer's solution  $\frac{1}{4}$  (NaCl 2.5 g L<sup>-1</sup>; KCl, 0.105 g L<sup>-1</sup>; CaCl<sub>2</sub>·2H<sub>2</sub>O, 0.120 g L<sup>-1</sup>; and 121 NaHCO<sub>3</sub>, 0.05 g  $L^{-1}$ ). Optical density of the bacterial suspensions was adjusted to 122 OD600 of 0.59–0.64 with Ringer's solution <sup>1</sup>/<sub>4</sub> and used as inoculum for the following 123 124 experiments.

#### 125 **Production of bacterial cellulose films**

Bacterial cellulose films were produced cultivating the bacterial strains in liquid
media in 150 mm diameter Petri dishes. 100 mL of HS liquid media were inoculated
with 250 μL of the bacterial suspension and incubated at 30°C for 10 days in static
conditions. After growth, the produced films were soaked in 1% NaOH, incubated at 60
°C for 2 h and washed with distilled water up to neutral pH. Bacterial cellulose films
were dried at room temperature over Whatman filters.

#### 132 Composites and bilayers biomaterials

BC was introduced in paper sheets by two different methods in order to obtain a composite or a bilayer. Two paper sheets were used in each case: commercial filter paper of 73 g m<sup>-2</sup> (Filtros Anoia 1305) or laboratory made paper sheets of 75 g m<sup>-2</sup> from 136 *Eucalyptus globulus* TCF (totally chlorine free) bleached pulp, PFI refined at 45°SR.

137 Eucalyptus pulp was supplied by ENCE (Pontevedra, Spain).

138 Composites of bacterial cellulose films and paper were produced growing the 139 bacterial strains on the surface of paper sheets layered on the top of solid media in 150 140 mm diameter Petri dishes. 500 µL of the bacterial suspension were mixed with 20 mL of 141 Ringer's solution <sup>1</sup>/<sub>4</sub> and inoculated in 150 mL of HS solid media covered with paper 142 sheets and incubated at 30°C for 10 days under static conditions. After growth, the 143 composites of paper sheets and bacterial cellulose were removed, treated with NaOH, 144 washed and dried as before at room temperature. Alternatively, they were also dried at 145 90°C for 5 min.

In the bilayers, bacterial cellulose films, once washed, were layered over paper
sheets and the resulting coated sheets were dried at room temperature or at 90°C as
above mentioned. In this case, only the BC films from *K. xylinus*, were used.

149 Paper characterization

### 150 Physical-mechanical properties

They were determined in accordance with the standards in brackets as follows: apparent density (ISO 534:2005), tensile strength index and elongation (ISO 1924-2:1994), burst strength index (ISO 2758:2001), wet zero-span index (ISO 15361:2000)

- and Bendtsen roughness (ISO 8791-2:2013).
- 155 Optical properties

Pulp brightness was determined according to ISO 2470–1. Specular Gloss was
determined according to ISO 8254-1:2009.

## 158 Barrier properties

- 159 Air permeance was measured with Bendtsen equipment (ISO 5636-5:2003).
- 160 Hydrophobicity was measured by the water contact angle (WCA) and water
- 161 impermeability by the water drop test (WDT). WCA was measured by using a
- 162 Dataphysics OCA15EC contact angle goniophotometer (Dataphysics, USA), using an
- 163 image capture ratio of 25 frames  $s^{-1}$ . Following the procedure described by Cusola et al.
- 164 2014 a 4  $\mu$ L water drop was delivered to the sample surface. At least 8 measurements

- 165 were made for each sample. Water drop test (WDT) was performed on each treated
- 166 paper specimen according to Tappi standard T835 om-08. Previously the paper sheets
- 167 were conditioned according to ISO 187. The WDT involved placing a drop of deionised
- 168 water on the surface of paper and recording the time needed for complete absorption,
- 169 which was signaled by vanishing of the drop specular gloss. Fifteen measurements per
- 170 treated paper sample were made and averaged.

#### 171 Scanning electron microscopy (SEM)

172 Surface and cross-sectional SEM pictures of the different films and biomaterials

173 obtained were taken on a JEOL JSM-6400 microscope (Japan). Samples were placed on

- the SEM sample holding stub by means of conductive double side sticky carbon film
- and coated with Au/Pd alloy prior to analysis.

## 176 **RESULTS**

#### 177 Bacterial cellulose films vs. papers from wood cellulose

Several bacterial strains were tested for bacterial cellulose (BC) production on the HS standard medium. The screening includes several newly isolated and also culture collection strains. Two of them, *Komagataeibacter xylinus* and *Gluconacetobacter sucrofermentans* were selected as the best producers in the culture conditions assayed.

182 The selected strains were grown for 10 days at 30 °C in liquid media on Petri 183 dishes of 15 cm diameter under static conditions. The bacterial growth produced surface 184 cellulose films that were recovered and treated with NaOH to eliminate microbial cells, 185 washed and dried at room temperature. Properties of the bacterial cellulose dried films 186 obtained were analyzed and compared to those of commercial filter paper or of paper 187 made from TCF eucalyptus pulp (Table 1). These two types of paper showed different 188 properties in accordance to their different composition. Eucalyptus paper was smoother, 189 had more density, higher physical properties and lower gloss than filter paper. 190 Moreover, it had better barrier properties to air and water. The properties of the BC 191 films produced by the two strains were quite similar and differed widely from those of 192 paper sheets. Although BC films had lower grammage than wood papers, their 193 mechanical properties were similar or even higher in some cases. This fact can be

194 explained by the higher density of films made of BC, due to a better conformability of

195 BC fibers. In fact, Chen et al. (2017) reported similar density values of films from

196 nanofibrillated cellulose with high strength properties, but in that case the nanocellulose

- 197 was obtained from different plants (Chen et al. 2017).
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**Table 1**. Physical, optical and barrier properties of bacterial cellulose films and papers from wood fibers,filter paper (Fp) and eucalyptus paper (Eu)

	Bacterial cellulose films		Papers from wood fiber	
	K. Xylinus	G. sucroferm.	Fp	Eu
Grammage (g m <sup>-2</sup> )	10.7±2.1	8.1±0.7	71.4±1.4	76.2±0.8
Thickness (µm)	9.7±1.3	9.3±1.3	154±4.9	115±1.0
Apparent density (g cm <sup>-3</sup> )	1.1±0.1	0.9±0.1	$0.5 \pm 0.0$	$0.7 \pm 0.0$
Tensile strength index $(N \cdot m g^{-1})$	$18.1 \pm 5.2$	$61.7 \pm 1.5$	$34.0\pm3.2$	$45.0\pm7.2$
Burst strength index (kN g <sup>-1</sup> )	$6.4\pm0.4$	$1.2\pm0.9$	$1.8 \pm 0.2$	$3.0 \pm 0.1$
Elongation (%)	$0.8\pm0.4$	ND	$2.0\pm0.5$	$2.7\pm0.2$
Wet Zero-Span index $(N \cdot m g^{-1})$	126±26	$114 \pm 4$	$110 \pm 1$	$106 \pm 3$
Gloss (%)*	$31.0\pm6.0$	32.5 ± 3.3	$17.0\pm0.3$	0.2±0.2
Brightness (%)*	$81.4\pm0.8$	$82.5\pm0.7$	$86.3 \pm 0.1$	$85.0\pm0.6$
Bendtsen roughness (mL min <sup>-1</sup> )*	$24 \pm 9$	$30 \pm 7$	$1823 \pm 211$	993± 54
Bendtsen Air Permeance $(\mu m (Pa \cdot s)^{-1})^*$	1.3±0.1	1.1±0.5	52.9±1.5	7.4±1.3
WDT (s)*	4121±300	4823±247	1.7±0.3	10.7±1.7
WCA (°)*	48.8±10.9	38.6±0.8	24.0±2.3	33.8±7.0

201 \* Properties measured in the upper face

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203 The more compact structure of BC provided dense films with a smoother surface 204 (lower Bendtsen roughness), and therefore higher gloss (Table 1). However, the most 205 remarkable difference with paper sheets was the strongly increased barrier properties to 206 air and water. Although air permeance was lower in eucalyptus paper than in filter 207 paper, it was significantly much lower in BC films. They also showed a remarkable 208 increased water drop test that raised from 10 s in eucalyptus paper to more than 4000 s 209 in BC films. The water contact angle showed also higher values in BC films. 210 Comparing with other reports in which nanocellulose from plant cellulose was used, it 211 was found that nanocellulose provide lower air permeability (Syverud and Stenius 2009; Gicquel et al. 2017; Herrera et al. 2017) and similar values of WCA (Beltramino et al.2015).

The results showed the high barrier properties of the films made of bacterial cellulose. The high resistance to water absorption and to air penetration of the BC dried films is an important trait that can be applied to enhance barrier properties of paper sheets, especially in food packaging, in order to replace petrol-based packaging by biodegradable products. For this reason, in order to provide these properties to final papers products, two kinds of biomaterials were made combining BC and paper sheets: composites and bilayer.

#### 221 Bacterial cellulose-paper composites

#### 222 Mechanical and optical properties of composites

To evaluate the contribution of BC to the properties of paper made from wood pulp, composites of the two types of cellulose were made. For this purpose, the BC producing strains were grown on the surface of paper sheets (filter or eucalyptus) soaked on the top of solid media in 150 mm petri dishes. The paper sheets covered with the bacterial growth were recovered and treated with alkali in the same conditions as above. They were dried at room temperature or, alternatively, at 90 °C and their physical properties analyzed (Table 2).

230 **Table 2.** Physical and optical properties of Composites made of filter (Fp) or eucalyptus (Eu) papers and

231 bacterial celluloses dried at room temperature

	Composite Fp		Compo	site Eu
	K. xylinus	G. sucroferm	K. xylinus	G. sucroferm
Tensile strength index (N·m g <sup>-1</sup> )	37.7 ± 1.5	39.5 ± 0.5	44.7 ± 2.7	46.5 ± 1.7
Burst strength index (kN g <sup>-1</sup> )	$1.8 \pm 0.1$	2.3 ± 0.4	$2.4 \pm 0.140$	2.7 ± 0.4
Elongation (%)	$1.8 \pm 0.1$	2.7 ± 0.2	2.2 ± 0.2	$3.5 \pm 0.254$
Wet Zero-Span index (N·m g⁻¹)	108 ± 4	101 ± 22	90 ± 1	75 ± 6
Gloss (%)*	31.9 ± 1.4	31.5 ± 3.8	23.3 ± 2.2	22.9 ± 1.6
Brightness (%)*	74.3 ± 0.5	79.0 ± 0.3	71.1 ± 0.2	79.2 ± 0.3
Bendtsen roughness (mL min <sup>-1</sup> )*	1372 ± 171	1374 ± 223	945 ± 50	826 ± 128

232 \* Properties measured in the upper face

Physical properties (grammage, thickness and apparent density) of papers were
not significantly affected by the addition of BC. In general, mechanical properties of the
BC-paper composites showed similar values or a small increase than control paper
sheets (Table 1). This increase was slightly higher with *G. sucrofermentans* than with *K. xylinus*.

239 Optical properties were determined on the upper face of the composites, that 240 covered by the BC (Table 2). Gloss is an important property in the printing paper 241 industry. Composites reached a notably higher gloss than their control samples, showing 242 similar increased values with the two types of BC. Santos et al. (2017) reported that 243 nonglossy papers can show a noticeable increment in their specular gloss when 244 reinforced with BC, in accordance with our results. Interestingly, a higher increase in 245 gloss than in papers coated by cellulose nanocrystals from biomass (Gicquel et al. 2017) 246 was obtained here with bacterial cellulose. Composites of filter paper showed also 247 remarkable increase of whiteness (data not shown). Brightness determination revealed a 248 decrease in this property in all composites. Composites with K. xylinus had the lowest 249 brightness values.

### 250 Barrier properties of composites

251 The BC-paper composite sheets showed a notably lower wettability than the 252 control paper sheets (Fig. 1), although not as low as the bacterial cellulose films (Table 253 1). WDT was determined on the two sides of the composites, the upper face, covered by 254 BC, and the down face of the sheets. The WDT values of the upper face of the 255 composites were remarkably higher in all samples (Fig. 1a). It was increased from 2-10 256 s in control paper sheets not covered by BC to values ranging from 414 to 1220 s in the 257 BC-pulp composites. Down face of the composite sheets showed much lower increase 258 in water drop test values, indicating higher wettability of this face of the composites, 259 probably because the lack of bacterial cellulose penetration among pulp fibers in this 260 side. Regarding the influence of the drying temperature on the properties of the 261 composites, a detrimental effect of temperature was found, as water drop value of 262 samples dried at room temperature was approximately two times that of parallel samples 263 dried at 90 °C.



Fig. 1. Barrier properties to water of Composites made of filter (Fp) or eucalyptus papers (Eu) and
bacterial celluloses dried at room or high temperature. a) WDT; b) WCA; c) Air Permeability

To evaluate the hydrophobicity of the composites, the water contact angle was also analyzed. The results showed an increased water contact angle of all composites, that exhibited up to 3 fold increase compared with control paper sheets (Fig 1b). Moreover, the differences between upper and down faces of the composites were minimized. The drying temperature also influenced in water contact angle, with higher results for the samples dried at room temperature.

274 The results found revealed that the composites of bacterial cellulose and paper 275 sheets have a diminished capacity of water absorption, indicating an increased barrier 276 property to water. To evaluate the barrier property to a different matter, air, the air 277 permeability was analyzed. Composites containing filter paper showed a high decrease 278 in permeability that diminished from the values corresponding to a high permeable 279 control filter (52.9  $\mu$ m/Pa·s) to a very closed paper (0.53-0.94  $\mu$ m/Pa·s) when the 280 composites were dried at room temperature. A lower effect was produced with 281 eucalyptus sheets (from 7.4 to 1.11  $\mu$ m/Pa·s). Drying at high temperature gave also less 282 permeable composites although permeability was decreased in lower extent, especially 283 with filter paper. The lower air permeability was probably due to the small BC 284 fragments filling the gaps between wood fibers and increasing the affinity between 285 them. Controlling the permeability of substances through the packaging is also very 286 important in food packaging in order to increase the shelf life of the product. In fact, 287 Tabarsa et al., 2017 also found a decrease in porosity combining BC and softwood 288 fibers, but mixing the fibers with BC before sheet formation.

### 289 K. xylinus vs. G. sucrofermentans

290 K. xylinus has been applied in previous works to increase the Young's modulus 291 of composites made with cellulose acetate butyrate (Gindl and Keckes 2004) or with 292 phenolic resins (Nakagaito et al. 2005), or to modify the surface of natural fibers to 293 improve composite properties (Pommet et al. 2008). It has been also used to increase the 294 physical properties of papers resulting from mixing the BC with wood fibers (Gao et al. 295 2011; Tabarsa et al. 2017; Xiang et al. 2017a), but in a different manner as in the 296 present paper and with different results. However, fewer works have been reported with 297 G. sucrofermentans. A similar composite was performed by Santos et al. (2015, 2016a,

b, 2017) in order to reinforce degraded papers. In this case, no variation in physicalproperties and a reduction of wettability was also found.

300 The results obtained in this research revealed that the two bacterial strains 301 provided different properties in some cases. In contrast with the similar barrier 302 properties of the BC films of the two producing bacteria, composites containing K. 303 xylinus cellulose gave higher values of water barrier properties (Fig.1) than composites 304 with G. sucrofermentans cellulose (increase of up to 1100 s of WDT and 77° the WCA 305 with the former vs. increase of 760 s WDT and 48° WCA with the latter in the case of 306 filter paper, and a similar behavior in eucalyptus paper). On the other hand, in both 307 paper supports, the two BC partners of the composites made a similar contribution to air 308 permeability, as similar values were found for K. xylinus and G. sucrofermentans 309 composites.

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#### 311 Bacterial cellulose-paper bilayer

#### 312 Mechanical and optical properties of bilayer biomaterial

313 The BC-pulp composites analyzed were produced by the direct growth of the 314 cellulose producing bacteria on the surface of paper sheets. The rational of this 315 methodology was that the fibers of bacterial cellulose would probably grow intermixed 316 among pulp fibers, making a compacted composite, which as we have shown, would 317 exhibit an increased resistance to fluid penetration. The good results obtained made us 318 to evaluate a different strategy to combine pulp and bacterial cellulose in a sheet. For 319 this purpose, previously produced BC films were layered on paper sheets and the bilayer 320 sheets were dried by the same procedure as above mentioned. We only used BC films 321 from K. xylinus, which gave best results as previously shown.

322 Similar to that previously obtained in composites, physical and mechanical 323 properties were not significantly affected in the bilayer material (Table 3). On the other 324 hand, gloss was strongly increased, even more than in the composite. Brightness was 325 decreased but in a lower extent than in the composite and roughness was strongly 326 decreased, especially in filter paper. A high smoothness is a required property in 327 printing applications and essential in printed electronics. It has been reported that other 328 kind of nanocelluloses from biomass applied on paper surface as coating treatment also 329 provide some smoothness increase (Brodin et al. 2014; Gicquel et al. 2017; Herrera et

al. 2017).

**Table 3**. Physical and optical properties of Bilayers made of filter (Fp) or eucalyptus (Eu) papers and

	Bilayer Fp	Bilayer Eu
Tensile strength index $(N \cdot m g^{-1})$	$30.8\pm1.7$	$39.4 \pm 1.6$
Burst strength index (kN g <sup>-1</sup> )	$2.0\pm0.0$	$2.3\pm0.2$
Elongation (%)	$1.9 \pm 1.6$	$1.9 \pm 0.2$
Wet Zero-Span index $(N \cdot m g^{-1})$	$95 \pm 4$	$97 \pm 2$
Gloss (%)*	$49.2 \pm 2$	$46.4\pm0.4$
Brightness (%)*	$80.0\pm1.0$	81.6±0.8
Bendtsen roughness (mL min <sup>-1</sup> )*	$680\pm158$	$517 \pm 41$

bacterial cellulose from *K. xylinus* dried at room temperature

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\* Properties measured in the upper face

#### 334 Barrier properties of bilayer biomaterial

335 The barrier properties of the bilayer sheets were determined on both sides (Fig. 336 2). Wettability of bilayer sheets was much lower than control paper sheets. Water drop 337 test was again notably increased to values around 490 to 300 s in filter and eucalyptus 338 papers, respectively. The values obtained were much lower than those of the 339 corresponding BC-paper composites. No significant differences were obtained at room 340 or high temperature. Similar to that observed in the composites, no effect was produced 341 in the down face of the bilayer. In agreement with the water drop values, the water 342 contact angle of bilayer sheets was also increased in the upper face to 86° and 44° in 343 filter and eucalyptus papers, respectively. By contrast with that obtained with the WDT, 344 the water contact angle was increased in the down face of the bilayer made with filter 345 paper, although in a lower extent than in the upper face (53°). The temperature used for 346 drying, room or hot, did not make an important difference in wettability of bilayers, it 347 was only reduced in the upper face of filter paper.

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**Fig. 2**. Barrier properties to water of Bilayer made of filter (Fp) or eucalyptus papers (Eu) and bacterial

351 cellulose from *K. xylinus* dried at room or high temperature. a) WDT; b) WCA; c) Air Permeability

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The barrier property to air, measured as air permeability, was strongly decreased in the bilayer biomaterials, especially in the case of filter paper. Similarly to that found in the composites no differences between the upper and lower face were observed, and no effect of the drying temperature was produced.

357 Therefore, interesting results were found concerning the barrier properties to air 358 and water with paper and BC. Nanocelluloses from plants instead of bacteria have also 359 been used to improve these properties. Several authors (Syverud and Stenius 2009; 360 Aulin et al. 2010; Lavoine et al. 2014b) obtained a complete reduction of air 361 permeability when nanofibrillated cellulose was applied as a surface layer on paper 362 sheets. However Lavoine et al. 2014a found that nanofibrillated cellulose did not 363 increase the barrier property to water. Lower knowledge exists about the barrier 364 properties that cellulose nanocrystals coated on papers may provide. Recently, Gicquel 365 et al 2017 reported that papers coated with cellulose nanocrystals can strongly reduce its 366 air permeability maintaining the mechanical properties. One of the problems associated 367 with coating with this kind of nanocellulose is that the surface obtained is brittle and the 368 coat is split along the substrate fiber (Gicquel et al. 2017).

#### 369 Composite vs. bilayer biomaterial

370 Two kinds of biomaterials (composite and bilayer) have been constructed 371 combining wood fibers and bacterial cellulose produced by K. xylinus. Physical 372 properties of papers were not adversely affected by the addition of bacterial cellulose in 373 any case. Previous works (Gao et al. 2010; Tabarsa et al. 2017; Xiang et al. 2017a) 374 reported an increase in physical properties in softwood or sugarcane bagasse fibers with 375 K. xylinus. Some of these authors also stated that the amount of bacterial cellulose 376 incorporated could affect the increases in physical properties. For example, Xiang et al. 377 (2017a) specified that BC has to be introduced at low doses (lower than 1%) whereas 378 Tabarsa et al.(2017) and Gao et al. (2010) found that physical properties of the sheets 379 increased with the bacterial cellulose dosage. According to the grammage increase (data 380 not shown), we determined that the amount of bacterial cellulose incorporated in our 381 biomaterials was around 15%, which is similar to that used in these papers. An 382 explanation of the different behavior found may be explained by the way in which BC

was introduced in vegetal fibers: in the previous works quoted, BC was disintegrated
and mixed with fibers before sheet formation. Moreover, the wood fibers used in our
work were refined, what probably made more difficult to increase the physical
properties. In fact, Surma-Slusarska et al. (2008) also obtained a reduction in some
physical properties when they combined BC and pine or birch fibers, obtaining a
bilayer. Mechanical properties can also be increased by the addition of nanofibrillated
cellulose as an additive in papermaking (Boufi et al. 2017).

Concerning brightness property, lower values were obtained in composites than
in bilayer. This could be related with the highest roughness of composites structures. In
fact, Gicquel et al. (2017) found that when the roughness increased, brightness
decreased in their study in which paper samples were coated with nanocellulose.
Moreover, Brodin et al. (2014) stated that the addition of nanofibrillated cellulose in the
paper reduced the light scattering coefficient and the brightness of the sheets.

BC provided smoother surfaces with higher gloss in the upper face of both
biomaterials. These properties were more improved in the bilayer biomaterial.
Smoothness is an important factor that determines the good paper printability. However,
barrier properties to water and air were much higher increased in composites. In the
composite made with filter paper WDT and WCA increased up to 1120 s and 77° with
BC whereas these increases were 480 s and 62° in the bilayer. Similarly, permeability
was decreased 98% in the composite vs. 96% in the bilayer.

403 The temperature used for drying the biomaterials (room or 90°C) had some 404 influence on the final properties, that was different in the composites or bilayer 405 materials. Whereas in the composites a detrimental effect in barrier properties was 406 produced by drying at high temperature, no significant effect of temperature was 407 produced in the bilayer materials. In both cases, the wettability was strongly reduced in 408 the lower faces, because the lack of bacterial cellulose penetration among pulp fibers in 409 this side, whereas the air permeability was not affected. The heterogeneous network 410 structure of composite, formed by vegetal fibers (macro-material) and bacterial 411 cellulose (nano-material) could be the reason of the different effect of temperature 412 drying in final properties. Before the drying treatment, the composite has two wet 413 materials with different size, and with different drying kinetics. According to the drying 414 theory of porous materials, when the drying temperature is higher, the evaporation rate

415 increases, that is, the drying kinetics is faster. Then the differences between the size and 416 the temperature during the drying treatment of the composite structure gives rise to a 417 different behavior of cellulosic fibers and bacterial cellulose, and therefore to a different 418 final dried structure.

The results show that the adhesion of bacterial cellulose films to the surface of paper sheets in the bilayer gives rise to novel sheets with decreased wettability, and decreased air permeability. However, the increase in barrier properties is much lower than that obtained when bacterial cellulose and paper fibers are more intensely interconnected in a composite.

### 424 Eucalyptus vs. filter paper

Values of barrier properties obtained in composites with each kind of paper support were very similar. However, the increases produced in each case were different. The increase in WDT was similar in both paper types (1120 s *vs.* 1210 s), whereas the WCA increase was slightly higher in filter paper (77° *vs.* 50° in eucalyptus). Finally, the air permeability decrease was also greater in filter paper (98%) than in eucalyptus (83%). These results suggest that the composition of the paper sheets used as support to hold the bacterial growth gave also some influence.

In the bilayer biomaterial, a similar effect than in composites was produced. The WDT and WCA increases were higher in filter paper (480 s and 62°) than in eucalyptus (290 s and 10°). Determination of air permeability showed that bilayers made of bacterial cellulose and filter paper exhibited a notable decrease of permeability (96%), while when the paper component of the bilayer was eucalyptus a lower effect was produced (71%). In fact, this effect may be explained by the lower initial barrier properties of filter paper, what made it easier to improve them.

#### 439 Scanning Electron Microscope analysis

Microscopic observation showed a different surface aspect of *K. xylinus* and *G. sucrofermentans* BC films, although both of them are formed by a dense net of thin
cellulose fibers. Those of *K. xylinus* showed longer size fibers while *G. sucrofermentans*films showed frequent short fibers. These differences may explain the different behavior
of bacterial cellulose in the composites, since barrier properties to water were more

increased with *K. xylinus*. Cross section of the BC films show that BC fibers are more
abundant and intensely connected at the surfaces, while they are more dispersed inside
the films (Fig. 3a-c).



G. sucrofermentans films

Composites Filter paper / G. sucrofermentans

449

451 **Fig. 3** SEM images of bacterial cellulose films from *G. sucrofermentans* (a, c) and *K. xylinus* (b).

452 Composite of filter paper and G. sucrofermentans (d)

452

464 SEM analysis of BC-paper composites showed how the two types of cellulose 465 fibers (bacterial and pulp) are interconnected. Paper fibers, of much thicker width, are 466 covered by thin BC fibers making a compact material (Fig 3d). Analysis of composite 467 surface shows it is covered by BC fibers that fill the space among pulp fibers making an 468 apparently smooth and closed surface, in accordance with the increase in barrier 469 properties found (Fig 4a, b). Cross section of the composites visualizes also the thin 470 layer of BC fibers from G. sucrofermentans growing mainly on the surface of the 471 composite (Fig 4c, d). A similar effect was found with the composite obtained from K. 472 xylinus (Fig 4e, f). The low thickness of the BC layer on paper sheets justifies that 473 thickness of composites or bilayers was not greatly modified. The images of composites 474 show a compact structure, which made the surface of biomaterials more hydrophobic 475 than the original paper surface.







466

468 Fig. 4 SEM images of control filter paper (a, c); composite of filter paper and *G. sucrofermentans* (b, d);
469 composite of filter paper and *K. xylinus* (e, f)

## 469 **Conclusions**

Hydrophobic and non-porous papers can be obtained combining wood cellulose papers with a natural, biodegradable material: bacterial cellulose. Results show that this effect depends on the bacterial strain used, on the kind of paper used and in the way BC is incorporated into paper supports. Thus, BC from *K. xylinus* that presented longer size fibers had stronger effect in reducing wettability of composites than *G. sucrofermentans* that showed frequent short fibers. Barrier properties were more increased in filter paper, probably due to its worse initial properties. Finally, although smoother surfaces with

- 476 higher gloss can be obtained in the bilayer in comparison with the composite, BC-paper
- 477 fibers are more intensely connected in a composite, providing higher barrier properties
- 478 to air and water than in a bilayer biomaterial.
- 479
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## **REFERENCES**

494	Aulin C, Gällstedt M, Lindström T (2010) Oxygen and oil barrier properties of microfibrillated cellulose
495	films and coatings. Cellulose 17:559-574 . doi: 10.1007/s10570-009-9393-y
496	Beltramino F, Roncero MB, Torres AL, et al (2016) Optimization of sulfuric acid hydrolysis conditions
497	for preparation of nanocrystalline cellulose from enzymatically pretreated fibers. Cellulose
498	23:1777-1789 . doi: 10.1007/s10570-016-0897-y
499	Beltramino F, Roncero MB, Vidal T, et al (2015) Increasing yield of nanocrystalline cellulose preparation
500	process by a cellulase pretreatment. Bioresour Technol 192:574-581 . doi:
501	10.1016/j.biortech.2015.06.007
502	Boufi S, González I, Delgado-Aguilar M, et al (2017) Nanofibrillated cellulose as an additive in
503	papermaking process. Cellul Nanofibre Compos Prod Prop Appl 154:153-173 . doi: 10.1016/B978-
504	0-08-100957-4.00007-3
505	Brodin FW, Gregersen ØW, Syverud K (2014) Cellulose nanofibrils: Challenges and possibilities as a
506	paper additive or coating material - A review. Nord Pulp Pap Res J 29:156–166 . doi:
507	10.3183/NPPRJ-2014-29-01-p156-166
508	Chawla PR, Bajaj IB, Survase S a., Singhal RS (2009) Microbial cellulose: Fermentative production and
509	applications ( Review ). Food Technol Biotechnol 47:107-124
510	Chen Y, Geng B, Ru J, et al (2018) Comparative characteristics of TEMPO-oxidized cellulose nanofibers
511	and resulting nanopapers from bamboo, softwood, and hardwood pulps. Cellulose 25:895 . doi:
512	10.1007/s10570-017-1553-x
513	Cusola O, Valls C, Vidal T, Roncero MB (2014) Rapid functionalisation of cellulose-based materials
514	using a mixture containing laccase activated lauryl gallate and sulfonated lignin. Holzforschung
515	68:631–639 . doi: 10.1515/hf-2013-0128
516	Gao W-H, Chen K-F, Yang R-D, et al (2010) Properties of Bacterial Cellulose and Its Influence on the
517	Physical Properties of Paper. Bioresour Vol 6, No 1 6:144–153
518	Gicquel E, Martin C, Garrido Yanez J, Bras J (2017) Cellulose nanocrystals as new bio-based coating
519	layer for improving fiber-based mechanical and barrier properties. J Mater Sci 52:3048-3061 . doi:
520	10.1007/s10853-016-0589-x
521	Gilbert HJ (2010) The Biochemistry and Structural Biology of Plant Cell Wall Deconstruction. Plant
522	Physiol 153:444-455 . doi: 10.1104/pp.110.156646
523	Hagiwara Y, Putra A, Kakugo A, et al (2010) Ligament-like tough double-network hydrogel based on
524	bacterial cellulose. Cellulose 17:93-101 . doi: 10.1007/s10570-009-9357-2
525	Herrera MA, Mathew AP, Oksam K (2017) Barrier and mechanical properties of plasticized and cross-
526	linked nanocellulose coatings for paper packaging applications. Cellulose 24:3969-3980. doi:
527	10.1007/s10570-017-1405-8
528	Klemm D, Heublein B, Fink HP, Bohn A (2005) Cellulose: Fascinating biopolymer and sustainable raw
529	material. Angew Chemie - Int Ed 44:3358-3393 . doi: 10.1002/anie.200460587

- Klemm D, Kramer F, Moritz S, et al (2011) Nanocelluloses: A new family of nature-based materials.
  Angew Chemie Int Ed 50:5438–5466. doi: 10.1002/anie.201001273
- Lavoine N, Bras J, Desloges I (2014a) Mechanical and Barrier Properties of Cardboard and 3D Packaging
   Coated with Microfibrillated Cellulose. J Appl Polym Sci 40106:1–11. doi: 10.1002/app.40106
- Lavoine N, Desloges I, Khelifi B, Bras J (2014b) Impact of different coating processes of microfibrillated
   cellulose on the mechanical and barrier properties of paper. J Mater Sci 49:2879–2893 . doi:
   10.1007/s10853-013-7995-0
- Lim GH, Lee J, Kwon N, et al (2016) Fabrication of flexible magnetic papers based on bacterial cellulose
  and barium hexaferrite with improved mechanical properties. Electron Mater Lett 12:574–579. doi:
  10.1007/s13391-016-6179-x
- Lin SP, Loira Calvar I, Catchmark JM, et al (2013) Biosynthesis, production and applications of bacterial
   cellulose. Cellulose 20:2191–2219 . doi: 10.1007/s10570-013-9994-3
- 542 Osong SH, Norgren S, Engstrand P (2016) Processing of wood-based microfibrillated cellulose and
   543 nanofibrillated cellulose, and applications relating to papermaking: a review. Cellulose 23:93–123 .
   544 doi: 10.1007/s10570-015-0798-5
- Pommet M, Juntaro J, Heng JYY, et al (2008) Surface modification of natural fibers using bacteria:
  Depositing bacterial cellulose onto natural fibers to create hierarchical fiber reinforced
  nanocomposites. Biomacromolecules 9:1643–1651. doi: 10.1021/bm800169g
- Quintana E, Valls C, Vidal T, Blanca Roncero M (2013) An enzyme-catalysed bleaching treatment to
   meet dissolving pulp characteristics for cellulose derivatives applications. Bioresour Technol

550 148:1-8 . doi: 10.1016/j.biortech.2013.08.104

- Quintana E, Valls C, Vidal T, Roncero MB (2015) Comparative evaluation of the action of two different
   endoglucanases. Part I: On a fully bleached, commercial acid sulfite dissolving pulp. Cellulose
   22:2067–2079 . doi: 10.1007/s10570-015-0623-1
- Santos SM, Carbajo JM, Gómez N, et al (2016a) Use of bacterial cellulose in degraded paper restoration.
   Part I: application on model papers. J Mater Sci 51:1541–1552. doi: 10.1007/s10853-015-9477-z
- Santos SM, Carbajo JM, Gómez N, et al (2016b) Use of bacterial cellulose in degraded paper restoration.
   Part II: application on real samples. J Mater Sci 51:1553–1561. doi: 10.1007/s10853-015-9477-z
- Santos SM, Carbajo JM, Gómez N, et al (2017) Paper reinforcing by in situ growth of bacterial cellulose.
   J Mater Sci 52:5882–5893 . doi: 10.1007/s10853-017-0824-0
- Santos SM, Carbajo JM, Quintana E, et al (2015) Characterization of purified bacterial cellulose focused
   on its use on paper restoration. Carbohydr Polym 116:173–181. doi: 10.1016/j.carbpol.2014.03.064
- Shah N, Ul-Islam M, Khattak WA, Park JK (2013) Overview of bacterial cellulose composites: A
  multipurpose advanced material. Carbohydr Polym 98:1585–1598. doi:
- 564 10.1016/j.carbpol.2013.08.018
- Surma-Slusarska B, Danielewicz D, Presler S (2008) Properties of Composites of Unbeaten Birch and
   Pine Sulphate Pulps with Bacterial Cellulose. Fibres Text East Eur 16:127–129
- 567 Syverud K, Stenius P (2009) Strength and barrier properties of MFC films. Cellulose 16:75–85 . doi:
- 568 10.1007/s10570-008-9244-2

569	Tabarsa T, Sheykhnazari S, Ashori A, Mashkour M (2017) Preparation and characterization of reinforced
570	papers using nano bacterial cellulose. Int J Biol Macromol 101:334-340 . doi:
571	10.1016/j.ijbiomac.2017.03.108
572	Tang L, Huang B, Lu Q, et al (2013) Ultrasonication-assisted manufacture of cellulose nanocrystals
573	esterified with acetic acid. Bioresour Technol 127:100-105 . doi: 10.1016/j.biortech.2012.09.133
574	Tuck CO, Pérez E, Horváth IT, et al (2012) Valorization of Biomass : Deriving More Value from Waste.
575	Science (80- ) 337:695–699
576	Wu SQ, Li MY, Fang BS, Tong H (2012) Reinforcement of vulnerable historic silk fabrics with bacterial
577	cellulose film and its light aging behavior. Carbohydr Polym 88:496-501 . doi:
578	10.1016/j.carbpol.2011.12.033
579	Xiang Z, Jin X, Quingguo L, et al (2017a) The reinforcement mechanism of bacterial cellulose on paper
580	made from woody and non-woody fiber sources. Cellulose 24:5147-5156 . doi: 10.1007/s10570-
581	017-1468-6
582	Xiang Z, Liu Q, Chen Y, Lu F (2017b) Effects of physical and chemical structures of bacterial cellulose
583	on its enhancement to paper physical properties. Cellulose 24:3513-3523 . doi: 10.1007/s10570-
584	017-1361-3
585	Yamanaka S, Watanabe K, Kitamura N, et al (1989) The Structure and Mechanical-Properties of Sheets
586	Prepared from Bacterial Cellulose. J Mater Sci 24:3141-3145 . doi: Doi 10.1007/Bf01139032
587	