

PERSPECTIVE

G protein-coupled receptor 37 (GPR37) emerges as an important modulator of adenosinergic transmission in the striatum

G protein-coupled receptor 37 (GPR37), also known as parkin associated endothelin-like (Pael) receptor, is an orphan G protein-coupled receptor, which suffers a defective parking ubiquitination in autosomal recessive Parkinson's disease promoting its endoplasmic reticulum aggregation and stress, neurotoxicity and neuronal death (Takahashi and Imai, 2003). Interestingly, we have demonstrated previously that GPR37 heteromerizes with adenosine A_{2A} receptor (A_{2A}R) in the striatum (Morató et al., 2017; Sokolina et al., 2017). In addition, we also reported some functional consequences of this direct interaction, whereby GPR37 deletion enhanced striatal A_{2A}R cell surface expression with a concomitant increase in A2AR agonist-mediated cAMP accumulation (Morató et al., 2017); accordingly, an enhancement of A2AR agonist-induced catalepsy and antagonist-induced locomotor activity was observed upon GPR37 deletion (Morató et al., 2017). Overall, it has been hypothesized that GPR37 might hold a chaperone-like activity controlling A_{2A}R cell surface targeting and function. However, the precise physiological function of GPR37 still is unidentified. The current findings now provide additional evidence for the role of GPR37 as a repressor of A_{2A}R function. Thus, while chronic A_{2A}R antagonist treatment (i.e., SCH58261, 1 mg/kg per day, intraperitoneal administration, 10 days) of mice lacking GPR37 did not affect the striatum-dependent cued learning, it enhanced locomotor sensitization (Morató et al., 2019). Moreover, chronic A_{2A}R blockade boosted striatal long-term depression (LTD) in corticostriatal synapses of GPR37-1- but not of wild type mice; this observation correlated well with the adenosinergic neurochemical modifications present in GPR37^{-/-} mice, namely an increased density of A2AR and decreased levels of adenosine (Morató et al., 2019). Overall, GPR37 emerged as key contestant controlling A_{2A}R function particularly upon chronic A_{2A}R blockade, thus delineating A_{2A}R-dependent long-term plastic changes in corticostriatal synapses.

Adenosine is an key neuromodulator in the central nervous system regulating two important brain processes, namely neurotransmission and neural metabolism. Indeed, adenosine modulates the basal levels and release of many neurotransmitters including glutamate, y-aminobutyric acid (GABA), dopamine and serotonin, thus fine-tuning synaptic transmission. Derived from the breakdown of intra- and extracellular adenosine triphosphate, adenosine is released upon some physiological and pathological stimuli, thus exerting its effects through four G protein-coupled adenosine receptors. In the brain, the density of A2AR is far greater in regions heavily innervated by dopaminergic terminals, namely in the striatum, nucleus accumbens and olfactory tubercle. Interestingly, the striatum is a large forebrain region involved in action initiation, motor skill learning and development and cued action-selection memory. Moreover, this structure could be involved both in reward-association memory and visual recognition memory. In Parkinson's disease, a progressive degeneration of midbrain dopamine neurons is associated with an aberrant striatal activity producing marked motor impairments, including bradykinesia, tremor, and rigidity. Indeed, dopamine replacement therapy

with levodopa is the "gold standard" Parkinson's disease treatment. While initially effective in treating motor deficits, most Parkinson's disease patients develop drug-induced involuntary movements (i.e., levodopa-induced dyskinesia). This clinical problem triggers the urgent need of identifying the circuit dysfunction resulting from dopamine loss and consequent levodopa replacement. Mostly, the information flows into the striatum through glutamatergic cortical terminals innervating medium spiny neurons. At the same time, corticostriatal synapses are modulated by the nigrostriatal dopaminergic system implicated in action control and learning (Figure 1). Conversely, there are two main GABAergic output circuits controlling locomotion: the direct and indirect pathways. Interestingly, although both pathways are not anatomically separated, they express a different set of GPCRs (Figure 2). Thus, the indirect-pathway neurons selectively express dopamine D₂ receptor (D₂R) and A_{2A}R, and the neurons from the direct pathway contain the dopamine D₁ receptor and muscarinic acetylcholine M₄ receptor. Importantly, intrinsic cholinergic interneurons, which represent only 1% of total striatal neurons, play an important role in modulating GABAergic neuron excitability by tuning their response to glutamatergic inputs through postsynaptic muscarinic acetylcholine M₁ receptor/muscarinic acetylcholine M₄ receptor. In addition, at the presynaptic level cholinergic interneurons exert a tonic inhibitory effect over incoming information through muscarinic acetylcholine M2 receptor/muscarinic acetylcholine M₄ receptor activation located on cortical and thalamic glutamatergic terminals.

Interestingly, many different A_{2A}R containing heteromers has been described within the brain, thus establishing molecular and functional receptor-receptor interactions allowing neurotransmission harmonization (Fuxe et al., 2014) (**Figure 2**). Precisely, in the dorsolateral striatum, the dopaminergic and adenosinergic transmission is tuned by $D_2R/A_{2A}R$ heteromers. Indeed, the control of motor function by adenosine is based on the ability of A_{2A}R to tightly control D₂R function, both at the level of intracellular signaling as well as by an heteromer-dependent A_{2A}R-D₂R negative allosteric modulation. Importantly, GPR37, through its chaperone-like activity, controls A_{2A}R cell surface targeting in vivo, thus limiting the plasma membrane density and function of A2AR (Morató et al., 2017). Therefore, its heteromerization with GPR37 may sculpt the ability of A2AR to modulate D₂R and therefore dopaminergic transmission. Indeed, GPR37 deletion prompts an enhanced A_{2A}R membrane trafficking and response to adenosinergic drugs (Morató et al., 2017, 2019). Thus, chronic blockade of A24R induced a sensitization of locomotor activity, which is associated with elevated dopamine concentrations and synaptic changes in the striatum. These striatal plastic changes powerfully regulate basal ganglia circuitry by setting the gain on cortical and thalamic inputs, thus formatting motor function. Indeed, long-lasting alterations in striatal plasticity have been proposed to play a key role in movement disorders such as Parkinson's disease, Huntington's disease and dystonia.

Interestingly, while long-term potentiation in the hippocampus and LTD in the cerebellum are well established models of synaptic plasticity, the molecular mechanisms underlying striatal plasticity remain elusive. Eventually, the study of synaptic plasticity in the striatum has been limited by the difficulty in triggering reliable, long-lasting long-term potentiation in striatal brain slices under physiological conditions. Certainly, the mechanisms governing LTD induction and expression vary across brain regions. Thus, in medium spiny neurons, low-frequency stimulation of corticostriatal afferent fibers leads to a

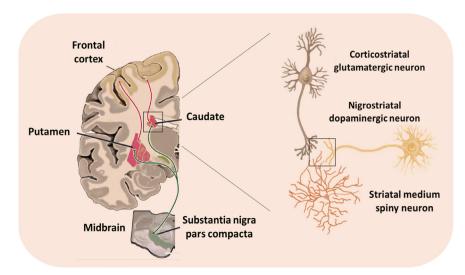


Figure 1 Striatal medium spiny neurons are modulated by corticostriatal and nigrostriatal neurons.

Collateral axons from glutamatergic cortical neurons (pink traces) establish synapses with the head of dendritic spines of medium spiny neurons, the most common neuron in the striatum. Simultaneously, dopaminergic afferents from the substantia nigra pars compacta (green traces) make synaptic contact with the neck of the medium spiny neuron dendritic spine.

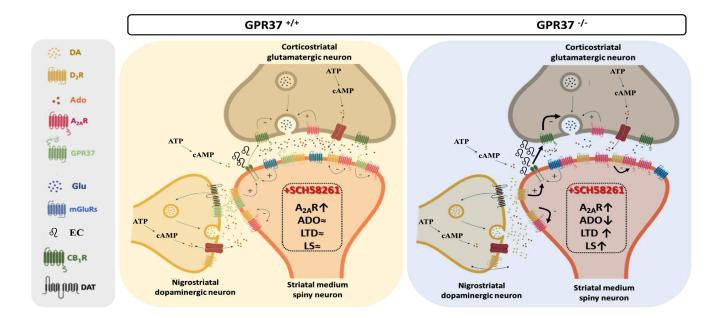


Figure 2 Indirect pathway striatal spiny module.

The medium spiny neuron dendritic spine, the glutamatergic terminal and the dopaminergic terminal constitute the so-called striatal spiny module (SSM). The segregation of DA and adenosine receptors within the two subtypes of medium spiny neurons suggests the existence of at least two subtypes of SSMs, which could be termed as enkephalin-SSM (*i.e.*, indirect pathway) and dynorphin-SSM (*i.e.*, direct pathway). Thus, while dynorphin-SSM predominantly express DA D_1 receptor (D_1 R), enkephalin-SSM express DA D_2 receptors (D_2 Rs) and D_2 Rs. The endocannabinoid (EC)-dependent long-term depression (LTD) model at enkephalin-SSM requieres activation of postsynaptic D_2 R enhances ECs production. ECs activate presynaptic cannabinoid 1 receptor (CB1R) which induce LTD. GPR37, by modulating D_2 R enhances ECs production and thus LTD. Chronic D_2 R antagonist treatment (*i.e.*, SCH58261, 1 mg/kg per day, intraperitoneal administration, 10 days) induced an increase of D_2 R density in the striatum of wild-type and GPR37- mice. Interestingly, under the same experimental conditions a reduction in striatal adenosine (Ado) levels and a potentiation of striatal LTD and locomotor sensitization was observed only in GPR37- mice (right panel). ATP: Adenosine triphosphate; cAMP: cyclic adenosine monophosphate; DA: dopamine; DAT: DA transporter; Glu: glutamate; LS: locomotor sensitization.

different form of LTD (Ronesi and Lovinger, 2005). This form of striatal long-term synaptic plasticity involves glutamate and dopamine. Indeed, the best-studied form of striatal plasticity is the homosynaptic endocannabinoids (EC)-dependent LTD (eCB-LTD) at glutamatergic synapses (**Figure 2**). Typically evoked by persistent low frequency stimulation, eCB-LTD relies in the activation of postsynaptic receptors couped to protein $G_{\alpha/11}$ (*i.e.*,

metabotropic glutamate type 5 receptors) triggering phospholipase C (PLC) activation and synthesis of ECs (2-arachidonoyl glycerol and anandamide) which diffuses presynaptically to activate CB1 cannabinoid receptors and suppress synaptic transmission (*i.e.*, lower glutamate release probability) (**Figure 2**) (Gerdeman et al., 2002). Interestingly, eCB-LTD is observed mainly at glutamatergic synapses onto indirect-pathway me-

dium spiny neurons (Kreitzer and Malenka, 2008), precisely where $D_2R/A_{2A}R$ heteromers are expressed and interact with metabotropic glutamate type 5 receptors (**Figure 2**) (Cabello et al., 2009). Furthermore, presynaptic $A_{2A}R$ are powerful regulator of CB1 receptor-mediated synaptic depression in corticostriatal synapses of the dorsal striatum and they also control metabotropic glutamate type 5 receptor function at these same synapses (Cunha, 2016). Likewise, post-synaptic $A_{2A}R$ also control these same receptor systems at postsynaptic compartments, as well as D_2R . This prompts several still unanswsered questions on the fine-tuning of LTD in the indirect-pathway by adenosine and dopamine, such as: i) how does $A_{2A}R$ blockade enhances LTD? ii) why does eCB-LTD requires D_2R activation? Indeed, dopamine depletion, D_2R antagonists and D_2R deletion prevent striatal LTD induction.

Although prosaposin has been postulated as ligand, GPR37 still is an orphan GPCR (i.e., not yet been linked to endogenous ligands). In addition, there is a lack of highly selective GPCR37 ligands allowing ex vivo and in vivo functional studies, thus currently research efforts are concentrated in GPR37-based drug discovery. Hence, experiments in mice lacking GPR37 are unique in providing some hints about its function. Also, the assessment of the GPR37 interactome revealing GPCR partners (i.e., A_{2A}R and D₂R) aid to understand the biological relevance of this orphan receptor particularly in the central nervous system (Sokolina et al., 2017). Indeed, since GPR37 deletion does not affect striatal LTD and hippocampal LTD (Lopes et al., 2015; Morató et al., 2019), we assume that GPR37 function per se do not play a role in synaptic plasticity. However, the proteomic studies revealing the ability of GPR37 to heteromerize with A_{2A}R suggested that this orphan receptor may function in a ligand-independent manner (i.e., chaperone-like) modulating A_{2A}R function (i.e., repressor) in different brain regions (i.e., striatum and hippocampus). It is important to highlight that GPR37-dependent physiological effects (i.e., increased striatal LTD and locomotor sensitization) were observed only upon chronic A_{2A}R blockade, thus reinforcing the role of GPR37 in long-term neuroadaptations mediated by the adenosinergic system. Also, these results consolidate the idea that A_{2A}R-mediated control of motor sensitization is a consequence of the ability of A_{2A}R to control corticostriatal plasticity. This selective impact of GPR37 ablation on long-term adaptive changes of A2AR function may be of particular relevance for striatal-related neurodegenerative diseases. In fact, there is an abnormal over function of A2AR coupled with perturbed corticostriatal plasticity at the onset of striatal-related neurodegenerative diseases, such as Parkinson's or Huntington's diseases (Cunha, 2016); this prompts considering a prominent role for GPR37 at the onset of these disorders, through its ability to control A2AR trafficking to the plasma membrane.

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References

Cabello N, Gandía J, Bertarelli DCG, Watanabe M, Lluís C, Franco R, Ferré S, Luján R, Ciruela F (2009) Metabotropic glutamate type 5, dopamine D2 and adenosine A2a receptors form higher-order oligomers in living cells. J Neurochem 109:1497-1507.

Cunha RA (2016) How does adenosine control neuronal dysfunction and neurodegeneration? J Neurochem 139:1019-1055.

Fuxe K, Borroto-Escuela DO, Romero-Fernandez W, Palkovits M, Tarakanov AO, Ciruela F, Agnati LF (2014) Moonlighting proteins and protein-protein interactions as neurotherapeutic targets in the G protein-coupled receptor field. Neuropsychopharmacology 39:131-155.

Gerdeman GL, Ronesi J, Lovinger DM (2002) Postsynaptic endocannabinoid release is critical to long-term depression in the striatum. Nat Neurosci 5:446-451.

Kreitzer AC, Malenka RC (2008) Striatal plasticity and basal ganglia circuit function. Neuron 60:543-554.

Lopes JP, Morató X, Souza C, Pinhal C, Machado NJ, Canas PM, Silva HB, Stagljar I, Gandía J, Fernández-Dueñas V, Luján R, Cunha RA, Ciruela F (2015) The role of parkinson's disease-associated receptor GPR37 in the hippocampus: functional interplay with the adenosinergic system. J Neurochem 134:135-146.

Morató X, Gonçalves FQ, Lopes JP, Jauregui O, Soler C, Fernández-Dueñas V, Cunha RA, Ciruela F (2019) Chronic adenosine A2A receptor blockade induces locomotor sensitization and potentiates striatal LTD IN GPR37-deficient mice. J Neurochem 148:796-809.

Morató X, Luján R, López-Cano M, Gandía J, Stagljar I, Watanabe M, Cunha RA, Fernández-Dueñas V, Ciruela F (2017) The Parkinson's disease-associated GPR37 receptor interacts with striatal adenosine A(2A) receptor controlling its cell surface expression and function in vivo. Sci Rep 7:9452-9452.

Ronesi J, Lovinger DM (2005) Induction of striatal long-term synaptic depression by moderate frequency activation of cortical afferents in rat. J Physiol 562:245-256.

Sokolina K, Kittanakom S, Snider J, Kotlyar M, Maurice P, Gandía J, Benleulmi-Chaachoua A, Tadagaki K, Oishi A, Wong V, Malty RH, Deineko V, Aoki H, Amin S, Yao Z, Morató X, Otasek D, Kobayashi H, Menendez J, Auerbach D, et al. (2017) Systematic protein-protein interaction mapping for clinically relevant human GPCRs. Mol Syst Biol 13:918.

Takahashi R, Imai Y (2003) Pael receptor, endoplasmic reticulum stress, and Parkinson's disease. J Neurol 250 Suppl 3:III25-29.

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