1 2	Isomeric and hybrid ferrocenyl/cyrhetrenyl aldimines: a new family of multifunctional Compounds†
3	
4	Juan Oyarzo, <sup>a</sup> Alejandra Acuña, <sup>a</sup> Hugo Klahn, <sup>*a</sup> Rodrigo Arancibia, <sup>b</sup>
5	Carlos P. Silva, <sup>c</sup> Ramón Bosque, <sup>d</sup> Concepción López, <sup>*d</sup>
6	Mercè Font-Bardía, <sup>e</sup> Carme Calvis <sup>f</sup> and Ramón Messeguer <sup>f</sup>
7	
8	
9	
10	
11	
12 12	
13 14	
15	a Instituto de Química, Pontificia Universidad Católica de Valparaíso, Casilla 4059, Valparaíso, Chile.
16	b Laboratorio de Química Inorgánica y Organometálica, Departamento de Química Analítica e
17	Inorgánica, Facultad de Ciencias Químicas, Universidad de Concepción, Concepción, Chile
18	c Departamento de Química de los Materiales, Facultad de Química y Biología, Universidad de Santiago
19	de Chile, Casilla 40, Correo 33, Santiago, Chile; Soft Matter Research and Technology Center, SMAT-
20 21	C, Santiago, Chile d Department de Químico Inorgànico i Orgànico (Secció de Químico Inorgànico) Escultat de Químico
21 22	Universitat de Barcelona, Martí i Francuès 1–11, 08028-Barcelona, Spain, e Unitat de Difracció de
23	Raigs-X, Centre Científics i Tecnològics (CCiT), Universitat de Barcelona, Solé i Sabaris 1–3, 08028
24	Barcelona, Spain
25	f Biomed Division, LEITAT Technological Centre, Parc Científic de Barcelona, Edifici Hèlix, Baldiri i
26	Reixach 13-21, 08028-Barcelona, Spain
27	
28 20	
29 30	
31	
32	
33	
34	
35	
36 27	
38	Hugo Klahn: hugo.klahn@pucy.cl
39	Concepción, Chile: conchi.lopez@qi.ub.es
40	
41	
10	

- 43 ABSTRACT:
- 44
- 45 The synthesis and characterization of two novel and isomeric hybrid ferrocenyl/cyrhetrenyl aldimines
- 46 [(n5-C5H5)Fe{(n5-C5H4)-CHvN-(n5-C5H4)}Re(CO)3] (1) and [(n5-C5H5)Fe{(n5-C5H4)-NvCH-(n5-
- 47 C5H4)}Re (CO)3] (2) are reported. Their X-ray crystal structures reveal that both adopt the E form.
- 48 However, molecules of 1 and 2 differ in the relative arrangement of the "Fe( $\eta$ 5-C5H5)" and "Re(CO)3"
- 49 units (anti in 1 and syn in 2). This affects the type of intermolecular interactions, the assembly of the
- 50 molecules and therefore their crystal architecture. Comparative studies of their electrochemical,
- 51 spectroscopic and photo-physical properties have allowed us to clarify the effect produced by the
- 52 location of the organometallic arrays (ferrocenyl or cyrhetrenyl) on electronic delocalization, the
- 53 proclivity of the metals to undergo oxidation and their emissive properties. Theoretical studies based on
- 54 Density Functional Theory (DFT) calculations on the two compounds have also been carried out in
- order to rationalize the experimental results and to assign the bands detected in their electronic spectra.
- 56 The cytotoxic activities of compounds 1 and 2 against human adenocarcinoma cell lines [breast (MCF7
- and MDA-MB-231) and colon (HCT-116)] reveal that imine 2 has a greater inhibitory growth effect
- than 1 and it is ca. 1.8 times more potent than cisplatin in the triple negative MDA-MB 231 and in the
- cisplatin resistant HCT-116 cell lines. A comparative study of their effect on the normal and non-tumour
- 60 human skin fibroblast BJ cell lines is also reported.
- 61
- 62
- 63

- 64 INTRODUCTION
- 65

66 Heterodimetallic compounds have attracted great interest in recent years. The presence of two proximal

- 67 metals with different environments, oxidation numbers and spin states may influence their mutual
- 68 cooperation, reactivity, electrochemical behaviour, photo-optical properties and also activities (i.e.
- 69 catalytic, biological).1–4 Besides this, the proper selection of two metal ions, their environments and
- their connectivity (i.e. by metal-metal bonds, ligands' scaffold or functional groups) may also produce
- 71 multifunctional compounds with outstanding relevance in new materials design (i.e. electrochemical
- 72 devices, sensors, molecular switches, etc.), photoelectronic technology, nanoscience, catalysis, biology
- 73 and medicine.5
- On the other hand, bioorganometallic chemistry has been one of the research areas with a greater, and
- also faster, development during the last decade.6,7 The great advances achieved so far as well as the
- 76 discovery of relevant applications of organometallic compounds in diagnosis, therapy and imaging are
- promoting the interest of a large number of scientists, making this area more attractive day after day.
- Among the huge variety of organometallic compounds, metallocenes (especially ferrocene derivatives)
- and three-legged half sandwich organometallic derivatives are the most promising candidates in
- 80 biotechnology, diagnosis and new drug design. Examples of their use as bioprobes for cellular imaging,
- 81 pharmaceutical sensors, molecular recognizers, detectors, and artificial metallo-enzymes have been
- 82 described, and the idea of using these organometallic complexes in new drug (or prodrug) discovery is
- becoming more and more fascinating and popular.8–11
- 84 The success of ferrocenyl-based molecules as antimalarial, antitumoral, anti-VIH, antibacterial, and
- antifungal agents and inhibitors8–11 has triggered the interest on novel ferrocene derivatives with
- 86 greater efficiencies, lower toxicities and minor side effects than the drugs used nowadays for the
- 87 treatment of these diseases. It has also allowed the extension of the strategies used for ferrocenes to the
- half-sandwich organometallic compounds. Those with fac-[M(CO)3] cores, such as cyrhetrene  $[Re(\eta 5 1)]$
- 89 C5H5)(CO)3] and cymantrene [Mn( $\eta$ 5-C5H5)(CO)3] derivatives, are attracting a great deal of interest
- 90 in new medicinal chemistry due to their high stability in air and water, lipophilicity, low toxicity,
- 91 properties (i.e. photo-physical or electrochemical) or biological activities.11–18
- 92 Cyrhetrene chemistry has undergone a fast and spectacular growth in the last five-years.11–18 The
- development of new hybrid compounds with the " $[Re(\eta 5-C5H4)(CO)3]$ " unit anchored on the
- 94 backbones of molecules of biological relevance is one of the most active and promising areas of research
- 95 in new medicinal chemistry. The compounds shown in Fig. 1 are representative examples to illustrate
- 96 the relevance of cyrhetrenyl derivatives in bioorganometallic chemistry. The amides and sulphonamides
- 97 (A and B in Fig. 1) are capable of pharmaceutical sensors, molecular recognizers, detectors, and
- 98 artificial metallo-enzymes have been described, and the idea of using these organometallic complexes in
- 99 new drug (or prodrug) discovery is becoming more and more fascinating and popular.8–11

- 100 The success of ferrocenyl-based molecules as antimalarial, antitumoral, anti-VIH, antibacterial, and
- antifungal agents and inhibitors8–11 has triggered the interest on novel ferrocene derivatives with
- 102 greater efficiencies, lower toxicities and minor side effects than the drugs used nowadays for the
- treatment of these diseases. It has also allowed the extension of the strategies used for ferrocenes to the
- half-sandwich organometallic compounds. Those with fac-[M(CO)3] cores, such as cyrhetrene  $[Re(\eta 5 1)]$
- 105 C5H5)(CO)3] and cymantrene [Mn( $\eta$ 5-C5H5)(CO)3] derivatives, are attracting a great deal of interest
- in new medicinal chemistry due to their high stability in air and water, lipophilicity, low toxicity,
- 107 properties (i.e. photo-physical or electrochemical) or biological activities.11–18 Cyrhetrene chemistry
- 108 has undergone a fast and spectacular growth in the last five-years.11–18 The development of new hybrid
- 109 compounds with the " $[Re(\eta 5-C5H4)(CO)3]$ " unit anchored on the backbones of molecules of biological
- 110 relevance is one of the most active and promising areas of research in new medicinal chemistry. The
- 111 compounds shown in Fig. 1 are representative examples to illustrate the relevance of cyrhetrenyl
- derivatives in bioorganometallic chemistry. The amides and sulphonamides (A and B in Fig. 1) are
- capable of inhibiting carbonic anhydrase enzymes.11d Several families of cyrhetrene conjugates with
- tamoxifen, hydroxytamoxifen (Fig. 1, C)12 and chloroquine (Fig. 1, D)13 have been prepared and
- evaluated as antitumoral or antiparasitic agents against malaria, leishmaniasis or trypanosomiasis.13,14
- 116 Compound D has remarkable activity (IC50 =  $0.9 \mu$ M) against Trypanosoma brucei and low toxicity to
- 117 normal human cells. Kowalski and co-workers have recently reported seven new hybrid
- 118 cyrhetrene/nucleobase derivatives, of which the uracil conjugates (Fig. 1, E) resulted to be the most
- 119 active.14
- 120 Other families of cyrhetrene derivatives containing crown ethers, chalcones and azoles as pendant
- arms15 or incorporating functional groups of biological relevance (i.e. thiosemicarbazones, azines,
- imines) have also been prepared and evaluated as antiparasitic agents.16,17 Imines (Fig. 1, F) (more
- 123 cytotoxic than their ferrocenyl analogues and nifurtimox) are amongst the most potent anti-
- 124 Trypanosoma cruzi agents reported so far.17 These findings suggested that the assembly of the
- 125 cyrhetrenyl unit and the imine group enhances their anti-Chagas activity. Besides this, the furyl
- derivative (Fig. 1, F) (with XvO) produces reactive oxygen species that may be relevant in view of their
- 127 potential antitumoral activity.17d Despite these findings, and the ongoing interest on (a)
- 128 ferrocenylimines as antitumoral drugs themselves or as ligands to achieve transition metal complexes
- 129 with enhanced cytotoxic activity (representative examples are shown in Fig. 2)18,19 and (b) cyrhetrenyl
- 130 derivatives with relevant photophysical properties and biological activities (or both simultaneously),12–
- 131 17 mixed ferrocenyl/cyrhetrenyl imines are still unknown.
- 132 Herein, we present the two novel aldimines R1-CHvN-R2, with R1 = ferrocenyl and R2 = cyrhetrenyl
- 133 (1) or vice versa (2) (Chart 1) as the first examples of small molecules containing both organometallic
- 134 fragments connected by the imine functionality. Experimental work and computational studies on
- compounds 1 and 2 clarify the effects produced by the interchange of the two organometallic arrays on
- their structures, stabilities and properties (electrochemical and photo-physical). Also, their effect on two

- 137 breast cancer cell lines (MCF7 and triple negative MDA-MB231), the HCT116 colon cancer cell line,
- and the non-tumoral human skin fibroblast BJ cell line has been studied.

#### 140 **RESULTS AND DISCUSSION**

141 142

#### 143 Synthesis and characterization of the compounds

144

#### 145 In the first attempt to achieve the synthesis of the aldimine (1), we decided to use the procedure described previously for the ferrocenylimines $[(\eta 5-C5H5)Fe{(\eta 5-C5H4)-CHvN-R1}]$ (R1 = phenyl or 146 147 benzyl)20.21 that consisted of the reaction of equimolar amounts of the aldehyde $[(\eta 5-C5H5)Fe](\eta 5-C5H5)Fe]$ 148 C5H4)-CHO}] and the corresponding amine H2N-R1 in refluxing benzene (or toluene) and using Dean 149 Stark apparatus to remove the benzene (or toluene)-water azeotrope formed. When the reaction was carried out using cyrhetrenylamine and toluene as solvents, the IR spectra as well as the results obtained 150 from thin layer chromatography (TLC) of the solution obtained after long refluxing periods (up to two 151 days) revealed the coexistence of small amounts of the desired aldimine (1) and 152 ferrocenecarboxaldehyde as the major product. This problem is similar to that found for related 153 ferrocenylimines $[(\eta 5-C5H5)Fe{(\eta 5-C5H4)-C(R2)vN-R1}]$ with bulky substituents (i.e. R1 = phenyl 154 155 rings and R2 = Ph or Me),21–23 which were finally obtained with the aid of activated alumina or 156 molecular sieves to fulfil the condensation process. In view of this we decided to explore whether the presence of molecular sieves could improve the process. In this case (Scheme S1, A<sup>+</sup>), the progress of 157

the reaction was monitored by IR and TLC and after 24 h, both revealed the absence of the aldehyde.

- 159 The 1H-NMR spectrum of the solid obtained after concentration confirmed the formation of imine 1.
- 160 This compound was finally purified by diffusion at -18 °C of a CH2Cl2 solution of the raw material
- 161 layered with n-hexane.
- 162 The preparation of imine 2, which can be visualized as arising from 1 by a simple interchange of the
- 163 ferrocenyl and cyrhetrenyl groups, was much easier than that of imine 1. This was achieved by the
- 164 treatment of  $[(\eta 5-C5H5)Fe{(\eta 5-C5H4)-NH2}]$  and  $[Re{(\eta 5-C5H4)-CHO}(CO)3]$  in a 1 : 1 molar ratio
- and in refluxing toluene (Scheme S1, B<sup>+</sup>) but using milder experimental conditions than for 1: shorter
- refluxing periods {12 h (for 2) versus 24 h (for 1)}, in the absence of molecular sieves and without the
- aid of Dean Stark apparatus.
- 168 The new aldimines were isolated in fairly good yields (80% and 85% for 1 and 2, respectively). They are
- air-stable solids at room temperature and exhibit high solubility in CH2Cl2, CHCl3, toluene, acetone
- and acetonitrile, but they are practically insoluble in hexane. As we will demonstrate later on (see
- 171 below), compound 1 hydrolyses slowly in CDCl3 solution at 298 K.
- 172 Compounds 1 and 2 were characterized by mass (HRMS and EI) and infrared spectra, X-ray diffraction
- and NMR. Their HRMS spectra showed a peak with the expected isotopic pattern at m/z = 547.9945 (for
- 174 1) and 547.9939 (for 2) that agree with the calculated value for their [M + H]+ cations (m/z = 547.9954).
- 175 The EI spectra of compounds 1 and 2 showed the peaks of the molecular ions and the fragments formed
- 176 by the successive loss of the CO ligands.

- 177 The common features observed in the IR spectra of 1 and 2 (Experimental section) are (a) the existence
- 178 of the typical intense bands of the cyrhetrenyl units in the range of 2020–1930 cm–1 that are ascribed to
- the stretching of the pendant CO groups and (b) the presence of another and less intense band at ca.
- 180 1615 cm-1. The position of this band is similar to the values reported for ferrocenyl20,21 and
- 181 cyrhetrenyl aldimines17 and is assigned to the stretching of the >CvN– functional group.
- 182 It is well known that (a) imines may adopt two different forms (E or Z) in solution as well as in the solid
- 183 state23 and (b) the presence of bulky substituents attached to the atoms of the >CvN- group may hinder
- the free rotation around the C-R1 and or N-R2 bonds giving rotameric species.24,25 In aldimines 1 and
- 185 2, the two substituents (ferrocenyl and cyrhetrenyl) are bulky and therefore, two different arrangements
- 186 of the "Fe( $\eta$ 5-C5H5)" and "Re(CO)3" units could be expected for the (E) and (Z) forms of the imine.
- 187 This is especially relevant in the solid state, because it affects not only the Fe…Re separation but also
- the arrangement of the rings, which could introduce significant changes on the assembly of the units in
- the crystal and the crystal architecture.
- 190 Fig. S1<sup>†</sup> shows a set of isomers (a–d) for compound 1. In the pair (a, b), the imine has the E form, while
- in c and d it adopts the Z form. The two isomers of each pair  $\{(a, c) \text{ and } (b, d)\}$  differ in the relative
- disposition of the two metal ions (Fe and Re) in relation to the main plane (herein after referred to as MP
- and defined by the two substituted "C5H4" rings and the connector (>CvN– group)). The two metal ions
- 194 may be located on the opposite sides of the MP plane {anti (in a and c)} or in the same side {syn (in b
- and d)}. Although this is not shown in Fig. S1, $\dagger$  a similar set of isomeric forms could also
- be expected for imine 2.
- 197 Good X-ray quality red monocrystals of  $[(\eta 5-C5H5)Fe{(\eta 5-C5H4)-CHvN-(\eta 5-C5H4)}Re(CO)3](1)$
- and  $[(\eta 5-C5H5)Fe{(\eta 5-C5H4)-NvCH(\eta 5-C5H4)}Re(CO)3]$  (2) were obtained by slow evaporation at
- 199 -18 °C of their CH2Cl2 solutions layered with n-hexane.
- 200

# 201 Description of the crystal structures of the new aldimines (1 and 2)

- 202 The crystal structures of compounds 1 and 2 (Fig. 3 and 4, respectively) confirmed the presence of the
- 203 two organometallic units (ferrocenyl and cyrhetrenyl) connected by the functional >CvN- group. A
- selection of bond lengths, bond angles and relevant angles between planes is presented in Table 1.
- 205 Bond lengths and angles of the ferrocenyl unit fall in the range expected for related
- ferrocenylaldimines; 20, 21, 26 the pentagonal rings are planar, and nearly parallel [tilt angles =  $1.3^{\circ}$  (in 1)
- and 1.1° (in 2) and they deviate by ca. 3.7° and 5.3° (in 1 and 2, respectively) from the ideal eclipsed
- 208 conformation. In both cases, (a) Re(I) exhibits the typical three legged piano stool geometry bound to
- three CO ligands and the substituted C5H4 ring in a  $\eta$ 5 fashion and (b) bond distances and angles of the
- 210 cyrhetrenyl unit are similar to those found in monosubstituted [ $Re(\eta 5-C5H4R)(CO)3$ ]
- **211** compounds.17,26,27
- The >CvN– bond length in compounds 1 and 2 is identical [C11–N1: 1.276(6) Å]. For 1 this value is
- similar to that of  $[(\eta 5-C5H5)Fe{(\eta 5-C5H4)-CHvN-R1}]$  (R1 = substituted phenyl rings),25,26 while in

- 214 2 this is clearly greater than that found in  $[Re\{(\eta 5-C5H4)-NvCHR4\}(CO)3]$  with R4 = 4-nitrofuryl
- 215 [1.266(15) Å].17c
- As shown in Fig. 3 and 4, the organometallic arrays are in a trans-disposition [torsion angles: C12–N1–
- 217 C11–C10: 175.9(4)° (in 1) and C10–N1–C11–C12: 179.72(16)° (in 2)] confirming that both aldimines
- adopt the E form in the crystals. However, the relative arrangement of "Re(CO)3" and " $Fe(\eta 5-C5H5)$ " is
- 219 markedly different: anti in 1 and syn in 2. On these bases, the crystal structure of compound 1
- corresponds to isomer a (Fig. S1<sup>†</sup>), but that of aldimine 2 matches isomer b (Fig. S1<sup>†</sup>). As a
- consequence of this, the distance Fe…Re in 1 (7.114 Å) is bigger than that in 2 (6.122 Å) and both
- clearly exceed the sum of their van der Waals radii [Fe: 2.19 Å and Re 2.35 Å].28 Moreover, in 1, the
- two C5H4 rings are less coplanar than in 2 (angle between their main planes are 12.6° and 7.7°,
- respectively). This is also relevant because it is well-known that deviations from co-planarity between
- aromatic rings are commonly associated with a decreese of electronic delocalization.
- 226 The different arrangement of the "Fe( $\eta$ 5-C5H5)" and "Re(CO)3" units in compounds 1 and 2 also
- 227 affects the assembly of the molecules in the crystals. In compound 1, a molecule sited at (x, y, z) is
- 228 connected by C–H··· $\pi$  interactions involving (a) the C2–H2 bond and the ring defined by the set of
- atoms [C12–C16] (Fig. S2<sup> $\dagger$ </sup>) of another unit at (-x, y, 12 z) and (b) the H13 atom and the C5H5 ring
- of the ferrocenyl group belonging to a molecule on (-x, -1 + y, 12 z). Additional C–H···O short
- contacts between the oxygen atoms of the hanging CO ligands and the hydrogen atoms of three
- proximal molecules (Fig. S2<sup>†</sup>) extend the assembly of the molecules in the crystals.
- In contrast with the results obtained for 1, in the crystals molecules of 2 are assembled by two co-
- operative  $\pi$  stacking interactions (Fig. S3<sup>†</sup>), involving the substituted ring of the ferrocenyl unit of a
- 235 molecule and the C5H4 ring of another and proximal one with a head-to-tail orientation, and vice versa
- 236 giving dimers (the distance between the centroids of the two rings being 3.498 Å). Additional
- 237 intermolecular C–H···O interactions between (a) the O1 atom of one molecule and the C8–H8 bond (of
- the ferrocenyl group) of a different unit and (b) the O3 atom the C5–H5 bond of the cyrhetrenyl group of
- another molecule fulfil the assembly of the dimers.
- 240

# 241 Solution studies

- 242 1H and 13C-NMR data for both compounds are presented in the Experimental section. In both cases, the
- 243 assignment of the signals detected in their spectra has been achieved with the aid of two-dimensional
- 244 [1H–1H] NOESY and [1H–13C] HMBC NMR experiments. In the 1H-NMR spectra of the two
- aldimines the resonance of the iminic proton appeared as a singlet at 8.29 ppm (for 1) or 8.18 ppm (for
- 246 2). The position of these signals agrees with that reported for ferrocenylimines  $[(\eta 5-C5H5)Fe{(\eta 5-$
- 247 C5H4)-CHvN-R1}] (with R1 = phenyl group)20,21,25 and for the cyrhetrenyl derivatives [Re{( $\eta$ 5-
- 248 C5H4)-CHvN-R1}],17 respectively, which adopted the E form in the solid state and also in CDCl3. This
- 249 indicated that imines 1 and 2 retained the (E)-form in solution (2D-NMR studies described below
- 250 confirmed this finding). At higher fields (4.0 ppm  $< \delta < 6.0$  ppm), the 1H-NMR spectra showed a set of

- 251 five signals: an intense singlet due to the protons of the C5H5 ring, and four triplets (of identical
- intensity) that correspond to the two types [(H2 and H5) and (H3 and H4)] of different protons of in
- each C5H4 unit. Additional 2D-NMR experiments ([1H–1H] NOESY and [1H–13C] HMBC) allowed
- us to fulfil the assignment of the signals.
- 255 The [1H–1H] NOESY spectra of freshly prepared solutions of 1 and 2 in CDCl3 at 298 K showed cross
- 256 peaks between the resonances due to the imine proton at around 8.1 ppm and those of the protons on the
- ortho sites of the two C5H4 rings [pairs (H2 and H5) and (H2 and H5)]. This confirmed the E form of
- the imines 1 and 2 in solution and also the identification of the signals due to the two types of protons of
- each ring.
- 260 13C{1H} NMR spectra of 1 and 2 in CDCl3 at 298 K exhibited a singlet in the low field region (ca.  $\delta \approx$
- 193 ppm) that corresponds to CO ligands. The resonance due to the imine carbon appeared at 166.6 ppm
- 262 (for 1) and at 148.9 ppm (for 2). The position of these signals is in good agreement with those reported
- for closely related addimines  $[(\eta 5-C5H5)Fe{(\eta 5-C5H4)-CHvN-R1}]$  (R1 = phenyl group)20,21,25 and
- for the cyrhetrenyl derivative type F shown in Fig. 1.17 The up-field shift of the resonance due to the
- imine carbon observed for 2 (ca. 18 ppm) in relation to that of 1 suggests that the interchange of the
- 266 ferrocenyl and cyrhetrenyl units produces a significant change in the electronic density of the >CvN-
- 267 moiety. In both cases the signals due to the ipso carbon atoms of the C5H4 rings (C1 and C1) exhibited
- low intensity, appeared in the range 78 ppm  $\leq \delta \leq 130$  ppm and their chemical shifts were clearly
- affected by the nature of the atom to which they are bound (Nimine or Cimine). At higher fields a set of
- 270 five additional signals were also observed, of which the most intense one corresponds to the carbon
- nuclei of the C5H5 ring and the remaining ones to the two pairs of carbon atoms [(C2 and C5) and (C3
- and C4)] of each C5H4 ring (see characterization data for the two compounds).
- 273 Study of the stability of compounds 1 and 2 in solution. The stability of the two aldimines in solution
- has also been investigated by comparing the 1H-NMR spectrum of a freshly prepared solution of the
  corresponding compound in CDCl3, CD3CN or DMSO-d6 with those registered after different periods
  of storage (t) at 298 K.
- 277 The 1H-NMR spectrum of the freshly prepared solution of 1 in CDCl3 at 298 K changed with time (Fig.
- 278 S4 $\dagger$ ) and after 4 h of storage additional signals with tiny intensity were also detected. For t = 18 h the
- changes became more evident and in this case the analyses of the resonances observed suggested the
- coexistence of the aldimine, ferrocenecarboxaldehyde and the amine in a relative abundance: 1.00 : 0.78
- 281 : 0.77. Thus, this indicates that compound 1 hydrolyzes slowly in CDCl3. In contrast with the results
- obtained for 1, no significant variations in the 1H-NMR spectrum of 2 were detected after long storage
- periods (up to five days) (Fig. S5<sup>†</sup>), thus indicating that 2 is more stable than 1 in CDCl3 and less prone
  to hydrolyze.
- 285 In order to check also the stability of the aldimines in the solvent used for the electrochemical studies
  - 286 (see the following section), a parallel study was carried out using acetonitrile-d3 (Fig. S6<sup>+</sup>). In this case

- no significant changes in their NMR spectra were detected after several hours of storage, indicating that
  both compounds are stable in this solvent at 298 K.
- As mentioned above compounds holding ferrocenyl arrays and organometallic transition metal
- 290 complexes with pianostool geometries and fac-[M(CO)3] cores are gaining importance due to their
- 291 potential biological activity and utility. In view of this and since for the biological studies described
- below, the first step consisted in the dissolution of the compound in DMSO, followed by subsequent
- dilutions with water, the stability of the new products in DMSO-d6 was also investigated by 1H-NMR.
- As shown in Fig. S7,<sup>†</sup> the narrow signals observed in the spectrum of the freshly prepared solution of 1
- in DMSO-d6 broaden with time. After 4 h, the spectrum showed new sets of signals of which one was
- due to ferrocenecarboxaldehyde, probably formed by hydrolysis of the imine group. In contrast with
- these results 1H-NMR studies of compound 2 (Fig. S8<sup>†</sup>) revealed that it is much more stable and robust
- than 1 in DMSO-d6. An additional study was carried out with compound 2 using a DMSO-d6 : D2O (4 :
- 1) mixture showing that it remained practically unaltered under these experimental conditions (298 K)
- 300 for several days. The 1H-NMR spectrum registered after 112 h of storage indicated the presence of 2 as
- the major product and small amounts of the degradation product (Fig. S9<sup>†</sup>).
- 302

### 303 Comparative study of their electrochemical, photo-optical and

- 304 biological activities
- 305 Electrochemical behaviour. The electrochemical properties of new isomeric ferrocenyl/cyrhetrenyl
- aldimines 1 and 2 were also investigated. As shown above, NMR studies confirmed that both aldimines
- 307 were stable in acetonitrile. The electrochemical studies were carried out by cyclic voltammetry of
- freshly prepared solutions (10-4 mol L-1) in acetonitrile with (Bu4N)[PF6] as the supporting
- electrolyte. All these experiments were carried out at a scan rate v = 250 mV s 1. Cyclic
- 310 voltammograms (hereinafter referred to as CVs) are shown in Fig. 5 and a summary of electrochemical
- 311 data for compounds under study is presented in Table 2.
- 312 The CVs of the new aldimines showed in the range -1.20 V  $\leq$  E  $\leq$  0.50 V (Fig. 5A) an oxidation peak
- 313 (I) with a directly associated reduction one in the reverse scan (I'), the intensity ratio (Ipa/Ipc) was close
- to 1 and the separation between the oxidation and reduction peaks  $\Delta E = (EIpa EI' pc was similar to$
- that obtained for ferrocene under identical experimental conditions. These findings agree with those
- expected for a simple reversible one electron-process.29 It is well-known that the proclivity of ferrocene
- derivatives to oxidize is strongly dependent on the nature of the substituents.9,30 In general, the
- 318 presence of electron withdrawing groups produces an increase of the Epa value, while donor groups
- 319 have the opposite effect.
- 320 For compound 1 the position of the wave and the anodic potential are similar to those for
- 321 ferrocenylaldimines of general formulae  $[(\eta 5-C5H5)Fe{(\eta 5-C5H4)-CHvN-R4}]$  and especially to those
- with R4 = CH2-C6H5 {EI pa = 0.20 V30} under identical conditions. This suggests that the effect
- produced by the  $Re(\eta 5-C5H5)(CO)3$  unit on the proclivity of Fe(II) to oxidize is similar to that of the

- benzyl group. In contrast with these results, for isomer 2, the wave shifts to the more cathodic region,
- thus indicating that the "-NvCH( $\eta$ 5-C5H4)Re(CO)3" unit is a stronger electron-donor group than "-
- 326 CHvN(η5-C5H4)Re(CO)3".
- 327 Besides this, the anodic potential (EI pa) of 2 is quite close to that of ferrocene itself, and therefore more
- prone to oxidize than 1. It is well-known that in biological systems the accessible redox potential
- 329 window ranges only from around -0.40 to +0.80 V versus the normal hydrogen electrode (NHE).31
- Compound 2 has a redox potential quite close to that of ferrocene [E(Fc/Fc+) = 0.5 V versus NHE] and
- in the range of the biological systems, while that of the aldimine 1 is higher and closer to the upper limit
- of the biological range. This differential behaviour may be important in view of their potential utility in
- drug design or for their use as biomarkers.
- 334 When the cyclic voltammograms were registered in a wider range of potentials [from -1.00 V to +1.60
- 335 V], an additional and poorly resolved oxidation peak (II in Fig. 5B) was observed. The potential of this
- peak [EIIpa = 1.29 V (for 1) and 1.42 V (for 2)] falls in the range reported for other cyrhetrene
- derivatives and has been assigned to the typical oxidation of Re(I) to Re(II).15a In the reverse scan (Fig.
- 5B), two (for 1) or three (for 2) additional reduction peaks were detected in the range  $-0.4 \text{ V} \le E \le 0.4$
- 339 V, of which one (I') corresponds to the reduction of Fe(III) formed during oxidation.
- 340 These studies provide conclusive evidence of the effect produced by the location of the ferrocenyl and
- 341 cyrhetrenyl arrays in the aldimines R1-CHvN-R2. The shifts detected in the position of anodic peaks
- 342 (Elpa and EII pa) for the two isomers suggest that the interchange of the two organometallic units
- produces a significant variation in the electronic distribution of charge. In both cases, the first anodic
- peak (I) corresponds to the oxidation of the Fe(II) centre, but for 2 it occurs at lower potentials than for
- 1, thus suggesting that the energy level of the HOMO of 2 is higher than that of 1. Computational
- 346 studies described below confirm this hypothesis.
- 347 Absorption and emission spectra. As mentioned in the Introduction section, one of the most attractive
- 348 properties of compounds holding fac-[M(CO)3] units arises from their potential luminescence that may
- allow their use as luminescent probes or sensors. Since the new aldimines contain this array we also
- investigated their photophysical properties. First, the UV-vis spectra of 10–4 mol L–1 solutions of
- compounds 1 and 2 in CH2Cl2 at 298 K were registered (Fig. 6 and Table 3). In both cases three (for 1)
- and four (for 2) intense absorption bands (with extinction coefficients,  $\varepsilon$  in the range of 10–3–10–4M–1
- 353 cm-1) were observed. The position of the band at lower energies ( $\lambda \approx 475$  nm) agrees with that of
- related ferrocenyl imines and hydrazones32 and has been attributed to a d-d transition of the Fe(II)
- 355 centre. Computational studies (see the following section) confirmed this assignment. In the UV-vis
- spectrum of complex 2 (Fig. 6), another absorption maximum at  $\lambda = 383$  nm was observed, which is
- 357 characteristic of cyrhetrene derivatives.11,32,33 For 1 this band is not observed in the spectrum
- 358 probably due to the presence of more intense bands at lower wavelengths that may mask it. In both cases
- two additional and more intense bands were also detected in the range of 220–320 nm.
- 360

- 361 Due to the increasing attractiveness of luminescent Re(I) complexes with "Re(CO)3" arrays,6,8,11,12
- 362 we also investigated the emissive properties of the new aldimines in CH2Cl2 at 298 K. Upon excitation
- at 477 nm, both complexes showed an emission band at 551 nm (Fig. 7A), which, according to the
- literature, 33 originates from a 3MLCT phosphorescence state.
- 365 As mentioned above, the UV-vis spectrum of compound 2 showed also an additional absorption band  $\lambda$
- 366 = 383 nm, and after excitation at 385 nm, the emission spectra of 2 (Fig. 7B) showed three bands in the
- range 400 nm  $\leq \lambda \leq$  500 nm. When the experiment was carried out under identical conditions, but using
- 1 instead of 2, the emission spectra showed a similar pattern (Fig. 7B). This suggests that the typical
- band of the "Re(CO)3" unit was not observed in the UV-vis spectrum. The spectrum of 1 was probably
- masked by the intense absorption at higher energies ( $\lambda = 314$  nm). Time-dependent DFT calculations
- 371 (see below) confirmed this hypothesis.
- 372 Computational studies. As mentioned above, multifunctional compounds with potential utility in new
- 373 materials and technological development are attracting a great deal of inter-est due to their potential
- utility in new materials design and technological devices.5 The new aldimines exhibit interesting
- photophysical and electrochemical properties and in addition it is well known that imines are valuable
- reagents in organic and organometallic synthesis.34 In view of these and in order to compare the
- stability of the isomers of 1 and 2 and to explain the effect produced by the interchange of the two
- 378 organometallic arrays in R1-CHvN-R2 on their stability, properties and reactivity of the new
- 379 compounds, computational calculations on both imines were carried out. Since (a) the X-ray crystal
- 380 structures and the NMR studies described above confirmed that imines 1 and 2 adopted the (E) form in
- the solid state and also in solution and, (b) the use of molecular models showed that for isomers (types c
- and d in Fig. S1<sup>†</sup>), with the imine in the syn (Z) form, the proximity of the C5H4 rings would introduce
- 383 strong steric hindrance that may reduce their stability, only types a and b isomers of the two aldimines
- 384 were used in the computational studies.
- All the calculations were carried out using the B3LYP hybrid functional35 and the LANL2DZ36a,b (for
- Fe and Re) and 6-31G\*36c,d (for the remaining atoms) basis set implemented in Gaussian 03
- software.37 Geometries of the E-isomers of compounds 1 and 2 with different arrangements of the
- 388 "FeCp" and "Re(CO)3" units (types a and b in Fig. S1<sup>†</sup>) were optimized without imposing any
- restriction. Final atomic coordinates of the optimized geometries are included in the ESI<sup>+</sup>
- 390 (Coordinates.xyz file). Bond lengths and angles of isomers 1a and 2b were consistent with those
- 391 obtained from X-ray crystal structures (described above).
- 392 A comparison of the calculated values of the total energy (ET) obtained for the pairs (1a and 1b) and (2a
- and 2b) (Table S1<sup>†</sup>) revealed that in vacuum, the ET values of isomers (1b and 2b) with the syn
- orientation of the Fe(Cp) and Re(CO)3 moieties were slightly smaller than of their anti analogues (1a
- and 2a, respectively) [ET (for 1a) ET (for 1b) = 1.4 kcal mol-1 and ET (for 2a) ET (for 2b) = 0.8
- kcal mol-1]. In CH2Cl2 the difference between the ET values of the two isomers 2a and 2b decreased to

- 397 0.2 kcal mol-1, while for the imine  $[(\eta 5-C5H5)Fe {(\eta 5-C5H4)-CHvN-(\eta 5-C5H4)}Re(CO)3]$  the total 398 energy of the anti isomer 1a was found to be 2.7 kcal mol-1 higher than that of 1b.
- 399 In order to get further information about the stability of the four isomers, their free energies were also
- 400 calculated (Table S1<sup>†</sup>). The results obtained in vacuum revealed that for 1 the anti isomer (1a) is 1.58
- 401 kcal mol-1 more stable than its syn analogue (1b), while for the aldimine 2, the trend is just the
- 402 opposite. Moreover, the free energies increase according to the sequence 1a < 2b < 2a < 1b, in vacuum,
- 403 thus indicating a decrease in the stability of the four isomers. When the calculations were carried out
- 404 taking into account the effect produced by the solvent (CH2Cl2), the calculated free energies followed
- 405 the same trend, showing again that isomers 1a and 2b (which are found in the crystal structures) are
- 406 more stable than their corresponding partners (1b and 2a, respectively) and the differences  $\Delta$ Gr, defined
- 407 as  $\Delta G$  (of isomer 1b, 2a or 2b)  $\Delta G$  (of 1a, the most stable isomer of the set), in CH2Cl2 were higher
- 408 than those in vacuum (Table 4). According to the calculation compound 1a is expected to be more stable
- 409 in CH2Cl2 than 2b. However, the NMR studies revealed that 1a hydrolysed slowly in CDCl3 and also in
- 410 DMSO-d6. This could be due to the presence of traces of HCl and/or residual water in the deuterated
- 411 solvents.
- Frontier orbitals [HOMO-1, HOMO, LUMO and LUMO+1] for the most stable isomer of each product
- 413 (1a, and 2b) are depicted in Fig. 8. The HOMO-1 of 1a and 2b is similar and located on the ferrocenyl
- unit. In both cases the HOMO is centered on >CvN- and the ferrocenyl unit, with a minor contribution
- 415 of the cyrhetrenyl moiety. The oxidation of the two aldimines involves this orbital. For 2b, its energy
- 416 level is higher than that of the HOMO of 1a (Fig. 8); therefore, the removal of one electron is expected
- 417 to require less energy than for 1a. This explains the differences observed in the cyclic voltammograms
- 418 shown in Fig. 5A and the shift of the first anodic peak (Elpa) to the more cathodic region (Table 2).
- 419 Furthermore, comparative analyses of the charge distribution on the metals and imine functional group
- 420 for isomers 1a and 2b (Table S2<sup>+</sup>) showed some interesting features. The interchange of the ferrocenyl
- 421 or cyrhetrenyl arrays in R1-CHvN-R2 of 1a (to give 2b) produces significant variations in the charges of
- 422 the ipso carbon atoms (C1 and C1'). In contrast, the Mulliken charges on nitrogen are practically
- 423 identical in both cases and the values are similar to those found in other ferrocenyl Schiff bases with rich
- 424 and versatile coordination ability to transition metal ions [i.e. Pd(II) and Pt(II)].18,25a This is relevant in
- 425 view of their potential use as metalloligands to achieve heterotrimetallic complexes containing Fe(II),
- 426 Re(I) and an additional metal ion Mm+ such as Pd(II) and Pt(II) among others. However, it should be
- 427 noted that in 2b, the imine nitrogen is not as accessible as in 1a due to steric hindrance.
- 428 In order to elucidate the origin of the bands detected in the UV-vis spectra in the range 300–500 nm, we
- 429 decided to undertake a study based on time-dependent DFT (TD-DFT) methodology38 to achieve the
- 430 assignments of the bands and to confirm the existence of an absorption band at around 380 nm not
- 431 observed in the UV-vis spectrum of 1. After the optimization of the geometries in vacuum, the
- 432 excitation energies and the oscillator strengths were calculated in CH2Cl2 solution (Table S3<sup>+</sup>). The

- calculated absorption spectra for the two pairs of isomers under study (1a, 1b) and (2a, 2b) are presented
  in Fig. S10.<sup>†</sup>
- 435 The computational results obtained revealed that the main absorption bands arise from a combination of
- 436 several monoelectronic transitions, of which those with greater contributions are presented in Table S3.†
- 437 For instance, the band that appears in the range 460–500 nm mainly involves three (for 1a) or two
- 438 monoelectronic transitions between MO mainly centred on Fe(II), while that observed in the
- 439 experimental UVvis spectrum of 2b at 383 nm results from the HOMO  $\rightarrow$ LUMO transition, and in both
- there is a contribution of the cyrhetrenyl unit. Although for 1a, the existence of an absorption band at
- 441 around 380 nm was not as evident as for 2a, it showed emission after excitation at the same wavelength
- 442 as for 2b ( $\lambda$ exc. = 385 nm). This suggested that 1a might also exhibit electronic transitions at around 380
- 443 nm. The computational studies confirmed this hypothesis and the presence of an absorption band at
- 444  $\lambda$ calc. = 387.2 nm that arises from two main monoelectronic transitions: the HOMO  $\rightarrow$  LUMO (as for
- 445 2b) and the HOMO-1  $\rightarrow$  LUMO+3.
- 446 The experimental UV-vis spectra (Fig. 6) also exhibited two intense and broad bands at lower
- 447 wavelengths ( $\lambda < 350$  nm), and according to these computational studies, they result from a large
- 448 number of monoelectronic transitions that take place in a narrow range of energies (Table S3 and Fig.
- 449 S10<sup>†</sup>), of which one of those with greater weight (81% for 1a and 82% for 2b) is the HOMO-2  $\rightarrow$
- 450 LUMO. For 1a this transition leads to an absorption band at  $\lambda$  calc. = 312 nm, while for 2b it appears
- 451 slightly shifted (ca. 11 nm) to lower energies ( $\lambda$ calc. = 323 nm).
- 452 Biological studies. Due to increasing interest on organometallic compounds in new drug design, and
- 453 especially in cancer therapy, in vitro studies on the effect produced by aldimines 1 and 2 on two human
- 454 breast cancer cell lines [MCF7 and MDA-MB231] and the cisplatin resistant HCT-116 colon cell line
- 455 were also carried out. In these studies cisplatin was also used as the positive control under identical
- 456 experimental conditions. A summary of the results obtained for the inhibition concentrations (IC50 in
- 457  $\mu$ M) is presented in Table 5 and Fig. 9 and viability plots are shown Fig. S11.<sup>†</sup>
- 458 The comparison of results reveals that (a) compound 1 is clearly less active than 2; and (b) aldimine 2
- 459 showed a cytotoxic effect similar to that of cisplatin in MCF7, but it resulted ca. two times more potent
- 460 on the triple negative (ER, PR and no HER2 over expression) MDA-MB231 cell line. In view of this,
- 461 we also evaluated their activity on the cisplatin resistant HCT-116 adenocarcinoma colon cell line. As
- shown in Table 5 and Fig. S12<sup>†</sup> compound 2 had a greater inhibitory growth effect than the reference
- 463 drug, while 1 did not show any relevant activity (IC50 > 30  $\mu$ M).
- 464 A parallel study of the antiproliferative effect of the compounds on the normal and non-tumoral human
- skin fibroblast BJ cell lines was also carried out. The results, presented in Table 5 and Fig. S12<sup>†</sup> show
- that the cytotoxicity of the compounds increases according to the sequence  $1 < \text{cisplatin} \le 2$ . Although
- 467 compound 2 has an inhibition growth potency on the BJ cell line quite similar to that of cisplatin, it is
- 468 particularly attractive due to (a) its remarkable stability in the solid state and also in solution; (b) its
- 469 cytotoxic potency that is comparable to (in MCF7) or even greater (in MDA-MB231 and HCT116) than

- 470 that of cisplatin; and (c) the fact that it does not contain Pt(II) and consequently might not produce the
- 471 typical and undesirable side effects of the conventional platinumbased drugs used clinically.39

- 473 CONCLUSIONS
- 474

Two new and isomeric aldimines R1-CHvN-R2 [with R1 = ferrocenyl and R2 = cyrhetrenyl (1) or vice 475 versa (2)] have been prepared and characterized in the solid state as well as in solution. The interchange 476 477 of the position of the two organometallic arrays in the R1-CHvN-R2 backbones produces significant 478 changes in their structures, the assembly of the molecules in the crystals, their stability, their 479 electrochemical and photophysical properties and also their biological activity. In particular, and despite 480 the formal similarity of 1 and 2 and the fact that both imines adopt the E form in the solid state, in 1 the "FeCp" and "Re(CO)3" arrays are in an anti disposition, while in 2, they are in syn. This affects the type 481 of assembly of the molecules and the intermolecular interactions. We have also proved that aldimine 2 is 482 less prone to hydrolyse and more proclive to undergo the first one electron oxidation process than 1. 483 484 The results obtained from the theoretical calculations have allowed us not only to compare (a) the 485 stability of the two isomers of these products with the imine in the E form and differing in the relative orientation of the "Fe(Cp)" and Re (CO)3 units in vacuum and in CH2Cl2, (b) the effect produced by 486 the interchange of the substituents on electronic delocalization and charge distribution and (c) the 487 488 different electrochemical behaviour, but also to assign the main monoelectronic transitions and the main 489 absorptions bands observed in their UV-vis spectra. 490 The new hybrid imines exhibit luminescence in CH2Cl2 at 298 K. These findings together with the results obtained from their in vitro studies on their antiproliferative effect on the three human cancer cell 491 lines [breast (MDA-MB231 and MCF7) and colon (HCT-116)] and non-tumoral BJ cells increase the 492 value of these products as a novel type of multifunctional compound, due to their electrochemical 493 494 behaviour, emissive properties or antitumor activity. Among the two new products presented here, compound 2, with remarkable stability in DMSO and also in DMSO-d6 : water mixtures, appears to be 495 496 an excellent candidate for further work mainly centred on its cytotoxic activity against other cancer cell 497 lines (i.e., lung, ovarian, etc.), its mechanism of action, and also its potential as an theranostic agent.40 498 Besides this, both imines have additional interest as building blocks and especially as metallo-ligands to 499 other transition metal ions Mm+ such as Pt(II), Pd(II), and Ru(II), to achieve heterotrimetallic 500 compounds with three metal centres [Fe(II), Re(I) and Mm+] and "potentially bioactive units", which are attractive not only in view of their potential interest in new drug design and development or 501 biotechnology, but also in other emerging fields, such as cooperative catalysis. 502 503

504 **EXPERIMENTAL** 

505

506

#### 507 **General remarks**

- 508 509  $[Re{(\eta 5-C5H4)-CHO}(CO)3]$  and the amines  $[Re{(\eta 5-C5H4)-NH2}(CO)3]$  and  $[{(\eta 5-C5H5)Fe(\eta 5-C5H4)-NH2}(CO)3]$ C5H4)-NH2}(CO)3] were prepared as described previously.41-43 Ferrocene (98%) and 510 511 errocenecarboxaldehyde (98%) were obtained from Aldrich and used as received. The solvents 512 (CH2Cl2, hexane and toluene) were obtained commercially and were purified using standard 513 methods.44 All manipulations were conducted under an N2 atmosphere using Schlenk techniques. 514 High resolution mass spectra (HRMS) were recorded at the Servei de Espectrometría de Masses (Univ. Barcelona) using a LC/MSD-TOF Agilent Technologies instrument and electron impact (EI) mass 515 spectra were obtained on a Shimadzu GC-MS spectrometer (70 eV) at the Laboratorio de Servicios 516 Analíticos (Pontificia Universidad Católica de Valparaíso). Infrared spectra of 1 and 2 were obtained 517 using a Nicolet 400 FTIR instrument with KBr pellets. UV-visible (UV-vis) spectra of  $1.0 \times 10-4$  M 518 519 solutions of the compounds in CH2Cl2 were recorded on a Cary 100 scan Varian UV spectrometer at 298 K. Emission spectra of CH2Cl2 solutions of 1 and 2 were obtained on a Horiba Jobin-Yvon SPEX 520 521 Nanolog-TM spectrofluorimeter at 298 K. 522 Routine 1H and 13C{1H} NMR spectra were recorded at 298 K on a Bruker Fourier 300 or a Mercury 523 400 MHz instrument. High resolution 1H-NMR spectroscopy and two dimensional NMR experiments were carried out using Bruker 400 Avance III HD equipment. Except where quoted, the solvent used for 524 525 NMR studies was CDCl3 (99.9%) and SiMe4 was the internal reference. The assignment of signals 526 detected in the 1H and 13C{1H} NMR spectra was achieved with the aid of two dimensional [1H–1H] 527 nuclear Overhauser effect spectroscopy (NOESY) and [1H-13C] heteronuclear multiple-bond 528 correlation spectroscopy (HMBC) experiments. NMR data are presented in the characterization section 529 of each compound. Chemical shifts ( $\delta$ ) are given in ppm and the coupling constants (J) in Hz, the 530 assignment of the resonances observed refers to the labelling patterns presented in Scheme S1<sup>+</sup> and the abbreviations for the multiplicities of the signals are s (singlet) and t (triplet). 1H-NMR studies of 531 compounds 1 and 2 in acetonitrile-d3 (Fig. S6<sup>+</sup>) and DMSO-d6 (Fig. S7 and S8<sup>+</sup>) at 298 K and of 1 in 532 DMSO-d6 : D2O (4 : 1) (Fig. S9<sup>+</sup>) were also undertaken in order to evaluate the stability of the 533 compounds in the solvents used in the electrochemical studies and in the biological studies. 534 535 536 **Preparation of the compounds** 537 538 Synthesis of compound [(n5-C5H5)Fe{(n5-C5H4)-CHvN-(n5-C5H4)}Re(CO)3] (1). To a solution of 539
- 540  $[Re{(n5-C5H4)-NH2}(CO)3 (122.7 mg, 3.5 \times 10-4 mol) in 20 mL dry toluene, [(n5-C5H5)Fe{(n5-C5H5)-C5H5}) = (n5-C5H5) = (n5-C5H$

- C5H4CHO)}] (75.0 mg,  $3.5 \times 10-4$  mol) and 4 Å molecular sieves (ca. 2.1 g) were added. The flask 541 542 containing the resulting mixture was connected to Dean-Stark apparatus and a condenser and then the reaction mixture was refluxed for 24 h. After this period, the solvent was removed under vacuum giving 543 a red solid that was later on crystallized from CH2Cl2/hexane at -18 °C to give orange crystals of 1 544 545 (yield: 153.2 mg,  $2.8 \times 10-4$  mol, 80%). Characterization data: Mass spectrum: HRMS (m/z): 547.9945, 546 calc. for: C19H15FeNO3Re: 547.9954; EIMS (based on 187Re) m/z: 547 [M+], 518 [M+-CO], 491 [M+-2CO], 463 [M+-3CO]. IR selected data (KBr; cm-1): 2010[v(CO)], 1931 [v(CO)] and 547 548 1613[v(CvN)]. 1H NMR (400 MHz) δ: 8.29 (s, 1 H, –CHvN–), 5.40 [t, 2H, 3J = 2.3, (H2 and H5)]; 5.23 549 [t, 2H, 3J = 2.3, (H3 and H4)], 4.71 [t, 2H, 3J = 1.9, (H2 and H5)], 4.52 [t, 2H, 3J = 1.9, (H3 and H4)] 550 and 4.24 (s, 5H, Cp). 13C NMR (75 MHz) δ: 194.5 (CO); 166.6 (>CHvN-); 129.4 (C1); 81.5 (C2 and C5); 78.9 (C1); 76.5 (C3 and C4); 72.2 (C2 and C5); 69.7 (Cp); and 69.5 (C3 and C4). 551 Synthesis of compound [(η5-C5H5)Fe{(η5-C5H4)-NvCH-(η5-C5H4)}Re(CO)3 (2). [Re{(η5-C5H4)-552 CHO}(CO)3] (75.0 mg, 2.1.  $\times$  10–4 mol) was added to a solution of [( $\eta$ 5-C5H5)Fe{( $\eta$ 5-C5H4)-NH2}] 553 (41.5 mg,  $2.1 \times 10-4$  mol) in 20 mL dry toluene. The mixture was refluxed for 12 h and after this 554 period, the solvent was pumped off giving a red solid, which was later on crystallized by slow 555 evaporation at -18 °C of a CH2Cl2 solution layered with n-hexane (yield: 96.2 mg,  $1.7 \times 10-4$  mol, 556 85%). Characterization data: Mass spectrum: HRMS (m/z): 547.9939, calc. for: C19H15FeNO3Re: 557 547.9954; EIMS (based on 187Re) m/z: 547 [M+], 518 [M+-CO], 491 [M+-2CO], 464 [M+-3CO]. IR 558 559 selected data (KBr; cm-1): 2015[v(CO)], 1932 [v(CO)] and 1610[v(CvN)]. 1H NMR (400 MHz)  $\delta$ :8.18 (s, 1H, -CHvN-), 5.93 [t, 2H, 3J = 2.2, (H2 and H5)], 5.40 [t, 2H, 3J = 2.2, (H3 and H4)], 4.49 [t, 2H, 3J 560 = 1.8, (H2 and H5)], 4.25 [t, 2H, 3J = 1.8, (H3 and H4)] and 4.20 (s, 5H, Cp). 13C NMR (75 MHz) δ: 561 193.2 (CO); 148.9 (>CvN-); 104.1 (C1); 101.1 (C1); 85.3 (C2 and C5), 84.70 (C3 and C4); 69.7 (Cp); 562 563 67.7 (C2 and C5) and 62.8 (C3 and C4).
- 564
- 565

# 566 Crystallography

- 568 A red prism-like specimen of 1 or a red plate-like crystal of 2 (sizes in Table 6) was selected and 569 mounted on a D8 Venture system equipped with a multilayer monochromator and a Mo microfocus ( $\lambda =$ 570 0.71073 Å). The frames were integrated with the Bruker SAINT software package using a narrow-frame 571 algorithm. The integration of the data using an orthorhombic (for 1) and a triclinic unit cell (for 2)
- 572 yielded a total of 21 377 (for 1) and 37 637 (for 2) reflections to a maximum  $\theta = 28.30^{\circ}$  (for 1) or 30.57°
- 573 (for 2) (0.75 Å and 0.70 Å resolution, respectively). For 1, 4125 were independent (average redundancy
- 574 5.182, completeness = 99.7%, Rint = 5.94%, Rsig = 4.85% and 3094 (75.01%) were greater than 2
- 575  $\sigma(F2)$ ), while for 2, 5064 reflections were independent (average redundancy 7.432, completeness =
- 576 99.8%, Rint = 3.06%, Rsig = 1.91% and 4758 (93.96%) were greater than 2  $\sigma$ (F2)). The final cell
- 577 constants and volumes for 1 and 2 (Table 6) are based upon the refinement of XYZ centroids of

- 578 reflections above 20  $\sigma$ (I). Data were corrected for absorption effects using the multi-scan method
- 579 (SADABS). The calculated minimum and maximum transmission coefficients (based on the crystal size)
- 580 were 0.5655 and 0.7457 for 1 and 0.5529 and 0.7461 for 2.
- 581 The structures were solved and refined using the SHELXTL software package45 using the Pbcn (with Z
- 582 = 8, for 1) and the P1<sup>-</sup> (Z = 2, for 2) space group for the formula unit C19H14FeNO3Re. The final
- anisotropic full-matrix least-squares refinement on F2 with 226 variables converged at R1 = 3.36 (for 1)
- and 1.50% (for 2) for the observed data, and wR2 = 6.01% (for 1) and 3.17% (for 2) for all data. The
- largest peak in the final difference electron density synthesis was 0.888 e Å–3 (in 1) and 0.691 e Å–3 (in
- 586 2) and the largest hole was -1.093 e Å-3 (in 1) and -1.031 e Å-3 (in 2) with RMS deviations of 0.196
- and 0.118 e Å–3 for 1 and 2, respectively. Further detail concerning the resolution and refinement of the
- two crystal structures is presented in Table 6.
- 589 CCDC 1576497 (for 1) and 1576498 (for 2)<sup>†</sup> contain the supplementary crystallographic data for this
  590 paper.
- 591
- 592

# 593 Electrochemical studies

594

595 Cyclic voltammetric (CV) studies were carried out at room temperature using a potentiostat (Metrohm

- 596 Autolab potentiostat) in a three-electrode cell. Each complex was dissolved in acetonitrile containing 0.1
- 597 mol L-1 tetrabutylammoniumhexafluorophosphate (Bu4N)[PF6], as the supporting electrolyte to give
- 598 10–3 mol L–1 final concentration. A platinum 2 mm working electrode and a platinum coil counter
- 599 electrode were used. The reference electrode contained a silver wire with 10 mM silver nitrate in
- 600 (Bu4N)[PF6] electrolyte solution.
- 601 The working electrode was polished with 0.3 and 0.05 μm alumina slurries, rinsed with distilled water
- $(18 \text{ M}\Omega \text{ cm})$  and acetone, and dried prior to use. All electrolyte solutions were thoroughly pre-purged
- using purified nitrogen gas before use. The measurements were carried out at a scan rate of 0.25 V s-1.
- 604 The ferrocene/ferricinium (Fc/Fc+) couple served as the internal reference and appeared at +89 mV (vs

Ag/Ag+) for each experiment.

606 607

#### 608 Computational studies

- 609
- 610 DFT calculations were carried out using Gaussian 03 software, 37 with the B3LYP functional.35 The
- basis set was chosen as follows: LANL2DZ36a,b for Fe and Re and 6-31G\*36c,d (including
- 612 polarization functions for non-hydrogen atoms) for C, N, O and H. All molecular structures were
- 613 optimized without symmetry constraints and characterized as minima by vibrational analysis. Solvent
- 614 effects have been included using the CPCM method.46

- 615 **Biological studies**
- 616

617 Cell culture. Breast cancer (MCF7 and MDA-MB231) cells (from European Collection of Cell Cultures,

- ECACC) and colon adenocarcinoma (HCT-116) cells (from the American Type Culture Collection)
- 619 were used in all the experiments. Cells were grown as a monolayer culture in minimum essential
- 620 medium (DMEM with L-glutamine, without glucose and without sodium pyruvate) in the presence of
- 621 10% heat-inactivated fetal calf serum, 10 mM of D-glucose and 0.1% streptomycin/penicillin under
- 622 standard culture conditions.
- 623 Cell viability assays. For these studies, the compounds were dissolved in 100% DMSO at 50 mM as
- stock solution; then, serial dilutions were prepared in DMSO (1 : 1) (in this way the DMSO
- 625 concentration in cell media was always the same); and finally, 1 : 500 dilutions of the serial dilutions of
- 626 the compounds in cell media were prepared. The assay was performed as described by Givens et al.47
- In brief, MDA-MB231 and MCF7 cells were plated at 5000 cells per well and 10 000 cells per well,
- respectively, in 100 Ml media in tissue culture 96 well plates (Cultek). After 24 h, the medium was
- 629 replaced by 100 μL per well of serial dilution of drugs. Each point concentration was run in triplicate.
- 630 Reagent blanks, containing media plus the colorimetric reagent without cells, were run on each plate.
- Blank values were subtracted from test values and were routinely 5–10% of uninhibited control values.
- The plates were incubated for 72 h. Hexosaminidase activity was measured according to the following
- 633 protocol: the media containing the cells were removed and the cells were washed once with PBS; 60 μL
- 634 substrate solution (p-nitrophenol-N-acetyl-β-D-glucosamide 7.5 mM [Sigma N-9376], sodium citrate
- 635 0.1 M, pH = 5.0, 0.25% Triton X-100) was added to each well and incubated at 37 °C for 1–2 hours;
- after this incubation time, a bright yellow color appeared; then, the plates could be developed by adding
- 637 90  $\mu$ L developer solution (Glycine 50 mM, pH = 10.4; EDTA 5 mM); and absorbance was recorded at
- 638 410 nm.
- 639 The human skin fibroblast cell line BJ was cultured in DMEM in the presence of 10% FBS, 12.5 mM
- 640 DE-glucose, 4 mM glutamine, 5 mN pyruvate and 0.5 streptomycin/penicillin. All the cells were
- 641 incubated under standard conditions (humidified air with 5% CO2 at 37 °C). The cells were passaged at
- 642 confluence by washing once with cation-free HBSS followed by a 3 minute incubation with trypsin ([0.5
- 643  $\mu g mL-1$ /EDTA [0.2  $\mu g mL-1$ ]) (Gibco-BRL, 15400054) solution in HBSS at 37 °C, and transferred to
- 644 its medium. Prior to seeding at a defined cell concentration, the cells were recovered from the medium
- by centrifugation and counted. For proliferation studies, the cells were plated at 5000 cells per well in
- 646 100 μL media in tissue culture 96 well plates (Cultek). After 24 h, the media were replaced by 100 μL
- 647 per well of serial dilution 1 : 2 of compounds 1 and 2.
- 648 For comparison purposes, a parallel study with cisplatin was also carried out under identical conditions.
- 649 Reagent blanks, containing media plus colorimetric reagent without cells, were run on each plate. Blank
- 650 values were subtracted from test values and were routinely 5-10% of uninhibited control values. The
- 651 plates were incubated for 72 h. Hexosaminidase activity was measured according to the following

- 652 protocol: the media containing the cells were removed and the cells were washed once with PBS; 60 μL
- of substrate solution (p-nitrophenol-N-acetyl-beta-D-glucosamide 7.5 mM [Sigma N-9376], sodium
- 654 citrate 0.1 M, pH 5.0, 0.25% Triton X-100) was added to each well and incubated at 37 °C for 1–2
- hours; after this incubation time, a bright yellow color appeared; then, the plates could be developed by
- adding 90 μL developer solution (Glycine 50 mM, pH 10.4; EDTA 5 mM); and absorbance was
- recorded at 410 nm.
- 658

# 659 ACKNOWLEDGEMENTS

- 660
- 661 H. K. and R. A. acknowledge FONDECYT-Chile (Projects 1150601 and 11130443), FONDEQUIP
- 662 EQM 130154 and D. I. Pontificia Universidad Católica de Valparaíso. J. O. is grateful to CONICYT-
- 663 PFCHA for a doctoral scholarship number 21170802 and D.I.-PUCV. C. P. S. is grateful to Postdoc
- 664 DICYT code 021740PI, Vicerrectoría de investigación, Desarrollo e investigación. This work was also
- supported by the Ministerio de Economia y Competitividad of Spain [grant number CTQ2015-65040-P
- 666 (subprograma BQU)].

668	REFERENCES
-----	------------

# (a) J. Pombeiro and J. A. McCleverty, Molecular Electrochemistry of Inorganic, Bioinorganic and Organometallic Compounds, Springer Science & Business Media, 2012; (b) D. M. Roundhill and J. P. Kackler, Optoelectronic properties of Inorganic Compounds, Springer Science & Business Media, 2013.

- 674 2 S. Di Bella, Chem. Soc. Rev., 2001, 30, 355–366.
- 675 3 (a) Homo and Heterobimetallic Complexes in Catalysis: Cooperative Catalysts, ed. P. Kalk,
  676 Springer, 2016; (b) M. H. Perez-Temprano, J. A. Casares and P. Espinet, Chem. Eur. J., 2012,
  677 18, 1864–1884.
- Selected articles on heterobimetallic compounds with outstanding biological activity: (a) Y. F.
  Mui, J. Fernandez-Gallardo, B. T. Elie, A. Gubran, I. Maluenda, M. Sanau, O. Navarro and M.
  Contel, Organometallics, 2016, 35, 1218–1227; (b) M. Wenzel, A. De Almeida, E. Bigaeva, P.
  Kavanagh, M. Picquet, P. Le Gendre, E. Bodio and A. Casini, Inorg. Chem., 2016, 55, 2544–
  2557.
- 5 L. Ouahab, Multifunctional Molecular Materials, Pan Sanford. Publish, Singapur, 2013.
- 684 6 Bioorganometallic Chemistry: Applications in Drug Discovery, Biocatalysys, and Imaging, ed.
  685 G. Jaouen and M. Le Salmain, Wiley-VCH, Weinheim, Germany, 2015.
- 686 7 For a general overview on the potential of organometallic compounds in drug design, medicinal chemistry and therapeutic uses see for instance: (a) B. Biersack and R. Schobert, Adv. Exp. 687 Med. Biol., 2016, 893, 211–224; (b) T. S. Morals and M. H. Garcia, Adv. Organomet. Chem. 688 689 Catal., 2014, 581-587; (c) P. Martins, M. Marqués, L. Coito, J. Pombeiro, P. Viana-Baptista and A. R. Fernandes, Anti-Cancer Agent Med. Ther., 2014, 14, 1199–1212; (d) D.-L. Ma, D. S.-H. 690 Chan and C.-H. Leung, Acc. Chem. Res., 2014, 47, 3613–3614; (e) B. Anilammert, 691 692 Pharmacology, 2012, 651-680; (f) G. Gasser and N. Metzler-Nolte, Curr. Opin. Chem. Biol., 2012, 16, 84–91; (g) N. Chavain and C. Biot, Curr. Med. Chem., 2010, 17, 2729–2745; (h) C. 693 Gaiddon and M. Pfeffer, Eur. J. Inorg. Chem., 2017, 1639–1654; (i) I. Omae, Coord. Chem. 694 Rev., 2014, 280, 84–95; (j) M. Patra, G. Gasser and N. Metzler-Nolte, Dalton Trans., 2012, 41, 695 696 6350-6358.

697 8 Selected contributions on the utility of organometallic compounds in cell imaging, as bioprobes,
698 selective biosensors or as radiopharmaceuticals: (a) K. K.-W. Lo, Inorganic and Organometallic
699 Transition Metal complexes with Biological Molecules and Living Cells, Academic Press, 2016;

700		(b) F. L. Thorp-Greenwood, R. G. Balasingham and M. P. Coogan, J. Organomet. Chem., 2012,
701		714, 12–21; (c) A. Monney and M. Albrecht, Coord. Chem. Rev., 2013, 257, 2420–2433; (d) I.
702		S. Butler, R. P. Kegne-Momo, G. Jaouen, C. Policar and A. Vessières, Appl. Spectrosc. Rev.,
703		2012, 47, 531–549; (e) Z. Lam. K. V. Kong, M. Olivo and W. K. Leong, Analyst, 2016, 141.
704		1569–1586
701		
705	9	(a) P. Stepnicka, Ferrocenes: Ligands, Materials and Biomolecules, Wiley-VCH, Weinheim,
706		Germany, 2008; (b) E. S. Phillips, Ferrocenes: Compounds, Properties, and Applications, Nova
707		Science Publishers, Hauppauge, 2011.
708	10	(a) F. A. Larik, A. Saeed, T. A. Fattah, U. Muqadar and P. A. Channar, Appl. Organomet.
709		Chem., 2016, 1–22; (b) C. Ornelas, New J. Chem., 2011, 35, 1973–1985; (c) S. S. Braga and A.
710		M. S. Silva, Organometallics, 2013, 32, 5626–5639; (d) D. Astruc, Eur. J. Inorg. Chem., 2017,
711		6–29.
712	11	For relevant and recent articles on the utility of Re(I)-carbonyl complexes as sensors, in live cell
713		imaging, or enzyme inhibitors see for instance: (a) L. J. Raszeja, D. Siegmund, A. L. Cordes, J.
714		Guldenhaupt, K. Gerwert, S. Hahn and N. Metzler-Nolte, Chem. Commun., 2017, 905–908; (b)
715		A. Ramdass, V. Sathish, V. Murugesan, P. Thanasekaran, S. Umapathy and S. Rajagopal, RSC
716		Adv., 2015, 5, 38479–38488; (c) D. Can, B. Spingler, P. Schmutz, F. Mendez, P. Raposinho, C.
717		Fernandes, F. Carta, A. Innocenti, I. Santos, C. T. Supuran and R. Alberto, Angew. Chem., Int.
718		Ed., 2012, 51, 3354–3357; (d) A. Leonidova, C. Mari, C. Aebersold and G. Gasser,
719		Organometallics, 2016, 35, 851–854: (e) A. Leonidova and G. Gasser, ACS Chem. Biol., 2014.
720		9 2180_2193
720		<i>y</i> , 2100–21 <i>y</i> .
721	12	S. Clede, F. Lambert, C. Sandt, Z. Gueroui, N. Delsuc, P. Dumas, A. Vessières and C. Policar,
722		Biotechnol. Adv., 2013, 31, 393–395.
723	13	(a) L. Glans, W. Hu, C. Jost, C. de Kock, P. J. Smith, M. Haukka, H. Bruhn, U. Schatzschneider
724		and E. Nordlander, Dalton Trans., 2012, 41, 6443–6450; (b) R. Arancibia, F. Dubar, B.
725		Pradines, I. Forfar, D. Dive, A. H. Klahn and C. Biot, Med. Chem., 2010, 18, 8085-8091.
726	14	K. Kowalski, L. Szczupak, S. Saloman, D. Steverding, A. Jablonski, V. Vrcek, A. Hildebrandt,
727		H. Lang and A. Rybarczyk-Pirek, ChemPlusChem, 2017, 82, 303-314.
728	15	(a) F. Godoy, A. Gómez, N. Agurto, M. Muñoz, R. Segura, C. P. Silva, J. Pavez, J. H. Zagal, A.
729		H. Klahn, M. Fuentealba, A. Ibañez and M. T. Garland, J. Organomet. Chem., 2015, 788, 42-
730		48; (b) R. Arancibia, C. Biot, G. Delanay, P. Roussel, A. Pascual, B. Pradines and A. H. Klahn,

731		J. Organomet. Chem., 2013, 723, 143–148; (c) P. Toro, A. H. Klahn, B. Pradines, F. Lahoz, A.
732		Pascual, C. Biot and R. Arancibia, Inorg. Chem. Commun., 2013, 35, 126–129.
733	16	(a) R. Arancibia, A. H. Klahn, M. Lapier, J. D. Maya, A. Ibañez, M. T. Garland, S. Carrere-
734		Kremer, L. Kremer and C. Biot, J. Organomet. Chem., 2014, 755, 1-6; (b) R. Arancibia, C.
735		Quintana, C. Biot, M. E. Medina, S. Carrere-Kremer, L. Kremer and A. H. Klahn, Inorg. Chem.
736		Commun., 2015, 55, 139–142.
737	17	(a) R. Arancibia, F. Godoy, G. E. Buono-Cuore, A. H. Klahn, E. Gutierrez-Puebla and A.
738		Monge, Polyhedron, 2008, 27, 2421–2425; (b) R. Arancibia, A. H. Klahn, G. E. Buono-Core, E.
739		Gutierrez-Puebla, A. Monge, M. E. Medina, C. Olea-Azar, J. D. Maya and F. Godoy, J.
740		Organomet. Chem., 2011, 696, 3238–3244; (c) R. Arancibia, A. H. Klahn, G. E. Buono-Core, D.
741		Contreras, G. Barriga, C. Olea-Azar, M. Lapier, J. D. Maya, A. Ibañez and M. T. Garland, J.
742		Organomet. Chem., 2013, 743, 49-54; (d) C. Echeverria, V. Romero, R. Arancibia, A. H. Klahn,
743		I. Montorfano, R. Armisen, V. Borgna, F. Simon and R. Ramirez-Tagle, BioMetals, 2016, 29,
744		743–749.
745	18	Selected articles on the antitumoral activity of different sorts of imines (including
746		ferrocenylaldimines) and their Pd(II) or Pt(II) complexes on cancer cell lines: (a) J. Albert, R.
747		Bosque, M. Crespo, J. Granell, C. López, R. Cortés, A. González, J. Quirante, C. Calvis, R.
748		Messeguer, L. Baldomà, J. Badía and M. Cascante, Bioorg. Med. Chem., 2013, 21, 4210-4217;
749		(b) C. López, R. Bosque, M. Pujol, J. Simó, E. Sevilla, M. Font-Bardía, R. Messeguer and C.
750		Calvis, Inorganics, 2014, 2, 620–648.
751	19	(a) D. Talancón, C. López, M. Font-Bardía, T. Calvet, J. Quirante, C. Calvis, R. Messeguer, R.
752		Cortés, M. Cascante, L. Baldomà and J. Badia, J. Inorg. Biochem., 2013, 118, 1-12; (b) R.
753		Cortés, M. Tarrado-Castellarnau, D. Talancón, C. López, W. Link, D. Ruiz, J. J. Centelles, J.
754		Quirante and M. Cascante, Metallomics, 2014, 6, 622-633. 20 C. López, J. Sales, X. Solans
755		and R. Zquiak, J. Chem. Soc., Dalton Trans., 1992, 2321–2328.
756	21	R. Bosque, C. López, J. Sales, X. Solans and M. Font-Bardía, J. Chem. Soc., Dalton Trans.,
757		1994, 735–745.
758	22	R. Bosque, C. López, J. Sales and X. Solans, J. Organomet. Chem., 1994, 483, 61-71.
759	23	See for instance: (a) Comprehensive Organic Chemistry, ed. D. Barton and W. D. Ollis,
760		Pergamon, Oxford, UK, 1979; (b) A. M. Belostotskii, Conformational Concept for Synthetic
761		Chemist's Use: Principles in Lab Exploitation, World Scientific, 2015.
762	24	C. López, R. Bosque, X. Solans and M. Font-Bardia, New J. Chem., 1996, 20, 1285–1292.

763	25	(a) S. Pérez, C. López, A. Caubet, R. Bosque, X. Solans, M. Font-Bardía, A. Roig and E.
764		Molins, Organometallics, 2004, 23, 224–236; (b) C. López, R. Bosque, S. Pérez, A. Roig, E.
765		Molins, X. Solans and M. Font-Bardía, J. Organomet. Chem., 2006, 691, 475-484.
766	26	(a) Cambridge Crystallographic Data Centre (CCDC), [http://www.ccdc.cam.uk/data (accessed
767		on June 2017)]; (b) C. R. Groom, I. J. Bruno, M. P. Lightfoot and S. C. Ward, Acta Crystallogr.,
768		Sect. B: Struct. Sci., Cryst. Eng. Mater., 2016, 72, 171–179.
769	27	T. Cautivo, A. H. Klahn, F. Godoy, C. López, M. Font-Bardía, T. Calvet, E. Gutierrez-Puebla
770		and A. Monge, Organometallics, 2011, 30, 5578–5589.
771	28	(a) A. Bondi, J. Phys. Chem., 1964, 68, 441–451; (b) S. S. Batsanov, Inorg. Mater., 2001, 37,
772		871–885.
773	29	E. R. Brown and J. R. Sandifer, in Physical Methods in Chemistry. Electrochemical Methods,
774		ed. B. W. Rossiter and J. F. Hamilton, Wiley, New York, USA, 1986, ch. 4, vol. 4.
775	30	R. Bosque, C. López and J. Sales, Inorg. Chim. Acta, 1996, 244, 141–145.
776	31	J. J. R. Fraústo da Silva and R. J. P. Williams, The Biological Chemistry of Elements: The
777		Inorganic Chemistry of Life, Oxford University Press, Oxford, UK, 2nd edn, 2001.
778	32	J. Gómez, N. Leiva, R. Arancibia, J. Oyarzo, G. E. Buono-Cuore, A. H. Klahn, V. Artigas, M.
779		Fuentealba, R. Bosque, G. Aullón, C. López, M. Font-Bardía and T. Calvet, J. Organomet.
780		Chem., 2016, 819, 129–137.
781	33	A. J. Lees, Photophysic of Organometallics, Springer, Heildelberg, Germany, 2010.
782	34	Comprehensive Coordination Chemistry II: from Biology to Nanotechnology, ed. J. A.
783		Mc.Cleverty and T. J. Meyer, Elsevier, Amsterdam, The Netherlands, 2003.
784	35	B3LYP. (a) A. D. J. Becke, Chem. Phys., 1993, 98, 5648–5652; (b) C. Lee, W. Yang and R. G.
785		Parr, Phys. Rev. B: Condens. Matter Mater. Phys., 1988, 37, 785-789.
786	36	(a) W. R. Wadt and P. J. Hay, J. Chem. Phys., 1985, 82, 284–298; (b) P. J. Hay and W. R. Wadt,
787		J. Chem. Phys., 1985, 82, 299–310; (c) P. C. Hariharan and J. A. Pople, Theor. Chim. Acta,
788		1973, 28, 213–222; (d) M. M. Francl, W. J. Pietro, W. J. Hehre, J. S. Binkley, M. S. Gordon, D.
789		J. DeFrees and J. A. Pople, J. Chem. Phys., 1982, 77, 3654–3665.
790	37	M. J. Frisch, G. W. Trucks, H. B. Schlegel, G. E. Scuseria, M. A. Robb, J. R. Cheeseman, J. A.
791		Montgomery, T. Vreven, K. N. Kudin, J. C. Burant, J. M. Millam, S. S. Iyengar, J. Tomasi, V.

792		Barone, B. Mennucci, M. Cossi, G. Scalmani, N. Rega, G. A. Petersson, H. Nakatsuji, M. Hada,
793		M. Ehara, K. Toyota, R. Fukuda, J. Hasegawa, M. Ishida, T. Nakajima, Y. Honda, O. Kitao, H.
794		Nakai, M. Klene, X. Li, J. E. Knox, H. P. Hratchian, J. B. Cross, V. Bakken, C. Adamo, J.
795		Jaramillo, R. Gomperts, R. E. Stratmann, O. Yazyev, A. J. Austin, R. Cammi, C. Pomelli, J. W.
796		Ochterski, P. Y. Ayala, K. Morokuma, G. A. Voth, P. Salvador, J. J. Dannenberg, V. G.
797		Zakrzewski, S. Dapprich, A. D. Daniels, M. C. Strain, O. Farkas, D. K. Malick, A. D. Rabuck,
798		K. Raghavachari, J. B. Foresman, J. V. Ortiz, Q. Cui, A. G. Baboul, S. Clifford, J. Cioslowski,
799		B. B. Stefanov, G. Liu, A. Liashenko, P. Piskorz, I. Komaromi, R. L. Martin, D. J. Fox, T.
800		Keith, M. A. Al-Laham, C. Y. Peng, A. Nanayakkara, M. Challacombe, P. M. W. Gill, B.
801		Johnson, W. Chen, M. W. Wong, C. Gonzalez and J. A. Pople, Gaussian 03 (Revision C.02),
802		Gaussian, Inc., Wallingford, CT, 2004.
803 804	38	M. E. Casida, C. Jamorski, K. C. Casida and D. R. Salahub, J. Chem. Phys., 1998, 108, 4439–4449.
805 806	39	See for instance: (a) D. Theile, Molecules, 2017, 22, 382; (b) S. Dasari and P. B. Tchounwou, Eur. J. Pharmacol., 2014, 364–378.
807 808	40	(a) A. Tiwari, H. K. Patra and J. W. Choi, Advances Theranostic Materials, Wiley, 2015; (b) X. Chen and S. Wong, Cancer Theranostics, Accademic Press, San Diego, USA, 2015.
809 810	41	D. Chong, D. Laws, A. Nafady, P. Costa, A. Rheingold, M. Calhorda and W. Geiger, J. Am. Chem. Soc., 2008, 130, 2692–2703.
811 812	42	J. Heldt, N. Fischer-Durand, M. Salmain, A. Vessiéres and G. Jaouen, J. Organomet. Chem., 2004, 689, 4775–4782.
813	43	D. Van Leusen and B. Hessen, Organometallics, 2001, 20, 224–226.
814	44	D. D. Perrin and W. L. F. Armarego, Purification of Laboratory Chemicals, Butterworth-
815		Heinemann, Oxford, UK, 4th edn, 1996.
816	45	G. M. Sheldrick, Acta Crystallogr., Sect. A: Found. Adv., 2015, 71, 3-8.
817	46	M. Cossi, N. Rega, G. Scalmani and V. Barone, J. Comput. Chem., 2003, 24, 669-681.
818 819	47	K. T. Givens, S. Kitada, A. K. Chen, J. Rothschiller and D. A. Lee, Invest. Ophthalmol. Visual Sci., 1990, 31, 1856–1862.

821	Legends to figures
822	
823	Figure. 1 Representative examples of cyrhetrenes containing appended bioactive units with relevant
824	biological activities.
825	
826	Figure.2 Examples of ferrocenylimines (A and B) with cytotoxic activity and their Pt(II) complexes (C-
827	E) with greater inhibitory growth potency than their corresponding parent ligands. IC50 values for D
828	and E in A549 (lung), MDA-MB231 (breast) or HCT116 (colon) cancer cell lines ranged from 1.5 $\mu M$
829	to 10 µM.
830	
831	Chart 1 Chemical formulae of the novel hybrid ferrocenyl/cyrhetrenyl aldimines prepared in this work
832	and the atom labelling scheme.
833	
834	Figure.3 The molecular structure of $[(\eta 5-C5H5)Fe\{(\eta 5-C5H4)-CHvN-(\eta 5-C5H4)Re(CO)3\}]$ (1).
835	
836	Figure.4. The molecular structure of $[(\eta 5-C5H5)Fe\{(\eta 5-C5H4)-NvCH-(\eta 5-C5H4)Re(CO)3\}]$ (2).
837	
838	<b>Figure.5</b> Cyclic voltammograms of the new aldimines 1 and 2, in the ranges of potentials: $-1.20 \text{ V} \le \text{E}$
839	$\leq$ 0.50 V (A) and -1.00 V $\leq$ E $\leq$ 1.60 V (B), together with the labelling system used for the observed
840	peaks.
841	
842	Figure.6 UV-vis spectra of 10–4 M solutions of aldimines 1 and 2 in CH2Cl2 at 298 K.
843	
844	Figure.7 Emission spectra of 10–4 M solutions of the aldimines 1 and 2 in CH2Cl2 at 298 K upon
845	excitation at $\lambda$ exc. = 477 nm (A) or 383 nm (B).
846	
847	Figure.8 Frontier orbitals of isomers 1a (left) and 2b (right) together with their energies and the values
848	of the HOMO–LUMO gap
849	
850	Figure.9 A comparative plot of the IC50 values ( $\mu$ M) of 2 and cisplatin against the three cancer cell
851	lines used in this study: MCF7 and MDA-MB231 (breast) and HCT-116 (colon) cancer cell lines.
852	



855





Hydroxytamoxifen



Chloroquine



Nucleobases



FIGURE 2



# 860

861





Platinum(II) complexes















FIGURE 4

















- **Table** 1 Selected bond lengths (in Å), bond angles (in deg.), and angles between relevant planes (in
- 907 deg.) of aldimines:  $[(\eta 5-C5H5)Fe{(\eta 5-C5H4)-CHvN-(\eta 5-C5H4)}Re(CO)3](1)$  and  $[(\eta 5-C5H5)Fe{(\eta 5-C5H5)}Fe{(\eta 5-C5H5)}Fe{($
- 908 C5H4)-NvCH-(η5-C5H4)}Re(CO)3] (2). Standard deviations are given in parenthesis

	1	2
Bond lengths		1.0
NI-C11	1.276(6)	1,276(6)
N1-C12	1.406(7)	-
N1-C10		1.430(2)
C10-C11	1.443(7)	-
C11-C12	_	1.463(3)
Rei-Ci7	1.912(6)	1,9145(19)
Rei-C18	1.916(5)	1.9223(19)
Rei-C19	1.910(5)	1.9170(18)
O1-C17	1.153(7)	1.150(2)
02-C18	1.150(6)	1.150(2)
03-C19	1.156(6)	1.148(2)
Fe1-C <sup>4</sup>	2.045(8)	2.046(4)
Rei-C	2.310(19)	2.309(7)
Bond angles		
NI-CII-C	121.6(5)	120.76(17)
C-Re-C <sup>d</sup>	89.6(5)	89.91(3)
Angles between main ;	d anes"	
I and II	1.3	1.1
II and II	12.6	7.7
Handly	11.6	5.4
III and IV	11.2	3.10

910

- 912 Table 2 The summary of electrochemical data [anodic (Epa)i, cathodic (Epc)i potentials and the
- 913 separation between peaks  $[\Delta E = (Epa)I (Epc)I']$  for the new addimines R1-CHvN-R2 (in V); intensity
- 914 ratio (Ipa/Ipc). Data were obtained at a scan rate v = 250 mV s-1 and referenced to the
- 915 ferrocene/ferricinium couple (Fc/Fc+) (for the identification of the peaks, see Fig. 5)
- 916

	R1	R <sup>2</sup>	Elpa	Epe	ΔE	Ipa/Ipe	Evz	Epa	Ep.	Epe
1	Ferrocenyl	Cyrhetrenyl	0.208	0.135	0.073	1.02	0.172	1.297	0.294	-
2	Owbetrenvil	Fermenul	0.057	-0.008	0.065	1.04	0.025	1 420	0.260	0 306

- 919 Table 3 Absorption and emission spectroscopic data for aldimines R1-CHvN-R2 (1 and 2) in CH2Cl2 at
- 920 298 K. Wavelengths,  $\lambda i$  (in nm), logarithms of the extinction coefficients {loge, in parenthesis ( $\epsilon$  in M-1
- 921 cm-1)}, and excitation and emission wavelengths [ $\lambda$ exc. and  $\lambda$ em., respectively]
- 922

Aldimines R <sup>1</sup> -CH=N-R <sup>2</sup>		Absorption spectroscopic data				Emission spectro scopic data				
	R1	R <sup>2</sup>	À,	à <sub>2</sub>	à,	λ	Jean.	Jen.	-2.0	
1	Ferro cenyl	Cyrhetrenyl	473(3.2)	4	234(4.4)	220(4.1)	477	551	432	457
2	Cythetrenyl	Fermenyl	475(3.3)	383(3.4)	314(4.1)	230(4.3)	473	549 405	431	456

"Shoulder at \$380 nm (see text).

924

**Table 4** Relative free energies ( $\Delta G$ ) of isomers, 1b, 2a and 2b in relation to the value obtained for the

927 most stable isomer (1a) in vacuum and in CH2Cl2

928

	$\Delta G_{v}$ (in keal mol <sup>-1</sup> )					
	1b	2a	2b			
In vacuum	1.58	1.32	0.64			
In CH <sub>2</sub> Cl <sub>2</sub>	3.26	2.26	1.94			

929

- **Table 5** Cytotoxic activities of the new aldimines (1 and 2) and cisplatin (IC50 values in  $\mu$ M) against
- the MCF7 (breast), triple negative MDA-MB231 (breast) and HCT-116 (colon) cell lines together with
- the values obtained in the human skin fibroblast BJ cell line. For comparison purposes, values obtained
- 934 for cisplatin under identical experimental conditions are also given

	ICao values <sup>a</sup>					
	1	2	Cisp latin			
MCF7	>30	12 ± 5.2	12 ± 2.8			
MDA-MB231	>30	7.4±1.5	13 ± 1.8			
HCT-116	>30	7.8±3.3	$14 \pm 1.0$			
BJ	71 ± 2	21 ± 2	23 ± 2			

"Data are shown as the mean values of two experiments performed in triplicate with the corresponding standard deviations.

936

- **Table 6** Crystal data and details of the refinement of the crystal structures of the new aldimines: [( $\eta$ 5-
- $C5H5)Fe{(\eta 5-C5H4)-CHvN-(\eta 5-C5H4)}Re(CO)3](1) and [(\eta 5-C5H5)Fe{(\eta 5-C5H4)-NvCH-(\eta 5-C5$
- 940 C5H4)Re(CO)3] (2)

	1	2
Empirical formula	C19H14FeNO3Re	C19H16FeNORRe
Formula weight	546.36	546.36
TK	100(2)	100(2)
Crystal sizes/mm × mm	0.120 × 0.087 ×	0.208 × 0.091 ×
× mm	0.047	0.051
A/A	0.71073	0.71073
Crystal system	Orthorhombic	Triclinic
Space group	Phen	Pi
alà	20.2799(7)	7.7538(3)
b/Å	7.3753(3)	10.0032(3)
cià	22.2740(9)	11.4058 (4)
a <sup>p</sup>	90	99.296(2)
(A)P	90	104.9040(10)
7/9	90	98.3590(10)
V/Å*	3331.5(2)	827.32(5)
Z	8	2
Dene/mg mm <sup>-2</sup>	2.3179	2.193
µ/mm-*	8.149	8.203
F(000)	2080	520
28 mange for data	From 2.008 to	From 2,158 to
collected/°	28,303	30.570
Index ranges	$-26 \le h \le 27$	$-11 \le h \le 11$
	$-8 \le k \le 9$	$-14 \le k \le 14$
	$-25 \le l \le 29$	$-16 \le l \le 16$
N. of reflections	21 377	37 637
(collected)		
N. of independent	4125, Rins = 0.0594	5064, Rins = 0.0306
reflections, Ring.		
N. of data	4125	5064
N. of parameters	226	226
Goodness of fit on F <sup>2</sup>	1.027	1.112
Final R indices $[I > 2d(I)]$	$R_1 = 0.0336$ , $wR_2 =$	R1 = 0.0150, wR2 =
	0.0582	0.0311
Final R indices (all data)	$R_1 = 0.0587, WR_2 =$	R1 = 0.0179, WR2 =
	0.0601	0.0317
Largest diff. peak and	0.888 and -1.093	0.691 and -1.031