1	Characterization and fate of EfOM during ozonation applied for effective
2	abatement of recalcitrant micropollutants
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12	ABSTRACT
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14	Alterations occurring in the effluent organic matter (EfOM) during ozonation could be
15	detrimental depending on the final application of the treated effluent. In this work, the

fate of EfOM in different ozonized wastewaters was assessed through the monitoring of

general water quality parameters and organic fractions determined through size-exclusion

chromatography combined with organic carbon detection (SEC-OCD) analysis. These

different components of EfOM were distinguished based on relative molecular weights

and assigned to fractions named as biopolymers, humic substances, building blocks and

low molecular weight neutrals and acids. The significant abatement (60-90%) of an

ozone-refractory micropollutant (MP) was employed as reference to simulate potential

scenarios in which also the presence of these species is wanted to be attenuated.

Ultraviolet absorbance at 254 nm (UVA<sub>254</sub>) and chemical oxygen demand (COD)

reductions ranged from 40 to 80% and from 10 to 45%, respectively, for ozone doses

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26 between 0.6 and 1.0 mM, depending on the organic matter content (both dissolved and 27 suspended) and alkalinity of the effluents. Dissolved organic carbon (DOC) analysis 28 showed 21-27% reductions in Membrane bioreactor (MBR) effluents, whereas for 29 conventional activated sludge (CAS) samples this value increased (6-35%) during the 30 oxidative treatment. This was attributed to the continuous solubilization of humic 31 substances, according to SEC-OCD results. Moreover, accumulation of lower molecular 32 weight fractions such as building blocks or acids was observed in all the tested effluents, 33 and attributed to the breakdown of largest EfOM fractions, mainly humic substances. 34 Relationships proposed in this work between humic substances evolution, water quality 35 (UVA254) and process parameters (immediate ozone demand (IOD), IOD-normalized hydroxyl radical exposure ( $\int -OH dt/IOD$ ) and transferred ozone dose (TOD)) might be 36 37 useful for EfOM variations estimations along ozonation.

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## 39 KEYWORDS

40

41 Wastewater ozonation, micropollutants oxidation, EfOM fractions, SEC-OCD, humic
42 substances

43

## 44 **1. Introduction**

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In a time in which water scarcity increasingly constitutes one of the most serious threats for human and environmental safety, enhanced wastewater treatment and reclamation consolidates as the strategy to follow if sustainability regarding this vital resource is wanted to be preserved [1–3]. Advanced treatment of effluents released into freshwater bodies minimizes the negative impacts (*i.e.*, pollution of the receiving aqueous

51 compartments) derived from this practice. On the other hand, reclaimed wastewater can 52 be employed for a variety of non-potable uses, including agricultural and municipal 53 irrigation, environmental applications, recreational activities or industrial processes [3,4]. 54 By means of these practices, significant volumes of freshwater are saved. However, the 55 use of this alternative water source in applications implying further human or animal 56 exposure is limited by the presence – among others– of organic micropollutants (MPs) 57 which, although in general not regulated, pose potential risks for living species [1]. 58 Considering this particular issue, ozonation and activated carbon are nowadays one of the 59 most recognized advanced treatment technologies for enhanced wastewater treatment and 60 reclamation purposes [5–11].

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62 So far, most studies dealing with ozonation of wastewater effluents mainly focus on the 63 fate of ozone-sensitive micropollutants and harmful oxidation byproducts - such as 64 bromate [12–14] – under such operational conditions (*i.e.*, low ozone doses). Higher 65 oxidant doses such as those required for ozone-resistant MPs abatement are in general not 66 considered in full-scale ozonation steps, although several organic compounds typically 67 present in wastewater effluents are recalcitrant to ozone and present toxic properties 68 which should encourage their effective abatement. On their part, changes in effluent 69 organic matter (EfOM) are traditionally set aside or studied, at most, through the variation 70 of general related parameters such as total and dissolved organic carbon (TOC and DOC, 71 respectively), chemical oxygen demand (COD) or ultraviolet absorbance at 254 nm 72 (UVA<sub>254</sub>) [15–20]. There is still, however, a lack of knowledge regarding the fate of the 73 different organic fractions during municipal wastewater ozonation. Deeper information 74 can be obtained by means of size-exclusion chromatography in combination with organic 75 carbon detection, SEC-OCD [21], which is able to separate and quantify different EfOM fractions according to their relative molecular weights and additional measurements such
as UVA<sub>254</sub> or organic nitrogen content.

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79 There are several publications in literature describing the impact of wastewater effluent 80 ozonation on EfOM fractions [22-31]. However, in some of these works the employed 81 ozone doses were either not properly quantified [22,23] or too high to be considered for 82 practical applications [24]. Moreover, changes in EfOM fractions are often available in a 83 very qualitatively way [25–27], or expressed as variations in the average molecular size 84 but not quantified in terms of organic carbon concentrations [28,29]. In other few works, 85 changes in EfOM fractions were not related to or discussed together with micropollutants 86 oxidation or the reduction of any other general parameters typically monitored in 87 wastewater treatment [30,31]. None of these studies discuss the fate of EfOM for various 88 wastewater sources presenting different water qualities.

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90 EfOM contained in reclaimed wastewater treated by ozonation can cause some technical 91 and environmental problems if further treatments are planned next. For instance, residual 92 organic matter (OM) could contribute to the formation of harmful disinfection by-93 products in a subsequent disinfection step, or be partly responsible for membrane fouling 94 in a filtration unit [32,33]. Also, EfOM can compete with target pollutants for adsorption 95 sites in activated carbon systems or avoid the separation of these chemicals from the water 96 matrix if these are bound to the organic matter surface [32,33]. Moreover, since prior to 97 be used this water need to be redirected to its final destination, EfOM - especially its 98 lower molecular weight fractions – can be a perfect substrate for bacterial growth in 99 distribution systems, or cause eutrophication in receiving water compartments if the final 100 use is related to surface or groundwater recharge [32].

102 This work aimed to contribute to the - still - scarce knowledge regarding the variation of 103 EfOM fractions during ozonation of wastewater effluents. Concretely, the objective of 104 the study was to assess changes taking place when ozone doses required for the effective 105 abatement of micropollutants are applied, including the elimination of ozone-resistant 106 species. Thus, a potentially realistic scenario was explored in this study. To do so, 107 ozonation experiments in semi-continuous mode where performed for six different 108 wastewater effluents presenting a wide range of water qualities and spiked with an ozone-109 resistant organic micropollutant as internal reference. The evolution of distinguishable 110 dissolved EfOM fractions was then followed by means of the SEC-OCD technique, 111 together with variations observed for other parameters typically discussed in practice, 112 such as UVA<sub>254</sub>, COD and DOC. Finally, some relationships between EfOM changes and 113 some effluent quality and ozonation parameters were discussed.

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## 115 **2. Materials and methods**

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# 117 2.1. Wastewater effluents

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Six different wastewater effluents coming from five wastewater treatment plants (WWTPs) in the province of Barcelona (Spain) were employed in this work. All of them treat municipal wastewater. Technical details of WWTPs from which they were collected are gathered in Table 1. All effluents samples were filtered through 25  $\mu$ m filter paper to remove coarse particles not belonging to the effluent (*e.g.*, dragged from the sample collection system) and avoid technical problems with the equipment used during ozonation experiments (*e.g.*, ozone sensor). All the effluent samples were kept at 4 °C
until they were used.

127

128 Table 1. List of wastewater effluent samples used in this work and technical details of WWTPs from which

129 they were collected. (PC: primary clarification; MBR: membrane bioreactor; UF: ultrafiltration membrane;

130 NR: nutrient removal; CAS: conventional activated sludge; IFAS: Integrated fixed-film activated sludge;

131 SC: secondary clarification).

Sample ID	WWTP location	Population equivalent [PE]	Design flow [m <sup>3</sup> d <sup>-1</sup> ]	Type of treatment
M1	Vacarisses	5280	1320	PC + MBR (UF) with NR
M2	Vallvidrera	5500	1100	PC + MBR (UF) with NR
M3	Gavà	192000	32000	PC + MBR (UF) with NR
C1	El Prat	2275000	420000	PC + CAS with NR + SC
C2	Gavà	192000	32000	PC + IFAS with NR + SC
C3	La Llagosta	358333	43000	PC + CAS + SC

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#### 133 2.2. Ozonation experiments

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Wastewater ozonation experiments were performed in a 750 mL semi-batch reactor, at a temperature of 20 °C and without pH adjustment. Ozone was generated by using a 301.19 lab ozonizer (Sander, Germany) and introduced into the reactor through a porous diffuser. A mechanical stirrer was used to provide a proper contact between liquid and gas phases. The gas flow rate and the ozone inlet concentration were maintained at 0.1 L min<sup>-1</sup> and 0.63 mmol L<sup>-1</sup>, respectively. The transferred ozone dose (TOD), which represents the

ozone consumption, was determined through continuous evaluation of the O<sub>3</sub> mass
balance in the gas phase [34]. Inlet and outlet ozone concentrations were measured by
two BMT 964 ozone analyzers (BMT Messtechnik, Germany). The residual
concentration of ozone in the aqueous phase was monitored by means of a Q45H/64
dissolved O<sub>3</sub> sensor (Analytical Technology, USA).

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Each wastewater effluent was spiked with 0.45 μM of the pesticide acetamiprid (ACMP), here employed as reference micropollutant because of its resistance to molecular ozone attack during ozonation, and then ozonized for 30 min under the mentioned operational conditions. Samples were withdrawn at known time intervals and kept at room conditions until complete consumption of dissolved ozone was achieved. Then, analyses for ACMP, typical physicochemical parameters (namely UVA<sub>254</sub>, DOC and COD) and EfOM fractions were conducted.

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Immediate ozone demand (IOD) of each wastewater sample was estimated in this work 155 156 as the minimum TOD required to detect dissolved ozone in the reaction medium [38]. 157 The initial hydroxyl radical (•OH) consumption rates, which provide information about 158 the amount of •OH available in the reaction medium for micropollutant oxidation [39], 159 were here estimated considering the sum of contributions by dissolved EfOM and 160 alkalinity. For EfOM contribution, a mean rate constant value between dissolved EfOM and •OH of  $2.1 \cdot 10^5$  (mg C L)<sup>-1</sup>s<sup>-1</sup> was used according to the work by Lee et al. [10]. 161 162 together with the DOC values determined for each effluent sample (see Table 3). 163 Consumption rate due to alkalinity was calculated according to the known rate constant of bicarbonate ion reaction with •OH ( $8.5 \cdot 10^6 \text{ M}^{-1}\text{s}^{-1}$  [40]) and the IC values determined 164

- 165 for each effluent sample (see Table 3). Thus, calculations were performed according to 166 the expression  $\bullet OH_{scavenging rate} = 2.1 \cdot 10^5 \cdot DOC + 8.5 \cdot 10^6 \cdot IC$ .
- 167

## 168 2.3. Analytical methods

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170 ACMP concentration was determined through HPLC-UV. Samples were previously 171 filtered through 0.45 µm polytetrafluoroethylene (PTFE) filters. The mobile phase 172 consisted of 70:30 volumetric mixtures of acetonitrile and ultrapure water at pH 3 173 (achieved by addition of H<sub>3</sub>PO<sub>4</sub>). Separation was achieved by means of a 25x0.46 cm (5 174 µm size packing) Sea18 column (Teknokroma, Spain). The flow rate and injection 175 volume were set at 1 mL min<sup>-1</sup> and 100  $\mu$ L, respectively. Detection was performed at 254 176 nm. UV absorbance at 254 nm was measured by means of a DR6000 UV Vis 177 spectrophotometer (Hach, USA) employing a quartz cuvette (path length: 1 cm). TOC, 178 DOC and inorganic carbon (IC) were measured by means of a Shimadzu TOC-VCSN 179 analyzer. COD was determined according to Standard Methods procedure 5220D [36]. 180 Nitrite  $(NO_2)$  concentration was determined by means of ion-exchange chromatography 181 with UV detection. For DOC, IC, UVA<sub>254</sub> and nitrite analyses, samples were previously 182 filtered through 0.45 µm PTFE filters. In order to characterize EfOM fractions after 183 applying particular ozone doses, the SEC-OCD technique from DOC-Labor was 184 employed. Detailed information regarding this technique can be found elsewhere [21,37]. 185 In summary, samples were previously filtered through 0.45 µm PTFE filters. The mobile 186 phase was a phosphate buffer (0.01 M, pH 7) containing 0.1 M of NaCl. Separation of 187 EfOM fractions by molecular weight was achieved by using a 25x2.2 cm column filled 188 with HW-50 resin purchased from Sigma-Aldrich (Germany). Operation flow-rate and injection volume were 1 mL min<sup>-1</sup> and 1 mL, respectively. As mentioned, the SEC-OCD 189

190 procedure consists of size exclusion chromatography (SEC) followed by organic carbon 191 detection. For calibration of humic substances molecular weights, Suwannee River 192 standard humic and fulvic acids were employed. Both were acquired from the 193 International Humic Substances Society. Based on the good agreement between the 194 chromatographic behavior of these standards and that of effluent samples, as well as to 195 additional UVA $_{254}$  measurements, the name humic substances (HS) was assigned to one 196 of the observed EfOM fractions. The nominal average molecular weights of fulvic and 197 humic acid standards (711 and 1066, respectively) could be used to determine the 198 molecular weight distribution of HS. The fraction with the highest molecular size (thus, 199 eluting first and before HS) received the name biopolymers (BP). The fraction observed 200 after HS elution was named as building blocks (BB), and was related to products coming 201 from the breakdown of humic substances. The names low molecular weight acids and 202 neutrals (LMWA and LMWN, respectively) were assigned to those fractions eluting last. 203 The slightly more hydrophobic character of LMWN compared to LMWA allowed 204 distinction between both fractions. A summary of the different fraction assignments can 205 be found in Table 2. It has to be noted that fraction assignments in this work were made 206 using the same fraction names firstly proposed by Huber and coworkers for 207 characterization of organic matter in natural waters [21,37], and then adopted by other 208 researchers for description of other types of water samples, including wastewater 209 effluents [39].

210

211 Table 2. Properties of dissolved EfOM fractions distinguished in SEC-OCD analyses.

Dissolved EfOM	Molecular	
fraction assignment	weight	Description

			Associated to amino s	ugars, polypeptides and	
	Biopolymers (BP)	> 20000	proteins. Do not a	bsorb UV radiation.	
	Humic substances		Mix of hydrophobic	humic substances and	
	(HS)	~1000	humic/fulvic acids, in	varying concentrations	
	Building blocks (BB)	300 - 450	Degradation intermedia	ates of humic substances	
	Low molecular weight		Short chain, non-acidi	c degradation products:	
	neutrals (LMWN)	< 300	alcohols, aldeh	ydes, ketones	
	Low molecular weight				
	acids (LMWA)	< 300	Final degradation	products of organics	
212					
213	3. Results and discussion	on			
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215	3.1. Effluent characteri	stics			
216					
217	The main physicochemic	al characteristi	cs of all effluent samples	are gathered in Table 3.	
218	Differences observed in t	he main quality	y parameters of tested eff	luents were attributed to	
219	particular WWTPs techn	ologies and op	perational conditions, es	pecially those regarding	
220	biological processes and	subsequent se	paration steps. They co	vered a relatively broad	
221	range of effluent qualities	s and therefore	represented the diverse r	ange of wastewaters that	
222	currently can be found in	practice.			
223					
224	Table 3. Effluent quality par	ameters. M and	C stands for MBR and CAS	effluents, respectively. All	
225	measurements were performed	l per triplicate (n=	3). Discrepancies between rep	licates were in all cases lower	
226	than 5%.				
Sample	WWTP UVA <sub>254</sub>	* TOC	DOC * COD	Turbidity IC *	NO2 <sup>-*</sup>
ID	pH location [m <sup>-1</sup> ]	[mg C L <sup>.1</sup> ]	$[mg C L^{-1}]$ $[mg O_2 L^{-1}]$	[NTU] [mg C L <sup>-1</sup> ]	[mg N L <sup>-1</sup> ]

M1	Vacarisses	7.9	17.9	6.7	6.6	14.9	0.6		87.4	0.9
M2	Vallvidrera	7.4	16.3	10.5	10.3	20.7	0.9		42.7	< 0.1
M3	Gavà	7.7	14.9	12.4	12.1	16.6	0.3		53.0	0.1
C1	El Prat	7.5	22.9	14.0	13.6	29.7	1.1		63.6	0.1
C2	Gavà	7.8	67.2	42.1	27.6	93.5	28.6	5	114.7	0.1
C3	La Llagosta	7.7	66.4	25.6	21.3	53.0	20.1		98.3	< 0.1
	227 * Samples v	vere filter	ed through (	).45 µm PTFI	E filters for the m	easurement o	f these para	meters.		
	228		-	·			-			
	220									
	229 <i>3.2. Ozo</i>	one demo	and, radice	al scavengi	ng and remove	l of the refe	erence col	mpound A	ACMP	
	230									
	231 Estimated	values	of IOD an	d •OH cons	sumption rates	are present	ed in Tak	Jo 4		
					sumption rates	are presen		nc 4.		
	232				sumption faces	are presen		<i>n</i> c 4.		
		stimated			on rates of tested	-			tions in	
	233 Table 4. E		IOD and •O	H consumpti	-	l effluent san	nples. EfON	A contribu		
	233Table 4. E234samples C2		IOD and •O	H consumpti	on rates of tested	l effluent san	nples. EfON	A contribu		
	233 Table 4. E 234 samples C2 Efflu	and C3 n	IOD and •O nay have bee <b>ple ID</b>	H consumpti en underestim M1	on rates of tested	l effluent san olved EfOM C1	nples. EfOM was taken i	Л contribu nto accour		
	233 Table 4. E 234 samples C2 Efflu IOD	and C3 n ent sam [mmol (	IOD and •O nay have bee <b>ple ID</b> D <sub>3</sub> L <sup>-1</sup> ]	H consumpti en underestim M1 0.29	on rates of tested nated, as only diss M2 M3	l effluent san olved EfOM C1 0.3	nples. EfOM was taken i C2 0.6	A contribu nto accour C3 0.4		
	233 Table 4. E 234 samples C2 Efflu IOD	and C3 n ent sam [mmol (	IOD and •O nay have bee <b>ple ID</b> D <sub>3</sub> L <sup>-1</sup> ]	H consumpti en underestim M1 0.29	on rates of tested nated, as only diss M2 M3 0.19 0.23	l effluent san olved EfOM C1 0.3	nples. EfOM was taken i C2 0.6	A contribu nto accour C3 0.4		
	233 Table 4. E 234 samples C2 Efflu IOD •OH cons 235	and C3 n ent sam [mmol ( sumptio	IOD and •O nay have bee <b>ple ID</b> D <sub>3</sub> L <sup>-1</sup> ] n rate [s <sup>-1</sup> ]	H consumpti en underestim M1 0.29 2.0·10 <sup>5</sup>	on rates of tested nated, as only diss M2 M3 0.19 0.23	l effluent san olved EfOM C1 0.3 0.3 0.3 $0.3 \cdot 10^5$	pples. EfON was taken i C2 0.6 $6.6 \cdot 10^5$	A contribution nto accourting $C3$ 0.4 $5.2 \cdot 10^5$	nt.	
	233 Table 4. E 234 samples C2 Efflu IOD •OH cons 235 236 Although	and C3 n ent sam [mmol ( sumptio	IOD and •O nay have bee ple ID O <sub>3</sub> L <sup>-1</sup> ] n rate [s <sup>-1</sup> ] ely low O	H consumption on underestimm M1 0.29 $2.0 \cdot 10^5$ 3 doses ( <i>e.</i>	on rates of tested hated, as only diss M2 M3 0.19 0.23 $2.5 \cdot 10^5 2.9 \cdot 10^{-5}$	l effluent san olved EfOM C1 0.3 0.3 $0.3 \cdot 10^5$ nol L <sup>-1</sup> or	nples. EfON was taken i C2 0.6 6.6·10 <sup>5</sup> 5-15 mg	A contribution nto accourted $C3$ 0.4 5.2 · 10 <sup>5</sup> $L^{-1}$ ) sho	nt. uld be	
	233 Table 4. E 234 samples C2 Efflu IOD •OH cons 235 236 Although 237 enough to	and C3 n ent sam [mmol ( sumptio relative o meet t	IOD and •O nay have bee <b>ple ID</b> D <sub>3</sub> L <sup>-1</sup> ] n rate [s <sup>-1</sup> ] ely low O he remova	H consumption en underestim M1 0.29 $2.0 \cdot 10^5$ 3 doses ( <i>e.</i> , al of organi	on rates of tester nated, as only diss M2 M3 0.19 0.23 $2.5 \cdot 10^5 2.9 \cdot 10^{-5}$ g., 0.1-0.3 mr	l effluent san olved EfOM C1 0.3 $0^5$ $3.3 \cdot 10^5$ nol L <sup>-1</sup> or with medi	nples. EfON was taken i C2 0.6 6.6·10 <sup>5</sup> 5-15 mg um or hig	A contribution nto accourting $C3$ 0.4 $5.2 \cdot 10^5$ $L^{-1}$ ) shough reactive	<sup>nt.</sup> uld be vity to	
	233 Table 4. E 234 samples C2 Efflu IOD •OH cons 235 236 Although 237 enough to 238 ozone [10	and C3 n ent sam [mmol ( sumptio relative o meet t 0,11,14,4	IOD and •O nay have bee <b>ple ID</b> O <sub>3</sub> L <sup>-1</sup> ] n rate [s <sup>-1</sup> ] ely low O he remova 41], this cr	H consumption on underestime M1 0.29 $2.0 \cdot 10^5$ 3 doses ( <i>e.</i> , al of organi- iteria could	on rates of tested nated, as only diss M2 M3 0.19 0.23 2.5 $\cdot 10^5$ 2.9 $\cdot 10^{-100}$ g., 0.1 $\cdot 0.3$ mm ic compounds	l effluent san olved EfOM C1 0.3	nples. EfON was taken i C2 0.6 6.6·10 <sup>5</sup> 5-15 mg um or hig as likely to	A contribution nto accourt C3 0.4 $5.2 \cdot 10^5$ L <sup>-1</sup> ) shough reactive po occur interpret of the second sec	uld be vity to n view	

basis of this hypothesis. ACMP reacts very slowly with ozone [35], and therefore only

required in a near future. In this study, we selected ACMP as reference compound on the

hydroxyl radicals generated through O<sub>3</sub> decay contribute to its degradation during
ozonation. If ozone-resistant MPs such as this pesticide are significantly removed from
wastewater effluents, a higher abatement of any other compound with higher sensitivity
towards ozone attack would be guaranteed. Furthermore, O<sub>3</sub> doses necessary for
disinfection purposes are generally lower than that for micropollutant oxidation [9,42].

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249 Measurements of the reference compound ACMP ([ACMP]<sub>0</sub>: 0.45 µM) at regular 250 reaction times allowed the obtaining of the degradation profiles presented in Fig. 1. Ozone 251 doses required to reach 80% abatement of ACMP differed significantly between samples. These were approximately between 0.4 and 0.8 mM (~19-38 mg  $L^{-1}$ ) for effluents M1-252 M3 and C1, whereas for effluents C2 and C3 such doses were about 1 mM (48 mg  $L^{-1}$ ) or 253 even higher. For an abatement level of 50%, doses between 0.3 and 0.5 mM (14-24 mg 254 255  $L^{-1}$ ) would instead be required for the less polluted effluents and between 0.6 and 0.8 Mm 256  $(29-38 \text{ mg L}^{-1})$  for samples C2 and C3.

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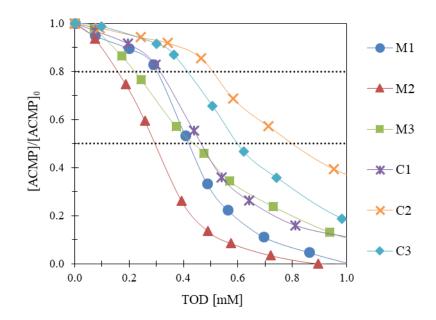


Figure 1. Abatement of the reference compound ACMP during semi-batch ozonation experiments withdifferent wastewater effluent samples.

The feasibility of ozonation steps implementation for MPs abatement (including ozoneresistant compounds) should be individually assessed for each wastewater source by means of more comprehensive procedures [43], especially for those effluents presenting a higher  $O_3$  demand and •OH scavenging rate (*e.g.*, C2 and C3). In any case, in this study we considered maximum ozone doses of about 1 mM as potentially practical for ozone applications to enhanced wastewater treatment and reclamation goals, and explored the fate of EfOM when subjected to these treatment conditions.

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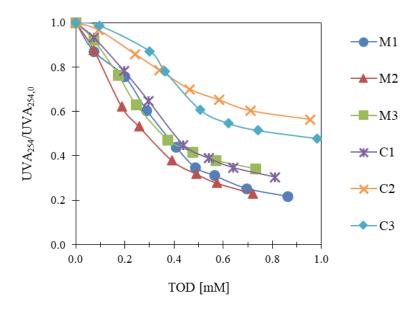
270 3.3. Changes of general EfOM descriptors (UVA254, COD and DOC) during ozonation

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272 Changes in UV absorbance at 254 nm (UVA254) during ozonation experiments are shown 273 in Fig. 2. Ozone typically reacts with electron-rich moieties of EfOM [39], resulting this 274 in a sustained decrease of the wastewater UV absorption along the process. Differences observed between effluents regarding UVA254 decrease can be related to their respective 275 276 water characteristics, which led to different availabilities of oxidants in the reaction 277 medium (see estimated IOD and •OH consumption rate values in Table 4). Thus, for C2 278 and C3 effluents, both containing important amounts of  $O_3$ -consuming [20] particulate 279 and colloidal matter (see the difference between TOC and DOC values as well as 280 turbidity, Table 3) and alkalinity (IC), the UVA<sub>254</sub> decrease at 30 min (TOD: 0.9-1.0 mM) 281 ranged between 40 and 50%, whereas for MBR effluents (M1-M3) this value was 282 determined to be considerably higher (70-80%) for a lower ozone consumption. In the 283 particular case of C1, its lower content in solid and colloidal matter (difference between TOC and DOC values: 0.4 mg C L<sup>-1</sup>; turbidity: 1.1 NTU) compared with C2 and C3 284 285 allowed a larger aromaticity reduction, more similar to that accounted for membrane

bioreactor effluents. Other organic matter descriptors such as COD, DOC, TOC (addressed in the following paragraphs) and turbidity varied along the oxidative treatment displaying a similar dependence with effluent characteristics. For instance, turbidity measurements at the end of treatment (*i.e.*, after 30 min of ozonation, which corresponded to the highest applied ozone dose), reductions of 70-80% for effluents M1-M3, 60% for C1 and about 30-40% in the case of samples C2 and C3 were registered.

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Figure 2. UV absorbance (at 254 nm) evolution during ozonation of wastewater effluents.

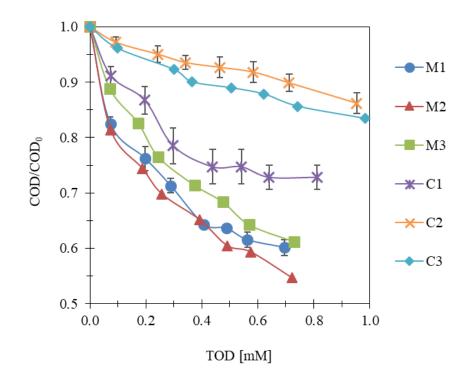
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296 Fig. 3 shows the COD/COD<sub>0</sub> profiles obtained for each water source, as a function of the 297 TOD. COD removals at the end of the treatment (TOD: 0.6-1.0 mM) ranged from 10 to 298 45%, depending on the tested sample. Again, clear differences were observed between 299 effluents, together with a consistent trend for this bulk parameter with respect to the 300 UVA254 evolution, previously described. According to this, for instance, the M2 effluent 301 - with relatively low content in organic matter and the lowest alkalinity among the studied 302 wastewaters – was the one for which the largest COD removal was observed, whereas C2 303 and C3 samples (highest content in both organic and inorganic carbon as well as in

304 colloidal and particulate matter) exhibited the lowest – and also the slowest – reduction for

# this parameter.

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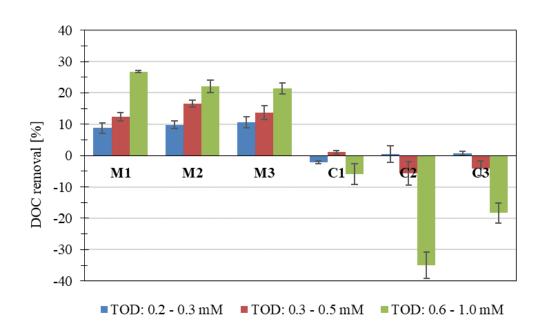
Figure 3. COD removal versus transferred ozone dose during semi-batch effluent ozonation experiments.

310 In general, lower overall reductions of COD (max. 45%) in comparison to  $UVA_{254}$  (up to 311 80%) were observed. It seems that the provided oxidation conditions were not strong 312 enough in order to achieve comparable levels of organic matter transformation in terms 313 aromaticity depletion and COD reduction.

Fig. 4 shows the percentage of DOC removal during effluent ozonation experiments, determined at three different ozone doses (TOD values of 0.2-0.3, 0.3-0.5 and 0.6-1.0 mM). For effluents M1, M2 and M3, all of them coming from MBR systems, the DOC concentrations after 30 min of treatment (TOD: 0.6-0.7 mM) were reduced by 27%, 22% and 21%, respectively. TOC measurements of the same samples (data not shown)

320 revealed almost identical removal values, which is logical given the fact that nearly all of 321 the organic matter was in dissolved form. In any case, organic carbon removals were in 322 comparison lower than those reached for COD or UVA<sub>254</sub> at the same consumed ozone 323 doses. In wastewater ozonation, the degree of mineralization is typically low, and other 324 water quality parameters related to the organic content – such as COD and UVA<sub>254</sub> – are 325 in general more affected during the process. This is reasonable, as DOC concentration 326 only is reduced when decarboxylation reactions are produced, that is, when the maximum 327 possible level of organic matter oxidation takes place [39].

328



329

Figure 4. DOC concentration removal in wastewater ozonation experiments at various transferred ozonedoses (TOD).

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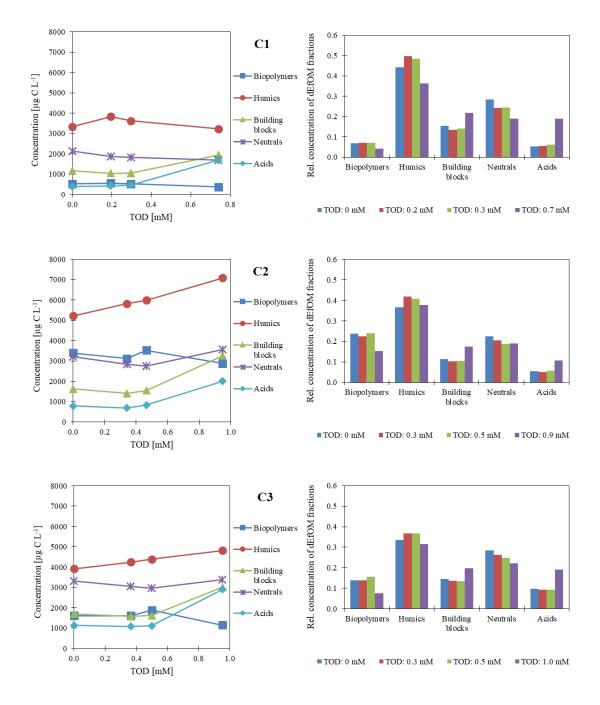
Concerning effluents C1, C2 and C3, the observed DOC removals at the end of ozonation experiments were negative in all cases. Water effluents coming from CAS systems usually contain residual amounts of suspended solids and colloids (see TOC, DOC and turbidity values of Table 3). Therefore, and in agreement with literature [44,45], it is possible that part of this non-dissolved material could be solubilized upon oxidation. For 338 the CAS effluents tested in the present study, this phenomenon was observed after 339 consumption of relatively high ozone doses (*i.e.*, 0.7-1.0 mM), rather than at low TOD 340 values. In fact, at O<sub>3</sub> doses between 0.2 and 0.5 mM only small changes in the DOC 341 concentration where registered, which could be indicative of a simultaneous oxidation-342 solubilization of, respectively, the dissolved and undissolved EfOM. An indicative of this 343 competition between degradation and solubilization may be the TOC removals measured 344 at the end of treatment, which were insignificant for samples C2 and C3 and only about 345 10% in the case of C1.

346

347 3.4. Evolution of EfOM fractions

348

349 Figs. 5 and 6 show the evolution of (dissolved) EfOM fractions and their relative 350 contribution to DOC during ozonation of wastewater effluents. At this point it has to be 351 recalled that EfOM fractions referred to in the following lines are due to fraction 352 assignments based on the chromatographic behavior of organic matter components in 353 wastewater effluent samples, according to the methods described for SEC-OCD analyses 354 in section 2.3. Samples coming from CAS processes presented larger percentages of 355 biopolymers than MBR effluents (7-25% vs 1-2%, respectively), which was already 356 expected given that ultrafiltration membranes employed in MBR units are able to retain 357 extracellular polymeric substances (EPSs). Differences were also observed in the humic substances concentration (2697-3319 and 3335-5217 µg C L<sup>-1</sup> for MBR and CAS, 358 359 respectively) which, in agreement with previous related studies concerning membrane 360 fouling in MBR units, suggests that a part of the largest constituents of this EfOM fraction 361 could also be retained during the separation process [24,46].



363

Figure 5. Evolution of dissolved EfOM fractions (left) and contribution to DOC (right) for ozonized CAS
effluents. Relative concentration of EfOM fractions in right column plots (y-axis) refer to DOC of
considered fraction divided by total DOC.

Regarding CAS effluents (Fig. 5), biopolymers were not significantly removed until higher doses of ozone were consumed [from 7 to 4% (C1), from 24 to 15% (C2) and from 14 to 8% (C3) of dissolved EfOM content for O<sub>3</sub> doses of 0.7-1.0 mM]. Especially

371 remarkable is the case of humic substances, which noticeably increased from the 372 beginning of the treatment, especially for C2 and C3 samples. The observed accumulation 373 of humic substances during the entire C2 and C3 ozonation time, together with the fact 374 that these two waters contain the highest fraction of suspended solids among all the 375 studied effluents may be linked. Thus, the non-soluble fraction of humic substances could 376 be solubilized when oxidized [24,37]. This affirmation would be experimentally 377 supported by the DOC measurements made at different ozone doses and early seen in this 378 study, which revealed significant increases in this parameter (35% and 18% for C2 and 379 C3 samples, respectively) for consumed ozone doses of 0.9-1.0 mM. Significantly higher 380 doses applied in related works for effluents containing suspended EfOM [24] could have 381 hinder a possible initial increase of this fraction followed by subsequent depletion for 382 higher oxidant dosages. This was in fact observed for sample C1, a CAS effluent 383 containing a lesser amount of suspended solids. Moreover, the continuous solubilization 384 of humic substances would provide an additional explanation to the low rates of UVA<sub>254</sub> 385 depletion found for C2 and C3 waters. BB remained almost unaltered until higher ozone 386 doses were applied, which means that humic substances were not being destroyed at those 387 oxidation extents. Only at O<sub>3</sub> doses of 0.7-1.0 mM, an enrichment in the BB contents 388 [from 15 to 22% (C1), from 11 to 17% (C2) and from 15 to 20% (C3)] was noticeable in 389 all EfOMs. Regarding LMWN, the concentration of this fraction increased at the end of 390 the treatment (i.e., for ozone doses of 0.7-1.0 mM) for samples C2 and C3, but only after 391 an initial – and also slight – reduction at the first stages of the process took place. On the 392 contrary, for effluent C1 a slightly decrease in this fraction concentration was observed 393 during the entire ozonation time. Again, differences between effluents C2 and C3, on one 394 hand, and effluent C1, on the other, appear to be well explained by the solubilization 395 process taking place in the first ones: the continuous introduction of humic substances to

396 the system would hypothetically lead to an accumulation of degradation intermediates 397 (*i.e.*, LMWN), contrarily to what typically happens in water matrices in which this re-398 dissolution of OM does not significantly take place (e.g., MBR effluents and also sample 399 C1). Despite the increase in LMWN observed for C2 and C3, a gradual impoverishment 400 and accumulation of this fraction in all the tested CAS effluents was registered [overall 401 changes in EfOM composition for consumed  $O_3$  doses of 0.7-1.0 mM: from 28 to 19% 402 (C1), from 23 to 19% (C2), from 28 to 22% (C3)]. Finally, a significant accumulation of 403 LMWA was observed for all three samples at ozone doses of 0.7-1.0 mM, leading to the 404 enrichment in this component of the corresponding EfOM compositions [from 5 to 19% 405 (C1), from 6 to 11% (C2) and from 10 to 19% (C3)]. This evolution was predictable, as 406 carboxylic acids present low reactivity towards ozone and the contribution of hydroxyl 407 radical oxidation in complex water matrices is usually expected to be low [39,47].

408

409 The evolution of EfOM fractions during ozonation of MBR effluents is shown in Fig. 6. 410 With no biopolymers nor suspended solids present in wastewater matrices, ozone 411 primarily attacked humic substances and LMWN, leading this to the gradual 412 accumulation of BB but particularly LMWA (from 4 to 18% (M1), from 3 to 20% (M2) 413 and from 5 to 8% (M3), for consumed ozone doses of 0.6-0.7 mM). The overall increase 414 of humic substances concentration observed for the M3 dissolved EfOM was the result 415 of the small changes that took place in the rest of fractions, together with a DOC reduction 416 of ca. 21% at the end of the treatment.

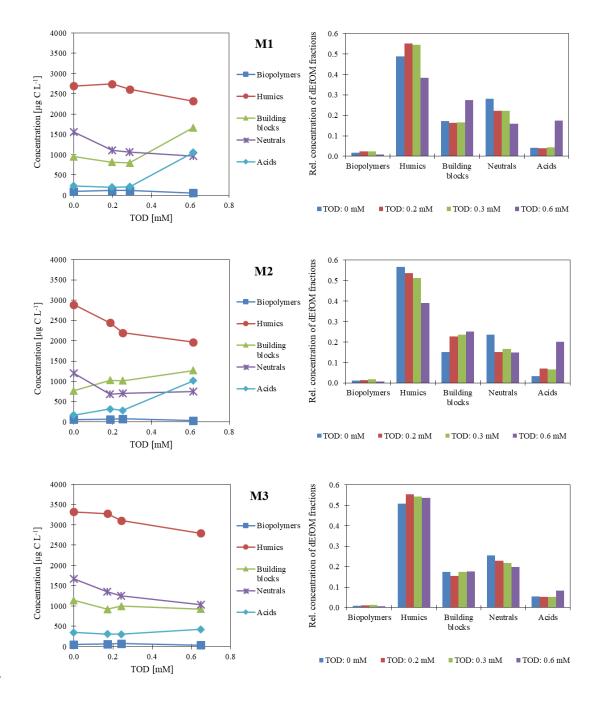




Figure 6. Evolution of dissolved EfOM fractions (left) and contribution to DOC (right) for ozonized MBR
effluents. Relative concentration of EfOM fractions in right column plots (y-axis) refer to DOC of
considered fraction divided by total DOC.

In view of the above results, it is clear that applying ozonation can change EfOM along
the treatment. These changes in turn, could cause negative impacts that require especial
mention: in first place, the cleavage of macromolecules and medium-size structures to

425 yield LMWA, which are not effectively mineralized by ozone, is known for leading to 426 the enhancement of water biodegradability [20,48]. Although this could be interesting if 427 a final biological treatment step is planned (*e.g.*, biological activated carbon), it could be 428 also detrimental if this water is discharged or directly reused instead: biodegradable 429 EfOM is a perfect substrate for bacterial growth in pipes, membranes or receiving water 430 compartments [49]. Regarding the biopolymers fraction, the observed reduction during 431 the process was in any of the cases higher than ca. 45%. For CAS effluents, containing a 432 significantly higher concentration of these components, the reduction percentage was still 433 lower. This means that the greatest part of biopolymers, partly responsible for membrane 434 fouling, for instance, remained unaltered in the water matrix. With respect to particulate 435 matter solubilization during the treatment, ozonizing secondary effluents containing 436 suspended solids seems to be not recommended. Finally, increasing DOC in an already 437 treated wastewater would be in first place inefficient for obvious reasons. In addition, this 438 detrimental effect could lead to other problems such as increased generation of 439 disinfection by-products in hypothetical post-treatment.

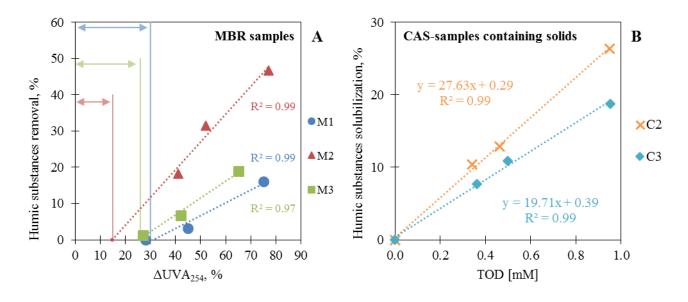
440

# 441 3.5. Relationship between process parameters and EfOM changes

442

Although UVA<sub>254</sub> is typically used as an estimate of the content in humic substances of EfOM [19,50–53], a strict correlation between these two factors has not been observed during ozonation experiments. Oxidation of humic substances initially occurs at the external part of the coil formed by these molecules, which in turn prevents its inner part to be destroyed by ozone and hydroxyl radicals [54]. According to the aromaticity model for humic substances proposed by Del Vecchio and Blough [55], the attack on the peripheral part of chemical structures of humic substances potentially leads to the 450 oxidation of electron-donating or -accepting subunits responsible for charge transfer 451 transitions. As charge transfer transitions between these structures are responsible for UV 452 absorption, this would be enough to reduce the sample UVA<sub>254</sub>. Similar observations 453 reported by studies employing chlorine as oxidizing agent [56], instead of ozone, support 454 this hypothesis. Likewise, the observed mismatch between COD and UVA<sub>254</sub> reduction 455 during ozonation experiments (see Figs. 1 and 2) could be partly related to the mentioned 456 steric impediment caused by the spatial configuration of humic substances.

457





459 Figure 7. Relationship between effluent quality and process parameters with humic substances
460 concentration in wastewater effluents ozonation. A) Humic substances removal in MBR effluent samples
461 versus % UVA<sub>254</sub> reduction; B) Humic substances solubilization versus transferred ozone dose in CAS
462 effluent samples containing significant amounts of suspended organic matter.

464 Recent works have shown how the measurement of spectroscopic parameters (UVA and 465 fluorescence removal) could be potentially used as on-line proxies for biodegradable 466 DOC generation during ozonation and other advanced tertiary treatments [49,57]. This 467 biodegradable fraction of EfOM corresponds to the presence of medium-weight BB, as 468 well as to LMWN and LMWA. Since all these species have their origin in the cleavage

of larger molecules of humic substances, it seems logical to put the focus on the
relationship between this fraction and some common effluent quality and process
parameters in order to anticipate potential changes during ozonation.

472

473 Fig. 7A shows a plot of the percentage of humic substances removal versus the UVA254 474 depletion, for samples coming from MBR units. As seen, a good relationship between 475 them appears to be possible after an initial lag stage in which any abatement of humic 476 substances is observed. In other words, a degree of UV absorbance reduction is observed 477 before measuring any depletion in the concentration of humic substances. This is also 478 indicated in Fig. 7A. The magnitude of this lag can be related with the oxidizing 479 conditions of the media, since a higher availability of oxidant species favors a more severe 480 oxidation of humic substances. As known, O<sub>3</sub> and •OH availability during ozonation will 481 depend on the effluent characteristics, which is mainly given by the contents in organic 482 and inorganic matter that readily consume these oxidants. Interestingly, the observed trend for IOD (*i.e.*, 0.29, 0.19 and 0.23 mmol  $O_3 L^{-1}$  for samples M1, M2 and M3, 483 484 respectively) agreed well with the trend observed for the lag values represented in Fig. 485 7A, which resulted to be: M2 > M3 > M1 (corresponding to 14.5, 25.5 and 31.1 % of 486 UVA<sub>254</sub> depletion, respectively). The slope of the humic substances-UVA<sub>254</sub> removal 487 correlation for these waters, which would give an idea about the process kinetics, also 488 followed the same trend (0.35, 0.76 and 0.47 for M1, M2 and M3, respectively).

489

490 Contrarily, the agreement between humic substances-UVA254 removal and IOD, on one
491 hand, and the observed lag phase and IOD, on the other, was not observed with the initial
492 •OH consumption rate values estimated for MBR effluents. In fact, the presence of •OH
493 in the reaction medium not only depends on the consumption rate of these species by the

494 water matrix but also on the capacity of consumed ozone to generate them. Therefore, a 495 better indicator of •OH availability to react with humic substances during the first stages 496 of ozonation process should also consider ozone consumption. A good option can be the use of the ratio between •OH exposure (*i.e.*,  $\int (-iOH) dt$ ) and IOD. Hydroxyl radical 497 498 exposure for an ozone consumption corresponding to the IOD value could be estimated 499 in this work through ACMP degradation data presented in Fig. 1, according to the 500 calculation procedure described elsewhere [34] and based on the use of an ozone-resistant 501 4.9.10<sup>-7</sup> s for samples M1, M2 and M3, respectively. As can be checked, the trend 502 503 followed by these values now match with the lag and slope values in Fig. 7A: the higher 504 the amount of hydroxyl radicals available per ozone dose, the stronger the oxidation 505 conditions, thus allowing a more significant (and faster) degradation of humic substances 506 by •OH during the initial stages of ozonation.

507

508 Finally, if it was the case that ozonation was applied to a CAS effluent containing 509 significant amounts of suspended matter, the percentage of humic substances 510 solubilization in the water matrix seems to be well correlated with the transferred ozone 511 dose (TOD). This is shown in Fig. 7B: the higher the oxidation extent, the higher the 512 number of hydrophilic moieties generated in the non-soluble fractions of EfOM, which 513 can then be solubilized into the water matrix. In addition, larger concentrations of solid 514 matter, which also contribute to a higher IOD value, seem to favor faster solubilization 515 kinetics, as shown in the correlation parameters obtained in experiments with samples C2 516 and C3.

517

520 Ozone application for the effective removal of micropollutants including ozone-resistant 521 species, which can be a potentially realistic situation in wastewater treatment in a near 522 future, from wastewater produced significant changes in EfOM concentration and quality 523 in all effluents tested. The extent of COD and UVA254 reduction agreed well with the 524 water quality of each effluent, being the most influencing factors the concentration of 525 both dissolved and particulate/colloidal matter and alkalinity. For CAS samples 526 containing relatively large amounts of suspended solids, an increase in the DOC 527 concentration was observed. This was attributed to the solubilization of non-dissolved 528 humic substances. The continuous introduction of this fraction (humic substances) in the 529 reaction medium resulted in the net accumulation of this component. Only for samples 530 coming from MBR systems, the sequential reduction of the largest fractions leading to an 531 accumulation of some of the lightest components, namely BB and LMWA, could be 532 clearly observed. Also for these waters, a net level of OM mineralization took place. In 533 general, an accumulation of low molecular weight acids at the end of the treatment was 534 registered. Correlations between some observed changes (variation in humic substances concentration), water quality (UVA<sub>254</sub>) and process parameters (IOD,  $\int \int \frac{\partial H}{dt} dt$ 535 536 TOD) were established. Changes in organic matter concentration and characteristics 537 derived from ozone application could be detrimental for water reuse purposes, depending 538 on the final application of the treated water. Thus, accumulation of lower weight fractions 539 could lead to bacterial growth or eutrophication, and the increase in humic substances 540 concentration upon oxidation of particular/colloidal matter is in any case detrimental 541 because of the general increase in the DOC content. Thus, a careful consideration of this 542 factor together with the water properties of the effluent to treat and the quality

543	requirements to achieve throughout the process (e.g., micropollutants removal) should be
544	properly assessed during the planning stage of an ozonation unit.

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547

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