



# Geochemistry, Geophysics, Geosystems

## TECHNICAL REPORTS: METHODS

10.1029/2020GC009042

### Key Points:

- A workbook deconvolves raw ICP-MS data to calculate concentrations of all the rare earth elements for multispiked samples
- Values of some common REE reference standards are evaluated
- Data quality using this approach is evaluated with seawater as an example

### Supporting Information:

- Supporting Information S1
- Data Set S1

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### Citation:

Wu, Y., Pena, L. D., Goldstein, S. L., Basak, C., Bolge, L. L., Jones, K. M., et al. (2020). A user-friendly workbook to facilitate rapid and accurate rare earth element analyses by ICP-MS for multispiked samples. *Geochemistry, Geophysics, Geosystems*, 21, e2020GC009042. <https://doi.org/10.1029/2020GC009042>

Received 17 MAR 2020

Accepted 29 AUG 2020

Accepted article online 4 SEPT 2020

## A User-Friendly Workbook to Facilitate Rapid and Accurate Rare Earth Element Analyses by ICP-MS for Multispiked Samples

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**Abstract** The rare earth elements (REEs) are widely used as geochemical tracers in the earth, planetary, and ocean sciences. Inductively coupled plasma-mass spectrometry (ICP-MS) has become the method of choice to analyze REE concentrations because it can rapidly measure the entire REE spectrum at the same time. This Technical Report presents a user-friendly “REE Calculation Workbook” in Microsoft Excel to be used for calculating REE abundances in samples equilibrated with a multielement REE spike. This Workbook can be conveniently used to calculate REE concentrations in natural samples for spiked and unspiked elements measured by ICP-MS. For the spiked elements, their concentrations are calculated using isotope dilution equations. Using these spiked elements as references, concentrations of the four mono-isotopic REE elements, and other REE elements that are treated as mono-isotopic elements (in our case, La and Lu), can be calculated. The REE Workbook can be easily set up for use with different REE spikes. Evaluation of our analytical quality using a quadrupole ICP-MS on 10-ml-sized seawater samples shows that our analyses are comparable to high-precision thermal ionization mass spectrometry (TIMS) studies, with much less time spent processing and analyzing, and with the added advantages of determining mono-isotopic elements. An important result is the clear demonstration of enrichments in Gd and Er compared to neighboring elements in seawater samples. In addition, we compare and evaluate commonly used reference standards BCR-1, Post-Archean Australian Shale (PAAS), and North American Shale Composite (NASC).

### 1. Introduction

The lanthanide series, also called the rare earth elements (REEs), are a group of 15 elements from lanthanum (La, atomic number 57) to lutetium (Lu, atomic number 71), in which the electrons are progressively filled in the 4f orbitals with increasing atomic number. As a result, the REEs have similar outer electronic configurations and thus show similar chemical characteristics. The decrease of ionic radius with increasing atomic number results in small but important differences in their behavior under different geological and cosmological conditions and leads to a wide variety of applications in natural samples as geochemical tracers. For example, the “REE pattern,” reflecting the degree of enrichment or depletion of light versus middle versus heavy REEs in rocks, can be used to constrain provenance or be applied to study partial melting, fractional crystallization, and magma mixing processes (e.g., Henderson, 1984). In seawater, REEs are useful tools to trace water mass mixing, redox processes, and lithogenic sources (e.g., Bertram & Elderfield, 1993; de Baar, Bacon, et al., 1985; de Baar et al., 1983; Elderfield & Greaves, 1982; German et al., 1995; Goldberg et al., 1963; Haley et al., 2014; Høgdahl et al., 1968; Lacan & Jeandel, 2001, 2004; Piper, 1974; Sholkovitz & Schneider, 1991; Zheng et al., 2016). While all the REEs are trivalent, europium (Eu) also has +2 and cerium (Ce) also has +4 charge under some natural conditions, which makes them additionally useful for evaluating redox. For example, Eu<sup>2+</sup> preferentially enters plagioclase feldspar compared to neighboring REE samarium (Sm) and gadolinium (Gd), and its enrichment or depletion relative to other REEs is used to trace processes associated with plagioclase fractionation (e.g., Drake & Weill, 1975; Henderson, 1984) and even crust-mantle recycling (e.g., Sobolev et al., 2000). The enrichment or depletion of Ce compared to

**Table 1**  
Abbreviations Used in This Paper

|                |                       |
|----------------|-----------------------|
| spk            | Spike                 |
| nat            | Natural               |
| meas           | Measured              |
| abc            | Acid blank correction |
| acid_blk       | Acid blank            |
| smpl           | Sample                |
| procedural_blk | Procedural blank      |
| cal            | Calibration           |
| intf           | Interference          |
| oxd_cor        | Oxide correction      |
| mb             | Mass bias             |
| spkfree        | Spike free            |
| sens           | Sensitivity           |
| std            | Standard              |
| smpl_spkfree   | Sample-spike free     |

La and praseodymium (Pr) or neodymium (Nd) is used to trace surface redox processes (e.g., Henderson, 1984; Piper, 1974). For example, Ce is highly depleted in the oxic seawater due to being scavenged in Mn oxides, which is shown as a negative Ce anomaly (e.g., German et al., 1995; Høgdahl et al., 1968; Sholkovitz & Schneider, 1991).

Total REE concentrations in natural samples range from tens of ppt ( $10^{-12}$  g/g) in seawater (global data are available in the GEOTRACES Intermediate Data Product 2017; Schlitzer et al., 2018) to hundreds of ppm ( $10^{-6}$  g/g) in samples of the Earth's crust (e.g., Taylor & McLennan, 1981; Wedepohl, 1995). The first measurements of REEs used X-ray spectrographic techniques to study REEs in meteorites (Noddack, 1935) and shales (Minami, 1935). Since the early 1960s, neutron activation analysis of REEs has been widely used for natural samples (e.g., Goldberg et al., 1963; Haskin & Haskin, 1966; Schmitt et al., 1960). Since the late 1960s, mass spectrometry has emerged as the method of choice to analyze REEs in natural samples, including thermal ionization mass spectrometry (TIMS, first reported by Schnetzler et al., 1967), secondary

ion mass spectrometry (SIMS, first reported by Crozaz & Zinner, 1985), and inductively coupled plasma-mass spectrometry (ICP-MS, first reported by Jenner et al., 1990; Shabani et al., 1990). All of the mass spectrometric approaches require deconvolution of the raw data (e.g., Barbey et al., 1995, for SIMS), which go back to the beginning of using mass spectrometers (e.g., Monteiro & Reed, 1969; Washburn et al., 1943).

A common approach for calculating the concentration of an element in a sample is to add a spike (enriched in an isotope of that element) to the sample and use isotope dilution (ID) analysis (e.g., Albarède, 1996; Dickin, 1995; Heumann, 1992; Webster, 1960) to calculate the sample concentration, which is based on knowing the precise weights of the sample and spike, the isotope ratios of the natural sample and the spike, and the isotope ratio of the spike-sample mixture. The first mass spectrometric analyses of REEs in the geosciences using multi-REE spike ID date back to the late 1960s (e.g., Arth & Hanson, 1975; Gast, 1968; Schnetzler & Philpotts, 1970; Schnetzler et al., 1967). Since then the approach has been used in studies to determine REE concentrations in all kinds of natural samples (Stracke et al., 2014) including rocks (e.g., Hanson, 1980; Hooker et al., 1975; Schnetzler et al., 1967), meteorites (e.g., Evensen et al., 1978; Masuda et al., 1973; Nakamura, 1974), ferromanganese sediments (e.g., Elderfield & Greaves, 1981; Elderfield et al., 1981), seawater (e.g., Behrens et al., 2016; Bertram & Elderfield, 1993; Elderfield & Greaves, 1982; Klinkhammer et al., 1983; Pahnke et al., 2012; Piepgras & Jacobsen, 1992; Rousseau et al., 2013; Sholkovitz & Schneider, 1991; van de Flierdt et al., 2012; Zheng et al., 2015), hydrothermal fluids (e.g., Michard & Albarède, 1986; Michard et al., 1983), river water (e.g., Goldstein & Jacobsen, 1988; Sholkovitz, 1995; Sholkovitz et al., 1999), marine particles (e.g., Cullen et al., 2001; Field & Sherrell, 1998; Sholkovitz et al., 1994), and geological reference materials (e.g., Baker et al., 2002; Kent et al., 2004; Krogh Jensen et al., 2003; Raczek et al., 2001; Willbold & Jochum, 2005), among others. Using the standard ID equation (Dodson, 1963, 1969) for two isotopes, given an element  $x$ , its concentration (grams of  $x$ /grams of sample) can be calculated as

$$[x] = \frac{\left(\frac{i}{j}x\right)_{\text{spk}} - \left(\frac{i}{j}x\right)_{\text{meas}}}{\left(\frac{i}{j}x\right)_{\text{meas}} - \left(\frac{i}{j}x\right)_{\text{nat}}} \times \frac{[jx]_{\text{spk}}}{jAb_{\text{nat}}} \times \left(\frac{m_{\text{spk}}}{m_{\text{smpl}}}\right) \times M_x,$$

where  $i$  is a non-enriched isotope in the spike,  $j$  is the enriched isotope in the spike,  $\left(\frac{i}{j}x\right)_{\text{spk}}$  is the isotope ratio in the spike,  $\left(\frac{i}{j}x\right)_{\text{meas}}$  is the isotope ratio in the measured sample-spike mixture,  $\left(\frac{i}{j}x\right)_{\text{nat}}$  is the natural isotope ratio,  $[jx]_{\text{spk}}$  is the molar concentration of  $jx$  in the spike (mol  $jx$ /g solution),  $jAb_{\text{nat}}$  is the natural fractional abundance of  $jx$  (i.e., the mole fraction of  $jx$  atoms in a natural sample),  $m_{\text{spk}}$  and  $m_{\text{smpl}}$  are spike and sample weights (g), and  $M_x$  is the atomic mass of element  $x$  (g/mol). Abbreviations used in this paper are shown in Table 1.

**Table 2**  
*Analyzed Isotopes of Each Element for Samples, Procedural Blank, Pure Pr Solution, and Mass Bias Solution*

| Element | Analyzed isotopes |     |     |     |
|---------|-------------------|-----|-----|-----|
| Ba      | 135               | 137 |     |     |
| La      | 138               | 139 |     |     |
| Ce      | 140               | 142 |     |     |
| Pr      | 141               |     |     |     |
| Nd      | 143               | 144 | 145 | 146 |
| Sm      | 147               | 148 | 149 |     |
| Eu      | 151               | 153 |     |     |
| Gd      | 155               | 157 | 160 |     |
| Tb      | 159               |     |     |     |
| Dy      | 161               | 163 |     |     |
| Ho      | 165               |     |     |     |
| Er      | 166               | 167 |     |     |
| Tm      | 169               |     |     |     |
| Yb      | 171               | 172 | 174 |     |
| Lu      | 175               | 176 |     |     |
| Hf      | 177               |     |     |     |

Since ICP-MS was developed in the early 1980s (Gray & Date, 1983), it has been increasingly used for rapid multielement analysis and has become the preferred method for REE analyses. For example, REE measurements using ID-TIMS are precise but time consuming: Fractions of the REEs need to be collected, loaded onto different filaments, and analyzed separately; a sample requires ~1–3 days; and only the spiked multi-isotopic elements are measured. With ID ICP-MS analysis, many studies analyze REEs in two fractions (light and heavy REEs). In this study, by ICP-MS the entire REE spectrum of 14 elements can be analyzed in a single fraction in ~10 min. Combining the use of multielement REE spikes and ICP-MS offers a convenient means to obtain high-precision REE analyses, and because the results are based on measurement of isotope ratios, their use avoids some of the pitfalls associated with reliance on signal intensities alone.

With the increasing number of labs performing REE analyses, in order to encourage the use of multielement REE spikes, help to ensure the accuracy of the results, and promote cross laboratory consistency, this Technical Report presents an *REE Calculation Workbook* (Data Set S1 in the supporting information) for convenient use by the community to calculate REE concentrations of samples equilibrated with a multielement spike and mea-

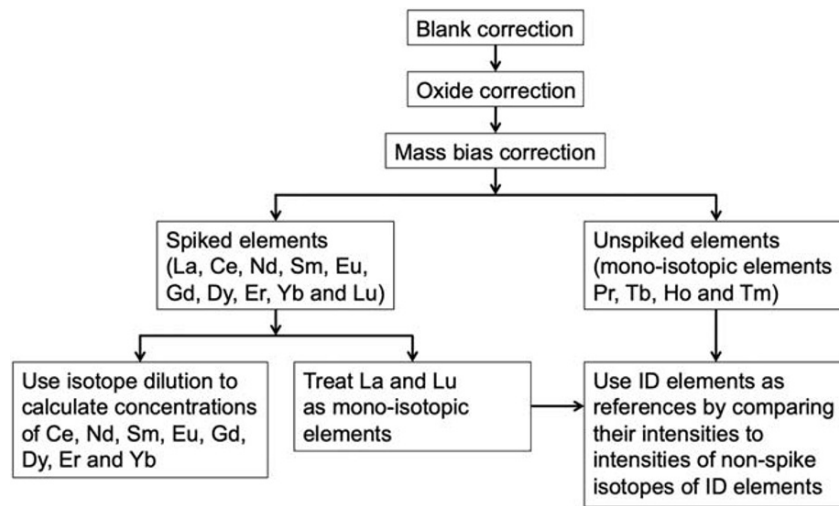
sured by ICP-MS. The Workbook is set up for the procedure used at the Lamont-Doherty Earth Observatory (LDEO) of Columbia University, in which concentrations of eight spiked elements are determined using ID, and the concentrations of the spiked elements are used as references to calculate concentrations of the mono-isotopic REE elements, and La and Lu. The Workbook can be easily modified for other combinations of spiked and nonspiked elements.

## 2. Explanation of the REE Calculation Workbook

The LDEO lab uses a multielement REE spike mixed by Robert D. Vocke and calibrated by Vocke and Steven B. Shirey in Professor Gilbert Hanson's lab at Stony Brook University. The spike is enriched in  $^{138}\text{La}$ ,  $^{142}\text{Ce}$ ,  $^{145}\text{Nd}$ ,  $^{149}\text{Sm}$ ,  $^{153}\text{Eu}$ ,  $^{155}\text{Gd}$ ,  $^{161}\text{Dy}$ ,  $^{167}\text{Er}$ ,  $^{171}\text{Yb}$ , and  $^{176}\text{Lu}$  (Table S1). An aliquot of the stock solution was later diluted by Diane K. McDaniel (the “DKM aliquot”), who remeasured the isotope ratios and calibrated the concentrations using gravimetric REE solutions made from metal ingots. At LDEO we use a further diluted split of the Stony Brook-DKM aliquot and its calibration for the isotope ratios and the relative concentrations. We recalibrated the absolute concentrations against a Nd standard from High-Purity™ Standards Inc., with a certified  $\pm 0.3\%$  error on the Nd concentration, using “reverse ID,” where the spike is treated as the “sample” and the natural Nd standard is treated as the “spike.”

This contribution uses seawater REE analyses as our example, although the Workbook can be used for any type of sample. We use seawater because it illustrates the possibility of obtaining high-quality analyses on very small samples, as discussed in section 5. LDEO methods for seawater REE analyses are described in Behrens et al. (2016). Each sample is spiked to aim for an ideal  $^{145}\text{Nd}/^{143}\text{Nd}$  ratio for the sample-spike mixture (~9.0, Table S1) that allows for analyses of all the multi-isotope REE elements except La and Lu. After the spike is added to the acidified seawater sample, it is allowed to equilibrate for at least 24 hr. The sample-spike mixture is then purified using a commercially available preconcentration unit that separates the REEs, the seaFast Automated Preconcentration System for Undiluted Seawater (Elemental Scientific Inc. or ESI, Omaha, Nebraska, USA). The seaFast processing time to obtain the REE fraction is ~15 min. We analyze the samples using a VG PlasmaQuad ExCell® quadrupole ICP-MS. The instrument is coupled to a desolvating introduction system (a CETAC Aridus™) that minimizes molecular oxide ion formation with a self-aspirated Apex ST PFA micro flow nebulizer (ESI). The mass spectrometer is optimized for sensitivity and operated in pulse counting mode with 10 ms of dwell time for all isotopes. The analyzing time for a sample is ~10 min for the entire REE spectrum. The isotopes analyzed for each element are listed in Table 2.

We perform blank corrections (section 2.1), oxide corrections (section 2.2), and mass bias corrections (section 2.3) for all REEs. Then we divide the REEs into three groups and calculate their concentrations for



**Figure 1.** Flow chart summarizing the steps of REE data reduction in the REE Calculation Workbook.

ID elements (section 2.4), mono-isotopic REE elements (section 2.5), and elements treated like the mono-isotopic REE elements (section 2.6). A flow chart for the REE data reduction is summarized in Figure 1.

### 2.1. Blank Correction

Background intensities of REEs are measured on the mass spectrometer using a 3% nitric acid solution. All intensities for the procedural blank (this section), a pure praseodymium (Pr) solution (section 2.2), a mass bias solution (section 2.3), and samples are corrected by subtracting the counts of the 3% nitric acid blank using  $I_{abc} = I_{raw} - I_{acid-blk}$ , where  $I_{raw}$  is the raw intensity,  $I_{acid-blk}$  is the intensity of the acid blank, and  $I_{abc}$  is the intensity after acid blank correction.

In our study, every batch of samples is accompanied by a procedural blank that is processed using the same method as samples. These procedural blanks provide information of how much signal comes from the whole sample processing procedure, an important step to ensure high-quality data. Sample intensities are corrected by subtracting the procedural blank using  $I_{smp} = I_{abc} - I_{procedural-blk}$ , where  $I_{abc}$  is the intensity after acid blank correction,  $I_{procedural-blk}$  is the intensity of the procedural blank after acid blank correction, and  $I_{smp}$  is the intensity of sample after procedural blank correction. In our study, procedural blanks ( $n = 5$ ) of 1%  $HNO_3$  typically represent  $\leq 0.9\%$  of the sample intensities except for Ce ( $\leq 1.7\%$ ).

### 2.2. Oxide Correction

To correct for molecular oxide formation and interference at REE isotope masses, we perform an oxide calibration analysis by measuring masses of M and MO in pure, single element solutions of barium (Ba) and REEs. We analyze Ba and all the REEs once to construct a calibration matrix and use them, along with a measurement of Pr and PrO during each measurement session to scale up or down the oxide corrections for the individual measurement session. The isotopes analyzed for each single element solution are listed in Table 3. The oxide calibration value is expressed as the ratio of  $\frac{^{MO}I_{cal}}{^M I_{cal}}$ , where  $^M I_{cal}$  is the intensity of mass M and  $^{MO}I_{cal}$  is the intensity of mass MO measured during the calibration. For an unknown sample, the oxide interference for any mass M is calculated as  $^{MO}I_{intf} = \frac{^{MO}I_{cal}}{^M I_{cal}} \times ^M I_{smp}$ , where  $^M I_{smp}$  is the measured intensity for any mass and  $^{MO}I_{intf}$  is the intensity of its oxide interference.

The oxide calibration values are not constant and can vary between different analytical sessions. Thus, oxide formation rates must be assessed and adjusted at every analytical session in which REEs are measured. However, we can assume that all the REEs show the same fractional change in oxide formation (e.g.,

**Table 3**  
Analyzed Isotopes for Each Pure and Single Element Solutions of Ba and REE for Oxide Calibration Analysis

| Analyzed elements and their oxides | Analyzed isotopes |     |     |     |
|------------------------------------|-------------------|-----|-----|-----|
| Ba                                 | 135               | 137 |     |     |
| BaO                                | 151               | 153 |     |     |
| La                                 | 139               |     |     |     |
| LaO                                | 155               |     |     |     |
| Ce                                 | 140               | 142 |     |     |
| CeO                                | 156               | 158 |     |     |
| Pr                                 | 141               |     |     |     |
| PrO                                | 157               |     |     |     |
| Nd                                 | 143               | 144 | 145 | 150 |
| NdO                                | 159               | 160 | 161 | 166 |
| Sm                                 | 147               | 149 | 150 |     |
| SmO                                | 163               | 165 | 166 |     |
| Eu                                 | 151               | 153 |     |     |
| EuO                                | 167               | 169 |     |     |
| Gd                                 | 155               | 156 | 158 | 160 |
| GdO                                | 171               | 172 | 174 | 176 |
| Tb                                 | 159               |     |     |     |
| TbO                                | 175               |     |     |     |
| Dy                                 | 161               |     |     |     |
| DyO                                | 177               |     |     |     |

Zhao et al., 2019; [http://minerva.union.edu/hollochk/icp-ms/ree\\_corrections.html](http://minerva.union.edu/hollochk/icp-ms/ree_corrections.html)). Therefore, we analyze the  $\frac{^{157}\text{PrO}}{^{141}\text{Pr}}$  ratio of a 100 ng/L Pr solution during each measurement session to determine the oxide correction

factor  $C = \frac{\left(\frac{^{157}\text{PrO}}{^{141}\text{Pr}}\right)_{\text{today}}}{\left(\frac{^{157}\text{PrO}}{^{141}\text{Pr}}\right)_{\text{cal}}}$ , where  $\left(\frac{^{157}\text{PrO}}{^{141}\text{Pr}}\right)_{\text{cal}}$  is measured during the oxide calibration analysis and  $\left(\frac{^{157}\text{PrO}}{^{141}\text{Pr}}\right)_{\text{today}}$  is measured each time we analyzed REEs.

Then the oxide interference is expressed as  $^{MO}I_{\text{intf}} = C \times \frac{^{MO}I_{\text{cal}}}{M_{\text{I}_{\text{cal}}}} \times M_{\text{I}_{\text{smp}}}$ .

The final true intensity without oxide interference is calculated as  $^{MO}I_{\text{Oxd}^{\text{cor}}}$

$$= ^{MO}I_{\text{smp}} - C \times \frac{^{MO}I_{\text{cal}}}{M_{\text{I}_{\text{cal}}}} \times M_{\text{I}_{\text{smp}}}$$

The above calculation is based on the assumption that the ratio of  $\frac{^{MO}I_{\text{cal}}}{M_{\text{I}_{\text{cal}}}}$  is

identical in a pure, single element solution and in a sample solution. However, there is a possibility of a temperature-dependent reaction such as  $\text{LaO} + \text{Ce} \rightleftharpoons \text{La} + \text{CeO}$  in the plasma, which can add additional oxide variables in the calculating equations and result in additional uncertainties to calculate elements (e.g., Yb) with oxide interference (e.g., GdO) from an element with similar isotopic abundance (e.g., Gd) (e.g., Albalat

et al., 2012). Therefore, it is very important to minimize this effect by minimizing the oxide formation in our analysis. In each REE measurement session, we adjust the gas flows to an optimal state to obtain the minimum oxide formation ( $0.0066 \pm 0.0017$ ,  $1\sigma$ , for the Pr monitor solution) so that the oxide correction is extremely small. Our high-quality data for natural samples used for international calibration (Behrens et al., 2016; Pahnke et al., 2012; van de Fliertdt et al., 2012) agree very well with results from different research groups in the world (section 5). In addition, the repeated analysis of samples yields very good long-term external reproducibility (section 5).

### 2.3. Mass Fractionation Correction

We use the exponential mass fractionation law  $R_c = R_m \times \left(\frac{m_i}{m_j}\right)^\beta$  (e.g., Maréchal et al., 1999; Russell et al., 1978; Wasserburg et al., 1981) to correct for mass fractionation, where  $R_c$  is the mass fractionation corrected isotope ratio in the sample,  $R_m$  is the measured isotope ratio in the sample,  $m_i$  is the atomic mass of the spike isotope, usually the one enriched in the spike, and  $m_j$  is the atomic mass of the reference isotope. To calculate  $\beta$ , the exponential law mass fractionation factor, we use a mass bias solution in which each REE has concentration of 100 ng/L, and we measure it during each REE measurement session.  $\beta$  is

expressed as  $\beta = \frac{\text{Ln}\left(\frac{R_n}{R_{mb}}\right)}{\text{Ln}\left(\frac{m_i}{m_j}\right)}$ , where  $R_n$  is the ratio in natural samples and  $R_{mb}$  is the measured ratio in the

mass bias solution.

The correction for instrumental mass fractionation has very little impact on the final results of REE concentrations. The differences between mass bias corrected values and nonmass bias corrected values are within 0.1% for La, Ce, Pr, and Nd, within 1% for Tb, Ho, Er, Tm, Yb, and Lu, and within 1–2% for Sm, Gd, and Dy, and 3–4% for Eu. We use the same  $\beta$  values for each analytical session. Based on 59 analytical sessions,  $\beta$  values ( $1\sigma$ ) are as follows: Nd,  $-0.9723 (\pm 0.9942)$ ; Sm,  $-1.0309 (\pm 0.7166)$ ; Eu,  $-1.5044 (\pm 0.8249)$ ; Gd,  $3.2681 (\pm 5.8387)$ ; Dy,  $-0.8947 (\pm 0.6918)$ ; Er,  $-0.1470 (\pm 1.6342)$ ; and Yb,  $-0.9719 (\pm 0.7546)$ .

#### 2.4. Concentrations of ID Elements

The concentrations of spiked elements are calculated using ID equations (section 1). Taking Nd as an example, we use a  $^{145}\text{Nd}$  spike, and the Nd concentration (g Nd/g sample) is calculated as

$$[\text{Nd}] = \frac{\left(\frac{^{143}\text{Nd}}{^{145}\text{Nd}}\right)_{\text{spk}} - \left(\frac{^{143}\text{Nd}}{^{145}\text{Nd}}\right)_{\text{meas}}}{\left(\frac{^{143}\text{Nd}}{^{145}\text{Nd}}\right)_{\text{meas}} - \left(\frac{^{143}\text{Nd}}{^{145}\text{Nd}}\right)_{\text{nat}}} \times \frac{[^{145}\text{Nd}]_{\text{spk}}}{^{145}\text{Nd}Ab_{\text{nat}}} \times \frac{m_{\text{spk}}}{m_{\text{smpl}}} \times M_{\text{Nd}}, \quad (1)$$

where  $\left(\frac{^{143}\text{Nd}}{^{145}\text{Nd}}\right)_{\text{spk}}$  is the isotope ratio in the spike after mass bias correction,  $\left(\frac{^{143}\text{Nd}}{^{145}\text{Nd}}\right)_{\text{meas}}$  is the isotope ratio in the measured sample-spike mixture after mass bias correction,  $\left(\frac{^{143}\text{Nd}}{^{145}\text{Nd}}\right)_{\text{nat}}$  is the natural isotope ratio (= 1.468),  $[^{145}\text{Nd}]_{\text{spk}}$  is the molar concentration of  $^{145}\text{Nd}$  in the spike (mol  $^{145}\text{Nd}$ /g solution),  $^{145}\text{Nd}Ab_{\text{nat}}$  is the natural abundance of  $^{145}\text{Nd}$  (i.e., the mole fraction of  $^{145}\text{Nd}$  atoms in a natural sample = 0.0829),  $m_{\text{spk}}$  and  $m_{\text{smpl}}$  are spike and sample weights (g), and  $M_{\text{Nd}}$  is the atomic mass of Nd (144.24 g/mol). Other elements calculated using the ID equation are Sm, Eu, Gd, Dy, Er, and Yb and henceforth will be called “ID elements.”

Since  $^{142}\text{Ce}$  has an isobaric interference from Nd,  $^{142}\text{Ce}$  in the sample is calculated by subtracting interference of  $^{142}\text{Nd}$  from the sample and the spike (Jones, 2010) using

$$^{142}\text{Ce}I_{\text{smpl}} = ^{142}\text{Total}I_{\text{smpl}} - ^{143}\text{Nd}I_{\text{smpl}} \times \left(\frac{^{142}\text{Nd}}{^{143}\text{Nd}}\right)_{\text{nat}} - ^{143}\text{Nd}I_{\text{spk}} \times \left(\frac{^{142}\text{Nd}}{^{143}\text{Nd}}\right)_{\text{spk}},$$

where  $^{142}\text{Total}I_{\text{smpl}}$  is the intensity of mass 142 in the sample,  $^{143}\text{Nd}I_{\text{smpl}}$  is the intensity of  $^{143}\text{Nd}$  in the sample,  $\left(\frac{^{142}\text{Nd}}{^{143}\text{Nd}}\right)_{\text{nat}}$  is the natural isotope ratio (= 2.230),  $^{143}\text{Nd}I_{\text{spk}}$  is the intensity of  $^{143}\text{Nd}$  in the spike (calculating equation given in Equation 2 in section 2.5), and  $\left(\frac{^{142}\text{Nd}}{^{143}\text{Nd}}\right)_{\text{spk}}$  is the isotope ratio in the spike. Then the Ce concentration is calculated using the ID Equation 1.

#### 2.5. Concentrations of Mono-Isotopic Elements

For the mono-isotopic elements Pr, Tb, Ho, and Tm, their concentrations are calculated by comparing their intensities to intensities of nonspike isotopes of ID elements. To calculate intensities of nonspike isotopes of ID elements, contributions from the spike need to be subtracted (Jones, 2010). Taking Nd as an example, the spike-free intensity for  $^{143}\text{Nd}$  is calculated as

$$^{143}\text{Nd}I_{\text{spkfree}} = ^{143}\text{Nd}I_{\text{total}} - ^{143}\text{Nd}I_{\text{spk}} = ^{143}\text{Nd}I_{\text{total}} - \frac{\left(\frac{^{143}\text{Nd}}{^{145}\text{Nd}}\right)_{\text{nat}} \times ^{145}\text{Nd}I_{\text{total}} - ^{143}\text{Nd}I_{\text{total}}}{\frac{\left(\frac{^{143}\text{Nd}}{^{145}\text{Nd}}\right)_{\text{nat}}}{\left(\frac{^{143}\text{Nd}}{^{145}\text{Nd}}\right)_{\text{spk}}} - 1}, \quad (2)$$

where  $^{143}\text{Nd}I_{\text{total}}$  is the intensity of  $^{143}\text{Nd}$  in total,  $^{143}\text{Nd}I_{\text{spk}}$  is the intensity of  $^{143}\text{Nd}$  in the spike,  $^{145}\text{Nd}I_{\text{total}}$  is the intensity of  $^{145}\text{Nd}$  in total,  $\left(\frac{^{143}\text{Nd}}{^{145}\text{Nd}}\right)_{\text{nat}}$  is the natural isotope ratio, and  $\left(\frac{^{143}\text{Nd}}{^{145}\text{Nd}}\right)_{\text{spk}}$  is the isotope ratio in the spike. Then the relationship between the intensities of the ID element and mono-isotopic element is established by comparing their sensitivities in the mass bias standard solution. Here sensitivity is defined as the ratio of the intensity of an isotope versus its concentration in a standard solution (e.g., Field & Sherrell, 1998; Kent et al., 2004; Willbold & Jochum, 2005; Willbold et al., 2003). As an example, for Nd, the sensitivity of  $^{143}\text{Nd}$  is calculated as  $^{143}\text{Nd}_{\text{sens}} = \frac{^{143}\text{Nd}I_{\text{std}}}{[\text{Nd}]_{\text{std}} \times ^{143}\text{Nd}Ab_{\text{nat}}}$ , where  $^{143}\text{Nd}I_{\text{std}}$  is the intensity of  $^{143}\text{Nd}$  in the standard solution,  $[\text{Nd}]_{\text{std}}$  is the Nd concentration in the standard solution (g Nd/g solution), and  $^{143}\text{Nd}Ab_{\text{nat}}$  is the fractional molar abundance of  $^{143}\text{Nd}$  in a natural sample (= 0.1217). For Pr, the sensitivity of  $^{141}\text{Pr}$  is calculated as  $^{141}\text{Pr}_{\text{sens}} = \frac{^{141}\text{Pr}I_{\text{std}}}{[\text{Pr}]_{\text{std}} \times ^{141}\text{Pr}Ab_{\text{nat}}}$ , where  $^{141}\text{Pr}I_{\text{std}}$  is the intensity of  $^{141}\text{Pr}$  in the

standard solution,  $[\text{Pr}]_{\text{std}}$  is the Pr concentration in the standard solution (g Pr/g solution), and  $^{141}\text{Pr}Ab_{\text{nat}}$  is the fractional molar abundance of  $^{141}\text{Pr}$  in a natural sample (= 1). Then the sensitivity ratio  $\frac{^{143}\text{Nd}_{\text{sens}}}{^{141}\text{Pr}_{\text{sens}}}$  in the standard solution is used to calculate the Pr concentration in the sample (g Pr/g sample) using

$$[\text{Pr}]_{\text{smpl}} = \frac{^{143}\text{Nd}_{\text{sens}}}{^{141}\text{Pr}_{\text{sens}}} \frac{^{141}\text{Pr}I_{\text{smpl}}}{^{143}\text{Nd}I_{\text{smpl}} - \text{spkfree}} \times \frac{[\text{Nd}]_{\text{smpl}} \times ^{143}\text{Nd}Ab_{\text{nat}}}{^{141}\text{Pr}Ab_{\text{nat}}}, \quad (3)$$

where  $^{141}\text{Pr}I_{\text{smpl}}$  is the intensity of  $^{141}\text{Pr}$  in the sample,  $^{143}\text{Nd}I_{\text{smpl}} - \text{spkfree}$  is the spike-free intensity for  $^{143}\text{Nd}$  in the sample,  $[\text{Nd}]_{\text{smpl}}$  is the Nd concentration in the sample (g Nd/g sample),  $^{143}\text{Nd}Ab_{\text{nat}}$  is the fractional abundance of  $^{143}\text{Nd}$  in a natural sample (= 0.1217), and  $^{141}\text{Pr}Ab_{\text{nat}}$  is the fractional abundance of  $^{141}\text{Pr}$  in a natural sample (= 1).

### 2.6. Concentrations of Multi-Isotope Elements Treated as Mono-Isotopic Elements

For La and Lu,  $^{138}\text{La}$  has isobaric interferences with  $^{138}\text{Ba}$  and  $^{138}\text{Ce}$  and  $^{176}\text{Lu}$  has isobaric interferences with  $^{176}\text{Yb}$  and  $^{176}\text{Hf}$ . In addition,  $^{138}\text{La}$  and  $^{176}\text{Lu}$  both have very low abundances (0.1% and 2.6%, respectively) in natural samples. Using the Stony Brook-DKM spike at LDEO, typically, only ~2% of the signal intensity on mass 138 comes from  $^{138}\text{La}$  and ~5% of the signal intensity of mass 176 comes from  $^{176}\text{Lu}$ . As a result, subtracting interferences adds major uncertainty to the ID calculations. For these reasons, after subtracting the  $^{139}\text{La}$  and  $^{175}\text{Lu}$  spike contributions, at LDEO we treat La and Lu as mono-isotopic elements, and their concentrations are calculated using equations for mono-isotopic elements.

Some multielement spikes have fewer enriched REE isotopes than the Stony Brook-DKM spike. In these cases, the Workbook can be easily modified so that they can be treated as mono-isotopic elements with their concentrations calculated using the equations for mono-isotopic elements.

## 3. Structure of the REE Calculation Workbook

The REE Calculation Workbook is downloadable as Data Set S1 in the supporting information. The worksheet “General Info” lists the isotopes measured during sample analysis and oxide calibration analysis. It also explains the parameters required for concentration calculations. Sample-related information is shown in blue, and spike-related information is shown in green. The worksheet “Raw Data” is used for entering the measured intensities for the blank solution, procedural blank, pure Pr solution, mass bias solution, and sample solution (in blue). The worksheet “Weights” is used for entering the sample (in blue) and spike (in green) weights for each sample. The worksheet “Spike” contains the concentrations of each isotope in the spike (in green), and the natural isotope ratios and atomic masses of these isotopes, isotopic compositions in the spike (in green) and in nature, and atomic masses of each REE element. The dilution factor (in green) is entered in the “Spike” worksheet to calculate the REE concentrations of the diluted spike. The worksheet “Isotope Abundances” contains the natural isotope abundances of Ba, each REE, and hafnium (Hf) (Holden et al., 2018). The worksheets “Blank Cor,” “Oxide Cor,” and “Mass Bias Cor” are used for blank correction (on 2.1), oxide correction (section 2.2), and mass bias correction (section 2.3), respectively. The worksheets “ID,” “Ce,” “Mono,” “La,” and “Lu” are used to calculate concentrations of ID elements (section 2.4), Ce (section 2.4), mono-isotopic elements (section 2.5), La (section 2.6), and Lu (section 2.6), respectively. The final results of calculated REE concentrations are shown in the worksheet “REEs.”

## 4. Evaluation of REE Standard Reference Values for BCR-1, NASC, and PAAS

In this contribution we show as an example some REE concentrations for some seawater samples calculated using the Workbook. Information for samples and spike were entered following the instructions in section 3. Seawater REEs are usually referenced to shale standards, based on the reasoning that in seawater these elements are mainly derived from weathering of the continental surface and shales represent admixtures of large portions of the continental surface. Also, shales, like average upper continental crust, show Eu depletions when compared to solar system and planetary reference values such as bulk silicate earth or chondrites (McDonough & Sun, 1995; Rudnick & Gao, 2003), and this is propagated into seawater; normalizing seawater to shales minimizes Eu anomalies in the seawater REE patterns. An important challenge is to

**Table 4**  
Reference Values of BCR-1, PAAS, and NASC Used in This Study and Their Sources

|    | BCR-1-G90 | BCR-1-A94 | BCR-1-J16 | PAAS-TM85 | PAAS-P12 | NASC-G84 | NASC-GJ88 |
|----|-----------|-----------|-----------|-----------|----------|----------|-----------|
| La | 24.9      | 25.0      | 25.46     | 38        | 44.56    | 31.1     | 34.0      |
| Ce | 53.7      | 53.6      | 53.94     | 80        | 88.25    | 66.7     | 66.7      |
| Pr | 6.80      | 6.90      | 6.765     | 8.9       | 10.15    | 7.90     | 7.93      |
| Nd | 28.8      | 28.6      | 28.68     | 32        | 37.32    | 27.4     | 30.1      |
| Sm | 6.59      | 6.55      | 6.603     | 5.6       | 6.884    | 5.59     | 5.80      |
| Eu | 1.95      | 1.92      | 1.957     | 1.1       | 1.215    | 1.18     | 1.16      |
| Gd | 6.68      | 6.82      | 6.725     | 4.7       | 6.043    | 5.40     | 5.12      |
| Tb | 1.05      | 1.05      | 1.063     | 0.77      | 0.8914   | 0.850    | 0.779     |
| Dy | 6.34      | 6.37      | 6.391     | 4.4       | 5.325    | 5.33     | 4.67      |
| Ho | 1.26      | 1.34      | 1.268     | 1.0       | 1.053    | 1.04     | 0.983     |
| Er | 3.63      | 3.71      | 3.658     | 2.9       | 3.075    | 3.21     | 2.73      |
| Tm | 0.560     | 0.545     | 0.5350    | 0.40      | 0.4510   | 0.500    | 0.414     |
| Yb | 3.38      | 3.39      | 3.377     | 2.8       | 3.012    | 3.06     | 2.67      |
| Lu | 0.510     | 0.497     | 0.4988    | 0.43      | 0.4386   | 0.456    | 0.406     |

*Note.* Concentrations are in ppm (they are also listed in pmol/kg in Table S3, often used in seawater studies). Sources: G90: Gladney et al. (1990); A94: Asmerom et al. (1994) (the multi-isotopic elements are from Goldstein & Jacobsen, 1988); J16: Jochum et al. (2016); TM85: Taylor and McLennan (1985) (the values with the exceptions of Tm and Lu are those of Nance & Taylor, 1976); P12: Pourmand et al. (2012); G84: Gromet et al. (1984); GJ88: Goldstein and Jacobsen (1988) (Lu is from Piepgras & Jacobsen, 1992; the mono-isotopic elements were measured at Harvard by quadrupole ICP-MS and are included here courtesy of S. B. Jacobsen).

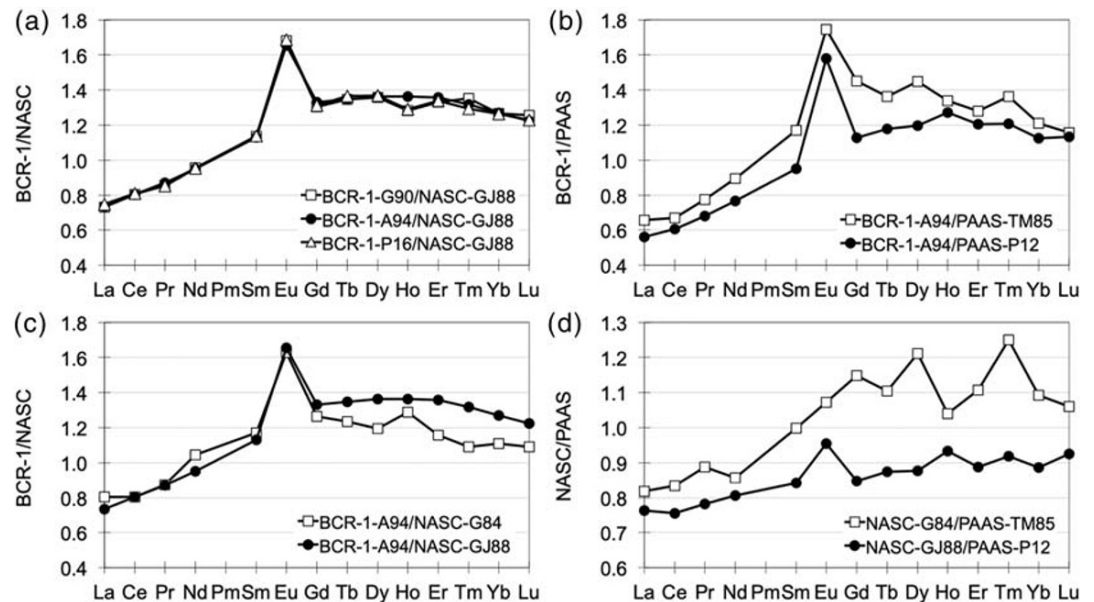
decide which reference values to use. The most used shale reference standards are Post-Archean Australian Shale (PAAS), whose values are averages of analyses of multiple samples (Nance & Taylor, 1976), and the North American Shale Composite (NASC), which is a sample composed of a mixture of 40 shale samples (Haskin et al., 1966). We have evaluated these reference standards to determine which values are best to use, and the USGS Columbia River basalt reference standard BCR-1, which as a mantle partial melt should show a smooth REE pattern for all the REEs except Eu. For BCR-1 we compare three sets of reference values, the preferred values of two compilations (Gladney et al., 1990; Jochum et al., 2016) and the analyses reported in Asmerom et al. (1994), henceforth BCR-1-G90, -J16, and -A94, respectively. For PAAS we use the classic values of Taylor and McLennan (1985) and the reevaluation by Pourmand et al. (2012), henceforth PAAS-TM85 and -P12, respectively. For NASC we use Gromet et al. (1984) and Goldstein and Jacobsen (1988), henceforth NASC-G84 and -GJ88, respectively. The values used are listed in Tables 4 (in ppm) and S2 (in pmol/kg, the units often used for seawater, and ppm). The BCR-1-A94 REE values use the data from Goldstein and Jacobsen (1988) for the ID elements and Asmerom et al. (1994) added the mono-isotopic elements. The PAAS-TM85 values (Taylor & McLennan, 1985) are the same as Nance and Taylor (1976) with the exceptions of Tm and Lu. The NASC-GJ88 values include a modification to Lu (in Piepgras & Jacobsen, 1992) and until now unpublished mono-isotopic elements measured along with Lu by quadrupole ICP-MS (S. B. Jacobsen, personal communication, May 18, 2020).

We compared all seven data sets (listed in Table 4) in order to evaluate the reference values. The distilled results are shown in Figure 2; while not all of the possible data combinations are shown, those that are shown confirm our conclusions. As the differences in chemical behavior between the REEs are primarily due to the systematic changes in cationic volume, we expect that more accurate analyses would result in smoother REE patterns, excepting Ce and Eu. Comparison of the three BCR-1 values with NASC-GJ88 (which we conclude below are the best shale reference values), on a linear scale to emphasize variability (Figure 2a), shows that BCR-1-A94 is smoother than the others; therefore, we use BCR-1-A94 as our BCR-1 reference. Comparing it to the PAAS reference values (Figure 2b) shows a smoother pattern for PAAS-P12. Comparing BCR-1-A94 to the NASC reference values (Figure 2c) shows a smoother value for NASC-GJ88. These indications that NASC-GJ88 and PAAS-P12 are the better NASC and PAAS reference values are seemingly confirmed by direct comparison of the shales (Figure 2d), where normalizing NASC-GJ88 and PAAS-P12 yields a smoother REE pattern than normalizing NASC-G84 and PAAS-TM85. Finally, comparing all four shale reference values to BCR-1-A94 (Figures 2b and 2c) shows that the NASC-GJ88 has the smoothest shale REE pattern. Thus, we recommend using NASC-GJ88 (with the appropriate modifications in Table 4), PAAS-P12, and BCR-1-A94, as reference values for these standards, and NASC-GJ88 as the preferred shale reference values.

## 5. Evaluation of Analytical Quality From Seawater Analyses

For seawater REE studies, the LDEO lab processes ~10 ml of seawater. Concentrations have ranged between 1–9 ppt (7–64 pmol/kg) for Nd, 0.2–2 ppt (1–14 pmol/kg) for Yb, and 0.3–3 ppt (2–21 pmol/kg) for Ce (Wu, 2019). Thus for the higher abundance REEs such as Nd, the total amount of element analyzed in a sample has ranged from ~10 to 90 pg, while for the lowest abundance REEs, Tm, the amount has ranged between ~0.5 and 5 pg. For quality control, the LDEO lab repeatedly analyzes seawater samples from a GEOTRACES intercalibration station Bermuda Atlantic Time Series (BATS; 31.7°N, 64.1°W) in the North Atlantic at 20 m ( $n = 18$ ) and 2,000 m ( $n = 16$ ) (Figures 3a–3d and Table S3). Internal measurement errors are <2%. As expected, the long-term uncertainty is much larger than internal measurement errors and is regarded as reflecting our real uncertainty associated with measuring REEs in natural seawater. By monitoring and reporting long-term external reproducibility, we ensure that the LDEO lab is consistently producing





**Figure 2.** Comparison of published REE abundances of USGS rock standard BCR-1 (a–c), North American Shale Composite (NASC) (a, c, d), and Post-Archean Australian Shale (PAAS) (b, d). (a) Relative REE abundances of BCR-1 values from Gladney et al. (1990), Asmerom et al. (1994), and Jochum et al. (2016) normalized to NASC values from Goldstein and Jacobsen (1988). BCR-1-A94 shows the smoothest pattern. (b, c) Comparison of PAAS values from Taylor and McLennan (1985) and Pourmand et al. (2012) to BCR-1-A94. PAAS-P12 shows the smoother pattern. (c) Comparison of NASC values from Gromet et al. (1984) and Goldstein and Jacobsen (1988) to BCR-1-A94. NASC-GJ88 shows the smoother pattern. (d) Comparison of NASC and PAAS reference values, those showing smoother patterns relative to BCR-1 (NASC-GJ88 and PAAS-P12) also show smoother patterns when the four sets of reference values are compared. Thus, we recommend using BCR-1-A94, PAAS-P12, and NASC-GJ88, and between the shale normalizations, we recommend NASC-GJ88. Values and citations are listed in Table 4. For NASC-GJ88, Lu uses the updated value in Piegras and Jacobsen (1992), and the mono-isotopic element abundances are courtesy of S. B. Jacobsen (personal communication, 2020).

high-quality data and that we realistically estimate the uncertainties that are achieved with the current technological setup.

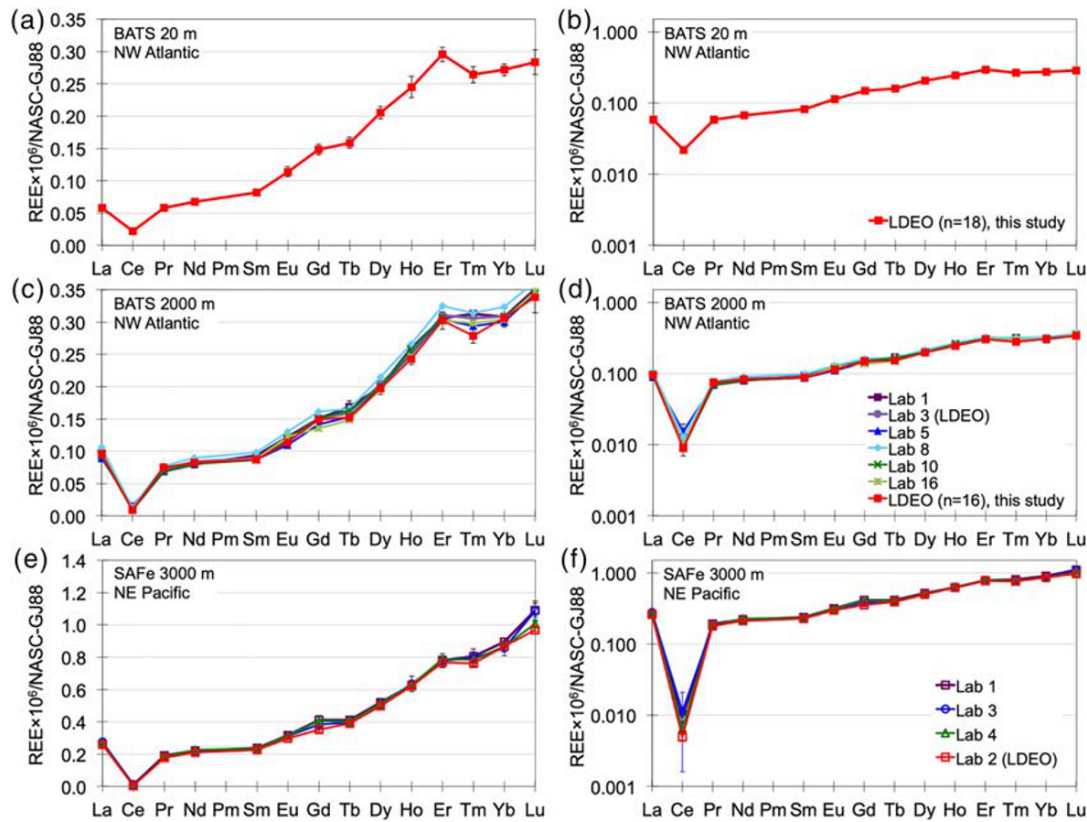
### 5.1. Repeated Analyses of BATS 20 and 2,000 m Seawater

For the BATS 20 and 2,000 m samples, long-term external reproducibilities ( $2\sigma$  RSD%) range from 1.5% to 6.2%, depending on the element, using a two standard deviation filter on the data. Those elements showing small errors of <2% to ~5% include high (10–23 pmol/kg, for La, Ce, Nd), intermediate (3–6 pmol/kg, for Pr in the 2,000 m sample, Sm, Dy, Er, Yb), and low (0.7–0.8 pmol/kg, for Ho in the 2,000 m sample, Tb, Tm) abundance elements; this includes elements treated as mono-isotopic (La, Pr, Tb, Ho, Tm) and as ID elements (the rest). Those showing higher errors of 5–6% include both intermediate (3–5 pmol/kg for Pr in the 20 m sample, Gd) and low (Eu, Ho in the 20 m sample, Lu; 0.7–1.5 pmol/kg) abundance elements, and also elements treated as mono-isotopic (Pr, Tb, Ho, Tm) and as ID elements (the rest).

We also monitor the long-term reproducibilities of REE ratios of the BATS 20 and 2,000 m seawater samples to evaluate the quality of our analysis. The long-term external errors of La/Nd, Pr/Nd, Sm/Nd, and Yb/Nd range from 4–5% ( $2\sigma$  RSD) for the BATS 20 m sample and 2–5% ( $2\sigma$  RSD) for the BATS 2,000 m sample. The errors for REE ratios are not systematically higher or lower than the errors of the individual elements (Table S3), indicating that these are random rather than systematic errors.

### 5.2. BATS and SAFe Seawater Intercalibrations

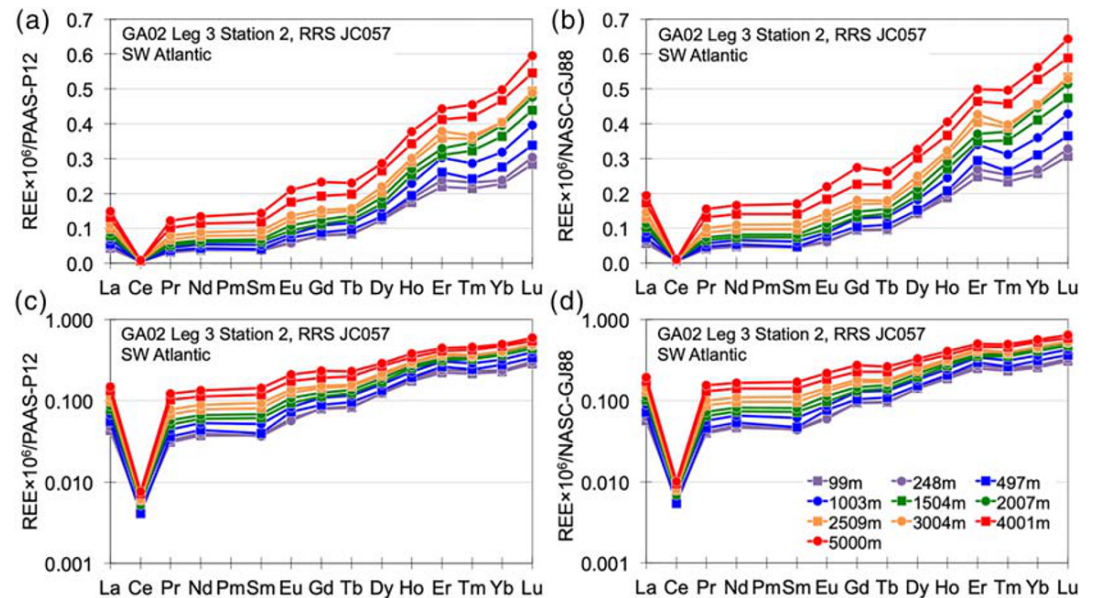
The LDEO lab participated in the international intercalibration of seawater REEs from the BATS GEOTRACES station (van de Flierdt et al., 2012) that included a sample from 2,000 m. In that study, seven labs contributed, and LDEO is Lab 3 (Figures 3c and 3d and Table S3). Overall agreement for six of the seven participants (one lab's results were clear outliers) ranged from 9–12% ( $2\sigma$  RSD) for La, Pr, Sm, Eu, Gd, and Tb



**Figure 3.** Average seawater REE concentrations from the BATS (Bermuda Atlantic Time Series) GEOTRACES intercalibration station (31.7°N, 64.1°W) at 20 and 2,000 m and the SAFe (Sampling and Analysis of Fe) GEOTRACES intercomparison station (30°N, 140°W) at 3,000 m, normalized to NASC-GJ88 (values from Goldstein & Jacobsen, 1988, with updates as explained in Table 4 note), on linear (a, c, e) and logarithmic (b, d, f) scales. For quality control, the LDEO lab repeatedly analyzes these BATS samples. (a, b) Average REEs for BATS 20 m sample ( $n = 18$ ) in this study. (c, d) REEs for BATS 2,000 m ( $n = 16$ ) in this study compared with the GEOTRACES intercalibration study (van de Flierdt et al., 2012), in which LDEO contributed data as Lab 3. (e, f) REEs for SAFe 3,000 m from the GEOTRACES intercomparison study (Behrens et al., 2016), in which LDEO contributed data as Lab 2. The error bars represent absolute two standard deviations from the mean values.

and 2–5% for all the others with the exception of Ce, where the  $2\sigma$  RSD was 44%. The range of concentrations for the group showing better agreement with 2–5%  $2\sigma$  RSD (0.8–17 pmol/kg) and worse agreement with 9–12%  $2\sigma$  RSD (0.8–24 pmol/kg) is similar, and both groups include spiked elements and those treated as mono-isotopic elements. The higher error for Ce occurred despite a moderately high concentration of ~5 pmol/kg. A potential reason for the higher error is a higher Ce blank level compared to the other REEs (4–10%, van de Flierdt et al., 2012). Another possibility could be a sample storage artifact resulting from variable Ce oxidation in the samples of different labs. For that study LDEO contributed two analyses of the REEs from 50 to 150 ml seawater samples. Our deviations from the average values are 3–7% for La, Ce, and Eu (25, 5, and 0.9 pmol/kg, respectively) and 0–2% for all other REEs (0.7–18 pmol/kg). It is important to note that a deviation from the average does not mean that our value is less correct. In that study, our lab was the only one that used a multielement spike.

The LDEO lab also participated in a later international intercomparison of seawater REEs from GEOTRACES station Sampling and Analysis of Fe (SAFe) in the North Pacific (30°N, 140°W) at 3,000 m (Behrens et al., 2016). In that study, four labs contributed, and ours is Lab 2 (Figures 3e and 3f and Table S3). Overall agreement ranged from 2–5%  $2\sigma$  RSD for all REEs (2–15 pmol/kg) except for La, Pr, Nd (6–7%  $2\sigma$  RSD; 10–65 pmol/kg), Gd and Lu (12–14%  $2\sigma$  RSD; 13 and 2 pmol/kg, respectively), and Ce (71%  $2\sigma$  RSD; ~4 pmol/kg). Our deviations from the average values are 1–4% for all REEs except for Gd and Lu (7–9%) and Ce (38%). In that study, our lab and the University of Oldenburg used different splits of the same (Stony Brook-DKM) spike. A third lab used a spike that has  $^{150}\text{Nd}$ ,  $^{172}\text{Yb}$ , indium, and rhenium. The fourth lab only used external standards.

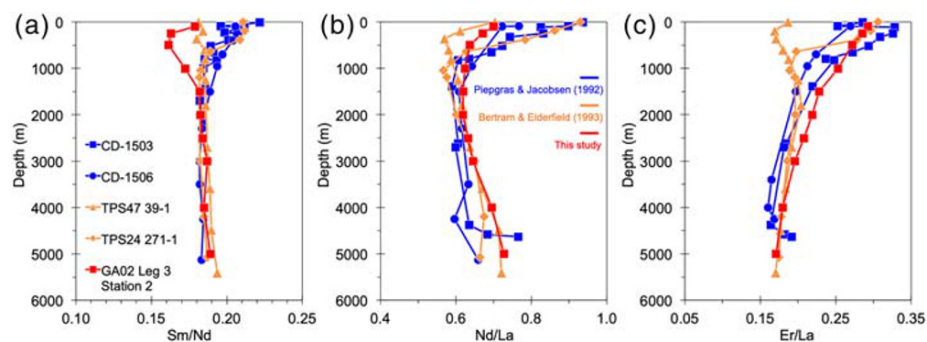


**Figure 4.** An example of REE results for seawater samples calculated using the REE Workbook. These seawater samples were collected between 99 and 5,000 m water depths from Station 2 (49.0°S, 48.9°W) of the GEOTRACES Southwest Atlantic Meridional Transect (RRS James Cook JC057, GA02 Leg 3) (Wu, 2019). (a–d) Relative abundances of REEs normalized to PAAS-P12 (Pourmand et al., 2012) (a, c) and NASC-GJ88 (Goldstein & Jacobsen, 1988) (b, d) on linear scales (a, b) and logarithmic scales (c, d). The depth profiles show typical seawater REE patterns, including negative Ce anomalies, HREE/LREE enrichment, and higher concentrations with increasing depth. Many of the samples clearly show positive deviations for Gd and Er.

### 5.3. A Seawater Station Depth Profile and Gd and Er Enrichments

The quality of the LDEO seawater results is illustrated in a depth profile (Figure 4 and Table S4) from the Southwest Atlantic (RRS James Cook JC057, GEOTRACES GA02 Leg 3, Station 2, 49.0°S, 48.9°W). The shale-normalized REE patterns are typical for seawater, with negative Ce anomalies and HREE/LREE enrichments. The absolute concentrations and the HREE/LREE enrichments both increase with increasing depth, which are also typical for seawater depth profiles. Scientific interpretations of the data are discussed in Wu (2019). We compare these results with two classic seawater REE studies that used multielement REE spikes and high-precision TIMS measurements (Bertram & Elderfield, 1993; Piepgras & Jacobsen, 1992) (Figure 5). The uncertainties reported by these studies are the internal measurement errors, better than 1% ( $2\sigma$ ) in Piepgras and Jacobsen (1992), and better than 1% in Bertram and Elderfield (1993) except for La (2.8%), Yb (3.4%), and Lu (1.6%) (they did not report whether the uncertainty estimates were at  $1\sigma$  or  $2\sigma$ ). Their high-precision results show smooth patterns of REE ratios such as Sm/Nd, Nd/La, and Er/La in depth profiles (Figure 5) that indicated much better data quality than previous studies that used neutron activation, where the REE ratio depth profiles show much more scattered patterns (Piepgras & Jacobsen, 1992, and Figure S1). Our results also show smooth patterns of REE ratios in depth profiles (Figure 5), indicating data quality comparable to theirs. The multielement spike ID approach thus yields high-quality data on both TIMS and quadrupole ICP-MS, with ICP-MS allowing for much faster analysis and the opportunity to analyze all the REEs at once.

A noteworthy characteristic of the Southwest Atlantic REE patterns is that they clearly show positive deviations at Gd and Er when normalized to shales (Figure 4), and although partly obscured by the Eu anomaly, they are also clearly present when normalized to BCR-1 (Figure S2). The Gd and Er positive deviations have been discussed as present in seawater REE patterns (e.g., de Baar, Brewer, et al., 1985; Kawabe et al., 1998; Masuda & Ikeuchi, 1979). Deviations of Gd and Er in particular from the otherwise smooth changes expected in REE behavior are consistent with “lanthanide tetrad effects,” resulting from increased stability at 1/4 (between Nd and Pm), 1/2 (Gd), and 3/4 (between Ho and Er) filled 4f electron orbitals (e.g., Kawabe, 1992). The tetrad effect divides the REEs into four tetrad groups (first: La, Ce, Pr, Nd; second: Pm, Sm, Eu, Gd; third: Gd, Tb, Dy, Ho; fourth: Er, Tm, Yb, Lu). Because Pm is absent in nature, it is



**Figure 5.** Depth profiles of some REE ratios for seawater samples collected from the GEOTRACES Southwest Atlantic Meridional Transect (RRS James Cook JC057, GA02 Leg 3, Station 2, 49.0°S, 48.9°W) (Table S4 and Wu, 2019) (red) measured by ICP-MS with a quadrupole mass analyzer, compared with the North Pacific seawater samples from Piegras and Jacobsen (1992) at stations TPS47 39-1 and TPS24 271-1 (orange) and the western Indian Ocean seawater samples from Bertram and Elderfield (1993) at stations CD-1503 and CD-1506 (blue), both measured by high-precision ID-TIMS. (a) Sm/Nd depth profile. (b) Nd/La depth profile. (c) Er/La depth profile. In all three studies, these ratios vary smoothly with depth (in contrast to other approaches, Figure S1), confirming the quality of the quadrupole ICP-MS data for spiked (Sm, Nd) and unspiked (La) elements.

difficult to investigate the first tetrad effect in seawater REE patterns. The identification of both the Gd and Er positive deviations benefit from measurement of the mono-isotopic elements Tb, Ho, and Tm (Figure 4), which are not an option using TIMS but are afforded by ICP-MS analysis. While the Er anomalies are recognizable in the Piegras and Jacobsen (1992) and Bertram and Elderfield (1993) TIMS data, with the neighboring mono-isotopic elements Ho and Tm missing, they require a four-element interpolation between Dy and Yb, making them less clear, while the ICP-MS data allow comparison with Ho and Tm. For identification of the Gd deviations, the ability to use the neighboring element mono-isotopic element Tb is critical, allowing for a three-element interpolation using Sm, as neighboring Eu cannot be used due to the possibility of a Eu anomaly.

## 6. Summary

This Technical Report contributes and explains an REE Calculation Workbook, in MS Excel, that reduces REE data for multielement spiked samples measured by ICP-MS, here offered for use by the community. The Report also compares and evaluates the commonly used reference standards BCR-1, PAAS, and NASC. The Workbook can be easily tailored for the needs of individual labs. It provides a convenient means for determining concentrations of all of the REEs in natural samples analyzed by ICP-MS, based on ID analysis for the spiked elements, and using ID elements as references for mono-isotopic elements and any multi-isotope elements treated as mono-isotopic elements. The data for the Stony Brook-DKM spike used at LDEO are in the REE Workbook and can be modified with concentrations and isotopic compositions of those spikes used in other labs. We show that the use of a multi-REE spike combined with ICP-MS analysis is a means to efficiently generate high-quality REE data, comparable to high-precision ID-TIMS, in about 10 min of analytical time rather than a couple of days, and including the unspiked elements. Thus, this Technical Report provides a framework to aid and encourage laboratories to adopt the multispike ID technique to measure REE concentrations by ICP-MS in natural samples.

## Data Availability Statement

Data in Figures 4 and 5 and Table S4 are also available through Wu (2019).

## References

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## Acknowledgments

This paper began as a short technical brief to contribute a Workbook to deconvolve the raw ICP-MS REE data, and we thank the reviewers Francis Albarède, Stein B. Jacobsen, and two other anonymous reviewers for their thorough reviews that have led to a much more comprehensive contribution. We thank reviewer S. B. Jacobsen for providing the mono-isotopic element data of NASC measured at Harvard and for suggesting that we might include it here. Interesting email discussions that included G. Hanson, M. F. Horan, J. A. Hurowitz, S. M. McLennan, E. T. Rasbury, S. B. Shirey, R. D. Vocke, and R. J. Walker illuminated the background and the calibration history of the Stony Brook multielement REE spike. We acknowledge support by NSF OCE-12-34687, NSF OCE-12-60514, and NSF OCE-14-59716 (National Science Foundation, USA), to S. L. G. and L. D. P. L. D. P. acknowledges support from Grant No. CTM2016-75411-R (MINECO, Spain). This is LDEO contribution number 8441.

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