

1 **Lignan exposure: A worldwide perspective.**

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18 **Keywords:** lignan, intake, biomarker, urine, plasma, serum, enterolignan

19 **Abbreviations:** 24-HDR, 24-h dietary recall; END, enterodiol; ENL, enterolactone; EPIC,
20 European Prospective Investigation into Cancer and Nutrition; FFQ, food frequency
21 questionnaire; LARI, lariciresinol; MATA, matairesinol; MEDI, medioresinol; PINO,
22 pinoresinol; SECO, secoisolariciresinol; SYRI, syringaresinol.

23 **ABSTRACT:**

24 Dietary lignans are phytoestrogens that are mostly found in plant-based foods, especially
25 whole grains, seeds, nuts, legumes and vegetables. An accurate assessment of lignan
26 exposure is crucial to evaluate their potential health benefits and to establish future
27 recommendations and dietary guidelines. This narrative review aimed to: (i) summarize the
28 pros and the cons of the current main assessment methods for lignan exposure —*i.e.*, dietary
29 questionnaires, food composition tables and biomarkers—, (ii) describe the individual
30 lignans more consumed from a worldwide perspective, as well as their main food sources,
31 (iii) determine the lignans concentrations in both urine and blood, and explore their
32 heterogeneity among countries, and finally (iv) discuss the main determinants of lignan
33 exposure.

34 INTRODUCTION

35 Chemistry and bioavailability

36 Lignans are secondary plant metabolites widely distributed in many plant-derived foods,
37 such as whole grains, seeds, nuts, legumes, vegetables, and drinks (*e.g.*, tea, coffee, or
38 wine) [1]. Lignans are bioactive compounds well-known by their ability to mimic or
39 modulate the action of endogenous estrogens [2]. Thus, they have been suggested to play a
40 role in the prevention of several chronic and hormone-related diseases such as
41 cardiovascular disease [1, 3], breast cancer [4, 5], osteoporosis [6], and menopausal
42 symptoms [7, 8]. Lignans are chemically polyphenolic compounds derived from two β - β -
43 linked phenylpropane (C6-C3) units. Based on the way in which oxygen is incorporated
44 into the skeleton and cyclization patterns, they can be classified into 8 subgroups: furans,
45 furofurans, dibenzylbutanes, dibenzylbutyrolactones, dibenzocyclooctadienes,
46 dibenzylbutyrolactols, aryltetralins, and aryl-naphthalenes. The most common lignans
47 consumed and for which the evidence has shown the most compelling benefits for health
48 are secoisolariciresinol (SECO), lariciresinol (LARI), pinoresinol (PINO), matairesinol
49 (MATA); although other lignans are also frequently consumed [*e.g.*, sesamol, sesamin,
50 syringaresinol (SYRI) and medioresinol (MEDI)] [9].

51 In nature, lignans are generally linked to other molecules, mainly as glycosylated
52 derivatives [10]. Lignan glycosides are absorbed in the gastrointestinal tract after being
53 metabolized by gut mucosa and/or colonic microbiota into lignan aglycones and further
54 converted into enterolignans [*i.e.*, enterolactone (ENL) and enterodiol (END)] [1, 11]. The
55 efficacy of this conversion depends on several factors, especially on the microbiota

56 composition and function, and differs considerably among individuals. In an *in vitro* fecal
57 microbiota metabolism system, 100% of LARI, 72% of SECO and 55% of PINO were
58 converted to END; while approximately half of END and 62% of MATA were transformed
59 to ENL [12]. Enterolignans, also called mammalian lignans, are efficiently absorbed and
60 conjugated to glucuronide and/or sulfates by enterocytes. Finally, enterolignans are
61 detected in blood (8-10h half-life) and excreted 30% through urine (residence time
62 approximately 24h) and 50% via enterohepatic circulation and feces [11]. Only small
63 amounts of LARI, MATA, PINO, SECO, and SYRI have been found in blood and urine
64 [13] (Figure 1).

65 In plant-derived foods, the richest sources of lignans are sesame seed oil (1,294 mg/100g),
66 flaxseed meal (867 mg/100g), and sesame seed meal (776 mg/100g), followed to a lesser
67 extent by whole grains and virgin olive oil (< 5mg/100g). The lignan content of other or
68 plant-derived foods is generally minimal with concentrations lower than 1 mg/100g [14].
69 Similarly, only negligible amounts of enterolignans have been detected in specific animal
70 foods (*i.e.*, milk, eggs, and derived products), which are produced by the intestinal bacterial
71 metabolism in the animals' guts after eating a diet rich in lignans [15]. A list of the top 25
72 richest foods of the main 6 individual lignans is shown in the Supplementary table 1.

73 **Exposure assessment**

74 In nutritional studies, lignan exposure has been assessed using either dietary questionnaires
75 or nutritional biomarkers. Both methodologies have advantages and disadvantages. On one
76 hand, dietary questionnaires [*e.g.*, food frequency questionnaires (FFQ), 24-h dietary
77 recalls (24-HDR), and food diaries] are inexpensive, easy to administer and can estimate a

78 lot of dietary data simultaneously, including dietary patterns, foods, nutrients and non-
79 nutrients [16]. On the other hand, dietary questionnaires are susceptible to random and
80 systematic reporting errors since they are based on subjects' memory and their ability to
81 estimate food portion sizes. Moreover, a food composition database is needed to convert
82 food consumption into lignan intake. Phenol-Explorer [17] is the most comprehensive
83 database on polyphenols that include all individual lignans (n~30) present in habitual foods.
84 Other studies have used other food composition databases from Canada [18], the
85 Netherlands [19], UK [20-22] and Finland [23]; although these only usually include the
86 four main individual lignans. The main limitations of using these databases are the large
87 amount of unknown values, the limited quantity of food items included, and the absence of
88 composition data on cooked foods. Thus, the estimation of lignan intake may be inaccurate
89 and tends to be underestimated. To improve accuracy of self-reported dietary estimates,
90 researchers are using new technologies, which are practical, have lower costs and burden
91 for both researchers and participants (*e.g.*, mobile phone applications) [24]. Moreover, they
92 are using databases that are regularly updated, allowing to increase the number of available
93 foods and individual lignans.

94 Nutritional biomarkers have become an alternative or complementary method for
95 estimating dietary intake. An ideal dietary biomarker would accurately reflect its dietary
96 intake and be specific, sensitive, and applicable to many populations. Their main advantage
97 is that they are objective, take into account bioavailability, and offer more accurate
98 assessment since they do not rely on subject's memory. In contrast, their disadvantages
99 include the requirement of biological samples, the complexity of the analytical
100 methodology, and the elevated cost [25]. During the last two decades, lignans and

101 especially enterolignans have been measured in blood and urine samples as potential
102 biomarkers of dietary lignans. Currently, the analytical method generally used is liquid
103 chromatography coupled to a tandem mass spectrometer (LC-MS/MS); although gas
104 chromatography GC-MS and time-resolved fluorescence immunoassay have also been
105 successfully used. These analytical methodologies allow us to have limits of detections
106 below 0.1 mg/L [26].

107 Concentrations of enterolignans in plasma and urine have been extensively investigated as
108 potential biomarkers of dietary lignan intakes. In a pooled analysis, urinary ENL levels
109 have been highly correlated with MATA and SECO intake ($r=0.78$), but not urinary END
110 ($r=-0.14$) [27]. However, in individual studies, correlations between lignan intake (sum of
111 MATA and SECO) and urinary enterolignans (sum of ENL and END) were moderate
112 ($r=0.40-0.46$) in 26 Canadian women [28] and low ($r=0.16-0.25$) in 195 adults from the
113 California Teachers Study [29]. Weak associations between lignan intake and plasma END
114 ($r=0.09$) and ENL ($r=0.18$) were observed in a Dutch study [30]. Similarly, correlations
115 between lignan intake and sum of plasma/serum enterolignans were low ($r=0.1-0.22$) [31].
116 These low correlations could be due to the constraints to accurately assess dietary lignan
117 intake (such as the aforementioned limitations of dietary questionnaires and food
118 composition databases) or to difficulties to analyze the lignan content in foods, particularly
119 in the extraction since they are usually bounded to dietary fiber [32]. It is also probable that
120 a low correlation may exist due to the high inter- and intra-individuality in the absorption,
121 metabolism and excretion of lignans or in the average lifetime of enterolignans in
122 biospecimens (plasma and urine) [11]. Despite of these results, concentrations of

123 enterolignans, especially in urine, are considered suitable and reliable alternative
124 measurements of lignan exposure.

125 **WORLDWIDE DIETARY LIGNAN INTAKE**

126 **Geographical differences in the intake of lignans and their food sources**

127 Due to differences in dietary patterns worldwide, lignan intakes vary considerably by
128 geographical region, with mean intakes mostly ranging from 0.2 to 6.4 mg/d in adults
129 (Table 1, Figure 2) [9, 33]. It is important to highlight that comparing results and estimates
130 across studies presents several challenges due to differences in the amount of individual
131 lignans included, and both the composition database and the dietary assessment method
132 used. However, some studies used similar methodologies that allow us to compare results
133 more easily.

134 *Europe*

135 Europe is the continent with more studies estimating the intake of lignans (Table 1). In
136 adults, the mean intake ranged from 0.2 mg/d to 5.2 in France [34] and Latvia [35],
137 respectively. Unsurprisingly, the highest intake of lignans (9.1 mg/d) was reached in a
138 vegetarian/vegan UK population, since lignan is almost exclusively found in plant-based
139 foods [9]. Despite the differences between studies, the existence of large multi-center
140 studies such as the European Prospective Investigation into Cancer and Nutrition (EPIC)
141 and the Healthy Lifestyle in Europe by Nutrition in Adolescents (HELENA) allows to
142 compare lignan intakes across Europe using the same methodology [9, 36, 37]. Data from
143 the EPIC study, that used Phenol-Explorer database, indicates that Mediterranean countries

144 have a higher intake than the non-Mediterranean ones [9, 36]. However, the HELENA
145 study, which used the Dutch database, showed a small decreasing north-to-south gradient
146 [37].

147 Data from studies using different methodology and databases indicates that the highest
148 lignan intake in Europe usually occurs in northern countries, including Scandinavian and
149 Baltic countries (Table 1). Considering the assessment of at least 6 individual lignans
150 (LARI, MATA, PINO, SECO, SYRI, and MEDI), the average of overall lignan intake
151 ranged between 2.3 mg/d and 5.2 mg/d. Intake estimates were lower (0.9-1.8 mg/d) if only
152 LARI, MATA, PINO, and SECO were considered. LARI, PINO and SECO were usually
153 the individual lignans more consumed, although SYRI was also common. The main food
154 sources of lignans in this region were whole grain cereals (especially rye, oat, and wheat),
155 bread, flaxseeds, and berries.

156 The mean intake of lignans in Central European countries, such as UK, Poland, Germany,
157 and the Netherlands, ranged between 0.6 [38] and 2.3 mg/d [9]. Most of the studies in this
158 region only assessed LARI, MATA, PINO, and SECO, and therefore, the intakes may be
159 slightly underestimated. In a Polish study [39] the mean intake of lignans was extremely
160 high (12.1 mg/day) due to a Phenol-Explorer error in the lignans content of some specific
161 vegetables [17] that were the main food sources in this Polish study (such as cucumber). In
162 Central European countries, LARI, PINO and SECO were the main individual lignans
163 consumed. Bread, seeds, and vegetables were the most common food sources of lignans in
164 this region.

165 Lastly, southern European countries, also referred as Mediterranean countries, had a highly
166 variable intake, ranging from 0.2 mg/day in France [36] to 4.3 mg/day in Greece [9].
167 France and Spain had relatively low intakes (0.2-2.1 mg/d), while Italy and Greece
168 generally had a high consumption (0.7-4.3 mg/d) [9, 36]. In an Italian study [40] the mean
169 intake was extremely high (80 mg/d). Although the authors did not provide any rationale
170 for such results, it is possible that this could be due to a processing error in the Eurofir-
171 eBASIS food composition database [41]. LARI, PINO and SECO were also the most
172 consumed individual lignans in this region; although depending on the study, the
173 proportions largely vary. These countries typically follow a Mediterranean dietary pattern,
174 where the main food sources of lignans are derived from olive oil, vegetables, fruits (mostly
175 citrus fruit), wine (predominantly red wine) and in a minor percentage bread and cereal
176 products.

177 *Americas*

178 In the US, there is also a great quantity of studies describing the lignan intake (Table 1).
179 Most of these studies used the Canadian database [18] which only contains data on the four
180 traditional individual lignans: LARI, MATA, PINO, and SECO. The mean intake of total
181 lignans ranged between 0.1 and 6.4 mg/d [42, 43] although in the majority of these studies,
182 their intake was <1mg/d. In this region the main food sources were tea and coffee, probably
183 due to a lower consumption of fruits, vegetables and whole grains compared to Europe. In
184 the US, SECO was clearly the most consumed individual lignan, followed by far by LARI
185 and PINO. In two Canadian studies, the intake of total lignans was slightly lower than in
186 the US, ranging from 0.2 to 0.4 mg/d [44, 45] and the main food sources were legumes,

187 seeds, cereals and grains, and berries. To date, only SECO and MATA were assessed in
188 Canada, which clearly underestimate total lignan intake.

189 To our knowledge, the existing data in Latin-American countries is limited to Mexico [33,
190 46] and Brazil [47-49]. The mean intake of total lignans was similar in both countries,
191 varying from 0.1 to 2.3 mg/d. A Brazilian study [47] was not included in the current
192 review, since its mean intake was exceptionally high 13.6 mg/d, possibly due to an error in
193 data calculation. As in Europe, SECO, LARI and PINO were the main contributors to total
194 lignans in this region. Main food sources were generally vegetables, fruits, nuts, seeds and
195 vegetable oils. However, there is a potential underestimation of lignan intakes in Latin
196 American countries due to the limited food composition data on some tropical foods [33],
197 such as mamey, zapote, papaya, sweet potato, nopal, guava, jicama, and prickly pears.
198 Those are frequently consumed in this region, but their lignan content is not available in
199 any food composition database yet.

200 *Other continents*

201 In Australia, two studies estimated the intake of total lignans in women only [50, 51]. Their
202 mean intake ranged from 0.7 to 2.7 mg/d. SECO was the major individual lignan consumed
203 and the main food sources were soy and linseed [51].

204 In Asian countries, lignan intake was estimated only in two Iranian-based [52, 53] and one
205 Korean-based [54] studies. In Iran, the mean intake of total lignans, including all individual
206 lignans, varied between 0.2 mg/d and 2.4 mg/d; whereas in Korea, including only MAT and
207 SECO, the mean intake was 1.5-1.8 mg/d. Data on main food sources was not available in
208 this region.

209 **Determinants of lignan intake**

210 Lignans were positively correlated to total energy intake [55]; therefore, participants
211 consuming more energy were more likely to be those with a higher intake of total lignans.
212 Although a Latvian study [35] showed a greater consumption of total lignans in men
213 compared to women; data from EPIC showed that women had a higher intake of lignans
214 after adjusting for total energy consumption (3.6 mg/d in women vs. 2.5 mg/d in men) [9].
215 Interestingly, one Korean study [54] observed slight differences between menopausal
216 statuses in women (1.8 mg/d in postmenopausal women vs. 1.5 mg/d in premenopausal
217 women). In the EPIC study [9], results indicated that lignan intake also increased with age.
218 For instance, young adults (35-44 years) had a lower intake of total lignans (2.8 mg/d) than
219 older adults (65-74 years; 3.5mg/d) [9]. In children and adolescents, the two available
220 European studies [36, 56] found that the mean intake was higher in adolescents (15-18
221 years) than in children (2-15 years), 0.98 - 1.10 mg/d vs. 0.61 - 1.00 mg/d, respectively.

222 The results by lifestyle factors and other sociodemographic variables are controversial. For
223 example, some studies showed that subjects with obesity had a higher intake of lignans [9,
224 36, 45, 57, 58-60] than individuals with normal weight; whereas in other studies occurred
225 the opposite [9, 35, 61-63]. Discrepancies were also observed comparing lignan intake by
226 educational level, smoking status, physical activity, and alcohol consumption.

227 **WORLDWIDE ENTEROLIGNANS CONCENTRATIONS**

228 **Geographical differences in total enterolignans concentrations**

229 Concentrations of lignan metabolites (END and ENL) in biospecimens, as potential
230 biomarkers of lignan intake, are useful indicators of lignan exposures across populations. In
231 order to straightforwardly compare concentrations of enterolignans, all estimates have been
232 converted into the same units (nmol/L) in Tables 2 and 3. These summarize the most
233 representative studies assessing urinary and blood (*i.e.*, serum or plasma) enterolignan
234 concentrations, respectively. Levels of urinary enterolignans were usually 100-fold higher
235 than those found in blood (serum or plasma). The mean urinary END concentrations
236 worldwide ranged from 38 [64] to 763 nmol/L [65] and for ENL from 148 [66] to 3,651
237 nmol/L [67] (Table 2, Figure 3). In the case of plasma and serum, END concentrations
238 varied between 0.2 nmol/L [68] and 7.0 nmol/L [69] while ENL levels ranged from 4.9
239 nmol/L [68] to 39.2 nmol/L [69]. Levels of enterolignans in plasma and serum were similar
240 (Table 3, Figure 3). Mean concentrations of END were between 2 to 13 times lower than
241 ENL in both urine and blood.

242 *Europe*

243 Few studies (n=8) have measured urinary enterolignans in Europe (Table 2). Northern
244 European countries tend to have the highest levels of enterolignans (ENL=768-3,267
245 nmol/L) [65, 70] followed by Central European countries (END= 204-288 and ENL=
246 2,414-3,333 nmol/L nmol/L) [71-75]. Data for Mediterranean countries were limited. There
247 is only one study from Italy, that reported a high urinary concentration (END=763 and
248 ENL=1,577 nmol/L) (76).

249 Most of the studies measuring enterolignan concentrations in blood specimens, of which 20
250 were conducted in plasma and 10 in serum, were performed in Central and Northern

251 European countries (Table 3). The lowest concentrations of END and ENL were 0.2
252 nmol/L and 4.9 nmol/L, respectively, in a UK-based study [68]; while the highest levels
253 were derived from a Dutch population: 7.0 nmol/L for END and 39.2 nmol/L for ENL [69].
254 Comparing studies that used the same analytical methodology, in general, concentrations in
255 Central European countries (*e.g.*, the Netherlands, Germany, UK) were slightly lower than
256 in Scandinavian countries [68, 77]. However, when all studies were considered
257 independently of lignan assessments, levels of enterolignans in central European countries
258 were very heterogeneous [68, 69]. The lowest mean enterolignan concentrations were
259 found in Mediterranean countries: 0.3 nmol/L for END and 6.7-7.8 nmol/L for ENL [77].
260 Italy was the Mediterranean country with the highest END (1.3 nmol/L) and ENL (9.1
261 nmol/L) concentrations in plasma [77], which is similar to intake estimations.

262 *Americas*

263 To our knowledge, only US data was available from both North and South American
264 continents, with the exception of a Jamaican study. In the US, several studies assessed
265 enterolignan concentrations in urine (n=15) (Table 2), plasma (n=2), and serum (n=2)
266 (Table 3). Both urinary END and ENL excretions varied considerably among US studies
267 from 38 [64] to 609 nmol/L [67] for END, and from 285 [64] to 3,651 nmol/L [67] for
268 ENL. Indeed, US populations included the worldwide minimum mean of END levels (285
269 nmol/L) and the worldwide maximum mean of ENL excretions (609 nmol/L). In the
270 Jamaican study, the mean intake of END was in the upper side of the interval of the US
271 studies (2,671 nmol/L) [78].

272 Similarly, a high variability in blood END levels was observed among US studies, ranging
273 between 1.5 nmol/L [79] and 6.0 nmol/L [80] while the range of mean levels for ENL was
274 narrower from 11.5 nmol/L [81] to 22.5 nmol/L [79].

275 *Asia*

276 To date, urinary concentrations of enterolignans in Asia were measured in Singapore [82],
277 Japan [66, 83], Vietnam [83], Cambodia [83] and India [83]. The mean of urinary END
278 concentrations varied from 60 nmol/L in Cambodia [83] to 245 nmol/L [83] in Vietnam.
279 For ENL, the highest mean value was found in Vietnam (1,678 nmol/L) [83] while the
280 lowest excretion was identified in a Japanese study (148 nmol/L) [66].

281 Several studies in East Asia (such as Japan, China, Korea and Vietnam) assessed
282 enterolignans in plasma and showed a relatively low variation in their mean concentrations
283 (~3-fold variation). Thus, END concentration means ranged from 2.0 nmol/L [84] to 5.6
284 [85] in the two Chinese studies. Mean ENL concentrations in blood samples were between
285 10.2 nmol/L [86] and 32.7 nmol/L [87] in Vietnam and Japan, respectively. In the study of
286 Liu et al. [84] median plasma concentrations of ENL (2.0 nmol/L) and END (16.4 nmol/L)
287 seem to be exchanged. Mean ENL concentrations in Korea were extremely high (177.8
288 nmol/L in women and 249.3 nmol/L in men), around 10-fold higher than values found in
289 any other study from other continents.

290 **Determinants of the total enterolignans concentrations**

291 Data from studies that analysed separately men and women showed that urinary
292 concentrations of enterolignans were slightly higher in women than in men [67, 70, 83],

293 with one exception [64]. Urinary ENL and END excretions were the highest in adults (20-
294 60 years), followed by the elderly (>60 years) and, finally, by adolescents (12-19 years)
295 [67]. This pattern according to age and sex is consistent with findings from dietary lignans
296 adjusted for energy intake. A Danish study suggested that smoking and higher BMI were
297 associated with lower concentrations of ENL [88]. No other information was found for
298 concentrations of entrolignans (in both urine and blood) and other determinants, such as
299 educational level and physical activity.

300 **STRENGTHS & LIMITATIONS**

301 *Dietary data*

302 The main limitation of this review was that each study used a different methodology to
303 estimate the lignan intake. Firstly, differences in both the type of dietary questionnaire
304 (FFQ, 24h dietary recall, history of diet) and the amount of food items included in the
305 questionnaire could complicate comparisons in the habitual estimation of individual foods,
306 particularly lignan-rich products. Although, the vast majority of studies used validated
307 FFQs; very few of these questionnaires were specifically validated for lignans. Secondly,
308 available food composition tables/databases were not complete. They have missing data on
309 several foods and, especially, on some individual lignans. Only Phenol-Explorer [17]
310 contains data on all commonly consumed lignans; while others only have data on two
311 (MATA and SECO) or four individual lignans (MATA, SECO, LARI, and PINO). These
312 four lignans are the most abundant ones accounting for at least 50% of total lignan intake in
313 Europe [9]. Thirdly, most of the presented studies were not representative of the entire
314 population, so the results may not be totally generalizable. However, the inclusion of

315 several medium-to-large size studies from the same geographical area enhances
316 generalizability. Fourth, studies evaluating reliability of enterolignans as biomarkers of
317 lignan intake are limited; especially those investigating all individual lignans, and
318 correlations were moderate for urinary concentrations [27-29] and low for plasma/serum
319 concentrations [31]. Therefore, inconsistent results have been observed comparing results
320 using dietary conventional dietary questionnaires and biomarkers. For example, a recent
321 meta-analysis showed no associations between dietary lignan intake and cancer outcomes;
322 while a higher concentration of serum/plasma ENL was inversely associated with overall
323 cancer survival [89].

324 *Biomarker data*

325 Variability in results due to differences in procedures and methods in the analysis of
326 concentrations of enterolignans in blood and urine were relatively minor, since all
327 analytical methodologies were validated. The main limitation was that the studies only
328 analyzed one sample per subject. It is well-known that enterolignans are relatively short-
329 term nutritional biomarkers [11] and therefore multiple measurements would be
330 recommended to estimate habitual exposure at individual level. However, the mean of a
331 single punctual measure in a large quantity of subjects was a suitable way to reflect the
332 habitual mean of lignan concentrations at population level. Another limitation was the
333 relatively small size of all studies and therefore the limited generalizability of the results.

334 **CONCLUSIONS**

335 Overall, common mean intakes of total lignans worldwide ranged from 1 to 5 mg/d, with a
336 higher intake in vegetarian populations (9.1 mg/d). There was a large heterogeneity in the

337 estimations of lignan intake across studies partially due to real differences among
338 geographical areas and populations and to differences between dietary assessment methods
339 used. Food sources also varied across regions, although the most typical ones were whole
340 grain cereal products, seeds, vegetables, and fruits.

341 As expected, similar trends and differences between regions were observed using dietary
342 and biomarker data. END concentrations were usually 10-fold lower than ENL levels in
343 both urine and blood. Results of enterolignans in plasma and serum were equivalent. END
344 and ENL concentrations in urine were approximately 100 times higher than in blood.

345 More food composition data is warranted in order to update current databases on lignans
346 and improve dietary intake estimations. Data from some regions, particularly in low- and
347 middle-income countries (Africa, Latin America, and some areas in Asia), was scarce or
348 null; therefore, further studies combining both dietary and biomarker data in these regions
349 are requested to improve data coverage globally.

350 Finally, an accurate estimation of lignan exposure is essential to better understand
351 associations between lignan intake and the risk of chronic diseases. In our opinion,
352 although, current estimations of dietary lignan intake are getting more precise, they are
353 often underestimated. Thus, concentrations of enterolignans in blood and urine are still
354 preferable to estimate lignan exposure in epidemiological studies. This data will be crucial
355 for setting and improving current dietary recommendations for populations.

356 **CONFLICT OF INTEREST**

357 The authors are not aware of any conflicts of interest.

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367 **REFERENCES**

- 368 1. Peterson J, Dwyer J, Adlercreutz H, Scalbert A, Jacques P, McCullough ML (2010)
369 Dietary lignans: physiology and potential for cardiovascular disease risk reduction.
370 *Nutr Rev* 68:571-603. doi: 10.1017/S0007114515005012
- 371 2. Kuiper GG, Lemmen JG, Carlsson B, Corton JC, Safe SH, van der Saag PT, van der
372 Burg B, Gustafsson JA (1998) Interaction of estrogenic chemicals and phytoestrogens
373 with estrogen receptor beta. *Endocrinology* 139:4252-4263.
- 374 3. Grosso G, Micek A, Godos J, Pajak A, Sciacca S, Galvano F, Giovannucci EL (2017)
375 Dietary flavonoid and lignan intake and mortality in prospective cohort studies:
376 Systematic review and dose-response meta-analysis. *Am J Epidemiol* 185(12):1304-
377 1316. doi: 10.1093/aje/kww207.
- 378 4. Touillaud MS, Thiébaud AC, Fournier A, Niravong M, Boutron-Ruault MC, Clavel-
379 Chapelon F (2007) Dietary lignan intake and postmenopausal breast cancer risk by
380 estrogen and progesterone receptor status. *J Natl Cancer Inst* 99(6):475-86. doi:
381 10.1093/jnci/djk096
- 382 5. Buja A, Pierbon M, Lago L, Grotto G, Baldo V (2020) Breast cancer primary
383 prevention and diet: An umbrella review. *Int J Environ Res Public Health* 17(13):4731.
384 doi: 10.3390/ijerph17134731
- 385 6. Ma ZP, Zhang ZF, Yang YF, Yang Y (2019) Sesamin promotes osteoblastic
386 differentiation and protects rats from osteoporosis. *Med Sci Monit* 25:5312-5320. doi:
387 10.12659/MSM.915529

- 388 7. Pruthi S, Qin R, Terstreip SA, et al (2012) A phase III, randomized, placebo-controlled,
389 double-blind trial of flaxseed for the treatment of hot flashes: North Central Cancer
390 Treatment Group N08C7. *Menopause* 19(1):48-53. doi:
391 10.1097/gme.0b013e318223b021
- 392 8. Adlercreutz H (2007) Lignans and human health. *Crit Rev Clin Lab Sci* 44:483-525.
- 393 9. Zamora-Ros R, Knaze V, Rothwell JA, et al (2016) Dietary polyphenol intake in
394 Europe: the European Prospective Investigation into Cancer and Nutrition (EPIC)
395 study. *Eur J Nutr* 55:1359-1375. doi: 10.1007/s00394-015-0950-x
- 396 10. Smeds AI, Eklund PC, Sjöholm RE, Willför SM, Nishibe S, Deyama T, Holmbom BR
397 (2007) Quantification of a broad spectrum of lignans in cereals, oilseeds, and nuts. *J*
398 *Agric Food Chem* 55:1337-1346.
- 399 11. Clavel T, Doré J, Blaut M (2006) Bioavailability of lignans in human subjects. *Nutr*
400 *Res Rev* 19:187-196. doi: 10.1017/S0954422407249704
- 401 12. Heinonen S, Nurmi T, Liukkonen K, Poutanen K, Wähälä K, Deyama T, Nishibe S,
402 Adlercreutz H (2001) In vitro metabolism of plant lignans: new precursors of
403 mammalian lignans enterolactone and enterodiol. *J Agric Food Chem* 49:3178-3186.
- 404 13. Nurmi T, Voutilainen S, Nyyssönen K, Adlercreutz H, Salonen JT (2003) Liquid
405 chromatography method for plant and mammalian lignans in human urine. *J*
406 *Chromatogr B Analyt Technol Biomed Life Sci* 798:101-110.
- 407 14. Pérez-Jiménez J, Neveu V, Vos F, Scalbert A (2010) Systematic analysis of the content
408 of 502 polyphenols in 452 foods and beverages: an application of the phenol-explorer
409 database. *J Agric Food Chem* 58:4959-4969. doi: 10.1021/jf100128b
- 410 15. Kuhnle GGC, Dell'Aquila C, Aspinall SM, Runswick SA, Mulligan AA, Bingham SA
411 (2008) Phytoestrogen content of foods of animal origin: dairy products, eggs, meat,
412 fish, and seafood. *J Agric Food Chem* 56:10099-10104. doi: 10.1021/jf801344x
- 413 16. Zamora-Ros R, Rabassa M, Llorach R, González CA, Andres-Lacueva C (2012)
414 Application of dietary phenolic biomarkers in epidemiology: past, present, and future. *J*
415 *Agric Food Chem* 60:6648-6657. doi: 10.1021/jf204742e
- 416 17. Neveu V, Perez-Jiménez J, Vos F, Crespy V, du Chaffaut L, Mennen L, Knox C, Eisner
417 R, Cruz J, Wishart D, Scalbert A (2010) Phenol-Explorer: an online comprehensive
418 database on polyphenol contents in foods. *Database (Oxford)* 2010:bap024. doi:
419 10.1093/database/bap024
- 420 18. Thompson LU, Boucher BA, Liu Z, Cotterchio M, Kreiger N (2006) Phytoestrogen
421 content of foods consumed in Canada, including isoflavones, lignans, and coumestan.
422 *Nutr Cancer* 54:184-201.

- 423 19. Milder IEJ, Arts ICW, van de Putte B, Venema DP, Hollman PCH (2005) Lignan
424 contents of Dutch plant foods: a database including lariciresinol, pinoresinol,
425 secoisolariciresinol and matairesinol. *Br J Nutr* 93:393-402.
- 426 20. Kuhnle GGC, Dell'Aquila C, Aspinall SM, Runswick SA, Joosen AMCP, Mulligan
427 AA, Bingham SA (2009) Phytoestrogen content of fruits and vegetables commonly
428 consumed in the UK based on LC-MS and ¹³C-labelled standards. *Food Chem*
429 116:542-554.
- 430 21. Kuhnle GGC, Dell'Aquila C, Aspinall SM, Runswick SA, Mulligan AA, Bingham SA
431 (2008) Phytoestrogen content of beverages, nuts, seeds, and oils. *J Agric Food Chem*
432 56:7311-7315. doi: 10.1021/jf801534g
- 433 22. Kuhnle GGC, Dell'aquila C, Aspinall SM, Runswick SA, Mulligan AA, Bingham SA
434 (2009) Phytoestrogen content of cereals and cereal-based foods consumed in the UK.
435 *Nutr Cancer* 61:302-309. doi: 10.1080/01635580802567141
- 436 23. Valsta LM, Kilkkinen A, Mazur W, Nurmi T, Lampi A-M, Ovaskainen M-L, Korhonen
437 T, Adlercreutz H, Pietinen P (2003) Phyto-oestrogen database of foods and average
438 intake in Finland. *Br J Nutr* 89 Suppl 1:S31-S38.
- 439 24. Illner A-K, Freisling H, Boeing H, Huybrechts I, Crispim SP, Slimani N (2012) Review
440 and evaluation of innovative technologies for measuring diet in nutritional
441 epidemiology. *Int J Epidemiol* 41:1187-1203. doi: 10.1093/ije/dys105
- 442 25. Potischman N (2003) Biologic and methodologic issues for nutritional biomarkers. *J*
443 *Nutr* 133 Suppl 3:875S-880S. doi: 10.1093/jn/133.3.875S
- 444 26. Peeters PHM, Slimani N, van der Schouw YT, et al (2007) Variations in plasma
445 phytoestrogen concentrations in European adults. *J Nutr* 137:1294-1300.
- 446 27. Pérez-Jiménez J, Hubert J, Hooper L, Cassidy A, Manach C, Williamson G, Scalbert A
447 (2010) Urinary metabolites as biomarkers of polyphenol intake in humans: a systematic
448 review. *Am J Clin Nutr* 92:801-809. doi: 10.3945/ajcn.2010.29924
- 449 28. French MR, Thompson LU, Hawker GA (2007) Validation of a phytoestrogen food
450 frequency questionnaire with urinary concentrations of isoflavones and lignan in
451 premenopausal women. *J Am Coll Nutr* 26(1):76-82. doi:
452 10.1080/07315724.2007.10719588
- 453 29. Horn-Ross PL, Barnes S, Lee VS, et al (2006) Reliability and validity of an assessment
454 of usual phytoestrogen consumption (United States). *Cancer Causes Control* 17(1):85-
455 93. doi: 10.1007/s10552-005-0391-6
- 456 30. Milder IEJ, Kuijsten A, Arts ICW, Feskens EJM, Kampman E, Hollman PC, Van 't
457 Veer P (2007) Relation between plasma enterodiol and enterolactone and dietary intake
458 of lignans in a Dutch endoscopy-based population. *J Nutr* 137:1266-1271.

- 459 31. Lin Y, Wolk A, Håkansson N, Peñalvo JL, Lagergren J, Adlercreutz H, Lu Y (2013)
 460 Validation of FFQ-based assessment of dietary lignans compared with serum
 461 enterolactone in Swedish women. *Br J Nutr* 109(10):1873-80. doi:
 462 10.1017/S000711451200387X
- 463 32. Liggins J, Grimwood R, Bingham SA (2000) Extraction and quantification of lignan
 464 phytoestrogens in food and human samples. *Anal Biochem* 287(1):102-9. doi:
 465 10.1006/abio.2000.4811
- 466 33. Zamora-Ros R, Biessy C, Rothwell JA, Monge A, Lajous M, Scalbert A, López-
 467 Ridaura R, Romieu I (2018) Dietary polyphenol intake and their major food sources in
 468 the Mexican Teachers' Cohort. *Br J Nutr* 120:353-360. doi:
 469 10.1017/S0007114518001381
- 470 34. Adriouch S, Kesse-Guyot E, Feuillet T, Touvier M, Olié V, Andreeva V, Hercberg S,
 471 Galan P, Fezeu LK (2018) Total and specific dietary polyphenol intakes and 6-year
 472 anthropometric changes in a middle-aged general population cohort. *Int J Obes (Lond)*
 473 42:310-317. doi: 10.1038/ijo.2017.227
- 474 35. Meija L, Söderholm P, Samaletdin A, Ignace G, Sikсна I, Joffe R, Lejnieks A,
 475 Lietuvietis V, Krams I, Adlercreutz H (2013) Dietary intake and major sources of plant
 476 lignans in Latvian men and women. *Int J Food Sci Nutr* 64:535-543. doi:
 477 10.3109/09637486.2013.765835
- 478 36. Wisnuwardani R, Henauw S, Androustos O, et al (2018) Estimated dietary intake of
 479 polyphenols in European adolescents: the HELENA study. *Eur J Nutr* 58:2345-2363.
 480 doi: 10.1007/s00394-018-1787-x
- 481 37. Tetens I, Turrini A, Tapanainen H, Christensen T, Lampe JW, Fagt S, Håkansson N,
 482 Lundquist A, Hallund J, Valsta LM (2013) Dietary intake and main sources of plant
 483 lignans in five European countries. *Food Nutr Res* 57. doi: 10.3402/fnr.v57i0.19805
- 484 38. Grosso G, Stepaniak U, Topor-Mądry R, Szafraniec K, Pająk A (2014) Estimated
 485 dietary intake and major food sources of polyphenols in the Polish arm of the HAPIEE
 486 study. *Nutrition* 30:1398-1403. doi: 10.1016/j.nut.2014.04.012
- 487 39. Witkowska AM, Zujko ME, Waśkiewicz A, Terlikowska KM, Piotrowski W (2015)
 488 Comparison of Various Databases for Estimation of Dietary Polyphenol Intake in the
 489 Population of Polish Adults. *Nutrients* 7:9299-9308. doi: 10.3390/nu7115464
- 490 40. Russo GI, Di Mauro M, Regis F, Reale G, Campisi D, Marranzano M, Lo Giudice A,
 491 Solinas T, Madonia M, Cimino S, Morgia G (2018) Association between dietary
 492 phytoestrogens intakes and prostate cancer risk in Sicily. *Aging Male* 21:48-54. doi:
 493 10.1080/13685538.2017.1365834
- 494 41. Plumb J, Pigat S, Bompola F, Cushen M, Pinchen H, Nørby E, Astley S, Lyons J, Kiely
 495 M, Finglas P (2017) eBASIS (Bioactive Substances in Food Information Systems) and
 496 Bioactive Intakes: Major Updates of the Bioactive Compound Composition and

- 497 Beneficial Bioeffects Database and the Development of a Probabilistic Model to Assess
498 Intakes in Europe. *Nutrients* 9:E320. doi: 10.3390/nu9040320
- 499 42. Carmichael SL, Cogswell ME, Ma C, Gonzalez-Feliciano A, Olney RS, Correa A,
500 Shaw GM (2013) Hypospadias and Maternal Intake of Phytoestrogens. *Am J Epidemiol*
501 178:434-440. doi: 10.1093/aje/kws591
- 502 43. Fink BN, Steck SE, Wolff MS, Kabat GC, Gammon MD (2006) Construction of a
503 flavonoid database for assessing intake in a population-based sample of women on
504 Long Island, New York. *Nutr Cancer* 56:57-66.
- 505 44. Cotterchio M, Boucher BA, Manno M, Gallinger S, Okey A, Harper P (2006) Dietary
506 Phytoestrogen Intake Is Associated with Reduced Colorectal Cancer Risk. *J Nutr*
507 136:3046-3053.
- 508 45. Morisset A-S, Lemieux S, Veilleux A, Bergeron J, John Weisnagel S, Tchernof A
509 (2009) Impact of a lignan-rich diet on adiposity and insulin sensitivity in post-
510 menopausal women. *Br J Nutr* 102:195-200. doi: 10.1017/S0007114508162092
- 511 46. Hernández-Ramírez RU, Galván-Portillo MV, Ward MH, Agudo A, González CA,
512 Oñate-Ocaña LF, Herrera-Goepfert R, Palma-Coca O, López-Carrillo L (2009) Dietary
513 intake of polyphenols, nitrate and nitrite and gastric cancer risk in Mexico City. *Int J*
514 *Cancer* 125:1424-1430. doi: 10.1002/ijc.24454
- 515 47. Nascimento-Souza MA, de Paiva PG, Pérez-Jiménez J, do Carmo Castro Franceschini
516 S, Ribeiro AQ (2018) Estimated dietary intake and major food sources of polyphenols
517 in elderly of Viçosa, Brazil: a population-based study. *Eur J Nutr* 57:617-627. doi:
518 10.1007/s00394-016-1348-0
- 519 48. Miranda AM, Steluti J, Fisberg RM, Marchioni DM (2016) Association between
520 Polyphenol Intake and Hypertension in Adults and Older Adults: A Population-Based
521 Study in Brazil. *PloS One* 11:e0165791. doi: 10.1371/journal.pone.0165791
- 522 49. Miranda AM, Steluti J, Fisberg RM, Marchioni DM (2016) Dietary intake and food
523 contributors of polyphenols in adults and elderly adults of Sao Paulo: a population-
524 based study. *Br J Nutr* 115:1061-1670. doi: 10.1017/S0007114515005061
- 525 50. Lahmann PH, Hughes MC, Ibiebele TI, Mulligan AA, Kuhnle GGC, Webb PM (2012)
526 Estimated intake of dietary phyto-oestrogens in Australian women and evaluation of
527 correlates of phyto-oestrogen intake. *J Nutr Sci* 1:e11. doi: 10.1017/jns.2012.11
- 528 51. Hanna KL, O'Neill S, Lyons-Wall PM (2010) Intake of isoflavone and lignan
529 phytoestrogens and associated demographic and lifestyle factors in older Australian
530 women. *Asia Pac J Clin Nutr* 19:540-549.
- 531 52. Sohrab G, Hosseinpour-Niazi S, Hejazi J, Yuzbashian E, Mirmiran P, Azizi F (213)
532 Dietary polyphenols and metabolic syndrome among Iranian adults. *Int J Food Sci Nutr*
533 64:661-667. doi: 10.3109/09637486.2013.787397

- 534 53. Sohrab G, Ebrahimof S, Hosseinpour-Niazi S, Yuzbashian E, Mirmiran P, Azizi F
535 (2018) Association of Dietary Intakes of Total Polyphenol and Its Subclasses with the
536 Risk of Metabolic Syndrome: Tehran Lipid and Glucose Study. *Metab Syndr Relat*
537 *Disord* 16:274-281. doi: 10.1089/met.2017.0140
- 538 54. Jang J-H, Yoon J-Y, Cho S-H (2007) Intake of dietary phytoestrogen and indices of
539 antioxidant and bone metabolism of pre- and post-menopausal Korean women. *Nutr*
540 *Res Pract* 1:305-312. doi: 10.4162/nrp.2007.1.4.30
- 541 55. Suzuki R, Rylander-Rudqvist T, Saji S, Bergkvist L, Adlercreutz H, Wolk A (2008)
542 Dietary lignans and postmenopausal breast cancer risk by oestrogen receptor status: a
543 prospective cohort study of Swedish women. *Br J Cancer* 98:636-640. doi:
544 10.1038/sj.bjc.6604175
- 545 56. Peñalvo JL, Moreno-Franco B, Ribas-Barba L, Serra-Majem L (2012) Determinants of
546 dietary lignan intake in a representative sample of young Spaniards: association with
547 lower obesity prevalence among boys but not girls. *Eur J Clin Nutr* 66:795-798. doi:
548 10.1038/ejcn.2012.45
- 549 57. Kilkinen A, Valsta LM, Virtamo J, Stumpf K, Adlercreutz H, Pietinen P (2003) Intake
550 of lignans is associated with serum enterolactone concentration in Finnish men and
551 women. *J Nutr* 133:1830-1833.
- 552 58. Pounis G, Di Castelnuovo A, Bonaccio M, Costanzo S, Persichillo M, Krogh V, Donati
553 MB, de Gaetano G, Iacoviello L (2016) Flavonoid and lignan intake in a Mediterranean
554 population: proposal for a holistic approach in polyphenol dietary analysis, the Moli-
555 sani Study. *Eur J Clin Nutr* 70:338-345. doi: 10.1038/ejcn.2015.178
- 556 59. Horn-Ross PL, John EM, Canchola AJ, Stewart SL, Lee MM (2003) Phytoestrogen
557 intake and endometrial cancer risk. *J Natl Cancer Inst* 95:1158-1164.
- 558 60. Schabath MB, Hernandez LM, Wu X, Pillow PC, Spitz MR (2005) Dietary
559 phytoestrogens and lung cancer risk. *JAMA* 294:1493-1504.
- 560 61. Milder IE, Feskens EJ, Arts IC, Bueno de Mesquita HB, Hollman PC, Kromhout D
561 (2005). Intake of the plant lignans secoisolariciresinol, matairesinol, lariciresinol, and
562 pinoresinol in Dutch men and women. *J Nutr* 135:1202-1207.
- 563 62. Hedelin M, Löf M, Andersson TM-L, Adlercreutz H, Weiderpass E (2011) Dietary
564 phytoestrogens and the risk of ovarian cancer in the women's lifestyle and health
565 cohort study. *Cancer Epidemiol Biomark Prev* 20:308-317. doi: 10.1158/1055-
566 9965.EPI-10-0752
- 567 63. Hedelin M, Löf M, Sandin S, Adami H-O, Weiderpass E (2016) Prospective Study of
568 Dietary Phytoestrogen Intake and the Risk of Colorectal Cancer. *Nutr Cancer* 68:388-
569 395. doi: 10.1080/01635581.2016.1152380

- 570 64. Rybak ME, Sternberg MR, Pfeiffer CM (2013) Sociodemographic and lifestyle
571 variables are compound- and class-specific correlates of urine phytoestrogen
572 concentrations in the U.S. population. *J Nutr* 143:986S-994S. doi:
573 10.3945/jn.112.172981
- 574 65. Uehar M, Arai Y, Watanabe S, Adlercreutz H (2000) Comparison of plasma and
575 urinary phytoestrogens in Japanese and Finnish women by time-resolved
576 fluoroimmunoassay. *Biofactors* 12:217-225.
- 577 66. Liu W, Tanabe M, Harada KH, Koizumi A (2013) Levels of urinary isoflavones and
578 lignan polyphenols in Japanese women. *Environ Health Prev Med* 18:394-400. doi:
579 10.1007/s12199-013-0338-6
- 580 67. Xu C, Liu Q, Zhang Q, Gu A, Jiang Z-Y (2015) Urinary enterolactone is associated
581 with obesity and metabolic alteration in men in the US National Health and Nutrition
582 Examination Survey 2001-10. *Br J Nutr* 113:683-690. doi:
583 10.1017/S0007114514004115
- 584 68. Perez-Cornago A, Appleby PN, Boeing H, et al (2018) Circulating isoflavone and
585 lignan concentrations and prostate cancer risk: a meta-analysis of individual participant
586 data from seven prospective studies including 2,828 cases and 5,593 controls. *Int J*
587 *Cancer* 143:2677-2686. doi: 10.1002/ijc.31640
- 588 69. Kuijsten A, Buijsman MN, Arts IC, Mulder PP, Hollman PC (2005) A validated
589 method for the quantification of enterodiols and enterolactone in plasma using isotope
590 dilution liquid chromatography with tandem mass spectrometry. *J Chromatogr B*
591 *Analyt Technol Biomed Life Sci* 822:178-184.
- 592 70. Krogholm KS, Bysted A, Brantsæter AL, Jakobsen J, Rasmussen SE, Kristoffersen L,
593 Toft U (2012) Evaluation of flavonoids and enterolactone in overnight urine as intake
594 biomarkers of fruits, vegetables and beverages in the Inter99 cohort study using the
595 method of triads. *Br J Nutr* 108:1904-1912. doi: 10.1017/S0007114512000104
- 596 71. Zamora-Ros R, Achaintre D, Rothwell JA, et al (2016) Urinary excretions of 34 dietary
597 polyphenols and their associations with lifestyle factors in the EPIC cohort study. *Sci*
598 *Rep* 6:26905. doi: 10.1038/srep26905
- 599 72. Ward H, Chapelais G, Kuhnle GG, Luben R, Khaw K-T, Bingham S. Lack of
600 prospective associations between plasma and urinary phytoestrogens and risk of
601 prostate or colorectal cancer in the European Prospective into Cancer-Norfolk study.
602 *Cancer Epidemiol Biomark Prev* 17:2891-2894. doi: 10.1158/1055-9965.EPI-08-0335
- 603 73. Low Y-L, Taylor JI, Grace PB, Dowsett M, Scollen S, Dunning AM, Mulligan AA,
604 Welch AA, Luben RN, Khaw KT, Day NE, Wareham NJ, Bingham SA (2005)
605 Phytoestrogen exposure correlation with plasma estradiol in postmenopausal women in
606 European Prospective Investigation of Cancer and Nutrition-Norfolk may involve diet-
607 gene interactions. *Cancer Epidemiol Biomark Prev* 14:213-220.

- 608 74. Grace PB, Taylor JI, Low Y-L, Luben RN, Mulligan AA, Botting NP, Dowsett M,
609 Welch AA, Khaw KT, Wareham NJ, Day NE, Bingham SA (2004) Phytoestrogen
610 concentrations in serum and spot urine as biomarkers for dietary phytoestrogen intake
611 and their relation to breast cancer risk in European prospective investigation of cancer
612 and nutrition-norfolk. *Cancer Epidemiol Biomark Prev* 13:698-708.
- 613 75. Low Y-L, Taylor JI, Grace PB, Dowsett M, Folkard E, Doody D, Dunning AM,
614 Scollen S, Mulligan AA, Welch AA, Luben RN, Khaw KT, Day NE, Wareham NJ,
615 Bingham SA (2005) Polymorphisms in the CYP19 gene may affect the positive
616 correlations between serum and urine phytoestrogen metabolites and plasma androgen
617 concentrations in men. *J Nutr* 135:2680-2686.
- 618 76. Durazzo A, Carcea M, Adlercreutz H, et al (2014). Effects of consumption of whole
619 grain foods rich in lignans in healthy postmenopausal women with moderate serum
620 cholesterol: a pilot study. *Int J Food Sci Nutr* 65:637-645. doi:
621 10.3109/09637486.2014.893283
- 622 77. Travis RC, Spencer EA, Allen NE, et al (2009) Plasma phyto-oestrogens and prostate
623 cancer in the European Prospective Investigation into Cancer and Nutrition. *Br J*
624 *Cancer* 100:1817-1823. doi: 10.1038/sj.bjc.6605073
- 625 78. Simon GA, Fletcher HM, Golden K, McFarlane-Anderson ND (2015) Urinary
626 isoflavone and lignan phytoestrogen levels and risk of uterine fibroid in Jamaican
627 women. *Maturitas* 82:170-175. doi: 10.1016/j.maturitas.2015.06.041
- 628 79. Zeleniuch-Jacquotte A, Adlercreutz H, Akhmedkhanov A, Toniolo P (1998) Reliability
629 of serum measurements of lignans and isoflavonoid phytoestrogens over a two-year
630 period. *Cancer Epidemiol Biomark Prev* 7:885-889.
- 631 80. Valentín-Blasini L, Blount BC, Caudill SP, Needham LL (2003) Urinary and serum
632 concentrations of seven phytoestrogens in a human reference population subset. *J Expo*
633 *Anal Environ Epidemiol* 13:276-282.
- 634 81. Xie J, Tworoger SS, Franke AA, Terry KL, Rice MS, Rosner BA, Willett WC,
635 Hankinson SE, Eliassen AH (2013) Plasma Enterolactone and Breast Cancer Risk in
636 the Nurses' Health Study II. *Breast Cancer Res Treat* 139:801-809. doi:
637 10.1007/s10549-013-2586-y
- 638 82. Talaei M, Lee BL, Ong CN, van Dam RM, Yuan JM, Koh WP, Pan A (2016) Urine
639 phyto-oestrogen metabolites are not significantly associated with risk of type 2
640 diabetes: the Singapore Chinese health study. *Br J Nutr* 115:1607-1615. doi:
641 10.1017/S0007114516000581
- 642 83. Kunisue T, Tanabe S, Isobe T, Aldous KM, Kannan K (2010) Profiles of
643 phytoestrogens in human urine from several Asian countries. *J Agric Food Chem*
644 58:9838-9846. doi: 10.1021/jf102253j

- 645 84. Liu J, Mi S, Du L, Li X, Li P, Jia K, Zhao J, Zhang H, Zhao W, Gao Y (2018) The
646 associations between plasma phytoestrogens concentration and metabolic syndrome
647 risks in Chinese population. *PloS One* 13:e0194639. doi:
648 10.1371/journal.pone.0194639
- 649 85. Morton MS, Chan PS, Cheng C, Blacklock N, Matos-Ferreira A, Abranches-Monteiro
650 L, Correia R, Lloyd S, Griffiths K (1997) Lignans and isoflavonoids in plasma and
651 prostatic fluid in men: samples from Portugal, Hong Kong, and the United Kingdom.
652 *Prostate* 32:122-128.
- 653 86. Ko K-P, Yeo Y, Yoon J-H, Kim C-S, Tokudome S, Ngoan LT, Koriyama C, Lim YK,
654 Chang SH, Shin HR, Kang D, Park SK, Kang CH, Yoo KY (2018) Plasma
655 phytoestrogens concentration and risk of colorectal cancer in two different Asian
656 populations. *Clin Nutr* 37:1675-1682. doi: 10.1016/j.clnu.2017.07.014
- 657 87. Morton MS, Arisaka O, Miyake N, Morgan LD, Evans BA (2002) Phytoestrogen
658 concentrations in serum from Japanese men and women over forty years of age. *J Nutr*
659 132:3168-3171.
- 660 88. Johnsen NF, Hausner H, Olsen A, Tetens I, Christensen J, Knudsen KE, Overvad K,
661 Tjønneland A (2004) Intake of whole grains and vegetables determines the plasma
662 enterolactone concentration of Danish women. *J Nutr* 134:2691-2697. doi:
663 10.1093/jn/134.10.2691
- 664 89. Micek A, Godos J, Brzostek T, et al (2021) Dietary phytoestrogens and biomarkers of
665 their intake in relation to cancer survival and recurrence: a comprehensive systematic
666 review with meta-analysis. *Nutr Rev.* 79(1):42-65. doi: 10.1093/nutrit/nuaa043
- 667 90. Nurmi T, Mursu J, Peñalvo JL, Poulsen HE, Voutilainen S (2010) Dietary intake and
668 urinary excretion of lignans in Finnish men. *Br J Nutr* 103:677-685. doi:
669 10.1017/S0007114509992261
- 670 91. Hedelin M, Klint A, Chang ET, Bellocco R, Johansson J-E, Andersson SO, Heinonen
671 SM, Adlercreutz H, Adami HO, Grönberg H, Bälter KA (2006) Dietary phytoestrogen,
672 serum enterolactone and risk of prostate cancer: the cancer prostate Sweden study
673 (Sweden). *Cancer Causes Control* 17:169-180.
- 674 92. Bhakta D, dos Santos Silva I, Higgins C, Sevak L, Kassam-Khamis T, Mangtani P,
675 Adlercreutz H, McMichael A (2005) A semiquantitative food frequency questionnaire
676 is a valid indicator of the usual intake of phytoestrogens by south Asian women in the
677 UK relative to multiple 24-h dietary recalls and multiple plasma samples. *J Nutr*
678 135:116-123.
- 679 93. Bhakta D, Higgins CD, Sevak L, Mangtani P, Adlercreutz H, McMichael AJ, dos
680 Santos Silva I (2006) Phyto-oestrogen intake and plasma concentrations in South Asian
681 and native British women resident in England. *Br J Nutr* 95:1150-1158.

- 682 94. Mulligan AA, Kuhnle GG, Lentjes MA, Scheltinga V van, Powell NA, McTaggart A,
683 Bhaniani A, Khaw KT (2013) Intakes and sources of isoflavones, lignans,
684 enterolignans, coumestrol and soya-containing foods in the Norfolk arm of the
685 European Prospective Investigation into Cancer and Nutrition (EPIC-Norfolk), from 7
686 d food diaries, using a newly updated database. *Public Health Nutr* 16:1454-1462. doi:
687 10.1017/S1368980012003904
- 688 95. Witkowska AM, Waśkiewicz A, Zujko ME, Szcześniewska D, Stepaniak U, Pająk A,
689 Drygas W (2018) Are Total and Individual Dietary Lignans Related to Cardiovascular
690 Disease and Its Risk Factors in Postmenopausal Women? A Nationwide Study.
691 *Nutrients* 10:865. doi: 10.3390/nu10070865
- 692 96. Linseisen J, Piller R, Hermann S, Chang-Claude J (2004) Dietary phytoestrogen intake
693 and premenopausal breast cancer risk in a German case-control study. *Int J Cancer*
694 110:284-290.
- 695 97. Boker LK, Van der Schouw YT, De Kleijn MJ, Jacques PF, Grobbee DE, Peeters PH
696 (2002) Intake of Dietary Phytoestrogens by Dutch Women. *J Nutr* 132:1319-1328.
- 697 98. Milder IE, Feskens EJ, Arts IC, Bueno-de-Mesquita HB, Hollman PC, Kromhout D
698 (2006) Intakes of 4 dietary lignans and cause-specific and all-cause mortality in the
699 Zutphen Elderly Study. *Am J Clin Nutr* 84:400-405.
- 700 99. Pérez-Jiménez J, Fezeu L, Touvier M, Arnault N, Manach C, Hercberg S, Galan P,
701 Scalbert A (2011) Dietary intake of 337 polyphenols in French adults. *Am J Clin Nutr*
702 93:1220-1228. doi: 10.3945/ajcn.110.007096
- 703 100. Lefèvre-Arbogast S, Gaudout D, Bensalem J, Letenneur L, Dartigues JF, Hejblum
704 BP, Féart C, Delcourt C, Samieri C (2019) Pattern of polyphenol intake and the long-
705 term risk of dementia in older persons. *Neurology* 90:e1979-e1988. doi:
706 10.1212/WNL.0000000000005607
- 707 101. Pellegrini N, Valtueña S, Ardigò D, Brighenti F, Franzini L, Del Rio D, Scazzina F,
708 Piatti PM, Zavaroni I (2010) Intake of the plant lignans matairesinol,
709 secoisolariciresinol, pinoresinol, and lariciresinol in relation to vascular inflammation
710 and endothelial dysfunction in middle age-elderly men and post-menopausal women
711 living in Northern Italy. *Nutr Metab Cardiovasc Dis* 20:64-71. doi:
712 10.1016/j.numecd.2009.02.003
- 713 102. Godos J, Marventano S, Mistretta A, Galvano F, Grosso G (2017) Dietary sources
714 of polyphenols in the Mediterranean healthy Eating, Aging and Lifestyle (MEAL)
715 study cohort. *Int J Food Sci Nutr* 68:750-756. doi: 10.1080/09637486.2017.1285870
- 716 103. Godos J, Bergante S, Satriano A, Pluchinotta FR, Marranzano M (2018) Dietary
717 Phytoestrogen Intake is Inversely Associated with Hypertension in a Cohort of Adults
718 Living in the Mediterranean Area. *Molecules* 23:E368. doi:
719 10.3390/molecules23020368

- 720 104. González S, Fernández M, Cuervo A, Lasheras C (2014) Dietary intake of
721 polyphenols and major food sources in an institutionalised elderly population. *J Hum*
722 *Nutr Diet* 27:176-183. doi: 10.1111/jhn.12058
- 723 105. Zamora-Ros R, Not C, Guinó E, Luján-Barroso L, García RM, Biondo S, Salazar R,
724 Moreno V (2013) Association between habitual dietary flavonoid and lignan intake and
725 colorectal cancer in a Spanish case–control study (the Bellvitge Colorectal Cancer
726 Study). *Cancer Causes Control* 24:549-557. doi: 10.1007/s10552-012-9992-z
- 727 106. Tresserra-Rimbau A, Medina-Remón A, Pérez-Jiménez J, (2013) Dietary intake and
728 major food sources of polyphenols in a Spanish population at high cardiovascular risk:
729 The PREDIMED study. *Nutr Metab Cardiovasc Dis* 23:953-959. doi:
730 10.1016/j.numecd.2012.10.008
- 731 107. Mendonça RD, Carvalho NC, Martin-Moreno JM, Pimenta AM, Lopes ACS, Gea
732 A, Martinez-Gonzalez MA, Bes-Rastrollo M (2019) Total polyphenol intake,
733 polyphenol subtypes and incidence of cardiovascular disease: The SUN cohort study.
734 *Nutr Metab Cardiovasc Dis* 29:69-78. doi: 10.1016/j.numecd.2018.09.012
- 735 108. Petrick JL, Steck SE, Bradshaw PT, Chow W-H, Engel LS, He K, Risch HA,
736 Vaughan TL, Gammon MD (2015) Dietary flavonoid intake and Barrett’s esophagus in
737 western Washington State. *Ann Epidemiol* 25:730-735. doi:
738 10.1016/j.annepidem.2015.05.010
- 739 109. Petrick JL, Steck SE, Bradshaw PT, Trivers KF, Abrahamson PE, Engel LS, He K,
740 Chow WH, Mayne ST, Risch HA, Vaughan TL, Gammon MD (2015) Dietary intake of
741 flavonoids and oesophageal and gastric cancer: incidence and survival in the United
742 States of America (USA). *Br J Cancer* 112:1291-1300. doi: 10.1038/bjc.2015.25.
- 743 110. Williams AM, Bonner M, Ochs-Balcom HM, Hwang H, Morrison C, McCann SE
744 (2015) Dietary Lignan Intake and Androgen Receptor Expression in Breast Tumors.
745 *Cancer Causes Control* 26:311-317. doi: 10.1007/s10552-014-0504-1
- 746 111. Waetjen LE, Leung K, Crawford SL, Huang M-H, Gold EB, Greendale GA (2013)
747 The Relationship Between Dietary Phytoestrogens and Development of Urinary
748 Incontinence in Midlife Women. *Menopause* 20:428-436. doi:
749 10.1097/gme.0b013e3182703c9c
- 750 112. Bandera EV, King M, Chandran U, Paddock LE, Rodriguez-Rodriguez L, Olson SH
751 (2011) Phytoestrogen consumption from foods and supplements and epithelial ovarian
752 cancer risk: a population-based case control study. *BMC Womens Health* 11:40. doi:
753 10.1186/1472-6874-11-40
- 754 113. Chang ET, Canchola AJ, Clarke CA, Lu Y, West DW, Bernstein L, Wang SS,
755 Horn-Ross PL (2011) Dietary phytochemicals and risk of lymphoid malignancies in
756 the California Teachers Study cohort. *Cancer Causes Control* 22:237-249. doi:
757 10.1007/s10552-010-9692-5

- 758 114. McCann SE, Thompson LU, Nie J, Dorn J, Trevisan M, Shields PG, Ambrosone
759 CB, Edge SB, Li HF, Kasprzak C, Freudenheim JL (2010) Dietary lignan intakes in
760 relation to survival among women with breast cancer: the Western New York
761 Exposures and Breast Cancer (WEB) Study. *Breast Cancer Res Treat* 122:229-235. doi:
762 10.1007/s10549-009-0681-x
- 763 115. Mervish NA, Teitelbaum SL, Pajak A, Windham GC, Pinney SM, Kushi LH, Biro
764 FM, Wolff MS (2017) Peripubertal dietary flavonol and lignan intake and age at
765 menarche in a longitudinal cohort of girls. *Pediatr Res* 82:201-208. doi:
766 10.1038/pr.2017.34
- 767 116. van der Schouw YT, Sampson L, Willett WC, Rimm EB (2005) The usual intake of
768 lignans but not that of isoflavones may be related to cardiovascular risk factors in U.S.
769 men. *J Nutr* 135:260-266.
- 770 117. Horn-Ross PL, Hoggatt KJ, Lee MM (2002) Phytoestrogens and thyroid cancer
771 risk: the San Francisco Bay Area thyroid cancer study. *Cancer Epidemiol Prev Biomark*
772 11:43-49.
- 773 118. McCann SE, Freudenheim JL, Marshall JR, Graham S (2003) Risk of human
774 ovarian cancer is related to dietary intake of selected nutrients, phytochemicals and
775 food groups. *J Nutr* 133:1937-1942.
- 776 119. de Kleijn MJ, van der Schouw YT, Wilson PW, Adlercreutz H, Mazur W, Grobbee
777 DE, Jacques PF (2001) Intake of dietary phytoestrogens is low in postmenopausal
778 women in the United States: the Framingham study. *J Nutr* 131:1826-1832.
- 779 120. Chávez-Suárez KM, Ortega-Vélez MI, Valenzuela-Quintanar AI, et al (2017)
780 Phytoestrogen Concentrations in Human Urine as Biomarkers for Dietary
781 Phytoestrogen Intake in Mexican Women. *Nutrients* 9:E1078. doi: 10.3390/nu9101078
- 782 121. Park S-Y, Wilkens LR, Franke AA, Le Marchand L, Kakazu KK, Goodman MT,
783 Murphy SP, Henderson BE, Kolonel LN (2009) Urinary phytoestrogen excretion and
784 prostate cancer risk: a nested case-control study in the Multiethnic Cohort. *Br J Cancer*
785 101:185-191. doi: 10.1038/sj.bjc.6605137
- 786 122. Hu Y, Song Y, Franke AA, Hu FB, van Dam RM, Sun Q (2015) A Prospective
787 Investigation of the Association Between Urinary Excretion of Dietary Lignan
788 Metabolites and Weight Change in US Women. *Am J Epidemiol* 182:503-511. doi:
789 10.1093/aje/kwv091
- 790 123. Reger MK, Zollinger TW, Liu Z, Jones J, Zhang J (2017) Association between
791 Urinary Phytoestrogens and C-reactive Protein in the Continuous National Health and
792 Nutrition Examination Survey. *J Am Coll Nutr* 36:434-441. doi:
793 10.1080/07315724.2017.1318722

- 794 124. Martínez Steele E, Monteiro CA (2017) Association between Dietary Share of
795 Ultra-Processed Foods and Urinary Concentrations of Phytoestrogens in the US.
796 *Nutrients* 9:E209. doi: 10.3390/nu9030209
- 797 125. Adlercreutz H, Fotsis T, Heikkinen R, Dwyer JT, Woods M, Goldin BR, Gorbach
798 SL (1982) Excretion of the lignans enterolactone and enterodiol and of equol in
799 omnivorous and vegetarian postmenopausal women and in women with breast cancer.
800 *Lancet* 2:1295-1299.
- 801 126. Miles FL, Navarro SL, Schwarz Y, Gu H, Djukovic D, Randolph TW, Shojaie A,
802 Kratz M, Hullar MAJ, Lampe PD, Neuhouser ML, Raftery D, Lampe JW (2017)
803 Plasma metabolite abundances are associated with urinary enterolactone excretion in
804 healthy participants on controlled diets. *Food Funct* 8:3209-3218. doi:
805 10.1039/c7fo00684e
- 806 127. Reger MK, Zollinger TW, Liu Z, Jones J, Zhang J (2016) Urinary phytoestrogens
807 and cancer, cardiovascular, and all-cause mortality in the continuous National Health
808 and Nutrition Examination Survey. *Eur J Nutr* 55:1029-1040. doi: 10.1007/s00394-015-
809 0917-y
- 810 128. Eichholzer M, Richard A, Nicastro HL, Platz EA, Linseisen J, Rohrmann S (2014)
811 Urinary lignans and inflammatory markers in the US National Health and Nutrition
812 Examination Survey (NHANES) 1999-2004 and 2005-2008. *Cancer Causes Control*
813 25:395-403. doi: 10.1007/s10552-014-0340-3
- 814 129. Valentín-Blasini L, Sadowski MA, Walden D, Caltabiano L, Needham LL, Barr DB
815 (2005) Urinary phytoestrogen concentrations in the U.S. population (1999-2000). *J*
816 *Expo Anal Environ Epidemiol* 15:509-523.
- 817 130. Sun Q, Wedick NM, Pan A, Townsend MK, Cassidy A, Franke AA, Rimm EB, Hu
818 FB, van Dam RM (2014) Gut microbiota metabolites of dietary lignans and risk of type
819 2 diabetes: a prospective investigation in two cohorts of U.S. women. *Diabetes Care*
820 37:1287-1295. doi: 10.2337/dc13-2513
- 821 131. Levine LD, Kim K, Purdue-Smithe A, Sundaram R, Schisterman EF, Connell M,
822 Devilbiss EA, Alkhalaf Z, Radoc JG, Buck Louis GM, Mumford SL (2019) Urinary
823 Phytoestrogens and Relationship to Menstrual Cycle Length and Variability Among
824 Healthy, Eumenorrheic Women. *J Endocr Soc* 4:bvz003. doi: 10.1210/jendso/bvz003
- 825 132. Stumpf K, Pietinen P, Puska P, Adlercreutz H (2000) Changes in serum
826 enterolactone, genistein, and daidzein in a dietary intervention study in Finland. *Cancer*
827 *Epidemiol Biomark Prev* 9:1369-1372.
- 828 133. Pietinen P, Stumpf K, Männistö S, Kataja V, Uusitupa M, Adlercreutz H (2001)
829 Serum enterolactone and risk of breast cancer: a case-control study in eastern Finland.
830 *Cancer Epidemiol Biomark Prev* 10:339-344.

- 831 134. Vanharanta M, Voutilainen S, Lakka TA, van der Lee M, Adlercreutz H, Salonen
832 JT (1999). Risk of acute coronary events according to serum concentrations of
833 enterolactone: a prospective population-based case-control study. *Lancet* 354:2112-
834 2115.
- 835 135. Kilkkinen A, Erlund I, Virtanen MJ, Alfthan G, Ariniemi K, Virtamo J (2006)
836 Serum enterolactone concentration and the risk of coronary heart disease in a case-
837 cohort study of Finnish male smokers. *Am J Epidemiol* 163:687-693.
- 838 136. Vanharanta M, Voutilainen S, Nurmi T, Kaikkonen J, Roberts LJ, Morrow JD,
839 Adlercreutz H, Salonen JT (2002) Association between low serum enterolactone and
840 increased plasma F2-isoprostanes, a measure of lipid peroxidation. *Atherosclerosis*
841 160:465-469.
- 842 137. Kilkkinen A, Stumpf K, Pietinen P, Valsta LM, Tapanainen H, Adlercreutz H
843 (2001) Determinants of serum enterolactone concentration. *Am J Clin Nutr* 73:1094-
844 1100.
- 845 138. Vanharanta M, Voutilainen S, Rissanen TH, Adlercreutz H, Salonen JT (2003) Risk
846 of cardiovascular disease-related and all-cause death according to serum concentrations
847 of enterolactone: Kuopio Ischaemic Heart Disease Risk Factor Study. *Arch Intern Med*
848 163:1099-1104.
- 849 139. Sonestedt E, Ivarsson MIL, Harlid S, Ericson U, Gullberg B, Carlson J, Olsson H,
850 Adlercreutz H, Wirfält E (2009) The protective association of high plasma
851 enterolactone with breast cancer is reasonably robust in women with polymorphisms in
852 the estrogen receptor alpha and beta genes. *J Nutr* 139:993-1001. doi:
853 10.3945/jn.108.101691
- 854 140. Stattin P, Bylund A, Biessy C, Kaaks R, Hallmans G, Adlercreutz H (2004)
855 Prospective study of plasma enterolactone and prostate cancer risk (Sweden). *Cancer*
856 *Causes Control* 15:1095-1102.
- 857 141. Lin Y, Wolk A, Håkansson N, Peñalvo JL, Lagergren J, Adlercreutz H, Lu Y (2013)
858 Validation of FFQ-based assessment of dietary lignans compared with serum
859 enterolactone in Swedish women. *Br J Nutr* 109:1873-1880. doi:
860 10.1017/S000711451200387X
- 861 142. Hultén K, Winkvist A, Lenner P, Johansson R, Adlercreutz H, Hallmans G (2002)
862 An incident case-referent study on plasma enterolactone and breast cancer risk. *Eur J*
863 *Nutr* 41:168-176.
- 864 143. Aarestrup J, Kyrø C, Knudsen KEB, Weiderpass E, Christensen J, Kristensen M,
865 Würtz AM, Johnsen NF, Overvad K, Tjønneland A, Olsen A (2013) Plasma
866 enterolactone and incidence of endometrial cancer in a case-cohort study of Danish
867 women. *Br J Nutr* 109:2269-2275. doi: 10.1017/S0007114512004424.

- 868 144. Eriksen AK, Kyrø C, Nørskov NP, Frederiksen K, Bach Knudsen K-E, Overvad K,
869 Landberg R, Tjønneland A, Olsen A (2019) Pre-diagnostic plasma enterolactone
870 concentrations are associated with lower mortality among individuals with type 2
871 diabetes: a case-cohort study in the Danish Diet, Cancer and Health cohort.
872 *Diabetologia* 62:959-969. doi: 10.1007/s00125-019-4854-9
- 873 145. Verheus M, van Gils CH, Keinan-Boker L, Grace PB, Bingham SA, Peeters PH
874 (2007) Plasma phytoestrogens and subsequent breast cancer risk. *J Clin Oncol* 25:648-
875 655.
- 876 146. Heald CL, Ritchie MR, Bolton-Smith C, Morton MS, Alexander FE (2007) Phyto-
877 oestrogens and risk of prostate cancer in Scottish men. *Br J Nutr* 98:388-396.
- 878 147. Piller R, Chang-Claude J, Linseisen J (2006) Plasma enterolactone and genistein
879 and the risk of premenopausal breast cancer. *Eur J Cancer Prev* 15:225-232.
- 880 148. Horner NK, Kristal AR, Prunty J, Skor HE, Potter JD, Lampe JW (2002) Dietary
881 determinants of plasma enterolactone. *Cancer Epidemiol Biomark Prev* 11:121-126.
- 882 149. Ko KP, Yeo Y, Yoon JH, et al (2018) Plasma phytoestrogens concentration and risk
883 of colorectal cancer in two different Asian populations. *Clin Nutr* 37:1675-1682. doi:
884 10.1016/j.clnu.2017.07.014
- 885

886 **FIGURE CAPTIONS**

887 **Figure 1.** Scheme of human bioavailability of dietary lignans.

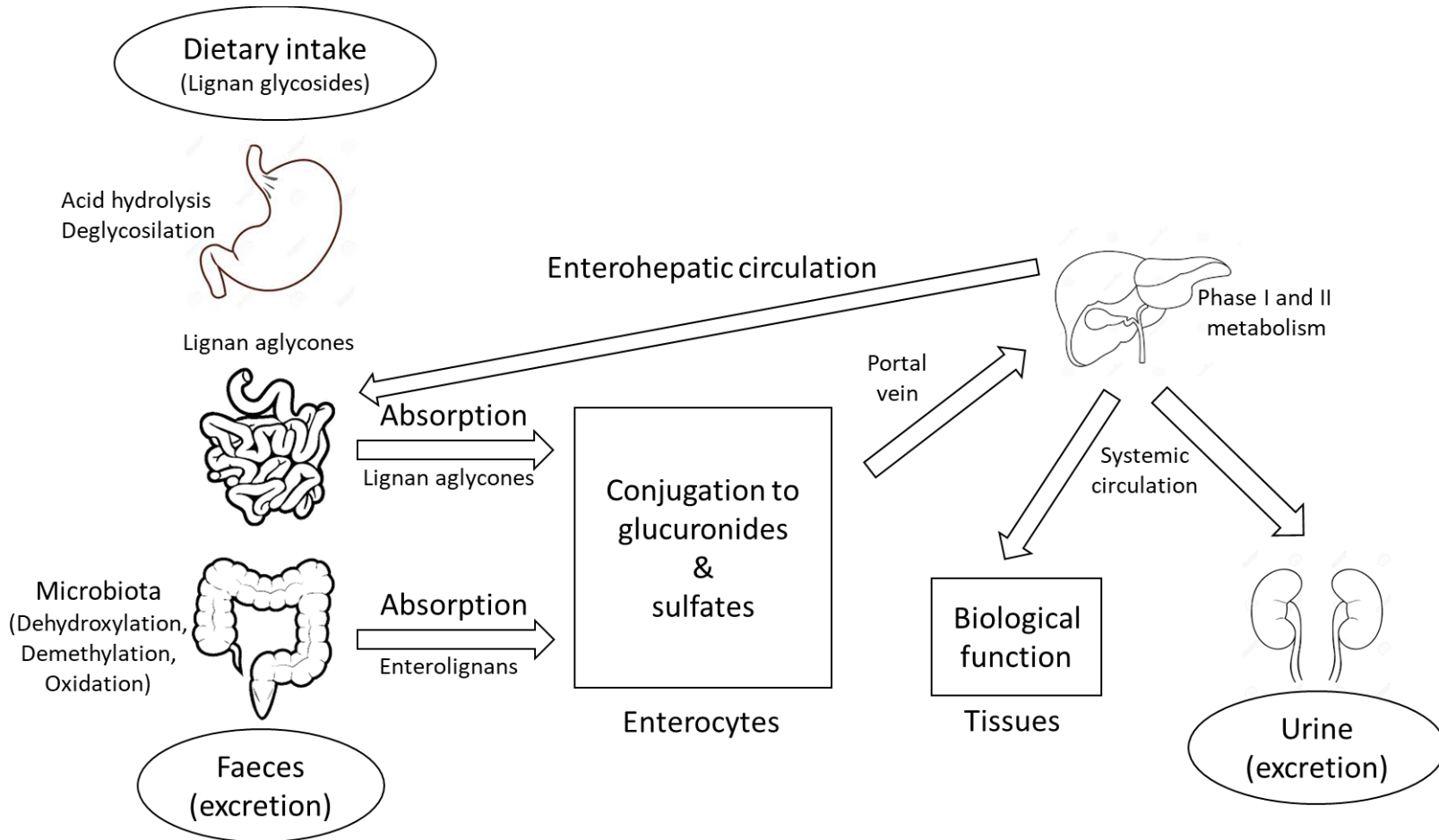
888 **Figure 2.** Mean of means/medians of total dietary lignan intake (mg/d) by country.

889 **Figure 3.** Mean of means/medians of urinary and blood enterolignan concentrations

890 (nmol/L) by country; A: urinary enterolactone, B: urinary enterodiols, C: blood

891 enterolactone, D: blood enterodiols.

892 Figure 1.

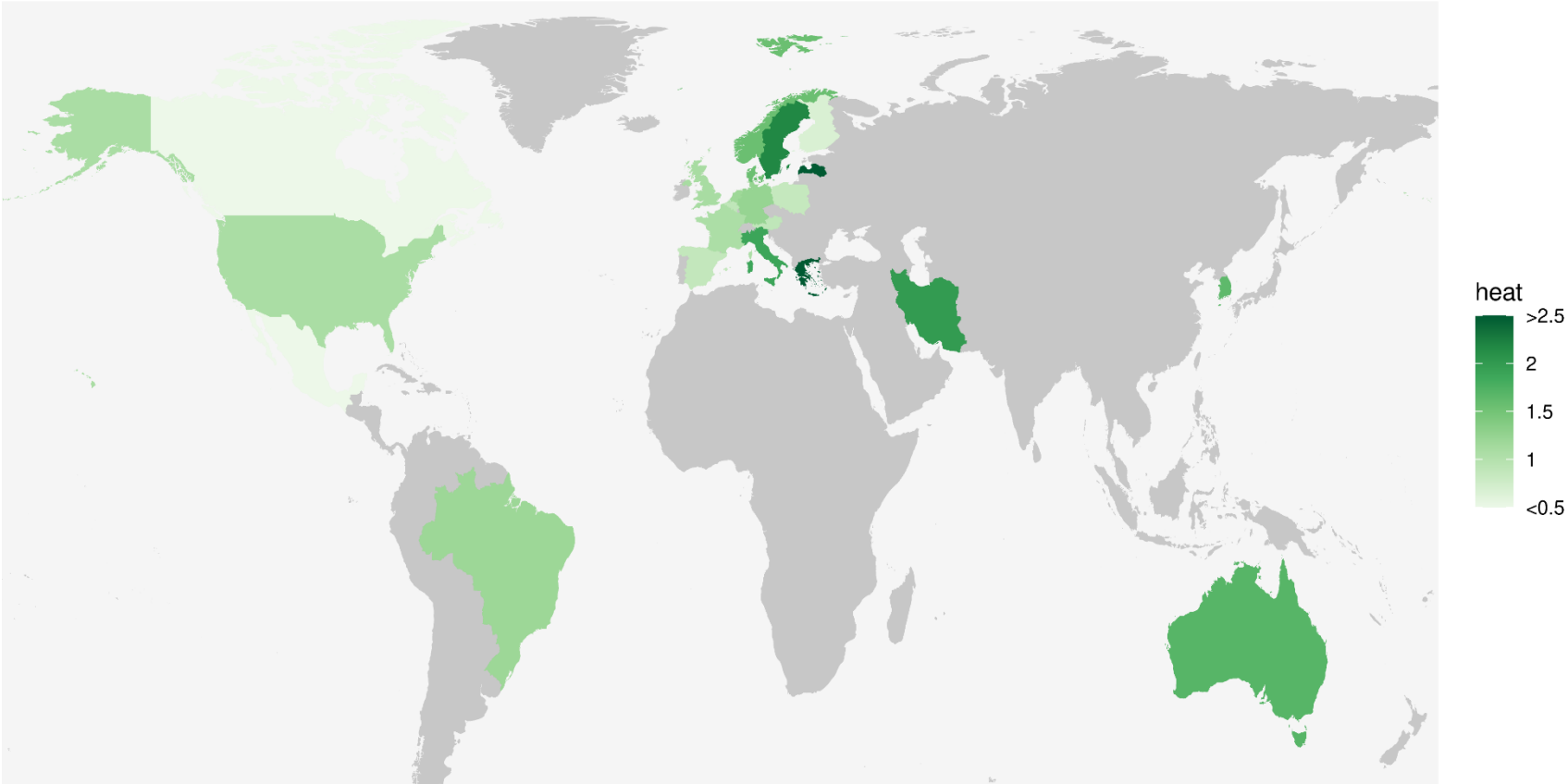


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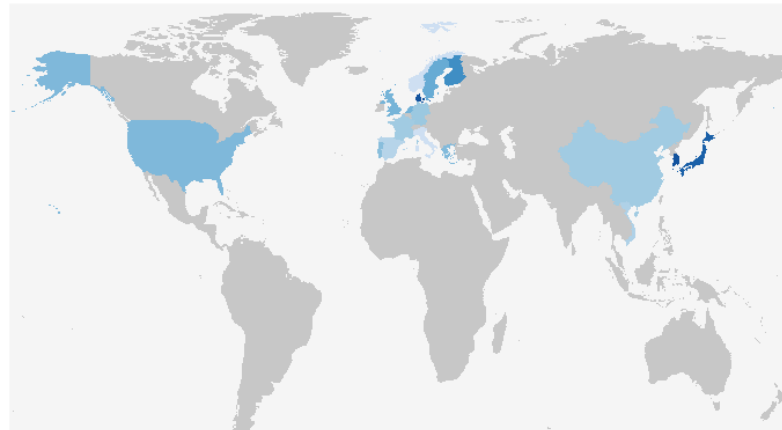
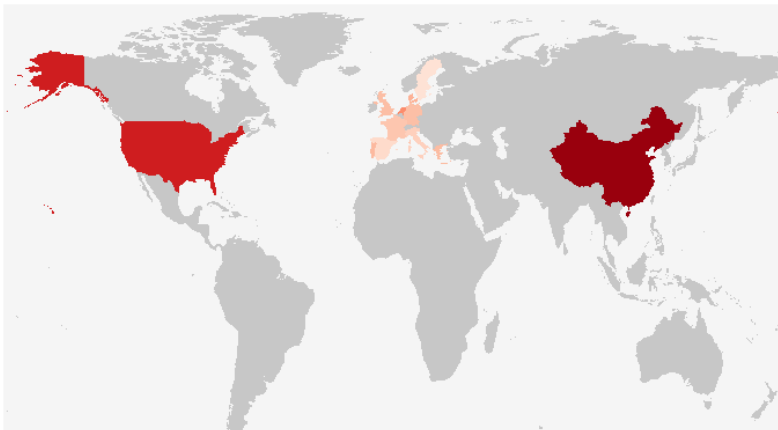
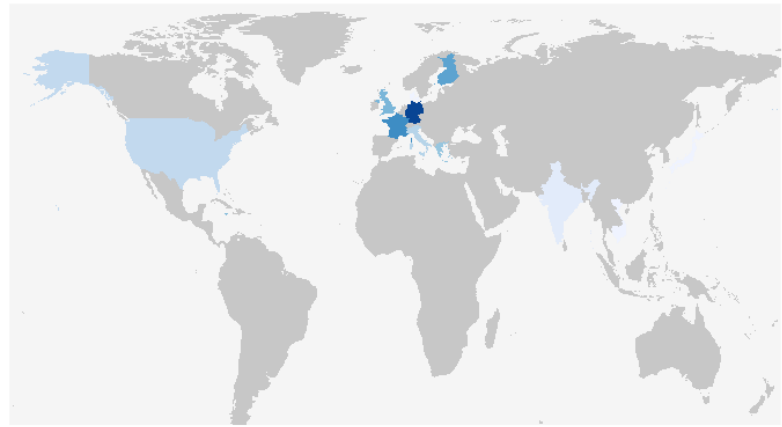
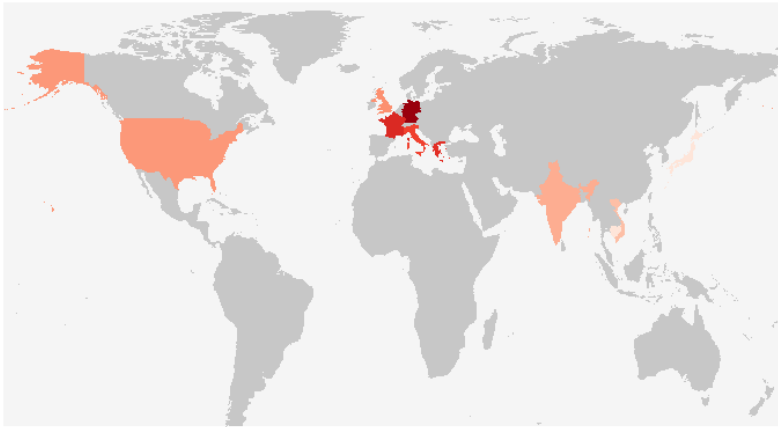
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896 Figure 2.



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898 Figure 3.



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902 Table 1. Characteristics of the studies included in the review of dietary lignan intake.

Author (Reference)	Year	Country	Population			Dietary survey	FCDB	TOTAL LIGNANS		
			N	Sex	Age (y)			Individual lignans	Intake (mg/d)	Food sources
Wisnuwarda-ni [36]	2006-2007	MED countries	915	F (53%)	12-18	2 x 24-HDR	Phenol Explorer	All ⁴	1.2 (0.0) ¹	Breads (71%), fruit (8%), vegetables (7%)
		non-MED countries	1,513						0.9 (0.0) ¹	Breads (58%), fruit (12%), vegetables (7%)
Zamora-Ros [9]	1995-2000	MED countries	11,285	F (64%)	35-74	24-HDR	Phenol Explorer	All ⁴ : SECO (18%), LARI (14%), Sesamol (12%), Sesamin (12%)	3.6 (0.1) ¹	Vegetable oils (26%), cakes & biscuits (20%), breads (12%)
		non-MED countries	24,443						2.3 (0.1) ¹	Breads (22%), spices (16%), seeds (16%), vegetable oils (11%)
		UK healthy	309						9.1 (0.9) ¹	Seeds (48%), vegetable oils (10%), vegetables (9%)
Tetens [37]	2000-2002	Denmark	2,463	F (53%)	25-64	7-DR	Dutch DB	LARI (43%), PINO (32%), SECO (22%), MATA (3%)	1.5 ²	Cereals (27-30%), fruit & berries (18-25%), coffee & tea (21%), vegetables (19-20%)
	2002	Finland	2,007	F (55%)	25-64	48-HDR			1.1 ²	Cereals (27-36%), fruit & berries (22-31%), vegetables (16-20%), coffee & tea (17%)
	1994-1996	Italy	1,268	F (54%)	25-64	7-DR			1.1 ²	Fruit & berries (42-46%), vegetables (26-28%), cereals (17%)
	1987-1990	Sweden	83,760	F (45%)	45-79	FFQ			1.8 ²	Cereals (26-42%), vegetables (18-30%), fruit & berries (15-23%), coffee & tea (18-19%)
	2000-2001	UK	1,724	F (56%)	19-64	7-DR			1.2 ²	Coffee & tea (30-32%), vegetables (23-25%), fruit & berries (15-20%),

										cereals (15-17%)
Kilkinen [57]	1997	Finland	1,359	M	25-63	24-HDR	Finish DB	MATA (73%), SECO (27%)	0.2 ²	Cereals (49%), fruits (25%), vegetables (12%)
			1,493	F	25-64			MATA (80%), SECO (20%)	0.2 ²	Fruits (39%), cereals (35%), vegetables (13%)
Nurmi [90]	1995	Finland	100	M	58(6) ¹	4-DR	Dutch DB	LARI (40%), PINO (38%), SECO (14%), MATA (7%)	1.2 (0.5) ³	Rye products, berries, coffee, tea, vegetable roots
Hedelin [63]	1991-1992	Sweden	48,268	F	30-49	FFQ	Finish DB II	LARI, MATA, PINO, SECO, SYRI, MEDI	2.3 (1.8-2.8) ³	Rye bread (57%), wheat bread (27%), cereals (8%)
Hedelin [62]	1991-1992	Sweden	46,977	F	30-49	FFQ	Finish DB II	LARI, MATA, PINO, SECO, SYRI, MEDI	2.3 (1.0-4.0) ³	Rye bread, wheat bread, cereals, berries
Suzuki [55]	1987-1990	Sweden	51,823	F	40-76	FFQ	Own DB	LARI, MATA, PINO, SECO	0.9 (0.7-1.0) ³	—
Hedelin [91]	2001-2002	Sweden	1,130	M	35-79	FFQ	Finish DB II	SECO (38%), SYRI (30%), PINO (15%), LARI (13%), MEDI (12%), MATA (1%)	4.9 ²	Flaxseed (36%), Rye bread (39%), wheat bread (15%)
Meija [35]	2009-2011	Latvia	172	M	40-75	FFQ	Canadian DB	SECO (58%), SYRI (22%), PINO (11%), LARI (6%), MATA (1%), MEDI (1%)	5.2 (6.4) ¹	Seed & rye bread (86%), flaxseed (7%);
			97	F					3.3 (4.4) ¹	Seed & rye bread (57%), flaxseed (35%)
Bhakta [92]	1995-1999	UK	108	F	25-75	≥9 x 24HDR	Finish DB II	SECO (93%), MATA (7%)	0.1 (0.1) ¹	Breads (75%), vegetables (9%), fruit & fruit juices (7%)
Bhakta [93]	1995-1999	UK (Asian)	221	F	<75	≥4 x 24HDR	Own DB	SECO (93%), MATA (7%)	0.1 (0.1) ¹	Breads (70%), vegetables (12%)
		UK (British)	49					SECO (93%), MATA (7%)	0.2 (0.1) ¹	Breads (60%), fruit & fruit juices (21%)
Mulligan [94]	1993-1997	UK	9,680	M	40-75	7d DR	Own DB	MATA, SECO, Shonanin	0.3 (0.2) ¹	Tea & coffee (33%), beer (12%), vegetables (9%)
			10,757	F					0.3 (0.1) ¹	Tea & coffee (37%), vegetables

										(12%), fruits (9%)
Grosso [38]	1993-1997	Poland	10,477	F (50%)	45-69	FFQ	Phenol Explorer	All ⁴	0.6 (12) ¹	Seeds (51%) tea (27%), dark bread (8%)
Witkowska [95]	2003-2014	Poland	1,683	F	>20	24-HDR	Dutch DB	SECO (45%), LARI (26%), PINO (26%), MATA (3%)	1.1 (4.4) ¹	Vegetables (38%), flaxseed (22%), tea (12%)
Witkowska [39]	2003-2005	Poland	6,661	F (53%)	20-74	24-HDR	Phenol Explorer	All ⁴	12.1 ²	Cucumber (41%), red cabbage (22%)
Linseisen [96]	1992-1995	Germany	666	F	43(6) ¹	FFQ	Own DB	SECO (94%), MATA (6%)	0.6 (0.3–1.3) ³	Nuts & seeds (75%), vegetables (7%), coffee (6%)
Boker [97]	1993-1997	Netherlands	17,140	F	50-69	FFQ	Dutch DB	SECO (93%), MATA (7%)	1.0 ²	Breads (41%), coffee & tea (23%), fruits (14%)
Milder [61]	1997-1998	Netherlands	4,661	F (55%)	≥19	2-DR	Dutch DB	LARI (43%), PINO (32%), SECO(24%), MATA(0.6%)	1.2 (2.1) ¹	Tea & coffee (37%), nuts & seeds (14%),
Milder [30]	1997-2002	Netherlands	306	F (56%)	19-75	FFQ	Dutch DB	LARI (47%), PINO (35%), SECO (18%), MATA (1%)	1.1 (0.4) ¹	Vegetables & black tea (>20%), whole-grain bread, fruits, wine.
Milder [98]	1985-1995	Netherlands	570	M	64-84	DH	Dutch DB	LARI (48%), PINO (36%), SECO (15%), MATA (1%)	1.0 (0.8-1.0) ³	Tea (28%), vegetables (27%), bread (14%)
Pérez-Jiménez [99]	1994-2001	France	4,942	F (47%)	35–60	6 x 24-HDR	Phenol Explorer	All ⁴	0.4 (0.2) ¹	Coffee (21%), refined wheat products (18%), whole-grain wheat products (16%)
Lefèvre-Arbogast [100]	1999-2000	France	1,329	F (62%)	≥65	24-HDR	Phenol Explorer	All ⁴	0.4 (0.3) ¹	Wine (65%), olive oil (12%), tea & infusion (9%), soy products (8%)
Adriouch [34]	1994-1996	France	3,903	F (47%)	35–60	≥6 x24-HDR	Phenol Explorer	All ⁴	0.2 (0.1) ¹	Bread (30%), red wine (29%), olive oil (15%), tea (9%)
Pellegrini [101]	2002-2003	Italy	242	F (38%)	60(8) ¹	3D-WR	Dutch DB	SECO (52%), LARI (27%), PINO (17%), MATA (3%)	0.7 (0.3) ¹	Red wine, fruits & vegetables (80%)
Pounis [58]	2005-	Italy	14,029	F (50%)	35 ²	FFQ	Eurofir-	—	80 (60-106) ³	Seasonal fruits (41%), grain & pod

	2010						eBASIS			vegetables (11%)
Godos [102]	2014–2015	Italy	1,947	F (33%)	>18	FFQ	Phenol Explorer	All ⁴	2.8 (2.6) ¹	Citrus fruits (44%), red orange (32%), garlic (11%)
Godos [103]	2014–2015	Italy	1,936	F (28%)	>18	FFQ	Phenol Explorer	All ⁴	1.4 (1.1–2.0) ¹	Citrus fruits, garlic, olive oil, bread
Russo [40]	2015–2016	Italy	340	M	>18	FFQ	Phenol Explorer	LARI (54%), PINO (34%), SECO (4%), MATA (1%)	3.1 (2.7) ¹	Cereals, fruits, vegetables, grains, nuts
González [104]	—	Spain	127	M	73(7) ¹	FFQ	Phenol Explorer	All ⁴	0.5 (0.3) ¹	Olive oil, white bread, & red wine (93%)
			177	F	77(6) ¹				0.4 (0.2) ¹	
Peñalvo [56]	1998–2000	Spain	3,438	F (57%)	2–24	24-HDR	Alignia DB	PINO (42%), SECO (17%), LARI (13%), MATA (1%)	0.8 (0.5–1.3) ³	Olive oil (27%), refined wheat bread (17%), whole-grain wheat bread (8%)
Zamora-Ros [105]	1996–1998	Spain	401	M	65 (12) ¹	FFQ	UK DB	SECO, MATA, LARI, PINO	0.7 (0.5–1.0) ³	Fruit (32%), vegetables (31%), cereals products (10%)
Tresserra-Rimbau [106]	2003–2009	Spain	7,200	M&F	55–80	FFQ	Phenol Explorer	All ⁴	0.9 (0.4) ¹	Olive oil (47%), virgin olive oil (25%), whole-grain wheat-flour bread (6%)
Mendonça [107]	1999–	Spain	17,065	F (61%)	20–89	FFQ	Phenol Explorer	All ⁴	0.6 (0.4) ¹	Olive oil, dried fruits, gazpacho, bread
Petrick [108]	1997–2000	US	183	M&F	20–80	FFQ	Canadian DB	SECO, MATA	0.6 (0.1) ¹	Coffee (31), wine (12), & citrus juice (9%)
Petrick [109]	1993–1995	US	662	M&F	30–79	FFQ	Canadian DB	SECO, MATA	0.07 (0.03) ¹	Coffee (35%), citrus juice (13%), wine (10%)
Williams [110]	2003–2008	US	216	F	55 (13) ¹	FFQ	Canadian DB	SECO, MATA, LARI, PINO	0.1 (0.1–0.2) ³	—
Carmichael [4]	1997–2005	US	3,118	F	~18–40	FFQ	UK DB	SECO (87%), MATA (13%)	0.2 (0.05–0.3) ³	Coffee & tea, alfalfa sprouts, flaxseed

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Waetjen [111]	2008	US	1,459	F	42-52	FFQ	Own DB	SECO, MATA, LARI, PINO	0.2 ²	—
Bandera [112]	2004-2008	US	391	F	>21	FFQ	Own DB	SECO (89%), LARI (6%), PINO (4%), MATA (1%)	1.0 ¹	—
Chang [113]	1995-2007	US	110,215	F	20-84	FFQ	Canadian DB	SECO, MAT, LARI, PINO	0.8 (0.3-1.3) ³	Vegetables, fruits, whole grains
McCann [114]	1996-2001	US	1,122	F	35-79	FFQ	Canadian DB	SECO (50%), PINO (21%) MATA (3%), LARI (3%)	0.2 (0.1) ¹	Whole-grain bread, peaches , orange juice, coffee ,onions, string beans, tea
Fink [43]	1996-1997	US	1,500	F	<65	FFQ	Own DB	SECO, MATA	6.4 (4.7) ¹	Tea (99%), strawberries (0.5%), whole grain products(0.3%)
Horn-Ross [59]	1996-1999	US	470	F	35-79	FFQ	Own DB	SECO (77.9%), MATA (16.9%),	0.2 (0.1-0.2) ³	—
Mervish [115]	2004-2014	US	1,044	F	6-8	24-HDR	Phenol Explorer	All ⁴	0.4 ²	Orange juice (35%), strawberries (17%), broccoli (8%)
van der Schouw [116]	1994	US	468	M	47-83	FFQ	US DB	SECO (97%), MATA (3%)	0.7 (0.5-0.9) ³	Tea & coffee (28%), alcoholic beverages (9%), cereals & grains (7%)
Horn-Ross [117]	1992-1998	US	558	F	20-74	FFQ	Own DB	SECO (71%), MATA (29%)	0.1 (0.1-0.2) ³	—
McCann [118]	1986-1991	US	696	F	40-85	FFQ	US DB	SECO, MATA	0.5 (0.3) ¹	Coffee, carrots, cucumbers, strawberries
Schabath [61]	1995-2003	US	1,735	F (49%)	—	FFQ	US DB	SECO, MATA	5.3 (3.4-9.7) ³	Coffee 52%, tea 30%, flaxseed (6%)
de Kleijn [119]	1991-1994	US	964	F	—	FFQ	US DB	SECO (97%), MATA (3%)	0.6 (0.4-0.8) ³	Other fruits (13%), cereals and grains (11%), berries (8%)

Cotterchio [44]	2001	Canada	1,890	F (47%)	20-74	FFQ	Own DB	SECO, MATA	0.2 (0.1–0.3) ³	Legumes, seeds, cereals/grains, berries, dried fruit, vegetables
Morisset [45]		Canada	115	F	≤70	FFQ	Canadian DB	SECO, MAT, LARI, PINO	0.4 (3.8) ¹	—
Chávez-Suárez [120]	2012-2017	Mexico	100	F	25-80	FFQ,	Own DB	SECO (73%), END (18%), ENL (7%), MATA (2%)	1.1 (1.6) ¹	—
						24-HDR		SECO (75%), END (16%), ENL (5%), MATA (4%)	0.4 (1.8) ¹	
Hernández-Ramírez [46]	2004-2005	Mexico	478	F (46%)	>20	FFQ	Own DB	LARI (54%), PINO (26%), SECO (20%), MATA (0.1%)	0.3 (0.2-0.5) ³	Vegetables, fruits, legumes
Zamora-Ros [33]	2006-2011	Mexico	106,466	F	>20	FFQ	Phenol Explorer	All4: LARI (46%), PINO (21%), SECO (18%)	0.1 (0.03-0.2) ³	Broccoli & cauliflower (11%), strawberries (9%), fruit-flavoured water (6%)
Nascimento-Souza [47]	2016	Brasil	620	F (70%)	60-98	24-HDR	Phenol Explorer	All4: LARI(50%)	13.6 (25.5) ¹	Orange (16%), broccoli (15%), flaxseed (15%)
Miranda [48]	2008 - 2009	Brasil	550	F (65%)	>12	24-HDR	Phenol Explorer	All ⁴	0.1 (0.1-0.2) ³	Sesame seed oil (71%), nuts (20%), sesame seeds (4%)
Miranda [49]	2008	Brasil	1,103	F (54%)	>20	24-HDR	Phenol Explorer	All ⁴	2.3(0.7) ¹	Cereals oil (71%), nuts (26%), olive oil (2%)
Lahmann [50]	2002-2007	Australia	2,078	F	18-79	FFQ	Canadian & UK DB	SECO (68%), LARI (12%), MATA (10%), PINO (8%)	0.7 (0.3) ¹	—
Hanna [51]		Australia	511	F	40-80	FFQ	AusNut DB		2.7 (3.0) ¹	Soy, linseed
Sohrab [52]	2006-2008	Iran	2,618	F (56%)	19-84	FFQ	Phenol Explorer	All ⁴	0.2 (0.1-0.3) ³	Nuts, whole grains
Sohrab [53]	1999	Iran	1,265	F (56%)	19-74	FFQ	Phenol Explorer	All ⁴	3.8 (2.4-5.7) ³	—

Jang [54]	2004	Korea	48	F PreM	40-51	24-HDR	US DB	END (34%), ENL (33%), SECO (28%), MATA (5%)	1.5 (0.3) ¹	—
			53	F PostM	41-57			END (36%), ENL (32%), SECO (25%), MATA (7%)	1.8 (0.5) ¹	—

903 Abbreviations: 24-HDR (24-hour dietary recall), DB (Database), DH (Dietary History), DR (Dietary record), F (Female), FCDB (Food
904 Composition DataBase), FFQ (Food Frequency Questionnaire), LARI (lariciresinol), M (Male), MATA (matairesinol), MED
905 (Mediterranean), MEDI (medioresinol), PINO (pinoresinol), PostM (Post-menopausal), PreM (Pre-menopausal), SECO
906 (secoisolariciresinol), SYRI (syringaresinol)

907 ¹⁻³Type of estimation: ¹mean (SD), ²mean, ³median (p25-p75)

908 ⁴All lignans, including: 1-AcetoxyPINO, 7-HydroxyMATA, 7-HydroxySECO, 7-OxoMATA, Anhydro-SECO, Arctigenin,
909 Conidendrin, CycloLARI, DimethylMATA, Episesamin, Episesaminol, IsohydroxyMATA, IsoLARI, LARI, LARI-sesquiligann,
910 MATA, MEDI, Nortrachelogenin, PINO, SECO, SECO di-O-glucoside, SECO-sesquiligann, Sesamin, Sesaminol, Sesamol,
911 Sesamolin, Sesamolol, SYRI, Todolactol A, Trachelogenin

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914 Table 2. Characteristics of the studies included in the review of urinary lignan excretions.

Author (Reference)	Data collection	Country	N	Sex	Age	Type of study	Urine	Analytical method	END (nmol/L)	ENL (nmol/L)
Zamora-Ros [71]	1995-1999	Europe ⁴	475	F (58%)	33-77	Cohort	24h	LC-MS/MS	247 ²	2,080 ²
Uehar [65]	—	Finland	126	F	24-65	C-S	24h	TR-FIA	—	3,267 ¹
Krogholm [70]	2002-2004	Denmark	84	M	38-63	Cohort	24h	LC-MS	—	768 ¹
			107	F			Overnight		—	696 ¹
							24h		—	1,050 ¹
							Overnight		—	970 ¹
Ward [72]	1993-1997	UK	828	M	45-75	NCC	—	GC-MS	204 ²	2,953 ²
			889	F (43%)					210 ²	3,333 ²
Low [73]	1993-1997	UK	125	F	45-76	Cohort	Spot	LC-MS	288 ¹	2,561 ¹
Grace [74]	1993-1997	UK	219	F	45-75	NCC	Spot	GC-MS	274 ¹	2,792 ¹
Low [75]	1993-1997	UK	267	M	45-75	Cohort	Spot	GC-MS	207 ¹	2,414 ¹
Durazzo [76]	—	Italy	13	F	48-58	CT	24h	LC-CEAD	763 ¹	1,577 ¹
									348 ¹	1,092 ¹
Park [121]	2001-2006	US	404	M	45-75	NCC	Spot	LC-MS/MS	—	1,313 ²
Hu [122]	1997-2010	US	1,111	F	25-55	Cohort	Spot	LC-MS	159 ¹	2,938 ¹
Reger [123]	1999-2010	US	6,009	F (52%)	>40	C-S	Spot	LC-MS/MS	248 ¹	2,041 ¹
Martínez Steele [124]	2009-2010	US	2,692	M/F	>6	C-S	Spot	LC-MS/MS	133 ²	728 ²
Adlercreutz [125]	—	US	10	F	58 ¹	C-S	24h	GC-MS	267 ³	2,120 ³
			10						213 ³	1,533 ³
			7						140 ³	693 ³
Miles [126]	2006	US	80	F (50%)	18-45	CT	24h	GC-MS	533 ¹	3,000 ¹
									267 ¹	1,933 ¹
Rybak [64]	2003-2006	US	2,873	M	≥20	C-S	Spot	LC-MS/MS	41 ¹	302 ¹
				F					38 ¹	285 ¹
Reger [127]	1999-2004	US	5,179	F (52%)	>18	C-S	Spot	LC-MS	133 ²	1,178 ²
Eichholzer [128]	1999-2004	US	2,028	F (49%)	>18	C-S	Spot	LC-MS	147 ²	1,507 ²
	2005-2008		2,628	F (48%)				LC-MS/MS	164 ²	1,683 ²
Xu [67]	2001-2010	US	694	M	12-19	C-S	Spot	LC-MS	278 ¹	2,246 ¹
			600	F	20-60				463 ¹	2,618 ¹
			1,273	M					552 ¹	2,950 ¹
			1,226	F					609 ¹	3,319 ¹

			578	M	>60				533 ¹	3,651 ¹
			584	F					386 ¹	2,907 ¹
Valentín-Blasini [129]	1999-2000	US	334	F (52%)	6-11	C-S	Spot	LC-MS	89 ³	802 ³
			757		12-19				84 ³	852 ³
			1,496		≥20				93 ³	758 ³
Valentín-Blasini [80]	1988-1994	US	199	F (61%)	20-58	C-S	Spot	LC-MS/MS	209 ¹	1,718 ¹
Sun [130]	1995-2001	US	452	F	53-79	NCC	Spot	LC-MS	123 ¹	2,506 ¹
			655		32-52				77 ¹	2,172 ¹
Kunisue [83]	2005-2009	US	10	M	24-63	C-S	24h	LC-MS/MS	43 ¹	738 ¹
			6	F	23-48				129 ¹	872 ¹
Levine [131]	2005-2009	US	471	F	18-40	Cohort	Spot	LC-MS/MS	94 ²	754 ²
Simon [78]	—	Jamaica	171	F	20-75	CC	Spot	TR-FIA	—	2,671 ²
Liu [66]	2000-2001	Japan	500	F	20-70	C-S	Spot	GC-MS	95 ³	148 ³
Uehar [65]	—	Japan	111	F	24-65	C-S	24h	TR-FIA	—	ND
Kunisue [83]	2005	Japan	15	M	22-54				126 ¹	1,376 ¹
			11	F	21-35				80 ¹	1,074 ¹
	2002	Vietnam	31	M	20-78				133 ¹	772 ¹
			32	F	21-73				245 ¹	1,678 ¹
	2006	Vietnam	14	M	21-74				80 ¹	503 ¹
			14	F	33-74				182 ¹	705 ¹
	2000	Cambodia	13	M	21-48				60 ¹	571 ¹
			24	F	21-46				86 ¹	671 ¹
	2005	India	16	M	27-62				179 ¹	1,141 ¹
			23	F	20-70				119 ¹	940 ¹
2006	India	18	M	26-55	255 ¹	1,443 ¹				
		24	F	20-48	205 ¹	1,342 ¹				
Talaei [82]	1999-2004	Singapore	564	F (58%)	45-74	NCC	Spot	LC-MS/MS	228 ¹	1,140 ¹

915 Abbreviations: CC (Case-Control), C-S (Cross-Sectional, CT (Clinical Trial), END (Enterodiol), ENL (Enterolactone), F (Female),
916 HCC (Hospital-based Case-Control), GC-MS (Gas Chromatography–Mass Spectrometry), LC-CEAD (Liquid Chromatography-
917 Coulometric Electrode Array Detector), LC-MS (Liquid Chromatography–Mass Spectrometry), M (Male), NCC (Nested Case-
918 Control), ND (Non Detected), PCC (Population-based Case-Control), TR-FIA (Sensitive Time-Resolved Fluoroimmunoassay).

919 ¹⁻³Type of estimation: ¹mean, ²median, ³geometric mean

920 ⁴France, Italy, Greece, and Germany

921 ENL and END concentrations have been converted into nmol/L from the original studies.

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924 Table 3. Characteristics of the studies included in the review of blood lignan concentrations.

Reference	Data collection	Country	N	Sex	Age	Type of study	Specimen	Methods	END (nmol/L)	ENL (nmol/L)	
Travis [71]	1992-2000	Europe ⁴	1,042	M/F	60.1 ¹	NCC	Plasma	LC-MS/MS	1.0 ²	12.4 ²	
Peeters [26]	1992-2012	Europe ⁴	1,344	F (51%)	54-55	Cohort	Plasma	LC-MS	1.0 ³	8.7 ³	
		UK healthy	70	F (49%)					3.6 ³	17.8 ³	
Pérez-Cornago [68]	1992-2000	Europe ⁴	1,042	M	59.6 ¹	NCC	Plasma	LC-MS/MS	1.0 ³	11.2 ³	
	1993-1997	UK	130	M	64.7 ¹				0.2 ³	4.9 ³	
	1981-1991	Finland, Norway, Sweden	2,209	M	46.5 ¹				TR-FIA	—	5.8 ³
	—	Sweden	1,664	M	60.0 ¹					—	9.6 ³
	1985-2017	Sweden	514	M	58.0 ¹					—	14.6 ³
Uehara [65]	—	Finland	87	F	24-65	C-S	Plasma	TR-FIA	—	25.0 ¹	
Stumpf [132]	1983	Finland	85	M/F	35-49	CT	Plasma	TR-FIA	—	19.5 ²	
Pietinen [133]	1990-1995	Finland	75	F PreM	25-75	PCC	Serum	TR-FIA	—	20.7 ¹	
			133	F PostM					—	28.9 ¹	
Vanharanta [134]	2005	Finland	167	M	42-60	NCC	Serum	TR-FIA	—	23.5 ¹	
Kilkinen [135]	1986-1999	Finland	420	M	50-69	Case-Cohort	Serum	GC-MS	—	18.1 ¹	
Vanharanta [136]	1995	Finland	100	M	58,6 ¹	CT	Serum	TR-FIA	—	16.6 ¹	
Kilkinen [137]	1997	Finland	1,168	M	25-64	C-S	Serum	TR-FIA	—	13.8 ²	
			1,212	F					—	16.6 ²	
Vanharanta [138]	1998-2000	Finland	1,889	M	42-60	Cohort	Serum	TR-FIA	—	17.1 ¹	

Hedelin [91]	2002	Sweden	1,130	M	67.8 ¹	PCC	Plasma	TR-FIA	—	24.0 ¹
Sonestedt [139]	1991-1996	Sweden	728	F	56.3 ¹	NCC	Plasma	TR-FIA	—	16.3 ²
Stattin [140]	2001	Sweden	525	M	59.9 ¹	NCC	Plasma	TR-FIA	—	15.0 ¹
Lin [141]	2003-2004	Sweden	135	F	55-75	Cohort	Serum	TR-FIA	—	23.2 ¹
Hultén [142]	1986-1994	Sweden	308	F	51.2 ¹	NCC	Plasma	TR-FIA	—	22.9 ¹
	1995-2000		185		58.1 ¹				—	20.4 ¹
Aarestrup [143]	1993-1997	Denmark	149	F	50-64	Case-Cohort	Plasma	TR-FIA	—	31.0 ¹
Eriksen [144]	1993-1997	Denmark	850	F (40%)	50-64	Case-Cohort	Plasma	LC-MS/MS	—	10.9 ²
Johnsen [88]	1993-1997	Denmark	857	F	50-64	NCC	Plasma	TR-FIA	—	38.0 ¹
Kuijsten [69]	—	Netherlands	3	F (25%)	28-53	C-S	Plasma	LC-MS	7.0 ¹	39.2 ¹
Milder [30]	1997-2002	Netherlands	637	F (55%)	19-75	PCC	Plasma	LC-MS/MS	1.4 ¹	11.3 ¹
Verheus [145]	1993-1997	Netherlands	87	F PreM	51.6 ¹	NCC	Plasma	LC-MS	0.6 ¹	8.9 ¹
			296	F PostM	58.6 ¹				0.6 ¹	8.9 ¹
Heald [146]	1998-2001	Scotland	483	M	50-74	PCC	Serum	GC-MS	—	16.2 ²
Bhakta [92]	1995-1999	UK	58	F	25-75	PCC	Plasma	TR-FIA	—	13.7 ¹
Ward [72]	1993-1997	UK	815	M	45-75	NCC	Serum	LC-MS	0.7 ²	18.1 ²
			877	F (43%)					0.3 ²	17.4 ²
Morton [85]	—	UK	36	M	41-74	C-S	Plasma	GC-MS	—	13.1 ¹
	—	Portugal	50		35-71				1.2 ¹	13.1 ¹
Low [73]	1993-1997	UK	109	F	45-76	NCC	Serum	GC-MS	1.3 ¹	12.4 ¹
Grace [74]	1993-1997	UK	187	F	45-75	NCC	Plasma	LC-MS	1.3 ¹	12.8 ¹
Low [75]	1993-1997	UK	267	M	45-75	Cohort	Plasma	LC-MS/MS	1.0 ¹	12.8 ¹
Xie [81]	1996-1999	US	802	F	25-42	NCC	Plasma	LC-MS	—	11.5 ²
Bhakta [93]	—	UK	40	F	25-75	PCC	Plasma	TR-FIA	—	28.5 ¹
		UK (Asian)	100						—	13.9 ¹

Piller [147]	1992-1995	Germany	237	F	≤50	PCC	Plasma	TR-FIA	—	9.7 ¹
Zeleniuch-Jacquotte [79]	1985-1991	US	60	F	34-65	Cohort	Serum	CG-MS	1.5 ²	21.2 ²
Valentín-Blasini [80]	1988-1994	US	199	F (61%)	20-58	C-S	Serum	LC-MS	6.0 ¹	11.9 ¹
Horner [148]	—	US	78	M	20-40	C-S	Plasma	TR-FIA	—	11.0 ³
			115	F						13.3 ³
Uehar [65]	—	Japan	111	F	40-60	C-S	Plasma	TR-FIA	—	13.3 ¹
Morton [87]	—	Japan	102	M	40-85	C-S	Plasma	GC-MS	—	32.7 ¹
			125	F	40-89				—	22.8
Morton [85]	—	China	53	M	31-85	C-S	Plasma	GC-MS	5.6 ¹	20.8 ¹
Liu [84]	2010-2012	China	264	F (71%)	35-60	NCC	Plasma	LC-MS	16.4 ²	2.0 ²
Ko [149]	1993-2004	Korea	206	F	60.4 ¹	HCC	Plasma	LC-MS	—	249.3 ¹
			185	M					—	177.8 ¹
	2003-2007	Vietnam	114	F	54.5 ¹				—	10.2 ¹
			92	M					—	10.4 ¹

925 Abbreviations: C-S (Cross-Sectional), CT (Clinical Trial), END (enterodiol), ENL (enterolactone), HCC (Hospital-based Case-
926 Control), GC-MS (Gas chromatography–mass spectrometry), LC-MS (Liquid Chromatography–Mass Spectrometry), NCC (Nested
927 Case-Control), PCC (Population-based Case-Control), PostM (Post-menopausal), PreM (Pre-menopausal), TR-FIA (sensitive time-
928 resolved fluoroimmunoassay)

929 ¹⁻³Type of estimation: ¹mean, ²median, ³geometric mean

930 ⁴Europe: Denmark, France, Germany, Greece, Italy, Netherlands, Spain, Sweden, UK

931 ENL and END concentrations have been converted into nmol/L from the original studies.