1	Kinetic study of colored species formation during paracetamol removal from
2	water in a semicontinuous ozonation contactor
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18	ABSTRACT
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20	Paracetamol aqueous solutions, when ozonized, acquired a strong red coloration
21	depending on the applied ozone dose and the initial pH of the aqueous solution. Then,
22	this color loses intensity and turns to yellow. Color formation is favored when operating
23	at initial $pH_0=12.0$ and ozone flow-rate 4.2 mg/min. A mechanism describing color
24	formation was proposed, being the main pathway involved an initial paracetamol
25	hydroxylation to yield 3-hydroxyacetaminophen followed by the formation of 2-amino-

5-hydroxyacetofenone. Then, these compounds are degraded to colored oxidation by-26 27 products. A model describing color evolution was also proposed, considering first-order kinetics for both color formation and degradation. The corresponding kinetic constant 28 values were determined to be $k_f=0.01$ (1/min) and $k_d=0.03$ pH-0.055 (1/min), 29 respectively. A relationship between aromaticity loss and color changes during the 30 reaction has been estimated considering the parameter $\alpha = k_A/k_f$, being $\alpha = 1.62 \text{ pH} + 3.5$ 31 32 and the first-order rate constant for aromaticity loss given by $k_A=0.0162 \text{ pH} + 0.035$ (1/min). 33

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35 KEYWORDS

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37 Chemical pathway; Color; Kinetic modelling; Ozone; Paracetamol; Initial pH

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39 **1. Introduction**

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41 Among the so called contaminants of emerging concern, pharmaceuticals stand out as 42 one of the most detected families of compounds in wastewater effluents, but also in surface waters and drinking waters (Mompelat et al., 2009; Richardson, 2009; Seifrtová 43 44 et al., 2009; Sun et al., 2014). These pollutants enter the water resources by means of industrial, agricultural and domestic residual effluents that are transported to wastewater 45 treatment plants (WWTPs), where many of them -because of their chemical properties-46 47 cannot be effectively eliminated by the conventional treatment technologies typically implemented in those facilities (Blair et al., 2015; Baalbaki et al., 2016). Current 48 research focus on the development of chemical processes able to degrade recalcitrant 49

chemicals from wastewater effluents and natural water compartments, in order to avoid
their presence in urban water distribution networks (Ziylan-Yavaş and Hince, 2018). In
this context, Advanced Oxidation Processes (AOPs) are considered a promising
alternative (Hollender et al., 2009; Rosario-Ortiz et al., 2010; Oller et al., 2011; Margot
et al., 2013; Lee et al., 2016).

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This work focused on the study of paracetamol (4-hydroxyacetanilide, 4-56 57 acetamidephenol or acetaminophen) degradation by means of ozonation. This pharmaceutical is employed as anti-inflammatory and analgesic (Skoumal et al., 2006) 58 59 and, despite human organism is capable of metabolizing up to a 90% of the consumed drug -excreting only a 3-5%- (Ellenhorn and Barceloux, 1988; McEvoy, 1992), the 60 worldwide amount of this chemical reaching WWTPs have been estimated to be in the 61 range of 292-585 ton/year (El Najjar et al., 2014). Moreover, this pollutant has been 62 often found in municipal sewage effluents at concentrations up to 65 µg/L (Villaroel et 63 al., 2014). Although the concentration of paracetamol detected in natural waters is 64 generally below the µg/L range (Skoumal et al., 2006), its presence may potentially 65 affect aquatic organisms (Santos et al., 2010) due to the ability of this chemical to 66 bioaccumulate (Radjenović et al., 2009; Deblonde and Hartemann, 2013; Valdés et al., 67 2014; Zenker et al., 2014). Studies reported in the bibliography show that AOPs are 68 69 usually effective treatments for degrading this kind of organic pollutants from water (Ay and Kargi, 2001). In addition, and due to the low concentrations of these substances 70 71 in water matrices, membrane technology can be used to concentrate micropollutants 72 prior to AOPs treatments reducing the volume of water to treat (Savchuk and Krizova, 73 2015).

Ozone is a powerful oxidant employed in drinking and wastewater treatment plants due 75 76 to its ability for microorganisms inactivation and both inorganic and organic contaminants transformation (Paraskeva and Graham, 2002; Rodríguez et al., 2008; 77 Gomes et al., 2017). However, the application of ozone presents some practical 78 limitations related to operational pH: under acidic conditions, O₃ is relatively stable and 79 directly reacts with pollutants, whereas for pH values corresponding to neutral and basic 80 conditions this oxidizing species decomposes to highly oxidizing radicals (*i.e.*, hydroxyl 81 radicals •OH) that enhance the process efficiency (Rodríguez et al., 2017). In addition, 82 the physicochemical properties (i.e., pKa and second-order rate constants with O₃ and 83 84 OH•) of micropollutants determine their removal efficiencies according to the medium 85 pH, which can be different depending on the wastewater origin. Therefore, it is interesting to perform a study that covers a wide range of operational pH. Furthermore, 86 87 it is necessary to ensure a reduction in the toxicity of the ozonized effluents, which is normally attributed to the toxic character of the transformation products generated 88 because of the treatment application (Wert et al., 2007; Lee and von Gunten, 2016; 89 Maya et al., 2018). Thus, in order to ensure the treatment effectiveness, it is necessary to 90 91 perform a complete study of the process assessing reaction kinetics, transformation 92 products and toxicity of the treated water (Cruz-Alcalde et al., 2017).

93

During organic matter ozonation, a reduction in the relative amount of aromatic rings and conjugated bonds contained in the water matrix is produced. Also, the number of electron acceptors (*i.e.*, carboxyl, carbonyl, hydroxyl, alcoxy) increases (Swietlik and Sikorska, 2004). Ozone preferently degrades organic molecules with low oxidation state (low O/C ratio) and a high degree of insaturation (high H/C ratio), yielding more saturated and oxygenated (alcohols, aldehydes, carbonyls, ketones and carboxylic acids) 100 (Reemtsma and These, 2005; von Gunten, 2003). Among them, the relative amount of 101 carboxylic acids generated during the process is generally much higher than that of 102 aldehydes or ketones (Nawrocki and Kasprzyk-Hordern, 2003). They typically are 103 compounds with short molecular chains (less than 5 C atoms), such as formic, acetic or 104 oxalic acid (Can and Gurol, 2003).

105

106 During paracetamol ozonation, aqueous solutions of this chemical acquire a red tonality 107 which gradually turns to yellow. In the present work, therefore, and since color is an important organoleptic parameter determining water quality (Villota et al., 2016), 108 109 degradation intermediates causing this change of tone in water have been identified. According to the transformation products detected through LC-MS, the oxidation 110 111 mechanism taking place simultaneously to the mentioned color changes in paracetamol 112 solutions has been proposed. Moreover, the influence of main operational parameters of ozonation process (*i.e.*, initial pH and applied ozone flow-rate) on color formation, 113 114 degree of mineralization and aromaticity, as well as on the mass transfer of ozone to the 115 aqueous phase, has been studied including some toxicity studies (El Najjar et al., 2014). 116 By this way, operational conditions leading to the formation of chromophoric 117 intermediate (quinone compounds) increasing water toxicity have been characterized (Mijangos et al., 2006). 118

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120 **2. Materials and methods**

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122 **2.1.** Chemicals and reagents

Paracetamol solutions [Pa]₀=50.0 mg/L were prepared by dissolving pure paracetamol 124 125 (Sigma-Aldrich 99.9%) in milli-Q water produced by a filtration system (Millipore, 126 USA). Assays were performed operating under different initial pH conditions (pH₀ between 3.0 and 12.0) in order to assess the effect of this parameter on color formation 127 during ozonation of paracetamol aqueous solutions. This range was selected to cover a 128 wide range of water properties. The initial pH was adjusted by adding a few drops of 129 130 concentrated HCl and NaOH solutions in order to avoid sample dilution. Although the operational pH has not been controlled during the reaction, its maximum variation has 131 been less than 0.5 pH units (Mijangos et al., 2006). Pure oxygen (≥99.999%) for ozone 132 133 production was supplied by Abelló Linde (Spain). The reproducibility of the ozonation 134 assays was considered based on the mean of the colour of the values measured, obtaining standard deviations less than 5% in all cases. 135

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137 2.2. Semicontinuous ozonation setup

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For each ozonation experiment, 1.0 L of paracetamol solution was prepared and added 139 140 into a jacketed reactor operating in semi-continuous mode. Experiments were performed 141 at a constant temperature of 25.0 °C, maintained by means of a thermostatic bath. 142 Ozone/oxygen gaseous mixtures were generated by means of a 301.19 Labor Ozonator 143 (Sander, Germany) and injected at the bottom of the reactor employing a metallic difusser (pore size: 10 µm). The ozone flow rate applied was in the range 4.2 - 25.0 144 mg/min. The medium was under stirring conditions in order to ensure homogeneity. The 145 146 inlet and outlet ozone concentration in the gas phase were meausured by means of two BMT 964 ozone analyzers (BMT Messtechnik GMBH, Germany), placed up and 147 148 downstream the contactor, respectively. More information about the experimental setup may be found elsewhere (Marcé et al., 2016). The dissolved ozone concentration was
measured by the Indigo colorimetric method (Bader and Hoigné, 1981; Greenberg et al.,
151 1999).

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153 **2.4. Analytical procedures**

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Paracetamol mineralization was estimated through total organic carbon (TOC, mgC/L) 155 156 measurements using a TOC-VCSN Shimadzu Analyzer. Color was determined using an UV/Vis spectrophotometer (DR 6000-Hach Lange) by direct measurement of the 157 158 samples absorbance at λ =455 nm (Mijangos et al., 2006). For aromaticity loss monitoring, UV absorbance measurements were performed at λ =254 nm. The samples 159 were analyzed by Liquid Chromatography-Mass Spectrometry (LC-MS) to elucidate the 160 paracetamol degradation pathways that induce high levels of color in the water during 161 the ozonation process. Samples were analyzed by HPLC coupled in series to a MS-TOF 162 (G3250AA by Agilent, USA) system and a UV detector (1100 Agilent). This 163 164 configuration allowed the comparison between chromatograms obtained through UV 165 and MS detectors, discarding this way interferences caused by artifacts.

MS data were collected in full scan mode (50-1500 m/z), employing negative 166 167 electrospray ionization. The column employed was a Teknokroma Mediterranea Sea 18 168 (250 mm \times 4.6 mm and 5 µm size packing). The mobile phase consisted of a 65:35 volumetric mixture of methanol and Milli-Q water acidified at pH 3.0 by the addition of 169 H₃PO₄. The flow rate was maintained at 0.3 mL min⁻¹ and the detection wavelength (for 170 171 the UV detector) was set to 200 nm and 243 nm. The spectrophotometer conditions were voltage of capillary 3500 V and shredder 125 V, being the limit of detection 172 (LOD) 0.25 µM. 173

175 **3. Results and discussion**

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177 **3.1. Effect of applied ozone flow-rate on color formation**

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179 During the ozonation of aqueous solutions of paracetamol, a general aromaticity loss in 180 the reaction medium is observed (Fig. 1). This fact confirms organic matter oxidation, leading to changes in molecular structures of species initially contained in the water 181 matrix. During ozonation treatment, the highest absorption observed in the UV 182 183 spectrum corresponds to the opening of aromatic rings causing the transformation of organic molecules (Xiong and Legube, 1991). This reaction is fast and it is 184 185 characterized by the decrease of UV absorbance measured at 254 nm (von Gunten, 186 2003).

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Results show a total loss of the aromaticity in the water produced when ozonation of 50.0 mg/L paracetamol solutions employing ozone flow rates higher than 7.5 mg O₃/min is applied. It is also observed that an increase of ozone flow-rate causes an increase in the rate of aromaticity reduction. However, this increase does not follow a linear tendency, which would indicate the existence of particular operation conditions leading to the activation of different degradation pathways through which the overall oxidation mechanism of paracetamol takes place.



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Fig. 1. Influence of the inlet ozone flow rate during paracetamol oxidation experiments
on aromaticity loss. Experimental conditions: [Pa]₀=50.0 mg/L; pH₀=12.0; T=25.0°C.

200 Simultaneously to aromaticity loss, color changes in oxidized waters also take place, 201 indicating the kinetic evolution of a reaction intermediate (Fig. 2). Initially, colorless 202 aqueous solutions containing paracetamol Color₀ (AU) gradually acquire a strong red 203 tonality, until the maximum color intensity Colormax (AU) is reached at a certain 204 reaction time t_{max} (min). Then, color acquires a yellowish tone with time-decreasing intensity until a residual value of this parameter is reached (Color_∞, AU). This fact 205 206 would indicate the simultaneous cleavage of paracetamol molecules and subsequent 207 formation of chromophoric groups -such as -C=O, -C=C y NO₂- in the respective 208 structures of the degradation products.

209

Observed changes in color show an asymmetric bell-shaped tendency, with thedecreasing part of this curve presenting a tail. This evolution can be explained by the

two stages taking place during ozonation of paracetamol (El Najjar et al., 2014). The 212 213 first of these regimes, characterized by exhibiting fast oxidation kinetics, occurs during 214 the first minutes of the process. During this period, in which a loss in water aromaticity 215 is observed, paracetamol is oxidized to yield different reaction intermediates, some of them containing chromophoric terminations responsible for color appearance in the 216 217 water matrix. Then, a slower ozonation stage starts to take place. At this point of the 218 process, reaction intermediates -including colored species are slowly degraded to smaller molecules, ultimately leading to the formation of colorless carboxylic acids (i.e., 219 220 oxalic, acetic and formic).

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In order to analyze the effect of ozone flow-rate on color formation and degradation 222 223 kinetics, the color signal (in color area units) as a function of the ozone flow-rate inlet 224 has been represented (see Fig. 2). These results allow checking how, regardless of the employed operational conditions, paracetamol ozonation always leads to intermediates 225 226 containing chromophoric groups in their molecular structures. However, colored species 227 formation is better observed under low inlet ozone flow-rates ([O₃]=4.2 mg/min). When 228 gradually increasing this dosing rate up to 25.0 mg/min, color curves with smaller areas 229 are registered due to faster formation and degradation of chromophoric reaction intermediates. 230



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Fig. 2. Effect of flow rate of ozone applied during paracetamol oxidation on the color appearance in water matrices. Experimental conditions: [Pa]₀=50.0 mg/L; pH₀=12.0;
T=25.0°C. Legend: Inlet flow rate of ozone [O₃]= • 4.2 mg/min, ○ 7.5 mg/min, ▲ 10.0 mg/min, ■ 15.0 mg/min, □ 25.0 mg/min.

To explain this phenomenon, it is necessary to analyze the reaction time –as a function of the volumetric ozone flow-rate– for which the maximum color intensity in water matrix is observed (t_{max} , min). This is shown in Fig. 2. This t_{max} parameter does not proportionally decrease with increasing ozone flow-rate. Instead, a maximum is observed for certain operational conditions, which would indicate that ozone does not exert a catalytic effect in oxidation but determines the degradation yield of paracetamol leading to colored species formation.

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The ozone flow-rate employed during the process produces a selective paracetamol oxidation to yield colored species through different degradation routes. The overall degradation mechanism does not consist of a unique degradation via leading to the 249 generation of a single compound. On the contrary, a mix of chemical species coexists in 250 the system. Therefore, the observed color kinetics would be caused by reaction 251 intermediates presenting different molecular structures. The time for which the 252 maximum color intensity is registered would indicate the generation of a group of 253 predominant colored species, as well as the oxidative level in which these compounds 254 are formed throughout the overall degradation mechanism.

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256 **3.2. Pathway of colored intermediates generation**

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258 Ozonized aqueous solutions of paracetamol ([Pa]₀=50.0 mg/L) corresponding to experiments with an ozone flow-rate of 4.2 mg/min and pH₀=12.0 as experimental 259 260 conditions, have been analyzed by LC-MS to determine the nature of intermediates 261 causing color in water. These are the operational conditions for which a higher color intensity remained in the residual water ozonized that makes possible to analyze the 262 263 compounds contained in the colored water. In this situation, therefore, the formed 264 chromophoric intermediates could be analyzed before their subsequent transformation 265 to colorless acids. On the basis of the obtained results, and considering the degradation 266 mechanisms of paracetamol reported in scientific literature, (He et al., 2018; Villota et al., 2018; Villota et al., 2016; Martignac et al., 2013; Moctezuma et al., 2012), an 267 scheme of those reactions causing color generation in ozonized waters has been 268 269 proposed (Fig. 3).

270

The nature of the detected ozonation by-products depends on the employed flow-rate of ozone. Previous works show that employing lower ozone dosages and high substrate concentrations leads to the detection of reaction intermediates containing from 4 to 6 C

atoms, whereas employing elevated ozone dosages favored the formation of smaller
molecules generally containing 1-3 C atoms (Brunet et al., 1982). For the current case
of study, therefore, the color observed during paracetamol ozonation operating at flowrates lower than the stoichiometric value, may be induced by intermediate species
exhibiting molecular structures with 4-6 carbon atoms.

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280 281

Fig. 3. Reaction intermediates generated during the ozonation of paracetamol aqueous solutions. Experimental conditions: $[Pa]_0=50.0 \text{ mg/L}$; $pH_0=12.0$; $[O_3]=4.2 \text{ mg/min}$; T=25.0°C.

Analysis on the colored reaction mixture would indicate the formation of some 286 287 transformation products such as hydroquinone, benzoquinone, p-aminophenol and p-288 nitrophenol. In accordance with the suggested mechanism, three reactions involving the initial attack of formed hydroxyl radicals (HO•) to finally yield ortho-meta- and para-289 290 hydroxylated compounds (with respect to the initial hydroxyl group of paracetamol) 291 would compete to simultaneously take place in the system. Existent works dealing with 292 this subject have reported that, during the initial stages of the oxidation process, 293 hydroxyl and amino groups present a high reactivity towards ortho- and para- positions, 294 due to the high electron-donating capacity exhibited by these moieties (Westerhoff et 295 al., 1999). Contrarily, reactions occurring at meta- positions are less favored and, 296 therefore, hydroxylation reactions at this point of the aromatic ring are considered to be 297 of minor relevance (Vogna et al., 2002).

298

Oxidation could be initiated via hydroxyl radical addition to the ortho-position of paracetamol, leading to the formation of 3-hydroxyacetaminophen. The predominant pathway, however, is the one initiated by the generation of the paracetamol isomer 2amino-5-hydroxyacetophenone. This reaction implies the migration of an acetyl group from the amino moiety of paracetamol in position para- (with respect to the hydroxyl group) to position ortho- (in the aromatic ring) with respect to the amino (-NH₂) group.

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The presence of p-aminophenol and p-nitrophenol in the colored mixture has been also detected. This fact would indicate a deacylation mechanism, subsequently leading to the generation of an amide group bonded to the aromatic ring (Bahnemann et al., 2007). Due to the fact that hydroxyl radicals mainly react through addition to the aromatic ring, p-nitrophenol would be generated by oxidation of p-aminophenol. First, the addition of HO' to the aromatic ring would be produced, followed by the elimination of a hydroxyl group. Then, a deprotonation of the formed radical cation would generate an easily oxidizable anilinyl radical, to finally yield p-nitrophenol (Martignac et al., 2013).

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During p-nitrophenol oxidation, two competitive chemical transformations could take place: an hydroxylation in ortho- position to finally yield nitrocatechol, on one hand, and a hydroxylation in para- position (with respect to the phenolic hydroxyl group) and later elimination of the nitro group, obtaining by means of this way hydroquinone as product (Moctezuma et al., 2007). Further hydroquinone oxidation would finally yield benzoquinone.

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322 3.3. Influence of initial pH and oxidant dosage on ozonized water coloration

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Color formation in ozonized water has been analyzed, considering both the pH_0 (Fig. 4) 324 325 and ozone flow-rate ([O₃] mg/min), (Fig. 5) employed during ozonation assays. The aim 326 of this study was to determine whether the degradation route causing color formation 327 consisted of a direct ozone oxidation of paracetamol or, on the contrary, could be 328 attributed to the unselective oxidation by hydroxyl radicals generated through ozone decomposition (pH_0 values higher than 6.0). This part of the work was performed 329 operating within an initial pH range of 3.0-12.0 (see Fig. 4). Normalized TOC (mg/L) 330 and absorbance at 254 nm ([Arom], AU) values, together with the color (Color_∞, AU) 331 332 and the ozone mass transfer coefficient (K_La, l/min) determined at the end of treatment have been all represented. The maximum color generated during ozonation experiments 333 334 has been also included.

The determination of ozone transfer parameters may be done according to the equations 336 337 of gas-liquid mass transfer (Rodriguez et al., 2017). Ozone consumption N₀₃ (mg/L min) can be expressed as the volumetric mass transfer coefficient K_La (1/min) 338 339 multiplied by the mass transfer driving force, according to Eq. 1. The volumetric mass transfer coefficient has been estimated considering the mass transfer taking place from 340 gas to liquid phase, being C_i (mol/L) the ozone concentration at the interphase, C_L 341 342 (mg/L) in the liquid phase and C_L^* in the liquid phase in equilibrium with the gas (mg/L). Note that in Eq. 1 K_La (1/min) is the global mass transfer coefficient, whereas 343 k_Ga and k_La represent the individual coefficients for gas and liquid phases, respectively. 344

345

346
$$N_{O_3} = K_L a (C_L^* - C_L) = k_G a H (C_L^* - C_i) = k_L a (C_i - C_L)$$
 (1)

347

Ozone mass transfer can be also expressed as a function of the ozone flow rate (Q_{O3} , L/min), the reaction volume (V, L) and the difference between the inlet and outlet ozone concentrations in the gas phase ($C_{G,in}$ and $C_{G,out}$ (mg/L), respectively) (see Eq. 2). It is worth to mention here that transferred ozone can be accumulated in the liquid phase or be consumed through two main mechanisms: direct oxidation reactions with the contaminant (R_{O3} , mg/L min), or O₃ self-decomposition leading to the generation of hydroxyl radicals (r_{O3} , mg/L min).

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356
$$N_{O_3} = \frac{Q_{O_3}}{V} (C_{G,in} - C_{G,out}) = K_L a (C_L^* - C_L) = \frac{dC_L}{dt} + R_{O_3} + r_{O_3}$$
 (2)

Ozone concentration in the liquid phase in equilibrium with gas phase can be estimated considering the Henry Law (Eq. 3), where H (atm/ozone molar fraction in the liquid) is the Henry constant and P_G (atm) is the partial pressure of ozone in the gas phase.

361

$$362 C_{\rm L}^* = \frac{P_{\rm G}}{\rm H} (3)$$

363

The Henry constant has been determined as a function of the liquid phase temperature (K) and pH, according to Eq. 4 (Roth and Sullivan, 1981).

366

367
$$H = 38 \cdot 10^{6} C_{OH^{-}}^{0.035} \exp\left(\frac{-2428}{T}\right)$$
(4)

368

Obtained results allow checking how both aromaticity loss and residual color of 369 370 ozonized waters do not experiment important variations with the medium pH₀. 371 However, the maximum color generated in water shows a strong dependence from pH: when operating in the initial pH range comprised between 3.0 and 6.0, this intensity 372 remains constant, whereas if this parameter is increased from 6.0 to 12.0, a potential rise 373 374 in its value is observed. To explain these results, it is necessary to consider the 375 Henderson-Hasselbach equation concerning the ionization curve of paracetamol. When 376 water pH is increased, the concentration of hydroxide anions in the system also 377 increases, and consequently does the ozone decomposition to hydroxyl radicals. This 378 fact causes an enhancement in the oxidizing capacity of the system, leading to a higher degradation efficiency of the pharmaceutical to yield colored transformation products. 379

On the other hand, paracetamol is a weak acid, with a pKa value equal to 9.38 381 382 (Dastmalchi et al., 1995). This is going to determine the reactivity of this compound. 383 When assays are performed within the operational pH interval comprised between 3.0 and 9.4, as the system is acidified, the number of non-dissociated or non-ionized 384 molecules is relatively larger. On the contrary, for pH values higher than 9.4 (under 385 basic conditions) the proportion of dissociated molecules increases. This fact could 386 387 explain the strong color rise observed when operating at a pH value of 12.0, since paracetamol molecules under these conditions exhibit a high degree of dissociation. 388 Because of this, this specie is degraded through oxidative routes conducting to the 389 390 generation of chromophoric structures that provide color to the ozonized water.

391

392 As indicated in the relationship by Roth and Sullivan, 1981, if the system pH is 393 increases this favors the ozone transfer to the reacting solution. However, obtained results in the current case of study suggest that the ozone transference to aqueous 394 395 solutions of paracetamol is maximum when the process is performed at a pH value of 396 6.0. This phenomenon is accompanied by a decrease in the total organic carbon (TOC, 397 mg/L). To explain this phenomenon, it is necessary to consider the nature of chemical 398 species that are present in the reaction medium, since they can either accelerate the decomposition of ozone to hydroxyl radicals (radical chain reactions) or slowing it 399 down. In this case, the pKa value for paracetamol (9.4) determines the process 400 401 performance. The more acidic is the media the higher is the number of non-dissociated 402 paracetamol molecules. On the contrary, at pH > pKa conditions, the number of unprotonated molecules becomes larger. This fact can considerably impact the ozone 403 404 reactivity with this pharmaceutical. In addition, when operating within a pH range

between 6.0 and 12.0, the ozone concentration in water increasingly diminishes due a
higher concentration of hydroxide (OH⁻) ions.

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This clearly affects the ozone transfer to the liquid phase, which is reflected in lower K_La values with increasing pH values. In addition to the effect exerted by the hydroxide ion, it is possible that, because of paracetamol mineralization, some inorganic species (mainly nitrogen anions) could be formed. These ions would act as ozone and radical scavengers, also affecting the ozone transfer to the liquid phase (Gottschalk et al., 2010; Sumegova et al., 2013).

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Fig. 4. Influence of initial pH on color (AU) formation, mass transfer coefficient K_La
(1/min), organic matter mineralization TOC/TOC₀ and aromaticity loss
([Arom]/[Arom]₀) during paracetamol ozonation. Experimental conditions: [Pa]₀=50.0
mg/L; [O₃]=4.2 mg/min; T=25.0°C; t=120 min.

The influence of the employed ozone dose was evaluated in ozonation assays carried 421 422 out at pH=12.0. As mentioned, these conditions potentiate the formation of colored structures (Fig. 5). In this case, aromaticity loss exhibits a strong dependence with 423 424 ozone dose: when working at ozone flow rate below 4.2 mg/min for a total reaction time of 2 h, the aromaticity reduction is less than 4%. If on the contrary, the oxidant flow rate 425 is increased up to 7.5 mg/min, an abrupt drop of this parameter (approximately a 95%) 426 427 is observed. The oxidant/substrate ratio corresponding to these experimental conditions, 428 57 mol O₃/mol C₈H₉NO₂, would therefore correspond to that stoichiometric relationship required in order to produce a reduction in the aromaticity of the paracetamol 429 430 molecules. If ozone flow-rate is further increased, the observed final percentage of aromatic species in solution is less than 5%. From this point, the depletion of this 431 432 residue follows a linear behavior with the ozone dose increment: for an ozone flow rate 433 of 1.0 L/min, for instance, water aromaticity reduction is 99%.





Fig. 5. Effect of ozone flow rate during paracetamol oxidation on color formation (AU), mass transfer coefficient K_La (1/min), organic matter mineralization TOC/TOC₀ and aromaticity loss ([Arom]/[Arom]₀). Experimental conditions: [Pa]₀=50.0 mg/L; $pH_0=12.0$; T=25.0°C; t=120 min.

Ozone transference to the aqueous solution increases with increasing inlet ozone flowrates, but only if operation pH is constant. This enhancement in the gas-liquid transfer leads to an increase of the water-color formation. This phenomenon appears to indicate that pH conditions define the selectivity ozonation mechanism, while the efficiency of paracetamol degradation leading to colored species formation would be more influenced by the applied ozone doses.

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It has been verified that a residual coloration lasts in ozonized waters when the provided
ozone flow-rate is 4.2 mg/min. That color is less intense when the inlet flow of ozone is
7.5 mg/min. This maximum intensity value of the residual color would indicate that,

when operating at ozone doses below the stoichiometric relationship of 1/57 (mol paracetamol/mol ozone), the amount of oxidant provided to the system would not be enough to produce those significant alterations in the aromatic structures of paracetamol subsequently leading to a noticeable reduction in water aromaticity. Because of this, an aromatic chromophoric residue –conferring that characteristic color to the medium– persists in the treated waters.

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- 459 **3.4. Kinetic model for water coloration**
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The kinetic model suggested in the present study relates the aromaticity loss of ozonized paracetamol solutions with changes in color observed in the water matrix. In order to model aromaticity loss, in first place, a mathematical model based on the scheme shown in Fig. 1 has been considered. Thus, it is assumed that paracetamol ozonation process leads to the generation of transformation products exhibiting less aromaticity than the parent compound. The aromaticity reduction rate is given by the corresponding firstorder kinetic constant k_A (1/min).

468

469 Paracetamol $\stackrel{k_{A}}{\rightarrow}$ deg radation intermediates (5) 470 With, 471 472 $k_{A:}$ first-order kinetic constant describing aromaticity loss in aqueous solutions

472 kA: Inst order kneele constant deserioning aromaterly loss in aqueous solution.
473 of paracetamol during ozonation process (l/min)

In accordance with Eq. 5, a mass balance corresponding to aromaticity loss during
paracetamol ozonation is considered (Eq. 6). It is also assumed that reduction of water
aromaticity follows first-order kinetics.

478

479
$$\frac{d \left[\text{Arom}\right]}{dt} = -k_{\text{A}} \left[\text{Arom}\right]$$
(6)

480

481 being,

482

483 [Arom]: water aromaticity (AU)

484 t: time (min)

485

By integrating the mass balance (Eq. 7), the kinetic equation corresponding toaromaticity loss can be obtained (Eq. 8):

488

489
$$\int_{Arom_0}^{Arom} \frac{d [Arom]}{[Arom]} = -k_A \int_{t=0}^{t} dt$$
(7)

490

491
$$[Arom] = [Arom]_0 \exp(-k_A t)$$
 (8)

492

In Fig. 6, both predicted and experimental results on aromaticity loss are shown, these allowing the estimation of the first-order kinetic constant describing aromaticity loss $(k_A, 1/min)$ as a function of the pH (Eq. 9). On the other hand, it is verified that aromaticity of ozonized samples at the end of the treatment does not depend on initial pH, estimating a mean value for this parameter in the order of 0.06 AU (Eq. 10) that it

has been depreciated in the proposed model. Table 1 gathers the values of the estimated 498 499 kinetic parameters.

500
501
$$k_{A} = 0.0162 \text{ pH} + 0.035$$
 (9)
502 $r^{2}=0.9301$
503
504 $\overline{[\text{Arom}]}_{r} = 0.06 (\text{AU})$ (10)

504
$$\overline{[\text{Arom}]}_{\infty} = 0.06 (\text{AU})$$

505



506

Fig. 6. Effect of initial pH on aromaticity loss for ozonized paracetamol solutions. 507 508 Experimental conditions: Pa₀=50.0 mg/L; [O₃]=25.0 mg/min; T=25.0°C.

509

Color changes taking place in water during paracetamol ozonation have been modelled 510 based on studies reported in the bibliography about the changes of color observed in 511 512 aqueous solutions of organic compounds oxidized by Fenton technologies (Villota et al., 513 2018), following the scheme shown in Eq. 11. As mentioned, colorless aqueous

solutions of this pharmaceutical acquire a strong reddish tonality that turns to yellow when waters exhibit their maximum degree of coloration (Color_{max}). The rate of color formation has been expressed as a function of the kinetic constant k_f (1/min). Then, color is gradually degraded until a residual tonality is reached (Color_∞). The velocity of color degradation is given by the corresponding first-order kinetic constant, k_d (1/min).

519

$$\begin{array}{cccc} & k_{\rm f} & k_{\rm d} \\ 520 & {\rm Color}_{\rm 0} & \rightarrow & {\rm Color}_{\rm max} & \rightarrow & {\rm Color}_{\infty} \end{array} \tag{11}$$

521 Being,

523	Color ₀ :	initial color of paracetamol aqueous solutions (AU)
524	Color _{max} :	maximum intensity presented by oxidized paracetamol solutions (AU)
525	Color∞:	color of oxidized paracetamol solutions at the end of treatment (AU)
526	k _{f:}	first-order kinetic constant for color formation during paracetamol
527		ozonation (l/min).
528	k _{d:}	first-order kinetic constant for color degradation during paracetamol
529		ozonation (l/min).
530	t ₀ :	initial time (min)
531	t _{max} :	time for which treated solutions show a maximum in color intensity (min).
532	t∞:	time corresponding to end of treatment (min)
533		
534	According t	o the reaction scheme proposed in Eq. 11, the mass balances corresponding
535	to color form	nation-degradation processes have been applied (see Eqs. 12-13).
536		

537
$$\frac{d\text{Color}}{dt} = k_{f} \left(\text{Color} + \text{Color}_{0} \right) \left[\int_{\text{Color}_{0}}^{\text{Color}_{\text{max}}} \right]$$
(12)

539
$$\frac{d\text{Color}}{dt} = -k_{d} \left(\text{Color} + \text{Color}_{\infty} \right) \Big|_{\text{Color}_{\text{max}}}^{\text{Color}_{\infty}}$$
(13)

540 Considering the fact that color evolves according to first-order kinetics, mass balances 541 above expressed have been both integrated to obtain explicit kinetic expressions for 542 color formation (Eq. 14) and degradation (Eq. 15).

543

544
$$\operatorname{Color} = 2 \operatorname{Color}_{0} \exp\left(k_{f} t\right) - \operatorname{Color}_{0} \qquad \begin{bmatrix} \operatorname{Color}_{\text{max}} \\ \operatorname{Color}_{b} \end{bmatrix}$$
(14)

545

546
$$\operatorname{Color} = (\operatorname{Color}_{\max} - \operatorname{Color}_{\infty}) \exp \left[-k_{d} \left(t - t_{\max}\right)\right] + \operatorname{Color}_{\infty} \left[\operatorname{Color}_{\max} \right]_{\operatorname{Color}_{\max}}^{\operatorname{Color}_{\infty}}$$
(15)

547

548 In Fig. 7, predictions made according to the proposed model are shown. Table 1 also 549 gathers the values of the estimated kinetic parameters. Obtained results allow concluding that the kinetic constant describing color formation (kf, 1/min) does not 550 551 depend on pH. In fact, a mean value of $k_f=0.01$ 1/min could be estimated (Eq. 16). 552 However, the kinetic constant for color degradation presents a linear dependency with operation pH (Eq. 17). This fact could be explained as follows: despite the treatment 553 554 conditions appear to be powerful enough to degrade paracetamol molecules to colored 555 transformation species, further degrading these intermediates to colorless byproducts 556 could require a more severe oxidation. Because of this, in order to increase the rate of 557 color degradation it is necessary to favor -during ozonation process- the generation of more oxidizing species (*i.e.*, hydroxyl radicals) by taking advantage of the pH effect. 558

560
$$\overline{\mathbf{k}_{\mathrm{f}}} = 0.01 \left(1/\min \right) \tag{16}$$

562
$$k_d = 0.03 \, \text{pH}_0 - 0.055$$
 (17)

 $r^2 = 0.9783$

The influence of operational pH on the color of the treated water has been also assessed (Eqs. 18-20). As can be seen, this color intensity gets larger with increasing operation pH due to the fact that a higher concentration of oxidizing species is generated, causing the degradation of paracetamol to chromophoric reaction intermediates. In a similar way, the residual color at the end of the treatment is less intense when the reaction is carried out under basic medium conditions. This is explained by an enhancement in the oxidation efficiency of the process, leading to the generation of more hydroxyl radicals per consumed ozone, which allows a further oxidation of colored species to colorless degradation products. Therefore, the time for which the oxidized water reaches its peak color intensity diminishes with increasing operational pH.

576
$$\operatorname{Color}_{0} = 0.0006 \,\mathrm{pH}_{0} + 0.006$$
 (18)

 $r^2 = 0.9818$

579
$$\operatorname{Color}_{\max} = 0.0016 \,\mathrm{pH}_0^2 - 0.0139 \,\mathrm{pH}_0 + 0.0378$$
 (19)

 $r^2 = 0.9995$

 $\text{Color}_{\infty} = 0.002 - 0.0004 \text{ pH}_0$

 $r^2 = 0.9831$

(20)

586 $r^2 = 0.9884$

587



588

Fig. 7. Influence of initial pH on water color formation during ozonation of paracetamol aqueous solutions. Experimental conditions: $Pa_0=50.0 \text{ mg/L}$; $[O_3]=25.0 \text{ mg/min}$; T=25.0°C.

592

According to the previous estimations, a parameter α (Eq. 23) expressing the relationship between formed color and aromaticity loss as a function of pH has been defined. To do so, the kinetic constants k_f (1/min) and k_A (1/min) have been both considered, as expressed in Eq. 22.

597

$$\mathbf{598} \qquad \mathbf{k}_{\mathrm{A}} = \alpha \, \mathbf{k}_{\mathrm{f}} \tag{22}$$

$$600 \qquad \alpha = 1.62 \text{ pH}_0 + 3.5 \tag{23}$$

Table 1. Estimated kinetic parameters. Experimental conditions: $[Pa]_0=50.0 \text{ mg/L};$ [O₃]=25.0 mg/min; T=25.0°C.

604

\mathbf{pH}_0	Color ₀	Color ₀ Color _{max}		Color∞ t _{max}		kd	[Arom]∞	kA
	(AU)	(AU)	(AU)	(min)	(1/min)	(1/min)	(AU)	(1/min)
3.0	0.0075	0.0105	0.0000	10	0.01	0.02	0.050	0.085
6.0	0.0100	0.0095	0.0003	6	0.01	0.15	0.070	0.110
9.0	0.0115	0.0400	0.0016	4	0.01	0.21	0.050	0.220
12.0	0.0130	0.0950	0.0023	2	0.01	0.30	0.060	0.210

605

606 **4. Conclusions**

The ozonation of paracetamol aqueous solutions produces in a first stage a strong red coloration at the same time that water aromaticity decreases. Then, a second stage takes place; color intensity diminishes until it reaches a residual yellow tonality. This color formation is favored when operating at $pH_0=12.0$ and ozone flow-rates applied in the range 4.2 mg/min. The ozonation of aqueous solutions of paracetamol leads to compounds such as p-aminophenol, p-nitrophenol, nitrocatechol, hydroquinone and benzoquinone.

614

The hydroxide ions activate ozone decomposition to hydroxyl radicals, as well as to the increasing degree of dissociation of paracetamol lead to the generation of chromophoric structures. Ozone transfer to water is maximum when operating at $pH_0=6.0$, being this explained by the presence, at higher pH values, of larger amounts of the hydroxide anion and a possible generation of inorganic salts acting as scavengers for ozone and hydroxyl radicals.

A first order kinetic for water color changes fits well the experimental results. The obtained kinetic constants for color formation and degradation were to be $k_f=0.01$ (1/min) and $k_d=0.03$ pH₀-0.055 (1/min), respectively. Finally, the aromaticity loss is related to color changes. The corresponding first-order kinetic constant for aromaticity is given by $k_A=0.0162$ pH₀+0.035 (1/min).

627

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629

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636 **References**

637

Ay, F., Kargi, F., 2011. Effects of reagent concentrations on advanced oxidation of
amoxicillin by photo-Fenton treatment. J. Environ. Eng-Asce. 137(6):472-480.
https://10.1061/(ASCE)EE.1943-7870.0000344.

641

Baalbaki, Z., Sultana, T., Maere T., Vanrolleghem, P., Metcalfe, C.D., Yargeau, V.,
2016. Fate and mass balance of contaminants of emerging concern during wastewater
treatment determined using the fractionated approach. Sci. Total Environ. 573:1147–
1158. https://doi.org/10.1016/j.scitotenv.2016.08.073.

- Bader, H. Hoigné, J., 1981. Determination of ozone in water by the indigo method.
 Water Res. 15:449-456. https://doi.org/10.1016/0043-1354(81)90054-3.
- 649

Bahnemann, W., Muneer, M., Haque, M.M., 2007. Titatium dioxide-mediated
photocatalyzed degradation of selected organic pollutants in aqueous suspensions,
Catal. Today 124:133–148. https://doi.org/10.1016/j.cattod.2007.03.031.

653

Blair, B., Nikolaus, A., Hedman, C., Klaper, R., Grundl, T., 2015. Evaluating the
degradation, sorption, and negative mass balances of pharmaceuticals and personal care
products during wastewater treatment. Chemosphere 134:395–401.
https://doi.org/10.1016/j.chemosphere.2015.04.078.

658

Brunet, R., Bourbigot, M.M., Dore, M., 1982. The Influence of the Ozonation Dosage
on the Structure and Biodegradability of Pollutants in water, and its Effect on Activated
Carbon Filtration. Ozone Sci. Eng. 4:15-32.
https://doi.org/10.1080/01919518208550935.

663

Can, Z.S., Gurol, M., 2003. Formaldehyde formation during ozonation of drinking
water. Ozone-Sci. Eng. 25:41-51. https://doi.org/10.1080/713610649.

666

667 Cruz-Alcalde, A., Sans, C., Esplugas, S., 2017. Priority pesticides abatement by
advanced water technologies: The case of acetamiprid removal by ozonation. Sci. Total
669 Environ. 599–600:1454–1461. https://doi.org/10.1016/j.scitotenv.2017.05.065.

670

671	Dastmalchi, S., Rashidi, M.R., Rassi, M., 1995. Simultaneous determination of the pKa						
672	and octanol/water partition coefficient of acetaminophen. J. School Pharm. Med. Sci.						
673	Univ. Tehran. 4:7-14.						
674							
675	Deblonde, T., Hartemann, P., 2013. Environmental impact of medical prescriptions:						
676	assessing the risks and hazards of persistence, bioaccumulation and toxicity of						
677	pharamceuticals. Public Health 127:312-317.						
678	https://doi.org/10.1016/j.puhe.2013.01.026.						
679							
680	El Najjar, N.H., Touffet, A., Deborde, M., Journel, R., Karpel Vel Leitner, N., 2014.						
681	Kinetics of paracetamol oxidation by ozone and hydroxyl radicals, formation of						
682	transformation products and toxicity. Sep. and Purif. Technol. 136:137-143.						
683	https://doi.org/10.1016/j.seppur.2014.09.004.						
684							
685	Ellenhorn, M.J., Barceloux, D.G. 1988. Medical Toxicology: Diagnosis and Treatment						
686	of Human Poisoning. Elsevier Science, New York.						
687							
688	Gomes, J., Costa, R., Quinta-Ferreira, R.M., Martins, R.C. 2017. Application of						
689	ozonation for pharmaceuticals and personal care products removal from water. Sci.						
690	Total Environ. 586:265-283. https://doi.org/10.1016/j.scitotenv.2017.01.216.						
691							
692	Gottschalk, C., Libra, J.A., Saupe, A., 2010. Ozonation of Water and Waste Water. A						
693	Practical Guide to Understanding Ozone and its Applications, second ed. Whiley- VCH,						
694	Weinheim. https://doi.org/10.1002/9783527628926.						
695							

696	Greenberg, A.E., Eaton, A.D., Clesceri, L.S., 1999. Method 4500-O3 (ozone residual)
697	Indigo colorimetric method, in: Stand. Methods Exam. Water Wastewater, 20th Ed.,
698	American Public Health Association.

- He, W.L., Wu, C.D., 2018. Incorporation of Fe-phthalocyanines into a porous organic
- 701 framework for highly efficient photocatalytic oxidation of arylalkanes. Appl. Catal. B-

702 Environ. 234:290-295. https://doi.org/10.1016/j.apcatb.2018.04.055.

703

704 Hollender, J., Zimmermann, S.G., Koepke, S., Krauss, M., Mcardell, C.S., Ort, C., 705 Singer, H., Von Gunten, U., Siegrist, H., 2009. Elimination of organic micropollutants 706 in a municipal wastewater treatment plant upgraded with a full-scale post-ozonation 707 followed filtration. Environ. Sci. Technol. 43:7862-7869. by sand 708 https://doi.org/10.1021/es9014629.

709

Lee, Y., Gerrity, D., Lee, M., Gamage, S., Pisarenko, A., Trenholm, R.A., Cononica, S.,

711 Snyder, S.A., von Gunten, U., 2016. Organic contaminant abatement in reclaimed water

by UV/H2O2 and a combined process consisting of O3/H2O2 followed by UV/H2O2:

713 Prediction of abatement efficiency, energy consumption and byproduct formation.

714 Environ. Sci. Technol. 50:3809–3819. https://doi.org/10.1021/acs.est.5b04904.

715

Lee, Y., von Gunten, U. 2016. Advances in predicting organic contaminant abatement
during ozonation of municipal wastewater effluent: Reaction kinetics, transformation
products, and changes of biological effects. Environ. Sci. Water Res. Technol. 2:421–
442. https://doi.org/10.1039/C6EW00025H.

721	Marcé, M., Domenjoud, B., Esplugas, S., Baig, S. 2016. Ozonation treatment of urban
722	and biotreated wastewaters: Impacts and modelling. Chem. Eng. J. 283:768-777.
723	https://doi.org/10.1016/j.cej.2015.07.073.

725	Margot, J.	, Kienle,	C., 1	Magnet,	A.,	Weil,	М.,	Rossi,	L.,	de .	Alencastro,	L.F.,	Abegglen,
-----	------------	-----------	-------	---------	-----	-------	-----	--------	-----	------	-------------	-------	-----------

C., Thonney, D., Chevre, N., Schearer, M., Barry, D.A., 2013. Treatment of
micropollutants in municipal wastewater: Ozone or powdered activated carbon? Sci.
Total Environ. 461–462:480–498. https://doi.org/10.1016/j.scitotenv.2013.05.034.

Martignac, M., Oliveros, E., Maurette, M.T., Claparols, C., Benoit-Marquie, F., 2013.
Mechanistic pathways of the photolysis of paracetamol in aqueous solution: an example
of photo-Fries rearrangement. Photoch. Photobio. Sci. 12(3):527-535.
https://doi.org/10.1039/c2pp25341k.

733

Maya, N., Evans, J., Nasuhoglu, D., Isazadeh, S., Yargeau, V., Metcalfe, C.D., 2018.
Evaluation of Wastewater Treatment by Ozonation for Reducing the Toxicity of
Contaminants of Emerging Concern to Rainbow Trout (Oncorhynchus mykiss).
Environ. Toxicol. and Chem. 37(1):274–284.

738

McEvoy, G.K., 1992. American Hospital Formulary Service—Drug Information, 92,
Am. Soc. Hospital Pharmacists, Inc., Bethesda, MD. https://doi.org/10.1002/etc.3952.

741

Mijangos, F., Varona, F., Villota, N., 2006. Changes in solution color during phenol
oxidation by Fenton reagent. Environ. Sci. Technol. 40:5538-5543.
https://doi.org/10.1021/es060866q.

- Moctezuma, E, Leyva, E, Aguilar, C.A., Luna, R.A., Montalvo, C., 2012. Photocatalytic
 degradation of paracetamol: Intermediates and total reaction mechanism. J. Hazard.
 Mater. 243:130-138. https://doi.org/10.1016/j.jhazmat.2012.10.010.
- 749
- Moctezuma, E., Leyva, E., Palestino, G., de Lasa, H., 2007. Photocatalytic degradation
 of methyl parathion: reaction pathways and intermediate reaction products, J.
 Photochem. Photobiol. A: Chem. 186:71–84.
 https://doi.org/10.1016/j.jphotochem.2006.07.014.

Mompelat, S., Le Bot, B., Thomas, O., 2009. Occurrence and fate of pharmaceutical
products and by-products from resource to drinking water. Environ. Int., 35:803-814.
https://doi.org/10.1016/j.envint.2008.10.008.

757

Nawrocki, J., Kasprzyk-Hordern, B., 2003. Comments on "Solid phase catalytic
ozonation process for the destruction of a model pollutant" by D.S. Pines and D.A
Reckhow - (Ozone Sci. Eng. 25 (2003); 25). Ozone-Sci. Eng. 25: 535-537.
https://doi.org/10.1080/01919510390481847.

762

Oller, I., Malato, S., Sánchez-Pérez, J.A., 2011. Combination of Advanced Oxidation
Processes and biological treatments for wastewater decontamination--a review. Sci.
Total Environ. 409(20):4141-66. https://doi: 10.1016/j.scitotenv.2010.08.061.

766

Paraskeva, P., Graham, N.J.D., 2002. Ozonation of municipal wastewater effluents.
Water Environ. Res. 74:569-581. https://doi.org/10.2175/106143002x140387.

Radjenović, J., Petrović, M., Barceló, D., 2009. Complementary mass spectrometry and
bioassays for evaluating pharmaceutical-transformation products in treatment of
drinking water and wastewater. Trac-Trend. Anal. Chem., 28:562-582.
https://doi.org/10.1016/j.trac.2009.02.006.

774

775 Rauert, C., Harner, T., Schuster, J.K., Eng, A., Fillmann, G., Castillo, L.E., Fentanes, 776 O., Villa Ibarra, M., Miglioranza, K.S.B., Moreno Rivadeneira, I-, Pozo, K., Aristizabal Zuluaga, B.H., 2018. Atmospheric Concentrations of New Persistent Organic Pollutants 777 and Emerging Chemicals of Concern in the Group of Latin America and Caribbean 778 779 (GRULAC) Region. Environ. Sci. Technol., 52(13):7240-7249. 780 https://doi.org/10.1021/acs.est.8b00995.

781

Reemtsma, T., These, A., 2005. Comparative investigation of low-molecular-weight
fulvic acids of different origin by SEC-Q-TOF-MS: New insights into structure and
formation. Environ. Sci. Technol. 39: 3507-3512. https://doi.org/10.1021/es0480466.

785

Richardson, S.D., 2009. Water analysis: emerging contaminants and current issues.
Anal. Chem., 81:4645-4677. https://doi.org/10.1021/acs.analchem.7b04577.

788

Rodríguez, A., Rosal, R., Perdigón-Melón, J.A., Mezcua, M., Agüera, A., Hernando,
M.D., Letón, P., Fernández-Alba, A.R., García-Calvo, E., 2008. Ozone-Based
technologies in water and wastewater treatment. Handbook Environ. Chem. 5:127-175.
https://doi.org/10.1007/698_5_103.

- Rodríguez, C., Lombraña, J.I., de Luis, A., Sanz, J., 2017. Oxidizing efficiency analysis
 of an ozonation process to degrade the dye rhodamine 6G. J. Chem. Technol.
 Biotechnol. 92:674–683. https://doi.org/doi:10.1002/jctb.5051.
- 797 Rosario-Ortiz, F.L., Wert, E.C., Snyder, S.A., 2010. Evaluation of UV/H2O2 treatment
- for the oxidation of pharmaceuticals in wastewater. Water Res. 44:1440–1448.
- 799 https://doi.org/10.1016/j.watres.2009.10.031.
- Roth J.A. and Sullivan D.E., 1981. Solubility of ozone in water. Ind Eng Chem Fundam
 20:137–140. https://doi.org/10.1021/i100002a004.
- 802
- 803 Santos, L., Araùjo, A.N., Fachini, A., Pena, A., 2010. Ecotoxicological aspects related
- to the presence of pharmaceuticals in the aquatic environment. J. Hazard. Mater.
 175:45-95. https://doi.org/10.1016/j.jhazmat.2009.10.100.
- 806
- Savchuk, N., Krizova, P., 2015. Membrane and AOP processes-their application and
 comparison in treatment of wastewater with high organics content. Desalin. Water
 Treat. 56:3247-3251. https://doi.org/10.1080/19443994.2014.980978.
- 810
- Seifrtová, M., Nováková, L., Lino, C., Pena, A., Solich, P., 2009. An overview of
 analytical methodologies for the determination of antibiotics in environmental waters.
 Anal. Chim. Acta 649:158-179. https://doi.org/10.1016/j.aca.2009.07.031.
- 814
- 815 Skoumal, M., Cabot, P-L., Centellas, F., Arias, C., R.M., Garrido, J.A., Brillas, E.,
- 816 2006. Mineralization of paracetamol by ozonation catalyzed with Fe^{2+} , Cu^{2+} and UVA
- 817 light. App. Catal., B 20:228-240. https://doi.org/10.1016/j.apcatb.2006.03.016.
- 818

819	Sumegova, L., Derco, J., Melicher, M., 2013. Influence of reaction conditions on the
820	ozonation process. Acta Chim. Slovaca 6:168-172. https://doi.org/10.2478/acs-2013-
821	0026.

Sun, Q., Lv, M., Hu, A., Yang, X., Yu, C.P., 2014. Seasonal variation in the occurrence
and removal of pharmaceuticals and personal care products in a wastewater treatment
plant in Xiamen, China. J. Hazard. Mater. 277:69-75.
https://doi.org/10.1016/j.jhazmat.2013.11.056.

827

Swietlik, J., Sikorska, E., 2004. Application of fluorescence spectroscopy in the studies
of natural organic matter fractions reactivity with chlorine dioxide and ozone. Water
Res. 38: 3791-3799. https://doi.org/10.1016/j.watres.2004.06.010.

831

Valdés, M.E., Amé, M.V., Bistoni, M.M, Wunderlin, D.A., 2014. Occurrence and
bioaccumulation of pharmaceuticals in a fish species inhabiting the Suquía River basin
(Córdoba, Argentina). Sci. Total Environ. 472:389-396.
https://doi.org/10.1016/j.scitotenv.2013.10.124.

836

Villaroel, E., Silva-Agredo, J., Petrier, C., Taborda, G., Torres-Palma, R., 2014.
Ultrasonic degradation of acetaminophen in water: Effect of sonochemical parameters
and water matrix. Ultrason. Sonochem. 21:1763-1769.
http://dx.doi.org/10.1016/j.ultsonch.2014.04.002.

841

Villota, N., Lomas, J.M., Camarero, L.M., 2016. Study of the paracetamol degradation
pathway that generates color and turbidity in oxidized wastewaters by photo-Fenton

- technology. J. Photochem Photobiol A Chem. 329:113–119.
 https://doi.org/10.1016/j.jphotochem.2016.06.024.
- 846

Villota, N., Lomas, J.M., Camarero, L.M., 2018. Kinetic modelling of water-color
changes in a photo-Fenton system applied to oxidate paracetamol. J. Photochem
Photobiol A Chem. 356:573–579. https://doi.org/10.1016/j.jphotochem.2018.01.040.

850

Vogna, D., Marotta, R., Napolitano, A., d'Ischia, M., 2002. Advanced oxidation
chemistry of paracetamol. UV/H₂O₂-induced hydroxylation/degradation pathways and
15N-aided inventory of nitrogenous breakdown products, J. Org. Chem. 67:6143–6151.
https://doi.org/10.1021/jo025604v.

855

von Gunten, U., 2003. Ozonation of drinking water: Part I. Oxidation kinetics and
product formation. Water Res. 37:1443–1467. https://doi.org/10.1016/S00431354(02)00457-8.

Wert, E.C., Rosario-Ortiz, F.L., Drury, D.D., Snyder, S.A., 2007. Formation of
oxidation byproducts from ozonation of wastewater. Water Res. 41:1481–1490.
https://doi.org/10.1016/j.watres.2007.01.020.

862

Westerhoff P., Debroux J., Aiken G., Amy G., 1999. Ozone induced changes in natural
organic matter (nom) structure. Ozone Sci. Eng. 21: 551-570.
https://doi.org/10.1080/01919512.1999.10382893.

Xiong F., Legube B., 1991. Enhancement of radical chain reactions of ozone in waters
in the presence of an aquatic fulvic acid. Ozone Sci. Eng. 13:349-363.
https://doi.org/10.1080/01919519108552471.

Zenker, A., Cicero, M.R., Prestinaci, F., Bottoni, P., Carere, M., 2014. Bioaccumulation
and biomagnification potential of pharmaceuticals with a focus to the aquatic
environment. J. Environ. Manage. 133:378-387.
https://doi.org/10.1016/j.jenvman.2013.12.017.

- Ziylan-Yavaş, A., Ince, N.H., 2018. Catalytic ozonation of paracetamol using
 commercial and Pt-supported nanocomposites of Al₂O₃: The impact of ultrasound.
- 877 Ultrason. Sonochem. 40(B):175-182. https://doi.org/10.1016/j.ultsonch.2017.02.017.