

GH LEVELS AND INSULIN SENSITIVITY ARE DIFFERENTLY ASSOCIATED WITH BIOMARKERS OF CARDIOVASCULAR DISEASE IN ACTIVE ACROMEGALY.

Journal:	<i>Clinical Endocrinology</i>
Manuscript ID:	CEN-2011-000719.R1
Manuscript Type:	4 Original Article - Americas
Date Submitted by the Author:	n/a
Complete List of Authors:	Boero, Laura; University of Buenos Aires, Faculty of Pharmacy and Biochemistry, INFIBIOC Manavela, Marcos; University of Buenos Aires, Endocrinology Division , "José de San Martín" Clinical Hospital Meroño, Tomás; University of Buenos Aires. Faculty of Pharmacy and Biochemistry, INFIBIOC CONICET Maidana, Patricia; University of Buenos Aires, Faculty of Pharmacy and Biochemistry, INFIBIOC Gómez Rosso, Leonardo; University of Buenos Aires. Faculty of Pharmacy and Biochemistry, INFIBIOC CONICET Brites, Fernando; University of Buenos Aires. Faculty of Pharmacy and Biochemistry, INFIBIOC CONICET
Key Words:	Atherosclerosis < Conditions: < Obesity/Lipids/Nutrition, Insulin resistance < Conditions: < Obesity/Lipids/Nutrition, Acromegaly < Conditions: < Pituitary

1
2
3 **GH LEVELS AND INSULIN SENSITIVITY ARE DIFFERENTLY ASSOCIATED WITH**
4
5 **BIOMARKERS OF CARDIOVASCULAR DISEASE IN ACTIVE ACROMEGALY.**
6
7

8
9 **Boero L^a, Manavela M^b, Meroño T^a, Maidana P^a; Gómez Rosso L^a; Brites F^a.**
10

11
12
13 ^a Department of Clinical of Biochemistry, Faculty of Pharmacy and Biochemistry, University of
14 Buenos Aires. INFIBIOC. CONICET.
15

16
17 ^b Endocrinology Division , “José de San Martín” Clinical Hospital, University of Buenos Aires.
18

19 **Abbreviated title:** GH and HOMA association with CVD markers
20

21 **Key words:** Acromegaly, Insulin resistance, atherosclerosis.
22

23 **Word count:** 3249. **Abstract,** 249 words, **Tables:** 4, **Figures:** 1.
24

25 **Disclosure statement:** The authors have nothing to disclose.
26

27 **Corresponding author:**
28

29 Dr. Laura Boero.
30

31 Department of Clinical of Biochemistry
32

33 School of Pharmacy and Biochemistry
34

35 University of Buenos Aires
36

37 Junín 956 (1113)
38

39 Buenos Aires, Argentina
40

41 Tel.: +54-11-5950-8654 // Fax: +54-11-4508-3645
42

43 E-mail: laura.boero@hotmail.com
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

Abstract

Context: acromegaly is characterized by GH excess and insulin resistance. It is not known which of these two disorders is responsible for the increased atherogenic risk in these patients.

Objective: to analyze the associations of GH and HOMA with biomarkers of cardiovascular disease and to compare the above mentioned variables between patients with active acromegaly and control subjects.

Design and setting: this open-cross-sectional study was conducted at a university hospital.

Patients: Twenty two outpatients were compared with sex and age matched controls subjects.

Main outcomes: included clinical features, hormonal status, markers of insulin resistance, lipoprotein profile, and biomarkers of cardiovascular disease.

Results: patients presented higher triglyceride ($1.22[0.69-5.73]$ vs $0.86[0.30-1.39]$ mmol/L, $p<0.001$), LDL-C (3.5 ± 0.9 vs 3.0 ± 0.7 mmol/L, $p<0.01$), apoB (0.98 ± 23 vs 0.77 ± 22 g/L, $p<0.01$), free fatty acid (0.69 ± 0.2 vs 0.54 ± 0.2 nmol/L, $p<0.05$), oxidized-LDL (120 ± 22 vs 85 ± 19 U/L, $p<0.001$), endothelin-1 (0.90 ± 0.23 vs 0.72 ± 0.17 ng/L, $p<0.05$) and lower HDL-C (1.2 ± 0.4 vs 1.5 ± 0.4 mmol/L, $p<0.01$) levels, and higher CETP activity (179 ± 27 vs 138 ± 30 %ml⁻¹.h⁻¹, $p<0.001$) than control subjects. VCAM-1 and hsCRP concentrations were not different. By multiple regression analyses, HOMA explained the variability of triglycerides (25%), HDL-C (30%), and CETP activity (28%), while GH independently predicted LDL-C (18%), oxidized-LDL (40%) and endothelin-1 levels (19%).

Conclusions: in active acromegalic patients, GH excess contributes to the development of an insulin resistant state and the interaction between both disturbances would be responsible for the appearance of atherogenic prooxidative and proinflammatory factors and markers. Insulin resistance would be preferably associated to an atherogenic lipoprotein profile and to high CETP activity, while high GH levels would independently predict the increase in LDL-C, ox-LDL and endothelin-1 levels in acromegaly.

Introduction

It has been well established that active acromegaly, a condition defined by the presence of excessive secretion of growth hormone (GH), is associated with increased mortality from cardiovascular disease [1]. In several studies, different atherogenic risk factors and biomarkers of cardiovascular disease were detected to be altered in acromegalic patients [2,3]. Most of the above mentioned studies attributed these alterations to GH increment. However, it must be noted that acromegaly is also associated with insulin resistance, carbohydrate intolerance and, in about 40% of the patients, with type 2 diabetes, conditions known to be directly involved in chronic inflammation and atherogenesis [4]. In fact, many years ago, Moller et al. [5] evidenced the presence of profound disturbances of not only glucose but also lipid metabolism.

Insulin resistance is considered to be a pivotal event in atherosclerosis through different pathways [6]. Therefore, it could be hypothesized that this disorder could play a pathophysiological role in the development of cardiovascular disease in acromegalic patients beyond GH increment.

The specific consequences of insulin resistance in acromegaly are not completely known. In non-acromegalic patients, lipid abnormalities, such as increased non-esterified fatty acid and triglyceride as well as low high density lipoprotein-cholesterol (HDL-C) levels, are frequently associated with insulin resistance, which is, in turn, closely related to hyperglycemia [7].

On the other hand, resistin, a peptide secreted by adipocytes and inflammatory cells, has been shown to be increased in patients with insulin resistance and early type 2 diabetes, populations at high risk for the development of diffuse and extensive patterns of atherosclerosis. Nevertheless, the role played by this protein in insulin resistance in humans is controversial [8,9].

The main objective of the present study was to analyze the associations of atherogenic risk factors and biomarkers of cardiovascular disease with markers of insulin resistance and parameters of the GH axis in a cross-sectional study. Secondly, we aimed to compare the above mentioned variables, including resistin levels, between a group of active acromegalic patients and age- and sex-matched control subjects.

Materials and methods

Subjects:

Twenty two adult patients with diagnosis of active acromegaly, who were not receiving any specific treatment for acromegaly, were consecutively recruited from the Endocrinology Service, Hospital de Clínicas “José de San Martín”, Buenos Aires, Argentina, during a period of 24 months. Patients were included in the present study when presenting typical clinical features, lack of suppression of GH during an oral glucose tolerance test (OGTT) and elevated insulin-like growth factor-1 (IGF-1) level for corresponding sex and age. Disease duration ranged between 2 and 10 years. Twenty two healthy subjects, sex- and age-matched with the patients, agreed to participate in this study and were employed as controls. Both patients and controls had normal renal, hepatic and thyroid functions and did not present diagnosis of any other endocrine disorder, none of them presented history of any cardiovascular event, and they were not under treatment with antioxidants or any drug known to affect carbohydrates, lipids or biomarkers of cardiovascular disease. None of the patients or control subjects was diabetic and only 3 acromegalic patients presented impaired glucose tolerance, evidenced by the OGTT. Informed consent was obtained from all participants and the protocol of this open transversal study was approved by the Ethical Committees from Faculty of Pharmacy and Biochemistry and from Hospital de Clínicas “José de San Martín”, University of Buenos Aires.

Study Protocol and Samples:

Body weight, height and waist circumference were registered. The latter was measured midway between the lateral lower rib margin and the superior anterior iliac crest. This measurement was performed with the subject in a standing position and always by the same investigator.

After a 12-hour overnight fast, venous blood was drawn from the antecubital vein. Aliquots were collected in clean tubes. Samples were centrifuged at 1500 x g, for 15 minutes; at 4° C. Serum was immediately employed for glucose and stored at 4° C for lipid, lipoprotein and non-esterified fatty acids measurements within 24 hours. Serum and plasma aliquots were also stored at -70° C for determination of GH, IGF-1, insulin growth factor binding protein-3 (IGFBP-3), insulin, resistin,

1
2
3 cholesteryl ester transfer protein (CETP), oxidized (ox) low density lipoprotein (LDL), endothelin-1,
4
5 vascular cell adhesion molecule (VCAM)-1 and C reactive protein (CRP).
6
7

8
9 *Analytical Procedures:*

10
11 Glucose, triglycerides and total cholesterol were quantified by standardized methods (Roche
12
13 Diagnostics, Mannheim, Germany) in a Hitachi 917 autoanalyzer. Within-run precision (CV) were 2
14
15 %, 1.3 % and 1.1 %, respectively. Between-day precision (CV) were 2.8 %, 2.4 % and 1.5 %,
16
17 respectively. Laboratory bias for triglycerides and total cholesterol was 1.1 % and -1.7 %,
18
19 respectively. LDL-cholesterol (LDL-C) level was determined as the difference between total
20
21 cholesterol and the cholesterol contained in the supernatant obtained after selective precipitation of
22
23 LDL with 10 g/L polyvinylsulphate in polyethylenglycol (M.W. 600; 2.5 % w/v; pH = 6.7) [10].
24
25 Within-run and between-day precisions (CV) were 4.7 % and 5.0 %, respectively. HDL was isolated
26
27 in the supernatant obtained following precipitation of apo B-containing lipoproteins with 40 g/L
28
29 phosphotungstic acid in the presence of magnesium ions [11]. Within-run and between-day precisions
30
31 (CV) were 3.2 % and 3.8 %, respectively. Laboratory bias was -2.0 %. Apo A-I and apo B were
32
33 evaluated by immunoturbidimetry (Roche Diagnostics Mannheim, Germany) in a Hitachi 917
34
35 autoanalyzer. Within-run and between-day precisions (CV) for apo A were 1.9% and 2.4% and for apo
36
37 B were 1.2 % and 2.1 %, respectively. Non-esterified fatty acids were determined by a colorimetric
38
39 method (Randox Laboratories Ltd.UK).
40
41
42

43
44 *Hormonal Parameters:*

45
46 Serum GH was measured by the ultrasensitive immunochemiluminometric assay (Access®,
47
48 Beckman Coulter TM, USA) with analytical sensitivity of 0.003 µg/L. Within-run and between-day
49
50 precisions (CV) were 12.3 % and 15.5 %, respectively. Serum IGF-1 and IGFBP-3 levels were
51
52 measured by solid phase chemiluminiscent enzyme immunoassay (Diagnostics Products Corp., Los
53
54 Angeles, CA, USA) in an Immulite 2000 with analytical sensitivity of 20 ng/ml and 0.1 µg/ml.
55
56 Within-run and between-day precisions (CV) for IGF-1 were below 5.4 % and 11.9%, respectively.
57
58
59
60

1
2
3 Measurements of IGFBP-3 were all carried out within the same assay. Within-run precision (CV) was
4 4.8 %. Insulin concentration was measured by microparticle enzyme immunoassay (MEIA)
5 (ABBOTT, Japan). Within-run and between-day precisions (CV) were 2.9 % and 4.4%, respectively.
6
7 Homeostasis model assessment (HOMA) was calculated as $[\text{Glucose (mmol/L)} \times \text{Insulin (U/ml)}] / 22.5$.
8
9

10 11 12 13 *Resistin:*

14
15 Resistin plasma levels were determined by monoclonal antibody-based enzyme-linked
16 immunosorbent assay (ELISA) following the manufacturer`s instructions (R & D Systems, USA).
17 Sample levels were calculated by analyzing standards with known concentrations of recombinant
18 molecules coincident with samples and plotting of signal vs. concentration. Within-run and between-
19 day precisions (CV) were less than 5.3 % and less than 9.2 %, respectively.
20
21
22
23
24
25

26 27 *CETP Activity:*

28
29 CETP activity was determined in serum samples following the general procedure previously
30 described [12] with a few modifications. Briefly, the ability of serum to promote the transfer of
31 tritiated cholesteryl esters from a tracer amount of biosynthetically labeled HDL₃ (³H-CE-HDL₃)
32 (NEN Life Science Products, Boston, USA) to serum apo B-containing lipoproteins was evaluated.
33 Samples were incubated with ³H-CE-HDL₃ (50 μmol/L cholesterol) and 1.5 mmol/L iodoacetate for
34 3h, at 37° C. After incubation, lipoproteins were separated by a selective precipitation method
35 employing 40 g/L phosphotungstic acid in the presence of magnesium ions [10]. Radioactivity was
36 measured both in the incubation mixture and in the supernatant containing the HDL fraction in a
37 Liquid Scintillation Analyzer (Packard 210TR; Packard Instruments, Meridian, CT). Measurements
38 were all carried out in duplicate within the same assay. Within-run precision (CV) was 4.9 %.
39
40
41
42
43
44
45
46
47
48
49

50 51 52 *Biomarkers of Cardiovascular Disease:*

53
54 Ox LDL was measured by a competitive enzyme-linked immunosorbent assay which employs
55 the monoclonal antibody 4E6 (Mercodia AB, Uppsala, Sweden). Measurements of ox LDL were all
56 carried out within the same assay. Within-run precision (CV) was 6.1 %. Endothelin-1 levels were
57
58
59
60

1
2
3 determined by monoclonal antibody-based enzyme-linked immunosorbent assay following the
4 manufacturer`s instructions, with few modifications (ELISA) (R & D Systems, USA). Within-run and
5 between-day precisions (CV) were 4.5 % and 5.5 %, respectively. VCAM-1 plasma levels were
6 determined by the monoclonal antibody-based enzyme-linked immunosorbent assay following the
7 manufacturer`s instructions, (ELISA) (R & D Systems, USA). Within-run and between-day precisions
8 (CV) were 3.5 % and 7.7 %, respectively. CRP concentration was determined by Tina-quant CRP
9 (Latex) high sensitive immunoturbidimetric assay (Roche Diagnostics Mannheim, Germany) in a
10 Hitachi 917 autoanalyzer. Within-run and between-day precisions (CV) were 0.4 % and 3.4 %,
11 respectively.
12
13
14
15
16
17
18
19
20
21
22

23 *Data and Statistical Analysis:*

24
25 The sample size required to detect significant differences between groups was estimated for
26 the main parameters at a significance level of 0.05 and 80% power. Data distribution was tested
27 employing the Shapiro-Wilk test. Results were expressed as mean \pm standard deviation (SD) for
28 normally distributed data and as median [range] for skewed data. In this last case, data were
29 normalized by log-transformation. Then, analysis of covariance was used including body mass index
30 (BMI) as a covariate. Partial correlations were performed between different parameters including BMI
31 as a fixed variable. Multiple linear regression analysis was performed to examine the variables
32 independently associated with atherogenic risk factors and biomarkers of cardiovascular disease. In
33 these analyses, GH and HOMA were always included as independent variables while triglycerides,
34 HDL-C, LDL-C, non-esterified fatty acids, CETP, ox LDL, endothelin-1, resistin, VCAM-1 and CRP
35 were alternatively included as the dependent variable and the rest as independent ones. Differences
36 were considered significant at $p < 0.05$ in the bilateral situation. For statistical analysis, INFOSTAT
37 software was used.
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

Results

In the present study, 22 patients with active acromegaly were studied in comparison with 22 sex- and age-well-matched control subjects. Clinical characteristics, hormonal parameters and biomarkers of insulin resistance from acromegalic patients and control subjects are shown in table I. In accordance with the well-known physical features of subjects with acromegaly, BMI and waist circumference were significantly increased in the patient group. Given this difference between both studied groups, all the results obtained were compared performing analysis of covariance, including BMI as a covariate. Deriving from the inclusion criteria, mean or median GH, IGF-1 and IGFBP-3 concentrations were also significantly elevated in the group of patients with active acromegaly. Furthermore, all the markers of insulin resistance, including non-esterified fatty acids, were significantly higher in acromegalic patients than in control subjects, except for resistin which showed no difference.

Acromegalic patients presented a more atherogenic lipoprotein profile than control subjects, consisting of significantly higher triglyceride (1.22 [0.69-5.73] vs. 0.86 [0.30-1.39] mmol/L, $p < 0.001$), LDL-C (3.5±0.9 vs. 3.0±0.7 mmol/L, $p < 0.01$) and apo B (0.98±0.23 vs. 0.77±0.22 g/L, $p < 0.01$) levels, and lower HDL-C (1.2±0.4 vs. 1.5±0.4 mmol/L, $p < 0.01$) concentration (Figure 1). Moreover, CETP activity, responsible for modulating lipoprotein composition in plasma, was significantly increased in acromegalic patients (179±27 vs. 138±30 $\% \cdot \text{ml}^{-1} \cdot \text{h}^{-1}$, $p < 0.001$).

Table II shows different biomarkers of cardiovascular disease from acromegalic patients and control subjects. Oxidized LDL, a pro-inflammatory and pro-atherogenic biomarker, and endothelin-1, the most potent constrictor of human vessels, were significantly increased in acromegaly. On the other hand, VCAM-1, a cell adhesion molecule of endothelial location that actively participates in the firm adhesion and extravasation of circulating leucocytes into the artery wall, and CRP, which has been implicated in multiple aspects of atherogenesis and plaque vulnerability, showed no differences between both groups.

When analyzing the associations between GH with different atherogenic risk factors and biomarkers of cardiovascular disease evaluated in the present study, direct correlations were observed with triglycerides, LDL-C, oxLDL, endothelin-1 and CETP activity, though the latter did not reach

1
2
3 statistical significance ($p < 0.06$) (Table III). On the other hand, HOMA was positively associated with
4 triglycerides, CETP activity and showed a non-significant tendency with oxLDL ($p < 0.06$), while it
5 also presented a negative correlation with HDL-C.
6
7

8
9 Multiple regression analyses were carried out in order to identify independent predictors of the
10 atherogenic risk factors and biomarkers of cardiovascular disease (Table IV). When triglyceride levels
11 were evaluated, HOMA explained a 25 % of its variability, while HOMA and GH together explained a
12 32 %. In the case of HDL-C and non-esterified fatty acids only HOMA and triglycerides appeared to
13 be independently associated, respectively. The variability of CETP activity was attributed in a 28 % to
14 HOMA and in a 37 % to both HOMA and resistin. GH was the only independent predictor of LDL-C
15 and endothelin-1 (18 and 19 %, respectively). Finally, in ox LDL analysis, GH alone and GH with
16 BMI were identified as the independent predictors explaining 45 and 70 %, respectively, while no
17 significant associations were detected for VCAM-1 or CRP.
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

Discussion

Acromegaly is a progressive chronic disease associated with high risk of cardiovascular disease [4] whose severity, according to our point of view, could be mainly attributed to the complex interaction between hormonal and metabolic disturbances present in affected patients. In fact, alterations in GH axis are most frequently associated to insulin resistance [13], two situations independently related to the development of cardiovascular disease. Our main findings point out that acromegalic patients show an increase in triglyceride levels and CETP activity and a reduction in HDL-C concentration mainly predicted by the insulin-resistant marker HOMA. Moreover, the increment observed in LDL-C, ox LDL and endothelin-1 levels would be mostly associated with GH excess, independent of other metabolic parameters.

It has been well documented that GH hypersecretion and insulin resistance are closely interconnected [14]. Accordingly, in the present study, acromegalic patients showed significantly higher glucose and insulin levels, as well as HOMA index than healthy controls. In fact, GH has potent effects on intermediary metabolism and, in particular, it has the capacity to antagonize insulin actions. However, the underlying mechanisms are not completely understood. One possible explanation could be the increase in free fatty acid flux from the adipose tissue, also observed in acromegalic subjects evaluated in this study, which could, in turn, impair insulin action at target tissues [15,16]. Given the controversial data available on resistin role in the development of insulin resistance, this parameter was evaluated in the studied population and neither statistically significant difference nor a relationship with GH were found in this study. Accordingly, Silha et al. also observed similar resistin levels in acromegalic patients and control subjects [17]. Although some studies showed an increase in resistin concentration in obesity and type 2 diabetes, most reports did not detect any correlation between resistin and body mass index or insulin resistance [18].

The causal coexistence of GH excess and insulin resistance has been clearly established [19-21]. However, it is still ignored whether the multiple metabolic abnormalities present in acromegalic patients are due to GH hypersecretion, to insulin resistance or to a combination of both of them.

1
2
3 As it has been previously shown [2], the group of acromegalic patients evaluated in this study
4 showed significantly increased triglyceride and apo B levels, and decreased HDL-C concentration, the
5 so called “atherogenic dyslipidemia” characteristic of insulin-resistant states [22]. Actually, in
6
7 multivariate analysis, HOMA index was able to independently predict the variations in triglyceride
8
9 and HDL-C levels, as well as in CETP activity, which was also higher in patients than in controls.
10
11

12
13 The increment detected in triglyceride concentration could be due to the increased flux of free
14 fatty acids from adipose tissue, mainly of visceral localization, to the liver which may be, in turn,
15 attributed to the high activity of hormone sensitive lipase (HSL) in insulin resistance [23]. It is very
16 well-known that in a frame of insulin resistance, the liver employs these free fatty acids for
17 triglyceride synthesis and that high activity of microsomal transfer protein enables their assembly in
18 triglyceride-rich very low density lipoprotein (VLDL) particles, afterwards poured into the circulation
19 [24]. Even if multivariate analysis pointed out HOMA as the most powerful predictor of triglyceride
20 levels, GH effects cannot be discarded. In adipose tissue, GH activates HSL [25]. In the liver, GH
21 stimulates free fatty acid uptake by inducing lipoprotein lipase / hepatic lipase expression, it promotes
22 lipogenesis, and it inhibits both lipolysis and fatty acid oxidation [23]. Overall, these actions facilitate
23 the intrahepatic storage of triglycerides. Then, GH could also contribute to VLDL assembly by
24 upregulating the expression of microsomal transfer protein [26].
25
26
27
28
29
30
31
32
33
34
35
36

37 In non-acromegalic subjects, insulin resistance has been largely shown to be implicated not
38 only in triglyceride increase but also in the induction of CETP activity and in the reduction of HDL-C
39 levels [27]. Beyond insulin resistance, it is noteworthy that these three parameters, “triglycerides”,
40 “CETP” and “HDL-C”, are closely interconnected. Triglyceride levels are known to upregulate CETP
41 which is responsible for interchanging triglycerides and cholesteryl esters between apo B-containing
42 lipoproteins and HDL, thus adding to HDL-C diminution. Furthermore, in hypertriglyceridemia, HDL
43 particles have been shown to be triglyceride-enriched and less efficient in the promotion of cell
44 cholesterol efflux, which finally contributes to reduce HDL cholesterol content [28].
45
46
47
48
49
50
51
52

53 Among the different atherogenic risk factors and biomarkers of cardiovascular disease
54 evaluated in this study, LDL-C, oxidized LDL and endothelin-1 levels were significantly elevated in
55
56
57
58
59
60

1
2
3 acromegalic patients in comparison to healthy controls. Interestingly, these three parameters were
4
5 independently predicted by GH levels and not by HOMA.
6

7 The increase in oxLDL levels has been already reported in acromegalic patients [29] and we
8
9 now report that this increase is independently predicted by GH concentration. In the literature, there is
10
11 weak support for GH to stimulate oxidative stress directly. Nevertheless, it must be taken into
12
13 consideration that acromegalic patients showed hypercholesterolemia, which is known to be associated
14
15 to oxidative stress [30]. Then, Yarman et al. [31] found higher thiobarbituric acid reactive substances
16
17 (TBARS) levels in a group of newly diagnosed acromegalic patients and Andersson et al. [32]
18
19 associated GH overexpression with a time- and vessel-specific deterioration in endothelial function,
20
21 initially caused by increased oxidative stress in GH transgenic mice. Moreover, biochemical studies
22
23 showed that ceruloplasmin, which was found to be elevated in acromegalic patients [29], is a potent
24
25 catalyst of LDL oxidation *in vitro*.
26

27 The other novel biomarker of cardiovascular disease, endothelin-1, which is the most potent
28
29 vasoconstrictor in humans, has been implicated in atherosclerotic and ischemic heart disease [33].
30
31 Both, our previous results [2] and those from Kirilov et al. [34] showed an increase in endothelin-1 in
32
33 active acromegaly. In the present study, an association between endothelin-1 and GH excess was
34
35 evidenced independently of other metabolic parameters as HOMA. Then, it could be assumed that in
36
37 active acromegaly the GH secretory status would be an important determinant of plasma endothelin-1
38
39 level [2,34].
40

41 Besides contributing to the characterization of high risk of cardiovascular disease in
42
43 acromegalic patients, results from the present study could also have a great impact on potential
44
45 therapeutic management of acromegaly by employing not only specific inhibitors of GH secretion /
46
47 action, but in combination with insulin sensitizer agents. Nevertheless, confirmation of the above
48
49 mentioned independent associations would be necessary by studying models in which GH excess
50
51 would not be accompanied by insulin resistance. However, up to our knowledge, given the role played
52
53 by GH in the genesis of insulin resistance, there are no available models to be explored. Another
54
55 limitation is the number of acromegalic patients studied, though this is related with the low prevalence
56
57 of active acromegaly with absence of other endocrine pathologies, specific therapy or treatment with
58
59
60

1
2
3 other drugs known to affect carbohydrate or lipid metabolism.
4

5 In conclusion, in active acromegalic patients, GH excess clearly contributes to the
6 development of an insulin resistant state and the complex interaction between both disturbances would
7 be responsible for the appearance of an atherogenic cluster containing prooxidative and
8 proinflammatory factors and markers. From our results, insulin resistance would be preferably
9 associated to an atherogenic lipoprotein profile (high triglyceride and low HDL-C levels) and to high
10 CETP activity, while high GH levels would independently predict the increase in LDL-C, ox LDL and
11 endothelin-1 levels in acromegalic patients.
12
13
14
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

For Peer Review

Acknowledgements

This work was supported in part by grants from CONICET (PIP 0931) and University of Buenos Aires (UBACYT 200200902017). The authors specially thank Siemens S.A. for their collaboration.

For Peer Review

Figure Legends

Fig. 1: Lipid, lipoprotein and apolipoprotein profile from acromegalic patients (n = 22) and control subjects (n = 22). Results were expressed as Mean±SD. TG, triglycerides; TC, total cholesterol; LDL, low density lipoprotein; HDL, high density lipoprotein; apo, apolipoprotein.

^a p<0.0001; ^b p<0.01; ^c p<0.05 vs. acromegalic patients.

For Peer Review

Figure. 1

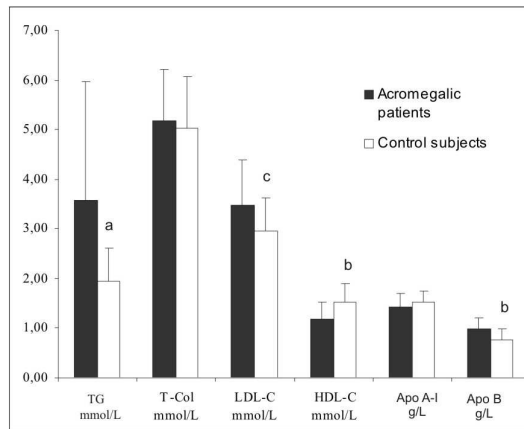


TABLE I: Clinical characteristics, hormonal parameters and biomarkers of insulin resistance from acromegalic patients and control subjects.

	Acromegalic Patients	Control Subjects
N	22	22
Women/Men	16/6	16/6
Age (years)	44±14	44±13
B.M.I. (kg/m²)	28 (21-43)	22 (19-28) ^a
Waist (cm)	95±10	86±8 ^a
GH (µg/L)	7.9 (1.9-55.0)	1.07 (0.05-8.6) ^a
IGF-1 (nmol/L)	92.8 (35.6-142.3)	20.5 (10.0-52.0) ^a
IGFBP-3 (mg/L)	7.6±1.63	4.4 (2.2-6.2) ^a
Glucose (mmol/L)	5.4±0.6	4.8±0.6 ^b
Insulin (pmol/L)	132.4 (40.2-391.8)	42.3 (13.6-122.7) ^a
HOMA	4.4 (1.1-13.5)	1.3 (0.5-3.8) ^a
NEFA (nmol/L)	0.69±0.2	0.54±0.2 ^c
Resistin (µg/L)	7.9 (2.7-15.8)	5.4 (2.7-33.6)

B.M.I, Body Mass; GH, Growth Hormone; IGF-1, Insulin-like Growth Factor I; IGFBP-3, IGF Binding Protein-3; HOMA, Homeostasis Model Assessment. NEFA: Non-Esterified Fatty Acids
Results were expressed as mean±S.D. or as median (range), depending on data distribution.

^a p<0.0001 ^b p<0.01 and ^c p<0.05 vs. acromegalic patients.

TABLE II: Biomarkers of cardiovascular disease from acromegalic patients and control subjects.

	Acromegalic patients (n =22)	Control subjects (n =22)
Oxidized LDL (U/L)	120±22	85±19 ^a
Endothelin-1 (ng/L)	0.90±0.23	0.72±0.17 ^b
VCAM-1 (µg/L)	37.8±13.3	38.0±7.4
CRP (mg/L)	0.25 (0.10-22.70)	0.85 (0.30-10.20)

VCAM-1, vascular cell adhesion molecule 1; CRP, C reactive protein. Results were expressed as mean±S.D. or as median (range), depending on data distribution.

^a p<0.001; ^b p<0.05 vs. acromegalic patients.

Table III: Correlations of GH and HOMA with different parameters in acromegalic patients and controls subjects (n=44).

	GH	HOMA
	r (p)	r (p)
TG	0.49 (<0.001)	0.35 (<0.05)
LDL-C	0.44 (<0.01)	0.04 (0.78)
HDL-C	-0.12 (0.45)	-0.54 (<0.001)
NEFA	0.13 (0.40)	0.13 (0.39)
CETP	0.33 (0.06)	0.44 (<0.01)
Ox LDL	0.72 (<0.0001)	0.41 (0.06)
Endothelin-1	0.43 (<0.05)	0.31 (0.09)

GH, Growth Hormone; HOMA, Homeostasis Model Assessment; TG, triglycerides; LDL-C, low density lipoprotein-cholesterol; HDL-C, high density lipoprotein-cholesterol; NEFA, non-esterified fatty acids; CETP, cholesteryl ester transfer protein; ox LDL, oxidized LDL. Correlations were performed including body mass index as fixed variable.

Table IV: Multiple regression analysis for the association of TG, HDL-C, NEFA, CETP, LDL-C, ox LDL and Endothelin-1 as dependent variable.

Dependent Variable	Significant Independent Variable	B	t	Significance <	R²
TG	Model 1: HOMA	0.52	3.8	0.01	0.25
	Model 2: HOMA	0.36	2.4	0.05	0.32
	GH	0.33	2.2	0.05	
LDL-C	GH	0.44	3.1	0.01	0.18
HDL-C	HOMA	-0.56	-4.28	0.0001	0.30
NEFA	TG	0.31	2.04	0.05	0.07
CETP	Model 1: HOMA	0.55	3.6	0.001	0.28
	Model 2: HOMA	0.57	4.0	0.0001	0.37
	Resistin	0.34	2.4	0.05	
Ox LDL	Model 1: GH	0.69	4.1	0.0001	0.45
	Model 2: GH	0.58	4.5	0.0001	0.70
	BMI	0.52	4.1	0.001	
Endothelin-1	GH	0.46	2.7	0.01	0.19

TG, triglycerides; LDL-C, low density lipoprotein-cholesterol; HDL-C, high density lipoprotein-cholesterol; NEFA, non-esterified fatty acids; CETP, cholesteryl ester transfer protein; ox LDL, oxidized LDL; GH, Growth Hormone; HOMA, Homeostasis Model Assessment; BMI, body mass index.

References

1. Bengtsson, B.A., Eden, S., Ernest, I., Oden, A. & Sjogren, B.(1988) Epidemiology and long-term survival in acromegaly. A study of 166 cases diagnosed between 1955 and 1984. *Acta Med Scand*, 223: 327-35.
2. Boero, L., Manavela, M., Gomez Rosso, L., Insua, C., Berardi, V., Fornari, M.C. & Brites, F.(2009) Alterations in biomarkers of cardiovascular disease (CVD) in active acromegaly. *Clin Endocrinol (Oxf)*, 70: 88-95.
3. Beentjes, J.A., van Tol, A., Sluiter, W.J. & Dullaart, R.P.(2000) Low plasma lecithin:cholesterol acyltransferase and lipid transfer protein activities in growth hormone deficient and acromegalic men: role in altered high density lipoproteins. *Atherosclerosis*, 153: 491-8.
4. Colao, A., Ferone, D., Marzullo, P. & Lombardi, G.(2004) Systemic complications of acromegaly: epidemiology, pathogenesis, and management. *Endocr Rev*, 25: 102-52.
5. Moller, N., Schmitz, O., Joergensen, J.O., Astrup, J., Bak, J.F., Christensen, S.E., Alberti, K.G. & Weeke, J.(1992) Basal- and insulin-stimulated substrate metabolism in patients with active acromegaly before and after adenomectomy. *J Clin Endocrinol Metab*, 74: 1012-9.
6. Theuma, P. & Fonseca, V.A.(2004) Inflammation, insulin resistance, and atherosclerosis. *Metab Syndr Relat Disord*, 2: 105-13.
7. Holland, W.L., Knotts, T.A., Chavez, J.A., Wang, L.P., Hoehn, K.L. & Summers, S.A.(2007) Lipid mediators of insulin resistance. *Nutr Rev*, 65: S39-46.
8. Jung, H.S., Park, K.H., Cho, Y.M., Chung, S.S., Cho, H.J., Cho, S.Y., Kim, S.J., Kim, S.Y., Lee, H.K. & Park, K.S.(2006) Resistin is secreted from macrophages in atheromas and promotes atherosclerosis. *Cardiovasc Res*, 69: 76-85.
9. Nagaev, I. & Smith, U.(2001) Insulin resistance and type 2 diabetes are not related to resistin expression in human fat cells or skeletal muscle. *Biochem Biophys Res Commun*, 285: 561-4.
10. Assmann, G., Jabs, H.U., Kohnert, U., Nolte, W. & Schriewer, H.(1984) LDL-cholesterol determination in blood serum following precipitation of LDL with polyvinylsulfate. *Clin Chim Acta*, 140: 77-83.
11. Warnick, G.R., Benderson, J. & Albers, J.J.(1982) Dextran sulfate-Mg²⁺ precipitation procedure for quantitation of high-density-lipoprotein cholesterol. *Clin Chem*, 28: 1379-88.
12. Lagrost, L., Gandjini, H., Athias, A., Guyard-Dangremont, V., Lallemand, C. & Gambert, P.(1993) Influence of plasma cholesteryl ester transfer activity on the LDL and HDL distribution profiles in normolipidemic subjects. *Arterioscler Thromb*, 13: 815-25.
13. Hansen, I., Tsalikian, E., Beaufriere, B., Gerich, J., Haymond, M. & Rizza, R.(1986) Insulin resistance in acromegaly: defects in both hepatic and extrahepatic insulin action. *Am J Physiol*, 250: E269-73.
14. Moller, N. & Jorgensen, J.O.(2009) Effects of growth hormone on glucose, lipid, and protein metabolism in human subjects. *Endocr Rev*, 30: 152-77.
15. Chen, X.L., Lee, K., Hartzell, D.L., Dean, R.G., Hausman, G.J., McGraw, R.A., Della-Fera, M.A. & Baile, C.A.(2001) Adipocyte insensitivity to insulin in growth hormone-transgenic mice. *Biochem Biophys Res Commun*, 283: 933-7.
16. Plockinger, U. & Reuter, T.(2008) Pegvisomant increases intra-abdominal fat in patients with acromegaly: a pilot study. *Eur J Endocrinol*, 158: 467-71.
17. Silha, J.V., Krsek, M., Hana, V., Marek, J., Jezkova, J., Weiss, V. & Murphy, L.J.(2003) Perturbations in adiponectin, leptin and resistin levels in acromegaly: lack of correlation with insulin resistance. *Clin Endocrinol (Oxf)*, 58: 736-42.
18. Antuna-Puente, B., Feve, B., Fellahi, S. & Bastard, J.P.(2008) Adipokines: the missing link between insulin resistance and obesity. *Diabetes Metab*, 34: 2-11.
19. Krag, M.B., Gormsen, L.C., Guo, Z., Christiansen, J.S., Jensen, M.D., Nielsen, S. & Jorgensen, J.O.(2007) Growth hormone-induced insulin resistance is associated with increased intramyocellular triglyceride content but unaltered VLDL-triglyceride kinetics. *Am J Physiol Endocrinol Metab*, 292: E920-7.

- 1
 - 2
 - 3
 - 4
 - 5
 - 6
 - 7
 - 8
 - 9
 - 10
 - 11
 - 12
 - 13
 - 14
 - 15
 - 16
 - 17
 - 18
 - 19
 - 20
 - 21
 - 22
 - 23
 - 24
 - 25
 - 26
 - 27
 - 28
 - 29
 - 30
 - 31
 - 32
 - 33
 - 34
 - 35
 - 36
 - 37
 - 38
 - 39
 - 40
 - 41
 - 42
 - 43
 - 44
 - 45
 - 46
 - 47
 - 48
 - 49
 - 50
 - 51
 - 52
 - 53
 - 54
 - 55
 - 56
 - 57
 - 58
 - 59
 - 60
20. Coculescu, M., Niculescu, D., Lichiardopol, R. & Purice, M.(2007) Insulin resistance and insulin secretion in non-diabetic acromegalic patients. *Exp Clin Endocrinol Diabetes*, 115: 308-16.
21. Resmini, E., Minuto, F., Colao, A. & Ferone, D.(2009) Secondary diabetes associated with principal endocrinopathies: the impact of new treatment modalities. *Acta Diabetol*, 46: 85-95.
22. Grundy, S.M.(2006) Atherogenic dyslipidemia associated with metabolic syndrome and insulin resistance. *Clin Cornerstone*, 8 Suppl 1: S21-7.
23. Vijayakumar, A., Novosyadlyy, R., Wu, Y., Yakar, S. & LeRoith, D.(2010) Biological effects of growth hormone on carbohydrate and lipid metabolism. *Growth Horm IGF Res*, 20: 1-7.
24. Verges, B.(2005) New insight into the pathophysiology of lipid abnormalities in type 2 diabetes. *Diabetes Metab*, 31: 429-39.
25. Richelsen, B., Pedersen, S.B., Kristensen, K., Borglum, J.D., Norrelund, H., Christiansen, J.S. & Jorgensen, J.O.(2000) Regulation of lipoprotein lipase and hormone-sensitive lipase activity and gene expression in adipose and muscle tissue by growth hormone treatment during weight loss in obese patients. *Metabolism*, 49: 906-11.
26. Ameen, C. & Oscarsson, J.(2003) Sex difference in hepatic microsomal triglyceride transfer protein expression is determined by the growth hormone secretory pattern in the rat. *Endocrinology*, 144: 3914-21.
27. Borggreve, S.E., De Vries, R. & Dullaart, R.P.(2003) Alterations in high-density lipoprotein metabolism and reverse cholesterol transport in insulin resistance and type 2 diabetes mellitus: role of lipolytic enzymes, lecithin:cholesterol acyltransferase and lipid transfer proteins. *Eur J Clin Invest*, 33: 1051-69.
28. Brites, F.D., Bonavita, C.D., De Geitere, C., Cloes, M., Delfly, B., Yael, M.J., Fruchart, J., Wikinski, R.W. & Castro, G.R.(2000) Alterations in the main steps of reverse cholesterol transport in male patients with primary hypertriglyceridemia and low HDL-cholesterol levels. *Atherosclerosis*, 152: 181-92.
29. Boero, L., Cuniberti, L., Magnani, N., Manavela, M., Yapur, V., Bustos, M., Rosso, L.G., Merono, T., Marziali, L., Viale, L., Evelson, P., Negri, G. & Brites, F.(2010) Increased oxidized low density lipoprotein associated with high ceruloplasmin activity in patients with active acromegaly. *Clin Endocrinol (Oxf)*, 72: 654-60.
30. Searle, A., Gomez-Rosso, L., Merono, T., Salomon, C., Duran-Sandoval, D., Giunta, G., Grant, C., Calvo, C., Lamperti, L., Brites, F. & Aguayo, C.(2011) High LDL levels are associated with increased lipoprotein-associated phospholipase A(2) activity on nitric oxide synthesis and reactive oxygen species formation in human endothelial cells. *Clin Biochem*, 44: 171-7.
31. Yarman, S., Ozden, T.A. & Gokkusu, C.(2003) The evaluation of lipid peroxidation and acute effect of octreotide on lipid peroxidation in patients with active acromegaly. *Clin Chim Acta*, 336: 45-8.
32. Andersson, I.J., Johansson, M.E., Wickman, A., Bohlooly, Y.M., Klintland, N., Caidahl, K., Gustafsson, M., Boren, J., Gan, L.M. & Bergstrom, G.(2006) Endothelial dysfunction in growth hormone transgenic mice. *Clin Sci (Lond)*, 110: 217-25.
33. Sainani, G.S., Maru, V.G. & Mehra, A.P.(2005) Role of endothelin-1 in genesis of coronary artery disease. *Indian Heart J*, 57: 121-7.
34. Kirilov, G., Zacharieva, S., Alexandrov, A.S., Lozanov, V. & Mitev, V.(2009) Increased plasma endothelin level as an endothelial marker of cardiovascular risk in patients with active acromegaly: a comparison with plasma homocysteine. *Methods Find Exp Clin Pharmacol*, 31: 457-61.