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Data and text mining

PPaxe: easy extraction of protein occurrence and interactions from the scientific literature

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Abstract

Motivation: Protein-protein interactions (PPIs) are very important to build models for understanding many biological processes. Although several databases hold many of these interactions, exploring them, selecting those relevant for a given subject, and contextualizing them can be a difficult task for researchers. Extracting PPIs directly from the scientific literature can be very helpful for providing such context, as the sentences describing these interactions may give insights to researchers in helpful ways.

Results: We have developed PPaxe, a python module and a web application that allows users to extract PPis and protein occurrence from a given set of PubMed and PubMedCentral articles; It presents the results of the analysis in different ways to help researchers export, filter and analyze the results easily. **Availability:** PPaxe web demo is freely available at https://compgen.bio.ub.edu/PPaxe. All the software can be downloaded from https://compgen.bio.ub.edu/PPaxe/download, including a command-line version and docker containers for an easy installation.

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Supplementary information: Supplementary data are available at *Bioinformatics* online.

1 Introduction

Protein-protein interactions (PPIs) play a major role in many biological processes, such as cancer, regeneration, and the development of many diseases (Ian et al., 2012); which makes the identification of these interactions a key point for the understanding of these processes. Databases such as iHOP (Hoffmann et al., 2005) have helped researchers to navigate the scientific literature using genes and proteins as drivers. Further tools exist in order to automatically retrieve interactions described in the scientific literature (Raja et al., 2013; Quan et al., 2014; Zhao et al., 2016); however, the accessibility to these tools for researchers is rather difficult, either because they can't be easily downloaded or because they lack a user-interface. Here we present **PPaxe**, a python module and a web application. superseding already developed code used for a previous analysis of the protein network of retinitis pigmentosa; that code was referred to as Sparser in the methods section of Boloc et al. 2015.

Implementation

PPaxe reads a text file with a list of PubMed identifiers and downloads all the necessary articles. **PPaxe** downloads either abstracts from Medline or full-text articles from PubMed Central, depending on the option provided by the user. On the web application, the user can also provide a wellformatted PubMed query, so that the tool can retrieve the list of publication identifiers directly from NCBI PubMed; or a plain-text file with the text to be analyzed.

PPaxe uses Stanford CoreNLP (Manning *et al.*, 2014) for name entity recognition (NER) of proteins and genes. Three datasets were used in order to train the The Stanford Named Entity Recognizer: AImed (Bunescu *et al.*, 2005), MedTag (Smith *et al.*, 2005) and BioInfer (Pyysalo *et al.*, 2007). First, each sentence was tokenized by Stanford CoreNLP; then, a Conditional Random Field (CRF) classifier was trained. Performance of the NER tagger was assessed by 2-fold crossvalidation. Once the NER is trained, **PPaxe** extracts all the co-occurring proteins in each sentence and for each pair of them, computes several features, as described on Suppl. Mat. Table S1, which will be used in order to identify pairs as interacting

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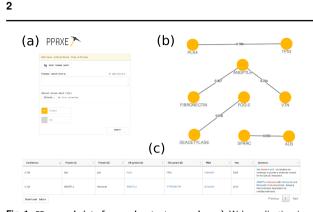


Fig. 1. PPaxe web interface and output examples. a) Web application input form; b) Graph visualization from the HTML report. c) Sentences containing PPIs retrieved from the analysis of four PubMed abstracts (PMIDs: 15640847, 20729546, 25196150, and 25211495), presented in the HTML output as a searchable table. The Confidence value (shown in the table and on the graph edges) corresponds to the normalized percentage of votes of the predictor (ranging from 0 to 1). Enlarged version available as Suppl. Mat. Figure S1.

or not. The prediction is based on a Random Forest Classifier from scikit-learn (Pedregosa *et al.*, 2011), trained over the annotated sentences of AImed, LLL-challenge (Nédellec, 2005) and BioInfer. A votes cut-off of 0.55 was selected to detect interacting proteins from sentences, based on the performance of the classifier on our evaluation (estimated using 10-fold cross-validation).

Finally, PPaxe produces several possible outputs: an HTML page with all the PPIs, the sentences in which they were found, a table with all the proteins found in the specified articles, a graph visualization made with the JavaScript library cytoscape.js (Franz *et al.*, 2015), and a PDF with a summary of the analyses. PPaxe is also distributed as two docker images, a command-line application and a web application, respectively; which can be retrieved from the downloads page. The web application docker image runs a localhost server on a customizable port. PPaxe python modules are available too.

Results and Discussion

Protein and gene name tagging by the Stanford CoreNLP was evaluated by 2-fold cross-validation on the datasets of AImed, MedTag and Bioinfer; which resulted in a precision of 74.5%, a recall of 70.0%, and an F1 of 72.1%. As PPaxe does not use any dictionary table in order to identify the proteins, it is able to tag protein symbols of any species, or even newly described proteins and genes. However, it cross-checks the identifiers against the aliases provided by the HUGO Gene Nomenclature Committee as a prior normalization step (Yates *et al.*, 2017).

PPaxe is able to retrieve protein-protein and genetic interactions without needing to define specific patterns or rules, which makes the application's use broader and more general than other previously mentioned approaches, such as PPInterFinder (Raja *et al.*, 2013). **PPaxe** considers only a narrow selection of features, namely POS composition, token distance, and keywords (see Suppl. Mat. Table S3), freeing **PPaxe** of syntactical parsing and the posterior processing of dependency trees. On an Intel Core i7 machine, **PPaxe** took $\sim 2''$ to download and analyze 10 articles, 16'' for 100, 2'36'' for 1,000, and 28'54'' for 10,000. An assessment of the interaction extraction performance and a validation over BioGRID (Stark *et al.*, 2006) are described in Suppl. Mat. evaluation section and is shown on Table S1 and Table S2. PPaxe facilitates the visualization of the results, without requiring any other additional software, thanks to the inclusion of an HTML report output. The output includes several summary tables, a dynamic visualization of the retrieved interactions, along with some statistics for the retrieved references (Fig.1).

In conclusion, yet some similar tools have already been developed, **PPaxe** allows researchers both to perform large-scale text-mining, but also to analyze small and focused sets. The newly implemented range of output options and reports should make **PPaxe** a valuable tool for researchers to retrieve and curate novel PPIs described in the scientific literature.

Acknowledgements

The authors are grateful to our beta tester users that played with the initial versions of the tool, especially to Rodrigo Arenas-Galnares, and to the referees for their useful comments.

Funding

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Supplementary materials

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PPaxe features

PPaxe uses three types of features: token distance measures between the entities in the sentence (e.g. from each candidate protein to each verb between them), POS tag composition (both between the candidate proteins and in their vicinity), and finally keyword occurrence in the sentences. This approach makes **PPaxe** capable of retrieving interactions that don't follow any particular structure or pattern, such as "*Protein-A binds Protein-B*", "*Protein-A and Protein-B interact.*", and so on. All features are listed on Supplementary Materials Table S3.

2 PPaxe evaluation

Performance of the PPaxe estimated over the three datasets using 10-fold cross-validation, compared to previously published tools, is shown on Supplementary Materials Table S1. *Quan et. al* I and II correspond to the unsupervised and the semi-supervised methods described in that article respectively. The performance of PPaxe when it comes to retrieving interactions varies from dataset to dataset. Overall, PPaxe performed better than a previously described method on the same datasets (Zhao *et al.*, 2016), slightly worse than a rule-based approach (Raja *et al.*, 2013), and better and similarly to an unsupervised and a semi-supervised method respectively (Quan *et al.*, 2014).

2.1 Performance metrics

$$Accuracy = \frac{tp+tn}{tp+tn+fp+fn}$$

$$Precision = \frac{tp}{tp+fp}$$

$$Recall = \frac{tp}{tp+fn}$$

$$F1 = 2 \cdot \frac{Precision \cdot Recall}{Precision + Recall}$$

where:

- tp is true positives.
- tn is true negatives.
- fp is false positives.
- fn is false negatives.

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2.2 BioGRID test case

PPaxe interaction extraction was compared against the interactions annotated in the BioGRID database. In order to do so, the following query was performed on PubMed:

protein protein interactions
 AND (hasabstract[text]
 AND "last 5 years"[PDat]
 AND Humans[Mesh])

That PubMed query returned 41,286 entries on October 17^{th} , 2018. We picked up the first 2,000 PubMed identifiers for posterior analyses, after sorting them by decreasing publication date, to ensure retrieving the most recent papers that may contain novel interactions.

PPaxe retrieved 104 interactions (92 unique). From those interactions, 31 contained interactions already annotated in BioGRID, defined as "BioGRID positive" (valid sentence defining interactions already supported by BioGRID) and "BioGRID negative" (wrong sentences defining an interaction contained in BioGRID). We manually annotated the remaining 73 interactions as "PPaxe positive" (those sentences with a correct protein-protein or genetic interaction, according to the sentence alone) or "PPaxe negative" (interactions not supported by the sentence). The results of this annotation can be seen in Table S2.

Even though the comparison against BioGRID showed that most interactions retrieved by PPaxe were not annotated in the database, the manual annotation revealed that most of them were in fact described in the sentence: 40 out of the 73 sentences were annotated as "PPaxe positive". By considering both "BioGRID positive" and "PPaxe positive" together, 65.38% of the interactions retrieved by PPaxe can be considered correctly retrieved, which is slightly smaller than the estimated precision of interaction retrieval computed by 10-fold cross validation shown in Table S1.

We can't consider this test as a full assessment of PPaxe performance due to several limitations of this approach. For instance, we don't have a gold standard neither for sentences, nor interactions; we can't consider BioGRID as a full validated gold standard, and the annotation of the sentences was performed after their selection. Moreover, the impact of the protein/gene tagger can't be considered separately from the interaction extraction procedure as it has been done in the cross-validation tests. In summary, that implies we can't estimate some of the performance metrics such as the Recall to compare those results with the cross-validation assessment. Finally, the large number of correctly retrieved interactions by PPaxe that were not annotated in BioGRID could be explained by either differences in gene symbol normalization (although both sets of protein symbols were normalized using the HGNC alias dictionary), by the use of non-official gene names in the articles, or by the fact that the interactions are yet to be included in BioGRID.

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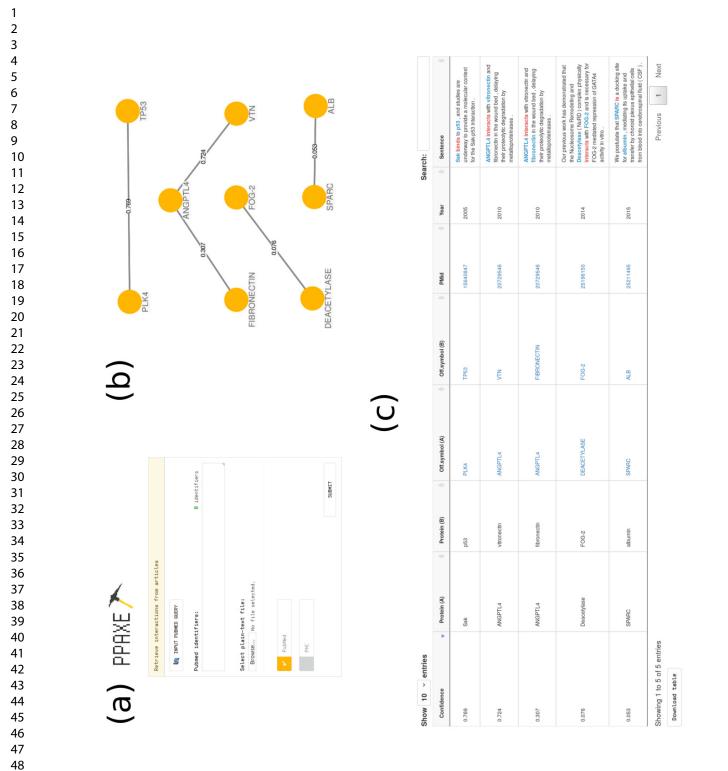


Figure S1: A larger view of manuscript Figure 1. a) PPaxe web application form, which allows users to both input PubMed identifiers directly or to write a PubMed query to retrieve the requested articles. b) Graph visualization made using Cytoscape.js from the HTML report of PPaxe. c) Sentences containing PPIs retrieved from the analysis of four PubMed abstracts (PMIDs: 15640847, 20729546, 25196150, and 25211495), presented in the HTML output as a searchable table. The Confidence value (shown in the table and on the graph edges) corresponds to the normalized percentage of votes of the predictor (ranging from 0 to 1).

Table S1: Assessment of PPaxe interaction extraction performance. PPaxe performance metrics were computed with a 10-fold cross-validation for each dataset. The same procedure was not applied to the other tools, due to the inability to get the corresponding software to run under the same conditions. Therefore, the metrics displayed on this table were retrieved from their respective articles. "All sets" correspond to the validation run on a merged dataset built from the other three (AImed + BioInfer + LLL).

	Tool	ACCURACY	PRECISION	RECALL	F1
	PPaxe	80.07	70.54	50.93	59.15
	Zhao et. al		50.51	63.38	56.12
AImed	Quan et. al I		44.80	71.40	55.10
	Quan et. al II	_	56.6	66.80	60.7
	Raja et. al		80.25	56.12	66.05
BioInfer	PPaxe	88.42	81.27	65.55	72.57
	Zhao et. al	- 5	53.89	72.9	61.63
LLL	PPaxe	72.96	76.98	87.39	81.86
	Zhao <i>et. al</i>		75.84	91.84	82.00
All sets	PPaxe	84.48	76.50	58.68	66.41

Table S2: **PPaxe extracted interactions when intersecting them with those in BioGRID.** The retrieved interactions were compared against the BioGRID database, and those not annotated in the database were manually curated. In blue, 28 interactions described in BioGRID ("BioGRID positive"); in yellow, 3 interactions described in BioGRID but annotated as incorrect after manual inspection ("BioGRID negative"); in green, 40 interactions not described in BioGRID but annotated as correct after manual curation ("PPaxe positive"); in red, 33 interactions not described in BioGRID annotated as incorrect ("PPaxe negative"). The "Conf." value corresponds to the normalized percentage of votes of the random forest classifier used by PPaxe.

PMID	INT A	INT B	Conf.	Sentence after tokenization
29899090	TRIM41 (TRIM41)	NP (ZNF384)	0.796	Here , we report that TRIM41 interacts with NP through its SPRY domain
29444082	S100B (S100B)	S100A1 (S100A1)	0.676	We found that S100B could interact with S100A1 via NMR 1H-15N HSQC titrations
29178343	GRP78 (HSPA5)	PRDM14 (PRDM14)	0.644	These results suggest that HSP90 and GRP78 interact with PRDM14 and participate in cancer regulation $\$
29220652	PARP1 (TIPARP)	ALC1 (MYL4)	0.627	Its engagement with PAR ylated PARP1 activates ALC1 at sites of DNA damage , but the underlying mechanism remains unclear $% \lambda =0.01$
29358401	DNAAF2 (DNAAF2)	SPAG1 (SPAG1)	0.564	FRET analysis of HEAT domain deletions and human mutations showed that HEATR2 interacted with itself and SPAG1 at multiple HEAT domains , while DNAAF2 interacted with SPAG1
30111544	REV7 (MAD2L2)	REV3 (REV3L)	0.556	Rev7 interacts with Rev3 by a mechanism conserved among HORMA proteins , whereby an open-to-closed transition locks the ligand underneath the "safety belt" loop
29328377	MYC (MYC)	CDK2 (CDK2)	0.518	The cyclindependent kinase inhibitor 1A (CDKN1A) , E2F transcription factor 1 (E2F1) , and MYC interacted with CDK2
29374759	VDR (VDR)	P53 (TP53)	0.480	VDR binding to p53 was confirmed by western blot analysis
30021902	CDC25A (CDC25A)	TBK1 (TBK1)	0.458	Further analysis indicated that Cdc25A can interact with TBK1 and reduce the phosphorylation of TBK1 at S172 , which in turn decreases the phosphorylation of its downstream substrate IRF3
29281729	IFITM3 (IFITM3)	LSD1 (KDM1A)	0.458	Our data suggest that the demethylation of IFITM3 by LSD1 is beneficial for the host to fight against RNA virus infection
29684085	KAT5 (KAT5)	CD4 (CD4)	0.449	The pro-latency effect of KAT5 is confirmed in a primary CD4 $+$ T cell latency model as well as cells from ART-treated patients
29295922	PAK4 (PAK4)	CDC42 ($CDC42$)	0.436	Using solution scattering we find that the full-length PAK4 het- erodimer with CDC42 adopts primarily a compact organization
29154191	CD11B (ITGAM)	RHO (RHOD)	0.413	Mechanistically , -synuclein bound to CD11b and subsequently activated Rho signaling pathway
29295922	CDC42 (CDC42)	PAK4 (PAK4)	0.387	These additional interactions modulate kinase activity and increase the binding affinity of CDC42 for full-length PAK4 compared with the CRIB domain alone
30021902	CDC25A (CDC25A)	IRF3 (IRF3)	0.382	Consistently , knockdown of Cdc25A upregulates the phosphory- lation of both TBK1-S172 and IRF3 in Sendai virus-infected or TBK1-transfected 293T cells
29925658	STAT1 (STAT1)	NSP2 (RTN2)	0.378	Chemically blocking CRM1-mediated nuclear export in the pres- ence of nsP2 additionally showed that nuclear translocation of STAT1 is not affected by nsP2
29218693	NCK2 (NCK2)	ITGB1 (ITGB1)	0.373	Co-IP showed that NCK2 can directly bind ITGB1 , but not VEGFA
29334217	P53 (TP53)	$\mathrm{HSP90}\ (\mathrm{HSP90AA1})$	0.360	The DNA-binding domain (DBD) of p53 is known to interact with the chaperone Hsp90 , but the role of other members of the chaperone network , including co-chaperones such as p23 , is unknown
29295922	CDC42 (CDC42)	PAK4 (PAK4)	0.351	We therefore show that the interaction of CDC42 with PAK4 can influence kinase activity in a previously unappreciated manner
29358401	HEATR2 (DNAAF5)	SPAG1 (SPAG1)	0.338	FRET analysis of HEAT domain deletions and human mutations showed that HEATR2 interacted with itself and SPAG1 at multiple HEAT domains , while DNAAF2 interacted with SPAG1

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PMID	INT A	INT B	Conf.	Sentence after tokenization
29232376	GCN5 (KAT2B)	CHE1 (BCHE)	0.329	In this study , we aimed to identify whether besides ADA3 , other components of the HAT modules of SAGA and ATAC complexed , human ADA2 and GCN5 also interact with Che-1 / AATF
29379028	NS1 (PTPN11)	TBK1 (TBK1)	0.324	This mutation enables NS1 binding to TBK1 and reduces TBK phosphorylation
29184850	CELLUGYRIN (SYNGR2)	CDTB (CDB2)	0.324	Furthermore , we demonstrate that cellugyrin is an intracellula binding partner for CdtB as demonstrated by immunoprecipitation
29512721	OSTEOCALCIN (BGLAP)	FOXO1 (FOXO1)	0.311	The HIF1induced expression of Runx2 and ALP may be com- pletely dependent on the expression levels of Foxo1 , and in tur , osteocalcin may be partially dependent on Foxo1 expression
29950413	SOX2 (SOX2)	STAT3 (STAT3)	0.307	IE1 mediates SOX2 depletion by targeting STAT3 , a critical up stream regulator of SOX2 expression
29281729	LSD1 (KDM1A)	IFITM3 (IFITM3)	0.302	We have found that LSD1 is recruited to demethylate IFITM3 a position K88 under IFN treatment
29212519	VEGF (VEGFA)	IGF1 (IGF1)	0.298	We found that a functional cooperation between HIF-1 and GPE is essential for the transcriptional activation of VEGF induced b IGF1
29178989	NETS (SPINK5)	FSAP (HABP2)	0.298	Taken together , NETs bind to FSAP , but do not activate pro- FSAP unless histones are released from NETs by DNAse
29743362	FUBP1 (FUBP1)	P53 (TP53)	0.284	Here we report that human adenovirus 5 coopts the cellular pro- tein FUBP1 to prevent the activation of the p53 stress respon- pathway that would block viral replication
29152905	L13A (RPL13A)	EIF4G (EIF4G1)	0.280	EPRS binds the GAIT element in target mRNAs , NSAP1 neg tively regulates mRNA binding , L13a binds eIF4G to block rib some recruitment , and GAPDH shields L13a from proteasom degradation
29899107	MDA5 (IFIH1)	IFN (IFNA1)	0.271	Although RIG-I has been recognized as the leading cytoplasm sensor against HCV for a long time , recent findings that MDA regulates the IFN response to HCV have emerged
29665350	PXR (NR1I2)	CYP3A4 (CYP3A4)	0.262	In conclusion , PXR activation and PXR-mediated induction CYP3A4 expression by PAs seem to be structure-dependent
29669839	GP (RNF130)	TETHERIN (BST2)	0.258	To our knowledge , these findings demonstrate for the first tim that GP can antagonize tetherin in infected cells and provide tool to study the impact of GP-dependent tetherin counteraction on EBOV spread
29232376	GCN5 (KAT2B)	AATF (AATF)	0.258	In this study , we aimed to identify whether besides ADA3 , oth components of the HAT modules of SAGA and ATAC complex , human ADA2 and GCN5 also interact with Che-1 / AATF
29660231	S100A4 (S100A4)	P53 (TP53)	0.253	Wnt $/$ -catenin targets , c-MYC and S100A4 were upregulate in p53 cells and were downregulated when plakoglobin was coepressed
29690653	VEGFA (VEGFA)	VEGFR2 (KDR)	0.218	Although VEGF-A ligands bind to both VEGFR1 and VEGFR , they primarily signal via VEGFR2 leading to endothelial c proliferation , survival , migration and vascular permeability
29925821	RHOA (RHOA)	RAC1 (RNASE1)	0.213	All genetic evidences indicate that in these disorders the Rho pathway is hyperactive while the Rac1 and cdc42 pathways a consistently hypoactive
29690653	VEGFA (VEGFA)	VEGFR1 (FLT1)	0.213	Although VEGF-A ligands bind to both VEGFR1 and VEGFI , they primarily signal via VEGFR2 leading to endothelial c proliferation, survival, migration and vascular permeability
29212519	GPER (GPER1)	VEGF (VEGFA)	0.204	We found that a functional cooperation between HIF-1 and GPE is essential for the transcriptional activation of VEGF induced IGF1
29444113	KCNQ1 (KCNQ1)	KCNE1 (KCNE1)	0.200	The results reveal that interactions between KCNQ1 with KCN causes a pore constriction in the former , which in-turn forms sm energetic barriers in the ion-permeation pathway
30111544	REV7 (MAD2L2)	REV1 (REV1)	0.196	We demonstrate that Rev7 uses the conventional HORMA dime ization interface both to form a homodimer when tethered by t two RBMs in Rev3 and to heterodimerize with other HORM domains , Mad2 and p31 Structurally , the Rev7 dimer can bi only one copy of Rev1 , revealing an unexpected Rev1/Pol arc tecture
29677136	NOP53 (NOP53)	RIGI (DDX58)	0.182	Cytoplasmic NOP53 interacts with the retinoic acid-inducil gene I (RIG-I) to remove its K63-linked ubiquitination, leadi to attenuation of type I interferon IFN-

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1 2	PMID	INT A	INT B	Conf.	Sentence after tokenization
3 4 5	29269420	L1 (IGKV116)	CENPA (CENPA)	0.182	We show that human CENP-N confers binding specificity through interactions with the L1 loop of CENP-A , stabilized by electrostatic interactions with the nucleosomal DNA
6 7	29690653	VEGFA (VEGFA)	VEGFR2 (KDR)	0.178	This review explores the molecular pharmacology of VEGF-A iso- forms at VEGFR2 in respect to ligand binding and downstream signalling
8 9	29167311	AND1 (WDHD1)	CTF4 (WDHD1)	0.178	AND-1 has maintained the trimeric structure of yeast Ctf4 , driven by its conserved SepB domain
10 11 12	29925658	NSP2 (RTN2)	IFN (IFNA1)	0.173	The research described here specifies where in the JAK/STAT signaling cascade the IFN response is inhibited and which protein domain of nsP2 is responsible for IFN inhibition
13 14 15	29207260	E2F1 (E2F1)	TEAD1 (TEAD1)	0.173	Further , we found that human E2F1 competes with YAP for TEAD1 binding , affecting YAP activity , indicating that this mode of cross-regulation is conserved
16 17	29444113	KCNE1 (KCNE1)	KCNQ1 (KCNQ1)	0.169	These findings correlate with the previous experimental reports that interactions of KCNE1 dramatically slows the activation of KCNQ1
18 19 20 21 22 23	29297316	CCR5 (CCR5)	CD4 (CD4)	0.169	It was found that the subnetworks formed by CCR5 and IFNAR1 and their neighbors were a fragments of two key pathways functioning during the course of tick-borne encephalitis : (1) the attenuation of interferon-I signaling pathway by the TBEV NS5 protein that targeted peptidase D; (2) proinflammation and tissue damage pathway triggered by chemokine receptor CCR5 interacting with CD4, CCL3, CCL4, CCL2
24 25	29669839	GP (RNF130)	TETHERIN (BST2)	0.160	Moreover , they provide the first evidence that GP can antagonize tetherin in the context of an infectious EBOV surrogate
26 27	29321315	RIGI (DDX58)	TRIM25 (TRIM25)	0.151	We report interactions between the Nipah virus V protein and both RIG-I regulatory protein TRIM25 and RIG-I
28 29 30	29471045	PRDM14 (PRDM14)	HOXA1 (HOXA1)	0.147	Here , we confirm PRDM14 is an interactor of HOXA1 and we identify the homeodomain of HOXA1 as well as the PR domain and Zinc fingers of PRDM14 to be required for the interaction
31 32	29367244	ATG12 (ATG12)	ATG5 (ATG5)	0.147	A close inspection of the HBV/autophagy cross talk revealed that the virus depended on Atg12 covalently conjugated to Atg5 $$
33 34	29760086	STIM1 (STIM1)	ORAI1 (ORAI1)	0.138	Store-operated Orai1 channels are activated through a unique inside-out mechanism involving binding of the endoplasmic reticulum Ca sensor STIM1 to cytoplasmic sites on Orai1
35 36 37	29549180	P53 (TP53)	MDM2 (MDM2)	0.133	In unstressed cells , p53 is normally held in check by MDM2 that targets p53 for transcriptional repression , proteasomal degradation , and cytoplasmic localization
38 39	29614078	GDNF (GDNF)	AP1 (JUND)	0.124	GDNF stimulates MAP kinase , activating the transcription factors SRF and AP-1 $$
40 41 42	29512721	ALP (SLPI)	FOXO1 (FOXO1)	0.116	The HIF1 induced expression of Runx2 and ALP may be completely dependent on the expression levels of Foxo1 , and in turn , osteocalcin may be partially dependent on Foxo1 expression
43 44	29734338	RIT1 (RIT1)	RAC1 (RNASE1)	0.111	We found RIT1 also to directly interact with the RHO GTP ases CDC42 and RAC1 , both of which are crucial regulators of actin dynamics upstream of PAK1
45 46 47	29232376	ADA2 (TADA2A)	AATF (AATF)	0.111	In this study , we aimed to identify whether besides ADA3 , other components of the HAT modules of SAGA and ATAC complexes , human ADA2 and GCN5 also interact with Che-1 / AATF
48 49	30021902	CDC25A (CDC25A)	TBK1 (TBK1)	0.107	These results demonstrate that Cdc25A inhibits the antiviral immune response by reducing the active form of TBK1 $$
50 51 52 53	29339503	BTN3A2 (BTN3A2)	BTN3A1 (BTN3A1)	0.107	Addressing this paradox , we show that BTN3A2 regulates the subcellular localization of BTN3A1 , including functionally important associations with the endoplasmic reticulum ($\rm ER$), and is specifically required for optimal BTN3A1-mediated activation of V9V2 T cells
54 55	29328377	E2F1 ($E2F1$)	CDK2 (CDK2)	0.107	The cyclindependent kinase inhibitor 1A (CDKN1A) , E2F transcription factor 1 ($\rm E2F1$) , and MYC interacted with CDK2
56 57	30181274	RAB5 (RAB5A)	$\begin{array}{c} \text{RABGAP5} \\ \text{(SGSM3)} \end{array}$	0.102	Thus , binding of Etf-2 to RAB5-GTP appears to delay RAB5 inactivation by impeding RABGAP5 localization to endosomes
58 59	29899144	LPL (LPL)	GPIHBP1 (GPI- HBP1)	0.102	Third , we show that LPL accumulates near capillary endothelial cells even in the absence of GPIHBP1
60	29949917	DDX6 (DDX6)	RIGI (DDX58)	0.098	These findings imply a novel function for DDX6 as an RNA co- sensor and signaling enhancer for RIG-I
					Continued on next page

PMID	INT A	INT B	Conf.	Sentence after tokenization
29949917	DDX6 (DDX6)	RIGI (DDX58)	0.098	Notably , DDX6 was found to bind viral RNA capable to stimul RIG-I $$
29712868	SOMATOSTATIN28 (SST)	INSULIN (INS)	0.098	To demonstrate the usefulness of this approach in revealing a ulatory pathways , we identify from among the contacted s the previously uncharacterized gene and show that it plays an portant role in controlling the effect of somatostatin-28 on insu- secretion
29269420	L1 (IGKV116)	CENPA (CENPA)	0.098	Mutational analyses demonstrate analogous interactions in , whare further supported by residue-swapping experiments involve the L1 loop of CENP-A
29450990	RBM10 (RBM10)	RBM5 (RBM5)	0.089	Splicing factor RBM10 and its close homologues RBM5 RBM6 govern the splicing of oncogenes such as Fas , NUM and Bcl-X $$
29321329	SAMHD1 (SAMHD1)	CYCLIN A2 (CCNA2)	0.089	Human SAMHD1 interacts with cell cycle regulatory proteins $clin A2$, $cyclin-dependent$ kinase 1 ($CDK1$), and $CDK2$
29205500	HSAMIR1270 (MIR1270)	TAP2 (TAP2)	0.089	Our findings support the hypothesis that hsa-miR-1270 suppret the production of TAP2 by binding to this SNP in the 3 ' - U of this gene
29281729	IFITM3 (IFITM3)	LSD1 (KDM1A)	0.087	However , infection by either Vesicular Stomatitis Virus (VS or Influenza A Virus (IAV) triggers methylation of IFITMS promoting its disassociation from LSD1
29297316	CCR5 (CCR5)	CCL4 (CCL4)	0.084	It was found that the subnetworks formed by CCR5 and IFNA and their neighbors were a fragments of two key pathways fit tioning during the course of tick-borne encephalitis : (1) attenuation of interferon-I signaling pathway by the TBEV 1 protein that targeted peptidase D; (2) proinflammation tissue damage pathway triggered by chemokine receptor CO interacting with CD4, CCL3, CCL4, CCL2
29734338	RIT1 (RIT1)	CDC42 (CDC42)	0.080	This effect was prevented by co-expression of RIT1 v dominant-negative CDC42 or RAC1 and kinase-dead PAK1
29505800	RAB7A (RAB7A)	ER (EREG)	0.076	Collectively , these findings reveal a new role of Rab7a in homeostasis , and indicate that genetic and pharmacological stress manipulation may restore ER morphology in Rab7a siler cells
29382845	TAX (CNTN2)	UPF1 (UPF1)	0.076	Tax interacts with the central helicase core domain of UPF1 might plug the RNA channel of UPF1 , reducing its affinity nucleic acids
29297316	CCR5 (CCR5)	CCL3 (CCL3)	0.076	It was found that the subnetworks formed by CCR5 and IFNA and their neighbors were a fragments of two key pathways fu- tioning during the course of tick-borne encephalitis : (1) attenuation of interferon-I signaling pathway by the TBEV protein that targeted peptidase D; (2) proinflammation tissue damage pathway triggered by chemokine receptor CC interacting with CD4, CCL3, CCL4, CCL2
29925821	RHOA (RHOA)	CDC42 (CDC42)	0.071	All genetic evidences indicate that in these disorders the R pathway is hyperactive while the Rac1 and cdc42 pathways consistently hypoactive
29669839	TETHERIN (BST2)	GP (RNF130)	0.071	However , tether in antagonism by GP has so far been dem strated only with virus-like particles , and it is unknown where GP can block tether in infected cells
30021902	CDC25A (CDC25A)	TBK1 (TBK1)	0.062	We demonstrated that Cdc25A reduces TBK1 activity and e sequently restrains the activation of IFN- transcription as we the antiviral status of nearby cells
29950413	STAT3 (STAT3)	SOX2 (SOX2)	0.062	IE1 mediates SOX2 depletion by targeting STAT3 , a critical stream regulator of SOX2 expression
29501414	PP1C (PPP1CC)	MYPT1 (PPP1R12A)	0.062	Br-BASG and TFM-BASG suppressed partially binding of P to MYPT1 in surface plasmon resonance based binding exp ments
29669839	GP (RNF130)	TETHERIN (BST2)	0.058	However , tether in antagonism by GP has so far been den strated only with virus-like particles , and it is unknown whe GP can block tether in infected cells
29614078	GDNF (GDNF)	SRF (SRF)	0.058	GDNF stimulates MAP kinase , activating the transcription tors SRF and AP-1
29571014	EIF4A3 (EIF4A3)	CASC2 (CASC2)	0.058	Knockdown of EIF4A3 reversed the effects of sanguinarine p CASC2 silencing

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1 2	PMID	INT A	INT B	Conf.	Sentence after tokenization
3 4 5 6 7 8	29297316	CCR5 (CCR5)	CCL2 (CCL2)	0.053	It was found that the subnetworks formed by CCR5 and IFNAR1 and their neighbors were a fragments of two key pathways func- tioning during the course of tick-borne encephalitis : (1) the attenuation of interferon-I signaling pathway by the TBEV NS5 protein that targeted peptidase D; (2) proinflammation and tissue damage pathway triggered by chemokine receptor CCR5 interacting with CD4, CCL3, CCL4, CCL2
9 10 11	29444113	KCNQ1 (KCNQ1)	KCNE1 (KCNE1)	0.049	The voltage-gated KCNQ1 potassium ion channel interacts with the type I transmembrane protein minK (KCNE1) to generate the slow delayed rectifier (IKs) current in the heart
12 13	29212935	MTOR (MTOR)	CD63 (CD63)	0.049	Increases in mTOR activation following CD63 knockout are coin- cident with the development of serum-dependent autophagic vac- uoles that are acidified in the presence of high LMP1 levels
14 15 16 17	29684085	BRD4 (BRD4)	TAT (TAT)	0.044	This unusual acetylation profile attracts Brd4 to suppress the in- teraction of Tat with the host super elongation complex (SEC) that is essential for productive HIV transcription and latency reversal
18 19	29450990	RBM10 (RBM10)	RBM6 (RBM6)	0.044	Splicing factor RBM10 and its close homologues RBM5 and RBM6 govern the splicing of oncogenes such as Fas , NUMB , and Bcl-X
20 21	29494137	CAM (KRIT1)	CSH2 (CSH2)	0.040	Because of the flexible nature of both CaM and cSH2 , multiple binding modes of the interactions are possible
22 23 24	29605296	ATF3 (ATF3)	JNK (MAPK8)	0.036	Mechanistically , TNFinduced Atf3 expression was significantly suppressed by the inhibition of the c-Jun N-terminal kinase (JNK) pathway
25 26 27	29925658	IFN (IFNA1)	STAT1 (STAT1)	0.031	This demonstrates that the C-terminal domain of nuclear nsP2 specifically inhibits the IFN response by promoting the nuclear export of STAT1
28	29705079	TLR2 (TLR2)	SAA1 (SAA1)	0.031	As TLR2 is known to be a functional receptor of SAA1 , a co- immunoprecipitation assay was performed
29 30	29375208	IFN (IFNA1)	HUH7 (MIR73HG)	0.031	HCV-1b core protein increased miR-93-5p expression and induced inactivation of the IFN signaling pathway in Huh7 cells
31 32	29269396	SP1 (SP1)	OSM (OSM)	0.031	Knockdown of Sp1 abrogated the expression and functionality of OSM
33 34	29614078	SRF (SRF)	EGR1 (EGR1)	0.022	SRF initiates an immediate transcriptional response , activating EGR1 and suppressing ER
35 36	29495654	NSP3 (SH2D3C)	PIN (DYNLL1)	0.018	The function of nsP3 has been more difficult to pin down and it has long been referred to as the more enigmatic of the nsPs
37 38 39	29328377	E2F TRAN- SCRIPTION FAC- TOR 1 (E2F1)	CDK2 (CDK2)	0.018	The cyclindependent kinase inhibitor 1A (CDKN1A) , E2F transcription factor 1 (E2F1) , and MYC interacted with CDK2
40 41	29184850	CELLUGYRIN (SYNGR2)	CDTB (CDB2)	0.018	We propose that cellugyrin plays a critical role in the internal- ization and translocation of CdtB to critical intracellular target sites
42 43 44 45	29203283	PD1 (SPATA2)	CDR2 (CDR2)	0.013	The residue-residue contact analysis further shows that PD-1 in- teracts with PD-L1 mainly by F and G strands while monoclonal antibodies like avelumab and BMS-936559 mainly interact with PD-L1 by CDR2 and CDR3 loops of the heavy chain
46 47 48	30181274	RAB5 (RAB5A)	RAB5 (RAB5A)	0.009	Ectopically expressed Etf-2-GFP also localized to inclusions and membranes of early endosomes marked with RAB5 and interacted with GTP-bound RAB5 but not with a GDP-bound RAB5
49 50 51	29895728	E2 (SNORA62)	E1 (SNORA73A)	0.004	These findings support a role for E2 beyond E1 recruitment in viral DNA replication , demonstrate a novel functional interaction in PV DNA replication , and further implicate cellular pol in PV DNA replication
52 53 54	29382759	MALT1 (MALT1)	BCL10 (BCL10)	0.001	Cooperative MALT1 interaction with BCL10 filaments observed under EM suggests immediate dimerization of MALT1 in the BCL10 filamentous scaffold
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Table S3: PPaxe features .

3		Table S3: PPaxe features .
4	Feature name	Description
5 6	TOK_DIST	Token distance between ProtA and ProtB.
7	TOTAL_TOK	Number of Tokens in sentence.
8	BETWEEN_VB_COUNT	Number of Tokens tagged as VB between ProtA and ProtB.
9	BETWEEN_VBD_COUNT	Number of Tokens tagged as VBD between ProtA and ProtB.
10 11	BETWEEN_VBG_COUNT	Number of Tokens tagged as VBG between ProtA and ProtB.
12	BETWEEN_VBN_COUNT	Number of Tokens tagged as VBN between ProtA and ProtB.
13	BETWEEN_VBP_COUNT	Number of Tokens tagged as VBP between ProtA and ProtB.
14 15	BETWEEN_VBZ_COUNT	Number of Tokens tagged as VBZ between ProtA and ProtB.
15	BETWEEN_VERB_MAXSCORE	Higher score for verbs between ProtA and ProtB.
17	BETWEEN_VERB_TOTALSCORE	Sum of verb scores between ProtA and ProtB.
18	BETWEEN_VERB_CLOSEST_DIST_A	Token distance to closer verb to ProtA located between ProtA and ProtB.
19 20	BETWEEN_VERB_FARTHEST_DIST_A	Token distance to farthest verb to ProtA located between ProtA and ProtB.
20	BETWEEN_VERB_CLOSEST_DIST_B	Token distance to closer verb to ProtB located between ProtA and ProtB.
22	BETWEEN_VERB_FARTHEST_DIST_B	Token distance to farthest verb to ProtB located between ProtA and ProtB.
23	ALL_VB_COUNT	Total number of Tokens tagged as VB in sentence.
24 25	ALL_VBD_COUNT	Total number of Tokens tagged as VBD in sentence.
26	ALL_VBG_COUNT	Total number of Tokens tagged as VBG in sentence.
27	ALL_VBN_COUNT	Total number of Tokens tagged as VBN in sentence.
28 29	ALL_VBP_COUNT	Total number of Tokens tagged as VBP in sentence.
30	ALL_VBZ_COUNT	Total number of Tokens tagged as VBZ in sentence.
31	ALL_VERB_MAXSCORE	Higher scoring verb in sentence.
32	ALL_VERB_TOTALSCORE	Sum of verb scores in sentence.
33 34	ALL_VERB_CLOSEST_DIST_A	Token distance to closer verb to ProtA in sentence.
35	ALL_VERB_FARTHEST_DIST_A	Token distance to farthest verb to ProtA in sentence.
36	ALL_VERB_CLOSEST_DIST_B	Token distance to closer verb to ProtB in sentence.
37 38	ALL_VERB_FARTHEST_DIST_B	Token distance to farthest verb to ProtB in sentence.
39	BETWEEN_POS_2AP	Number of Tokens tagged as 2AP between ProtA and ProtB.
40	BETWEEN_POS_1AP	Number of Tokens tagged as 1AP between ProtA and ProtB.
41	BETWEEN_POS_COMMA	Number of Tokens tagged as COMMA between ProtA and ProtB.
42 43	BETWEEN_POS_LRB	Number of Tokens tagged as LRB between ProtA and ProtB.
44	BETWEEN_POS_RRB	Number of Tokens tagged as RRB between ProtA and ProtB.
45	BETWEEN_POS_DOT	Number of Tokens tagged as DOT between ProtA and ProtB.
46 47	BETWEEN_POS_COLON	Number of Tokens tagged as COLON between ProtA and ProtB.
47	BETWEEN_POS_CC	Number of Tokens tagged as CC between ProtA and ProtB.
49	BETWEEN_POS_CD	Number of Tokens tagged as CD between ProtA and ProtB.
50	BETWEEN_POS_DT	Number of Tokens tagged as DT between ProtA and ProtB.
51 52	BETWEEN_POS_EX	Number of Tokens tagged as EX between ProtA and ProtB.
53	BETWEEN_POS_FW	Number of Tokens tagged as FW between ProtA and ProtB.
54	BETWEEN_POS_IN	Number of Tokens tagged as IN between ProtA and ProtB.
55	BETWEEN_POS_JJ	Number of Tokens tagged as JJ between ProtA and ProtB.
56 57	BETWEEN_POS_JJR	Number of Tokens tagged as JJR between ProtA and ProtB.
58	BETWEEN_POS_JJS	Number of Tokens tagged as JJS between ProtA and ProtB.
59	BETWEEN_POS_LS	Number of Tokens tagged as LS between ProtA and ProtB.
60	BETWEEN_POS_MD	Number of Tokens tagged as MD between ProtA and ProtB.

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2	Feature name	Description
3	BETWEEN_POS_NN	Number of Tokens tagged as NN between ProtA and ProtB.
4 5	BETWEEN_POS_NNP	Number of Tokens tagged as NNP between ProtA and ProtB.
6	BETWEEN_POS_NNPS	Number of Tokens tagged as NNPS between ProtA and ProtB.
7	BETWEEN_POS_NNS	Number of Tokens tagged as NNS between ProtA and ProtB.
8	BETWEEN_POS_PDT	Number of Tokens tagged as PDT between ProtA and ProtB.
9 10	BETWEEN_POS_POS	Number of Tokens tagged as POS between ProtA and ProtB.
10	BETWEEN_POS_PRP	Number of Tokens tagged as PRP between ProtA and ProtB.
12	BETWEEN_POS_PRP_DOLAR	Number of Tokens tagged as PRP_DOLAR between ProtA and ProtB.
13	BETWEEN_POS_RB	Number of Tokens tagged as RB between ProtA and ProtB.
14 15	BETWEEN_POS_RBR	Number of Tokens tagged as RBR between ProtA and ProtB.
15	BETWEEN_POS_RBS	Number of Tokens tagged as RBS between ProtA and ProtB.
17	BETWEEN POS_RP	Number of Tokens tagged as RP between ProtA and ProtB.
18	BETWEEN_POS_SYM	
19 20		Number of Tokens tagged as SYM between ProtA and ProtB.
20 21	BETWEEN_POS_TO	Number of Tokens tagged as TO between ProtA and ProtB.
22	BETWEEN_POS_UH	Number of Tokens tagged as UH between ProtA and ProtB.
23	BETWEEN_POS_VB	Number of Tokens tagged as VB between ProtA and ProtB.
24	BETWEEN_POS_VBD	Number of Tokens tagged as VBD between ProtA and ProtB.
25 26	BETWEEN_POS_VBG	Number of Tokens tagged as VBG between ProtA and ProtB.
20	BETWEEN_POS_VBN	Number of Tokens tagged as VBN between ProtA and ProtB.
28	BETWEEN_POS_VBP	Number of Tokens tagged as VBP between ProtA and ProtB.
29	BETWEEN_POS_VBZ	Number of Tokens tagged as VBZ between ProtA and ProtB.
30 31	BETWEEN_POS_WDT	Number of Tokens tagged as WDT between ProtA and ProtB.
32	BETWEEN_POS_WP	Number of Tokens tagged as WP between ProtA and ProtB.
33	BETWEEN_POS_WP_DOLAR	Number of Tokens tagged as WP_DOLAR between ProtA and ProtB.
34	BETWEEN_POS_WRB	Number of Tokens tagged as WRB between ProtA and ProtB.
35 36	ALL_POS_2AP	Number of Tokens tagged as 2AP in sentence.
30 37	ALL_POS_1AP	Number of Tokens tagged as 1AP in sentence.
38	ALL_POS_COMMA	Number of Tokens tagged as COMMA in sentence.
39	ALL_POS_LRB	Number of Tokens tagged as LRB in sentence.
40	ALL_POS_RRB	Number of Tokens tagged as RRB in sentence.
41 42	ALL_POS_DOT	Number of Tokens tagged as DOT in sentence.
43	ALL_POS_COLON	Number of Tokens tagged as COLON in sentence.
44	ALL_POS_CC	Number of Tokens tagged as CC in sentence.
45 46	ALL_POS_CD	Number of Tokens tagged as CD in sentence.
40 47	ALL_POS_DT	Number of Tokens tagged as DT in sentence.
48	ALL_POS_EX	Number of Tokens tagged as EX in sentence.
49	ALL_POS_FW	Number of Tokens tagged as FW in sentence.
50 51	ALL_POS_IN	Number of Tokens tagged as IN in sentence.
52	ALL_POS_JJ	Number of Tokens tagged as JJ in sentence.
53	ALL_POS_JJR	Number of Tokens tagged as JJR in sentence.
54	ALL_POS_JJS	Number of Tokens tagged as JJS in sentence.
55 56	ALL_POS_LS	Number of Tokens tagged as US in sentence.
50 57	ALL_POS_MD	Number of Tokens tagged as MD in sentence.
58	ALL_POS_NN	Number of Tokens tagged as NN in sentence.
59	ALL-POS_NNP	Number of Tokens tagged as NNP in sentence.
60	ALL_POS_NNPS	Number of Tokens tagged as NNPS in sentence.
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Bioinformatics

2	Feature name	Description
3	ALL_POS_NNS	Number of Tokens tagged as NNS in sentence.
4 5	ALL_POS_PDT	Number of Tokens tagged as PDT in sentence.
6	ALL_POS_POS	Number of Tokens tagged as POS in sentence.
7	ALL_POS_PRP	Number of Tokens tagged as PRP in sentence.
8	ALL_POS_PRP_DOLAR	Number of Tokens tagged as PRP_DOLAR in sentence.
9 10	ALL_POS_RB	Number of Tokens tagged as RB in sentence.
10	ALL_POS_RBR	Number of Tokens tagged as RBR in sentence.
12	ALL_POS_RBS	Number of Tokens tagged as RBS in sentence.
13	ALL_POS_RP	Number of Tokens tagged as RP in sentence.
14 15	ALL_POS_SYM	Number of Tokens tagged as SYM in sentence.
16	ALL_POS_TO	Number of Tokens tagged as TO in sentence.
17	ALL_POS_UH	Number of Tokens tagged as UH in sentence.
18	ALL_POS_VB	Number of Tokens tagged as VB in sentence.
19 20	ALL_POS_VBD	Number of Tokens tagged as VBD in sentence.
21	ALL_POS_VBG	Number of Tokens tagged as VBG in sentence.
22	ALL_POS_VBN	Number of Tokens tagged as VBN in sentence.
23	ALL_POS_VBP	Number of Tokens tagged as VBP in sentence.
24 25	ALL_POS_VBZ	Number of Tokens tagged as VBT in sentence.
26	ALL_POS_WDT	Number of Tokens tagged as WDT in sentence.
27	ALL-POS-WP	Number of Tokens tagged as WP in sentence.
28 29	ALL-POS-WP-DOLAR	Number of Tokens tagged as WP_DOLAR in sentence.
30	ALL_POS_WF_DOLAR ALL_POS_WRB	Number of Tokens tagged as WRB in sentence.
31	BETWEEN_PROTA_COUNT	Number of times ProtA appears between ProtA and ProtB.
32	BETWEEN_PROTB_COUNT	Number of times ProtB appears between ProtA and ProtB.
33 34	ALL_PROTA_COUNT	Number of times ProtA appears in sentence.
35	ALL_PROTB_COUNT	Number of times ProtB appears in sentence.
36	KEYWORD_COUNT_acetylate	Number of times 'acetylate' appears between ProtA and ProtB.
37	KEYWORD_COUNT_activate	Number of times 'activate' appears between ProtA and ProtB.
38 39	KEYWORD_COUNT_activate	Number of times 'acylate' appears between ProtA and ProtB.
40	KEYWORD_COUNT_amidate	Number of times 'amidate' appears between ProtA and ProtB.
41	KEYWORD_COUNT_amdate	Number of times 'assemble' appears between ProtA and ProtB.
42 43		
45 44	KEYWORD_COUNT_attach	Number of times 'attach' appears between ProtA and ProtB.
45	KEYWORD_COUNT_bind	Number of times 'bind' appears between ProtA and ProtB.
46	KEYWORD_COUNT_biotinylate	Number of times 'biotinylate' appears between ProtA and ProtB.
47 48	KEYWORD_COUNT_block	Number of times 'block' appears between ProtA and ProtB.
49	KEYWORD_COUNT_brominate	Number of times 'brominate' appears between ProtA and ProtB.
50	KEYWORD_COUNT_carboxylate	Number of times 'carboxylate' appears between ProtA and ProtB.
51	KEYWORD_COUNT_catalyze	Number of times 'catalyze' appears between ProtA and ProtB.
52 53	KEYWORD_COUNT_cleave	Number of times 'cleave' appears between ProtA and ProtB.
55	KEYWORD_COUNT_complex	Number of times 'complex' appears between ProtA and ProtB.
55	KEYWORD_COUNT_conjugate	Number of times 'conjugate' appears between ProtA and ProtB.
56	KEYWORD_COUNT_contact	Number of times 'contact' appears between ProtA and ProtB.
57 58	KEYWORD_COUNT_couple	Number of times 'couple' appears between ProtA and ProtB.
59	KEYWORD_COUNT_cysteinylate	Number of times 'cysteinylate' appears between ProtA and ProtB.
60	KEYWORD_COUNT_demethylate	Number of times 'demethylate' appears between ProtA and ProtB.
-	KEYWORD_COUNT_dephosphorylate	Number of times 'dephosphorylate' appears between ProtA and ProtB.

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Feature name Description

KEYWORD_COUNT_dimerise KEYWORD_COUNT_dimerize EYWORD_COUNT_disassemble KEYWORD_COUNT_discharge KEYWORD_COUNT_dissociate YWORD_COUNT_down-regulate YWORD_COUNT_downregulate KEYWORD_COUNT_farmesylate KEYWORD_COUNT_formylate **KEYWORD_COUNT_hydroxilate** EYWORD_COUNT_hydroxylate KEYWORD_COUNT_inactivate KEYWORD_COUNT_induce KEYWORD_COUNT_inhibit KEYWORD_COUNT_interact KEYWORD_COUNT_mediate KEYWORD_COUNT_methylate KEYWORD_COUNT_modify $KEYWORD_COUNT_modulate$ EYWORD_COUNT_multimerise EYWORD_COUNT_multimerize EYWORD_COUNT_myristoylate KEYWORD_COUNT_myristylate KEYWORD_COUNT_nitrosylate **KEYWORD_COUNT_overexpress** EYWORD_COUNT_palmitoylate KEYWORD_COUNT_palmitylate YWORD_COUNT_phosphorylate KEYWORD_COUNT_precipitate KEYWORD_COUNT_promote KEYWORD_COUNT_pyruvate KEYWORD_COUNT_regulate KEYWORD_COUNT_repress KEYWORD_COUNT_stimulate KEYWORD_COUNT_substitute KEYWORD_COUNT_sumoylate KEYWORD_COUNT_suppress EYWORD_COUNT_transactivate EYWORD_COUNT_ubiquitinate YWORD_COUNT_ubiquitinylate KEYWORD_COUNT_up-regulate KEYWORD_COUNT_upregulate

Number of times 'dimerise' appears between ProtA and ProtB. Number of times 'dimerize' appears between ProtA and ProtB. Number of times 'disassemble' appears between ProtA and ProtB. Number of times 'discharge' appears between ProtA and ProtB. Number of times 'dissociate' appears between ProtA and ProtB. Number of times 'down-regulate' appears between ProtA and ProtB. Number of times 'downregulate' appears between ProtA and ProtB. Number of times 'farnesylate' appears between ProtA and ProtB. Number of times 'formylate' appears between ProtA and ProtB. Number of times 'hydroxilate' appears between ProtA and ProtB. Number of times 'hydroxylate' appears between ProtA and ProtB. Number of times 'inactivate' appears between ProtA and ProtB. Number of times 'induce' appears between ProtA and ProtB. Number of times 'inhibit' appears between ProtA and ProtB. Number of times 'interact' appears between ProtA and ProtB. Number of times 'mediate' appears between ProtA and ProtB. Number of times 'methylate' appears between ProtA and ProtB. Number of times 'modify' appears between ProtA and ProtB. Number of times 'modulate' appears between ProtA and ProtB. Number of times 'multimerise' appears between ProtA and ProtB. Number of times 'multimerize' appears between ProtA and ProtB. Number of times 'myristoylate' appears between ProtA and ProtB. Number of times 'myristylate' appears between ProtA and ProtB. Number of times 'nitrosylate' appears between ProtA and ProtB. Number of times 'overexpress' appears between ProtA and ProtB. Number of times 'palmitoylate' appears between ProtA and ProtB. Number of times 'palmitylate' appears between ProtA and ProtB. Number of times 'phosphorylate' appears between ProtA and ProtB. Number of times 'precipitate' appears between ProtA and ProtB. Number of times 'promote' appears between ProtA and ProtB. Number of times 'pyruvate' appears between ProtA and ProtB. Number of times 'regulate' appears between ProtA and ProtB. Number of times 'repress' appears between ProtA and ProtB. Number of times 'stimulate' appears between ProtA and ProtB. Number of times 'substitute' appears between ProtA and ProtB. Number of times 'sumoylate' appears between ProtA and ProtB. Number of times 'suppress' appears between ProtA and ProtB. Number of times 'transactivate' appears between ProtA and ProtB. Number of times 'ubiquitinate' appears between ProtA and ProtB. Number of times 'ubiquitinylate' appears between ProtA and ProtB. Number of times 'up-regulate' appears between ProtA and ProtB. Number of times 'upregulate' appears between ProtA and ProtB.