

Metadata of the chapter that will be visualized online

Chapter Title	Mitochondrial Gymnastics in Retinal Cells: A Resilience Mechanism Against Oxidative Stress and Neurodegeneration	
Copyright Year	2019	
Copyright Holder	Springer Nature Switzerland AG	
Corresponding Author	Family Name	Mirra
	Particle	
	Given Name	Serena
	Suffix	
	Division	Departament de Genètica, Microbiologia i Estadística, Facultat de Biologia
	Organization/University	Universitat de Barcelona
	Address	Barcelona, Spain
	Division	Centro de Investigación Biomédica en Red de Enfermedades Raras (CIBERER)
	Organization/University	Instituto de Salud Carlos III
	Address	Barcelona, Spain
	Email	serena.mirra@ub.edu
Corresponding Author	Family Name	Marfany
	Particle	
	Given Name	Gemma
	Suffix	
	Division	Departament de Genètica, Microbiologia i Estadística, Facultat de Biologia
	Organization/University	Universitat de Barcelona
	Address	Barcelona, Spain
	Division	Centro de Investigación Biomédica en Red de Enfermedades Raras (CIBERER)
	Organization/University	Instituto de Salud Carlos III
	Address	Barcelona, Spain
	Division	

Organization/University

Institut de Biomedicina de la
Universitat de Barcelona, IBUB-
IRSJD

Address

Barcelona, Spain

Email

gmarfany@ub.edu

Abstract

Inherited retinal dystrophies (IRDs) are a broad group of neurodegenerative disorders associated with reduced or deteriorating visual system. In the retina, cells are under constant oxidative stress, leading to elevated reactive oxygen species (ROS) generation that induces mitochondrial dysfunction and alteration of the mitochondrial network. This mitochondrial dysfunction combined with mutations in mitochondrial DNA and nuclear genes makes photoreceptors and retinal ganglion cells more susceptible to cell death. In this minireview, we focus on mitochondrial dynamics and their contribution to neuronal degeneration underlying IRDs, with particular attention to Leber hereditary optic neuropathy (LHON) and autosomal dominant optic atrophy (DOA), and propose targeting cell resilience and mitochondrial dynamics modulators as potential therapeutic approaches for retinal disorders.

Keywords (separated
by “ - ”)

Retinal dystrophies - Oxidative stress - Mitochondrial dynamics - Retinopathies
- Neurodegeneration

Mitochondrial Gymnastics in Retinal Cells: A Resilience Mechanism Against Oxidative Stress and Neurodegeneration

Serena Mirra and Gemma Marfany

Abstract

Inherited retinal dystrophies (IRDs) are a broad group of neurodegenerative disorders associated with reduced or deteriorating visual system. In the retina, cells are under constant oxidative stress, leading to elevated reactive oxygen species (ROS) generation that induces mitochondrial dysfunction and alteration of the mitochondrial network. This mitochondrial dysfunction combined with mutations in mitochondrial DNA and nuclear genes makes photoreceptors and retinal ganglion cells more susceptible to cell death. In this minireview, we focus on mitochondrial dynamics and their

contribution to neuronal degeneration underlying IRDs, with particular attention to Leber hereditary optic neuropathy (LHON) and autosomal dominant optic atrophy (DOA), and propose targeting cell resilience and mitochondrial dynamics modulators as potential therapeutic approaches for retinal disorders.

Keywords

Retinal dystrophies · Oxidative stress · Mitochondrial dynamics · Retinopathies · Neurodegeneration

84.1 IRDs and Mitochondria

IRDs are a large group of diseases characterized by retinal cell degeneration and death. Among retinal cells, photoreceptors and ganglion cells receive the stressful impact of light photons and excess intraocular pressure; thus resilience mechanisms have to be switched on to promote cell survival. Correct mitochondrial metabolism and dynamics are essential for retinal cells and mutations in either mtDNA or in nuclear genes involved in mitochondrial function having a high impact on cell survival. Since progressive attrition of photoreceptors and retinal ganglion cells leads to blindness, the correct preservation of mitochondrial function and dynamics is essential for retinal homeostasis.

S. Mirra (✉)

Departament de Genètica, Microbiologia
i Estadística, Facultat de Biologia, Universitat de
Barcelona, Barcelona, Spain

Centro de Investigación Biomédica en Red de
Enfermedades Raras (CIBERER), Instituto de Salud
Carlos III, Barcelona, Spain
e-mail: serena.mirra@ub.edu

G. Marfany (✉)

Departament de Genètica, Microbiologia
i Estadística, Facultat de Biologia, Universitat de
Barcelona, Barcelona, Spain

Centro de Investigación Biomédica en Red de
Enfermedades Raras (CIBERER), Instituto de Salud
Carlos III, Barcelona, Spain

Institut de Biomedicina de la Universitat de
Barcelona, IBUB-IRSJD, Barcelona, Spain
e-mail: gmarfany@ub.edu

AUT

47 Mitochondria are essential organelles that
 48 supply energy to the cell through oxidative phos-
 49 phosphorylation (OXPHOS) and are also essential in
 50 calcium buffering, cell cycle control and regula-
 51 tion of apoptosis. Mitochondrial activity gener-
 52 ates 1–5% ROS in physiological conditions
 53 (Nissanka and Moraes 2018). Severe alterations
 54 in mitochondrial function due to physiological
 55 and environmental cues generate loss of mito-
 56 chondrial membrane potential ($\Delta\Psi_m$), decreased
 57 OXPHOS, mitochondrial DNA (mtDNA) dam-
 58 age and ROS-induced ROS vicious circle (Bae
 59 et al. 2011). These changes are associated to
 60 remodelling of the mitochondrial network, and
 61 these mitochondrial dynamics include fusion, fis-
 62 sion, transport, interorganellar communication
 63 and mitochondrial quality control (Cid-Castro
 64 et al. 2018). Although numerous studies have
 65 implicated ROS in neuronal death, their role in
 66 the pathophysiology of optical neuropathies
 67 remains to be further investigated. Leber heredi-
 68 tary optic neuropathy (LHON) and dominant
 69 optic atrophy (DOA) are two prototypic inherited
 70 ocular disorders related to mitochondrial dys-
 71 function. Despite their contrasting genetic basis,
 72 they share overlapping pathological features due
 73 to the particular vulnerability of retinal ganglion
 74 cells (RGCs) to perturbed mitochondrial
 75 function.

76 84.2 Mitochondrial Localization 77 within the Retina

78 The retina is among the most metabolically active
 79 tissues in the body due to its high oxygen demand.
 80 Allocation of mitochondria in high energy
 81 requirements regions is essential for cellular
 82 function. In the outermost layer of the vertebrate
 83 retina, rod and cone photoreceptors display
 84 highly differentiated outer segments, packed with
 85 membranous disks/folds where photoreception
 86 and the phototransduction cascade occur. The
 87 most distal tips of rod and cone outer segments
 88 closely interact with the retinal pigment epithe-
 89 lium (RPE), which continually supports photore-
 90 ceptor function by endorsing the outer segments
 91 feeding and renewal. To meet the high demand of

energy, photoreceptor cells display a high con- 92
 centration of mitochondria in the outer part of the 93
 inner segment, whereas in RPE cells mitochon- 94
 dria are located at the basal region. Significantly, 95
 in the mammalian inner retina, mitochondria are 96
 particularly concentrated in the unmyelinated 97
 proximal axons of RGC compared to myelinated 98
 segment of the optic nerve. This particular distri- 99
 bution is required to guarantee the energy supply 100
 for the generation of an action potential that con- 101
 tinuously propagates along these axonal regions. 102
 This high demand of energy together with a com- 103
 plex dendritic arborization underlies RGC sus- 104
 ceptibility to respiratory chain dysfunction, 105
 oxidative stress and, eventually, apoptosis (Ito 106
 and Di Polo 2017). 107

108 84.3 Mitochondrial Dynamics: 109 Fusion and Fission

Mitochondrial fusion requires the apposition of 110
 two adjacent organelles, followed by outer and 111
 inner membrane fusion. Fusion is mediated by 112
 the conserved dynamin-related GTP proteins 113
 mitofusins (Mfn1 and Mfn2) and the optic domi- 114
 nant atrophy 1 protein (OPA1). Mfns are distrib- 115
 uted evenly on the outer mitochondrial membrane. 116
 Loss of Mfns impairs mitochondrial fusion, and 117
 consequently, mitochondrial length is reduced. 118
 OPA1 is anchored to the mitochondrial inner 119
 membrane and interacts with Mfns to form pro- 120
 tein complexes that couple the fusion between 121
 the outer and inner membranes. *Opa1*-deficient 122
 mice display RGC mitochondrial fragmentation, 123
 dendritic atrophy prior to visual impairment and 124
 neuronal loss (Williams et al. 2010). 125
 Mitochondrial elongation confers resistance to 126
 apoptotic stimuli, and a network of fused mito- 127
 chondria has been described in many senescent 128
 postmitotic cell types. How does fusion protect 129
 mitochondrial function? Mitochondrial fusion 130
 contributes to mitochondrial homeostasis by 131
 enabling the exchange of mtDNA, substrates, 132
 metabolites or specific lipids contents between 133
 organelles (Hoitzing et al. 2015). 134

Mitochondrial fission instead is mediated by 135
 the adaptor FIS1 protein and the dynamin-related 136

GTPase DRP1. FIS1 is spread diffusely throughout the outer mitochondrial membrane and recruits DRP1 from the cytosol. DRP1 assembles into spiral filaments around mitochondria tubules, constricting mitochondria through conformational changes. Interestingly, the tubules of the endoplasmic reticulum (ER) wrap around and squeeze mitochondria at the early stage of division, facilitating DRP1 recruitment to complete fission (Friedman et al. 2011). Of note, an excess of mitochondrial fission can represent the first step of apoptosis. Following extensive cellular stress or damage, the pro-apoptotic Bcl-2 family member Bax translocates to mitochondria and accumulates in concentrated foci that colocalize with DRP1 and Mfns. This process mediates pore formation in outer mitochondrial membranes, which facilitates the release of cytochrome C from mitochondria and downstream caspase activation.

In vivo models of retinal detachment showed that DRP1 activation can be induced by exogenous ROS, triggering mitochondrial fission previous to apoptotic cascade activation. Moreover, DRP1 inhibition results in a neuroprotective effect by suppressing mitochondrial fission and apoptosis (She et al. 2018). Consistently, mitochondrial stress induces mitochondrial fragmentation by increasing DRP1 in the retina of glaucomatous D2 mice and in cultured RGC in vitro. And increase in OPA1 expression and DRP1 inhibition blocks mitochondrial fission, with a subsequent reduction of oxidative stress and an increase of RGC survival (Ju et al. 2010; Kim et al. 2015).

84.4 Mitophagy: Mitochondria as the Main Course

Mitochondria have multiple quality control mechanisms to ensure mitochondrial integrity, and alterations in this quality control have been extensively associated to neurodegenerative diseases (Pickles et al. 2018). Fission sequesters irreversibly damaged or fusion-incompetent mitochondria and results in their subsequent elimination by mitophagy, the autophagy-

mediated degradation of mitochondria (Shutt et al. 2012). The best-studied mitophagy pathway involves PINK1 and Parkin, genes associated to rare genetic forms of Parkinson's disease (McWilliams and Muqit 2017). When mitochondria lose their membrane potential, the mitochondrial protein PINK1 recruits the E3 ubiquitin ligase Parkin from the cytosol to dysfunctional mitochondria, where it ubiquitinates mitochondrial proteins for proteasomal degradation and promotes the engulfment of mitochondria by autophagosomes (Lazarou et al. 2015). These studies on PINK1/Parkin-dependent mitophagy were performed in vitro, and contribution of this pathway in vivo is yet to be determined.

Parkin is widely expressed in the murine retina, particularly in the RGCs (Esteve-Rudd et al. 2010). Conversely, PINK1 protein expression in the whole retina is very low, suggesting that retinal basal mitophagy occurs independently of PINK1 (McWilliams et al. 2018). Parkin overexpression stabilizes mitochondrial membrane potential and decreases glutamate cytotoxicity and apoptosis in RGCs (Hu et al. 2017). Also, Parkin expression is upregulated in murine model of hypertensive glaucoma, where its overexpression partially restores dysfunction of mitophagy in RGCs (Dai et al. 2018). All together these data point out the protective role of Parkin-mediated mitophagy against several damages potentially leading to optic neurodegeneration.

84.5 Mitochondrial Transport

As mentioned, mitochondrial transport and distribution to regions with high energy demand is crucial in neurons. The kinesin superfamily proteins (KIFs) and cytoplasmic dynein are the main motor proteins that transport mitochondria towards the microtubule positive and negative ends, respectively. In mammals, kinesin contacts the molecular adaptors Trak1 and Trak2, which in turn bind the GTPases Miro1 and Miro2 (Lopez-Domenech et al. 2018). Miro and Trak proteins interact with the machinery involved in fusion and fission, thus connecting mitochondrial dynamics and trafficking processes. Moreover,

227 the PINK/Parkin complex targets Miro for degra- 272
 228 dation, and as a consequence, kinesin is released 273
 229 from mitochondria, thus leading to a redistribu- 274
 230 tion of damaged mitochondria (Wang et al. 2011). 275

231 In contrast to mature dendrites, mitochondria 276
 232 are highly dynamic in RGC axons. In early stages 277
 233 of a glaucoma mice model, the number of trans- 278
 234 ported mitochondria in RGC decreased, and 279
 235 axons were devoid of mitochondria before RGC 280
 236 death (Takahara et al. 2015). Several evidences 281
 237 highlight the importance of cytoskeleton integ- 282
 238 rity for the correct mitochondrial motility along 283
 239 RGC processes (Tang 2018). Moreover, new 284
 240 players regulating mitochondrial trafficking in 285
 241 neurons have been recently described. Among 286
 242 them, the Miro-interacting mitochondrial protein 287
 243 *Armcx1* enhanced mitochondrial transport in 288
 244 adult RGC and promoted axonal regeneration 289
 245 after injury (Cartoni et al. 2016). Nonetheless, 290
 246 the molecular role of motor and adaptor proteins 291
 247 mediating mitochondrial transport in RGC 292
 248 remains to be further elucidated. 293

249 84.6 Mitochondrial Optic 294 250 Neuropathies 295

251 Primary mitochondrial disorders (PMD) are 296
 252 associated to pathogenic mtDNA or nuclear gene 297
 253 mutations, whereas secondary mitochondrial dis- 298
 254 orders (SMD) are mainly due to nongenetic 299
 255 causes, e.g. environmental factors or pharmaco- 300
 256 logical toxins. In Leber hereditary optic neurop- 301
 257 athy (LHON), several mtDNA mutations lead to 302
 258 dysfunction in the mitochondrial complex I, 303
 259 causing accumulation of ROS and cell death in 304
 260 the RGC cells. About 90% of LHON cases are 305
 261 caused by point mutation in *MT-ND1*, *MT-ND4*, 306
 262 *MT-ND4L* or *MT-ND6* genes (Kim et al. 2018), 307
 263 but it remains unclear how these genetic altera- 308
 264 tions lead to the specific features of LHON. 309

265 DOA is an autosomal dominant PMD charac- 310
 266 terized by progressive blindness with degenera- 311
 267 tion of RGC and the optic nerve, with a prevalence 312
 268 of 1:35,000 people worldwide. Approximately 313
 269 50–60% of DOA patients carry mutations in the 314
 270 nuclear *OPA1* gene, which regulates mitochon- 315
 271 dria fusion and OXPHOS and is involved in cell 316

death and mtDNA maintenance. A different set of 272
 OPA1 mutations causes “DOA-plus” phenotypes 273
 with mtDNA instability, deafness and movement 274
 disorders in addition to traditional DOA symp- 275
 toms (Pilz et al. 2017). 276

Other mitochondrial dysfunction syndromes 277
 with marked optic neuropathy are Charcot- 278
 Marie-Tooth disease (when caused by *MFN2* 279
 mutations), Friedreich Ataxia and Costeff syn- 280
 drome, although the mechanisms that trigger 281
 RGC death in the two latter are far from under- 282
 stood (Carelli et al. 2017). 283

84.7 Future Perspectives 284

Several therapeutic strategies have been tested 285
 over the years to prevent mitochondrial 286
 dysfunction-related neurodegeneration. 287
 Modification of ROS production or inhibition of 288
 caspase apoptotic pathway has been both 289
 employed in clinical trials, but these strategies 290
 failed to prevent neurodegeneration. 291

Targeting mitochondrial dynamics means to 292
 intervene between the triggering event (ROS gen- 293
 eration) and the terminal phase (cell death); thus 294
 it may represent an effective approach to prevent 295
 progressive degeneration. Screening compounds 296
 targeting the mitochondrial fission/fusion 297
 machinery and the mitochondrial quality control 298
 system would impact in the recovery of a 299
 “healthy” mitochondrial network and, as a conse- 300
 quence, improve the neurological phenotype. 301
 However, until now, no novel therapeutic strategy 302
 specifically targeting mitochondrial dynamics 303
 has been developed. 304

On the other hand, retinal cells deploy several 305
 pathways to deal with oxidative stress, which are 306
 most likely interconnected. Studying IRD genes 307
 that encode key protein sensors and modulators 308
 that play a role in the crosstalk between thefor- 309
 mation of lipid droplets (e.g. *MTTP*, *TTPA*, 310
CLN3, *PNPLA6*), mRNA stress granules (e.g. 311
CERKL), mitochondrial dynamics (e.g. *MFN2*, 312
SLC25A46, *OPA3*, *OPA8*) and autophagy (e.g. 313
DRAM2) will accrue knowledge on survival ver- 314
 sus apoptosis fate decisions in retinal cells and 315
 may offer new scenarios for therapeutic targets. 316

317 We propose that future drug and gene therapies
318 addressed to reduce mitochondrial fission,
319 increase mitochondrial fusion and favour other
320 resilience cell mechanisms would favour retinal
321 cell survival, preventing or halting the progres-
322 sion of the retinal degeneration.

323 References

324 Bae YS, Oh H, Rhee SG et al (2011) Regulation of reac-
325 tive oxygen species generation in cell signaling. *Mol*
326 *Cells* 32:491–509

327 Carelli V, La Morgia C, Ross-Cisneros FN et al (2017)
328 Optic neuropathies: the tip of the neurodegeneration
329 iceberg. *Hum Mol Genet* 26(R2):R139–R150

330 Cartoni R, Norsworthy MW, Bei F et al (2016) The
331 mammalian-specific protein Armcx1 regulates mito-
332 chondrial transport during axon regeneration. *Neuron*
333 92:1294–1307

334 Cid-Castro C, Hernandez-Espinosa DR, Moran J (2018)
335 ROS as regulators of mitochondrial dynamics in neu-
336 rons. *Cell Mol Neurobiol* 38:995–1007

337 Dai Y, Hu X, Sun X (2018) Overexpression of parkin pro-
338 tects retinal ganglion cells in experimental glaucoma.
339 *Cell Death Dis* 9:88

340 Esteve-Rudd J, Campello L, Herrero MT et al (2010)
341 Expression in the mammalian retina of parkin and
342 UCH-L1, two components of the ubiquitin-proteasome
343 system. *Brain Res* 1352:70–82

344 Friedman JR, Lackner LL, West M et al (2011) ER
345 tubules mark sites of mitochondrial division. *Science*
346 334:358–362

347 Hoitzing H, Johnston IG, Jones NS (2015) What is the
348 function of mitochondrial networks? A theoretical
349 assessment of hypotheses and proposal for future
350 research. *BioEssays* 37:687–700

351 Hu X, Dai Y, Sun X (2017) Parkin overexpression protects
352 retinal ganglion cells against glutamate excitotoxicity.
353 *Mol Vis* 23:447–456

354 Ito YA, Di Polo A (2017) Mitochondrial dynamics,
355 transport, and quality control: a bottleneck for reti-
356 nal ganglion cell viability in optic neuropathies.
357 *Mitochondrion* 36:186–192

358 Ju WK, Kim KY, Duong-Polk KX et al (2010) Increased
359 optic atrophy type 1 expression protects retinal gan-
360 glion cells in a mouse model of glaucoma. *Mol Vis*
361 16:1331–1342

Kim KY, Perkins GA, Shim MS et al (2015) DRP1 inhibi- 362
tion rescues retinal ganglion cells and their axons by 363
preserving mitochondrial integrity in a mouse model 364
of glaucoma. *Cell Death Dis* 6:e1839 365

Kim US, Jurkute N, Yu-Wai-Man P (2018) Leber heredi- 366
tary optic neuropathy-light at the end of the tunnel? 367
Asia Pac J Ophthalmol (Phila) 7(4):242–245 368

Lazarou M, Sliter DA, Kane LA et al (2015) The ubiquitin 369
kinase PINK1 recruits autophagy receptors to induce 370
mitophagy. *Nature* 524:309–314 371

Lopez-Domenech G, Covill-Cooke C, Ivankovic D et al 372
(2018) Miro proteins coordinate microtubule- and 373
actin-dependent mitochondrial transport and distribu- 374
tion. *EMBO J* 37:321–336 375

McWilliams TG, Muqit MM (2017) PINK1 and Parkin: 376
emerging themes in mitochondrial homeostasis. *Curr*
377 *Opin Cell Biol* 45:83–91 378

McWilliams TG, Prescott AR, Montava-Garriga L et al 379
(2018) Basal mitophagy occurs independently of 380
PINK1 in mouse tissues of high metabolic demand. 381
Cell Metab 27:439–449. e435 382

Nissanka N, Moraes CT (2018) Mitochondrial DNA dam- 383
age and reactive oxygen species in neurodegenerative 384
disease. *FEBS Lett* 592:728–742 385

Pickles S, Vigie P, Youle RJ (2018) Mitophagy and qual- 386
ity control mechanisms in mitochondrial maintenance. 387
Curr Biol 28:R170–r185 388

Pilz YL, Bass SJ, Sherman J (2017) A review of the 389
mitochondrial optic neuropathies: from inherited to 390
acquired forms. *J Optom* 10:205–214 391

She X, Lu X, Li T et al (2018) Inhibition of mitochondrial 392
fission preserves photoreceptors after retinal detach- 393
ment. *Am J Pathol* 188:1713–1722 394

Shutt T, Geoffrion M, Milne R et al (2012) The intracel- 395
lular redox state is a core determinant of mitochondrial 396
fusion. *EMBO Rep* 13:909–915 397

Takahara Y, Inatani M, Eto K et al (2015) In vivo imag- 398
ing of axonal transport of mitochondria in the diseased 399
and aged mammalian CNS. *Proc Natl Acad Sci U S A*
400 112:10515–10520 401

Tang BL (2018) Miro-working beyond mitochondria and 402
microtubules. *Cell* 7:18 403

Vives-Bauza C, Przedborski S (2010) PINK1 points 404
Parkin to mitochondria. *Autophagy* 6:674–675 405

Wang X, Winter D, Ashrafi G et al (2011) PINK1 and 406
Parkin target Miro for phosphorylation and degrada- 407
tion to arrest mitochondrial motility. *Cell* 147:893–906 408

Williams PA, Morgan JE, Votruba M (2010) Opa1 defi- 409
ciency in a mouse model of dominant optic atrophy 410
leads to retinal ganglion cell dendropathy. *Brain*
411 133:2942–2951 412

Author Queries

Chapter No.: 84 459489_1_En_84_Chapter

Queries	Details Required	Author's Response
AU1	Please confirm the affiliation details for both authors.	
AU2	Reference "Vives-Bauza and Przedborski (2010)" was not cited anywhere in the text. Please provide in text citation or delete the reference from the reference list.	

Uncorrected Proof