

2 **ABSTRACT**

3 **Introduction:** *Streptococcus agalactiae* or group B streptococci (GBS) are the main
4 etiologic agent of early neonatal sepsis in developed countries. This microorganism
5 belongs to the gastrointestinal tract microbiota from where it can colonize the vagina and
6 be vertically transmitted to child before or at birth and subsequently cause infection in the
7 newborn. Approximately 50% of newborns from pregnant women harboring GBS
8 become colonized, with 1-2% developing early neonatal infection if no preventive
9 intervention is performed. **The aim of this study was to characterize and compare the**
10 **serotype, virulence factors and antimicrobial resistance of GBS isolates from**
11 **pregnant women and newborns from several hospitals in Catalonia.**

12 **Methods:** We analyzed 242 GBS strains including 95 colonizers, 68 pathogenic strains
13 isolated from pregnant women and 79 strains isolated from neonates **with sepsis** to
14 determine serotype, virulence and antimicrobial resistance.

15 **Results:** Distribution of the serotypes was different among the three groups, being
16 serotypes Ia and II significantly more frequent among colonizing strains ($p=0.001$ and
17 0.012 , respectively). The virulence factors *bca* and *scpB* were significantly more frequent
18 among neonatal strains than pathogenic or colonizing strains ($p=0.0001$ and 0.002 ,
19 respectively). Pathogenic strains were significantly more resistant to erythromycin,
20 clindamycin and azithromycin than the non-pathogenic counterparts.

21 **Conclusions:** Taking into account that neonatal sepsis is an important problem
22 worldwide, surveillance of the epidemiology, antimicrobial resistance and virulence of
23 **GBS at local level** could provide great knowledge about these microorganisms and help

24 to improve treatment **and to prevent the neonatal invasive infection caused by this**
25 **microorganism.**

26

27 **RESUMEN**

28 **Introducción:** *Streptococcus agalactiae* o estreptococos del grupo B (SGB) es el
29 principal agente etiológico de la sepsis neonatal temprana en los países desarrollados.
30 Este microorganismo pertenece a la microbiota del tracto gastrointestinal desde donde
31 puede colonizar la vagina y ser transmitido verticalmente al niño antes o al nacer y
32 posteriormente causar infección en el recién nacido. Aproximadamente el 50% de los
33 recién nacidos de mujeres embarazadas que albergan GBS se colonizan, con 1-2%
34 desarrollando infección neonatal temprana si no se realiza intervención preventiva. **El**
35 **objetivo de este estudio fue caracterizar y comparar serotipos, factores de virulencia**
36 **y la resistencia a los antimicrobianos de aislamientos de SGB de mujeres**
37 **embarazadas y neonatos procedentes de varios hospitales de Cataluña.**

38 **Métodos:** Se analizaron 242 cepas de GBS incluyendo 95 colonizadoras, 68 cepas
39 patógenas aisladas de mujeres embarazadas y 79 cepas aisladas de neonatos **con sepsis**
40 para determinar serotipo, virulencia y resistencia antimicrobiana.

41 **Resultados:** La distribución de los serotipos fue diferente entre los tres grupos, siendo
42 los serotipos Ia y II significativamente más frecuentes entre las cepas colonizadoras ($p =$
43 $0,001$ y $0,012$, respectivamente). Los factores de virulencia *bca* y *scpB* fueron
44 significativamente más frecuentes entre las cepas neonatales que entre las patógenas o
45 colonizadoras ($p = 0,0001$ y $0,002$, respectivamente). Las cepas patógenas fueron
46 significativamente más resistentes a eritromicina, clindamicina y azitromicina que las no
47 patógenas. **Conclusiones:** Teniendo en cuenta que la sepsis neonatal es un problema
48 importante a nivel mundial, la vigilancia de la epidemiología, la resistencia a los

49 antimicrobianos y la virulencia del SGB **a nivel local** podría proporcionar un gran
50 conocimiento de estos microorganismos y ayudar a mejorar el tratamiento **y la**
51 **prevención de la infección invasiva causada por este microorganismo.**

52 **Keywords:** *Streptococcus agalactiae*, virulence, serotypes, antimicrobial resistance.

53

54 INTRODUCTION

55 *Streptococcus agalactiae* (group B streptococcus [GBS]) is an opportunistic pathogen that
56 colonizes the gastrointestinal and genitourinary tract of healthy people and is responsible
57 for severe diseases in susceptible hosts¹. Moreover, this specie is a leading cause of
58 invasive infection in newborns, causing early neonatal sepsis in developed countries. In
59 Spain, it has been reported that 10-18.5% of pregnant women are colonized by GBS².

60 Colonization of the vagina by GBS in pregnant women is of great importance because it
61 increases the risk of infection, due to it can be vertically transmitted to the child before or
62 at birth, causing infection in the newborn. Infection by GBS may arise from a prematurely
63 ruptured amniotic membrane which becomes infected or from infected amniotic fluid
64 (chorioamnionitis) or preterm delivery in a mother colonized by these bacteria, presenting
65 a much higher risk of infecting the offspring due to immaturity of the immune system³.

66 Approximately 50% of infants from GBS-colonized women become colonized, with 1-
67 2% of these newborns developing early or late neonatal infection if no preventive
68 intervention is performed⁴. To avoid this enhanced risk of vertical transmission, the Center
69 of Disease Control (CDC-1996) recommended the screening of all pregnant women at 35-
70 37 weeks of gestation to determine possible GBS colonization⁵. Penicillin and ampicillin
71 are the antibiotics of choice for treatment or intrapartum prophylaxis of GBS, followed by

72 first-generation cephalosporins. In the case of patients allergic to β -lactams, clindamycin,
73 erythromycin and vancomycin are the antimicrobials of choice^{6, 7}.

74 When implemented, the use of these prophylactic measures results in a decrease in the
75 incidence of infection by GBS⁸. In Spain, an example of the success of these prophylactic
76 measures was demonstrated in a study carried out in 10 hospitals in Barcelona (Spain). It
77 was found that the incidence of GBS as a cause of neonatal sepsis was reduced from
78 1.92/1,000 newborns in 1994 to 0.26/1,000 newborns in 2001 ($p < 0.001$)².

79 Ten GBS serotypes have been classified (Ia, Ib and II to IX), according to recognized
80 capsular polysaccharides that are considered to be a major virulence factor in invasive
81 disease caused by this microorganism¹⁰. Serotypes Ia, Ib, III and V are the most frequently
82 involved in invasive infection¹¹. Additionally, GBS can develop different pathogenic
83 mechanisms that allow colonization and invasion in different niches within the host being
84 able to avoid and suppress the host immune response.

85 The aim of this study was to characterize and compare the serotype, virulence factors and
86 antimicrobial resistance of GBS **isolates** from pregnant women and newborns from
87 several hospitals in Catalonia.

88

89 **MATERIALS AND METHODS**

90 **Clinical sample and bacterial isolates.**

91 Two hundred forty-two *S. agalactiae* **isolates** were collected **from 242 patients** from 5
92 Catalan hospitals (Hospital Clínic, Vall d'Hebron, San Joan de Déu, Parc Taulí and
93 CatLab) from 2010 to 2016. Among these **isolates**, 95 were obtained from vaginal, rectal-
94 vaginal and endocervical swabs from asymptomatic pregnant women at 35 to 37 weeks
95 of gestation and categorized as colonizing; 68 were obtained from urine, blood, amniotic

96 fluid, and endometrial and placental samples from symptomatic women and categorized
97 as pathogenic **isolates**; and 79 **isolates** were isolated from blood, cerebrospinal and
98 amniotic fluid samples of newborns with neonatal sepsis. **Samples arriving to the**
99 **different laboratories of Microbiology were cultured in Granada agar and selective**
100 **Todd-Hewitt broth specific for GBS isolation.** Suspected colonies were confirmed by
101 MALDI-TOF.

102 **Serotyping**

103 Determination of the capsular type or serotype was carried out by a multiplex-PCR using
104 a set of primers described previously⁴.

105 **Virulence profile**

106 Different virulence determinants, including *bac* (encoding alpha subunit of the C
107 protein), *bca* (encoding beta subunit of the C protein), *cyl(E)* (encoding cytolysin-
108 haemolysin), *rib* (Alp family protein Rib), *hyl(B)* (encoding hyaluronidase protein),
109 *pep(B)* (encoding oligopeptidase protein), *scp(B)* (encoding an invasin), *fbs(B)* (encoding
110 fibrinogen-binding protein mediating invasivity), *spb(1)* (encoding a protein involved in
111 invasivity and adherence), and *bib(A)* (encoding an adhesin) were investigated by PCR
112 using specific primers (Table 1).

113 **Antimicrobial susceptibility testing and macrolide resistance phenotype**

114 The minimal inhibitory concentrations (MICs) were determined using the E-test method
115 according to the protocols established by ¹². The antibiotics tested were: erythromycin,
116 clindamycin, azithromycin, ampicillin, penicillin G and vancomycin. The MLSB
117 resistance phenotype (macrolide–lincosamide–streptogramin B) of the isolates was
118 determined by the double disc diffusion test as described previously¹². The strain
119 *Streptococcus pneumoniae* ATCC 49619 was used as the quality control.

120 **PCR determination of the macrolide resistance phenotype**

121 The presence of the *erm(B)*, *erm(C)*, *erm(A)* (Subclass *erm(TR)*) and *mef(A/E)* genes
122 was determined by PCR amplification using the primers described in Table 1. The PCR
123 products were visualized by electrophoresis on 1% agarose gels and sent for sequencing
124 to Beckman Coulter Genomics (United Kingdom).

125 **Statistical analysis**

126 Data related to serotypes, virulence and antimicrobial susceptibility of the 3 study groups
127 were analyzed using contingency tables and Chi-square (χ^2) tests. A P value < 0.05 was
128 considered statistically significant. The analyses were carried out with the StataCorp.
129 2015. Stata Statistical Software: Release 14. College Station, TX: StataCorp LP.
130 Correlations of all variables have been determining by the Pearson's correlation
131 coefficient.

132

133 **RESULTS**

134 Two hundred forty-two GBS **isolates** were included in the study. The **isolates** were
135 divided into 3 groups: colonizers 95/242 (39.3%) (collected from asymptomatic pregnant
136 women), pathogenic 68/242 (28.1%) (collected from symptomatic pregnant women), and
137 neonatal 79/242 (32.6 %) (collected from infected neonates).

138 Serotype III was the most prevalent (33.9%) followed by serotypes II, Ia, V, IV, Ib, and
139 VII (22.7 %, 14.9%, 10.7%, 8.7%, 8.3% and 0.8% of isolates, respectively). However,
140 serotypes VI, VIII, IX and X were not detected (Table 2). Among the colonizing **isolates**,
141 the most frequent serotypes found were serotype II (31.6%), III (26.3%) and Ia (17.9%),
142 while serotypes III, II and IV (38.2%, 22.1% and 13.2%, respectively) were the most

143 prevalent among the pathogenic **isolates**. In the case of the neonatal **isolates**, serotypes
144 III, Ia and II (39.2%, 22.8% and 12.7%, respectively) were the most frequently found.
145 Serotype Ia was significantly more frequent among colonizing and neonatal **isolates** than
146 among pathogenic **isolates** ($p=0.001$). On the other hand, serotype II was significantly
147 more frequent among colonizing **isolates** ($p=0.012$).

148 Regarding the presence of virulence factors, *cyl*(**E**) (90.5%), followed by *scp*(**B**) (75.6%),
149 *rib* (62.1%) and *bca* (43.4%) (Table 2) were the most frequently found among the
150 **isolates**. The virulence factors *pep* and *bib*(**A**) were only found among the neonatal
151 **isolates** (26.6% and 17.7%, respectively). On the other hand, the *bca* and *scp*(**B**) genes
152 were significantly more frequent among neonatal **isolates** (68.3% and 87.3%, $p=0.0001$
153 and 0.002, respectively) and the *fb*s(**B**) gene was more frequently found among colonizing
154 (39.1%) and pathogenic **isolates** (39.7%) than among neonatal **isolates** (20.2%) ($p=0.01$).
155 Significant relationships were observed between the presence of several virulence
156 determinants and serotypes. The *bac* gene was significantly more frequent among
157 serotype Ib ($p=0.0002$), *rib* gene among serotype III ($p=0.0001$), *spb*(**1**) gene among
158 serotype II ($p=0.0001$), and the *bib*(**A**) gene was significantly more frequent among
159 serotype V ($p=0.049$).

160 All the **isolates** of this study were susceptible to penicillin, ampicillin and vancomycin.
161 However, 21.5% were resistant to azithromycin ($MIC>2\ \mu\text{g/mL}$), 20.7% to erythromycin
162 ($MIC>1\ \mu\text{g/mL}$), and 17.6% to clindamycin ($MIC>1\ \mu\text{g/mL}$). The erythromycin,
163 azithromycin and clindamycin resistant isolates were significantly more frequent among
164 the pathogenic than among the colonizing and neonatal **isolates** ($p=0.002$, 0.002 and
165 0.001, respectively) (Table 2).

166 Different MLSB phenotypes were observed among the **isolates**; 17.8% of strains showed
167 constitutive resistance to clindamycin (cMLSB), 3.3% showed inducible resistance

168 (iMLSB) and 0.3% presented the M phenotype. Among the resistant **isolates**, the most
169 prevalent gene found was the *erm(B)* gene (40/52, 77 %). 12/52 (23%) **unrelated isolates**
170 showed the combination of genes of *erm(A)-erm(B)-mef(A/E)*; 5/52 (9.7 %) **isolates**
171 presented the *erm(A)-erm(C)* genes; and 2/52 (3.8 %) **isolates** presented only the
172 *mef(A/E)* gene (Table 3).

173

174 Correlation between serotypes, presence/absence of virulence factors and resistance to
175 antibiotics was analysed. No relationship was observed with antimicrobial resistance.
176 Among colonizing **isolates**, the presence of *bca* gene was strongly associated with the
177 presence of *bac* gene (Pearson coefficient =0.80).

178

179 **DISCUSSION**

180 *S. agalactiae* or group B *Streptococcus* (GBS) remains one of the most important
181 etiological causes of neonatal sepsis. However, the real burden of GBS infection in
182 newborns is likely underestimated because some cases are not adequately diagnosed,
183 especially in developing countries ¹³.

184 The present study included 242 GBS **isolates** collected from five hospitals in Barcelona.
185 Among the GBS **isolates** studied, four serotypes (III, II, Ia and V) were the most frequent
186 in this study, being serotype II **significantly more prevalent** among colonizing **isolates**
187 and serotype Ia among colonizing and neonatal **isolates**. Our results are somewhat similar
188 to those from other countries in which serotype III was the most frequently found among
189 the GBS strains studied ^{14,15}. Another recent report showed that serotype V was the most
190 frequent in France (44.1%) followed by serotype III ¹⁶. In this sense, studies on the
191 distribution of the GBS serotype have shown that GBS varies both geographically and

192 over time, being serotypes Ia, III, and V more frequently involved in neonatal invasive
193 disease, adult infections and maternal carriage ^{17,18}.

194 GBS presented several virulence genes that allow these microorganisms to cause
195 infection and even to cross intact membranes and arrive to the fetus. Recent studies have
196 been focused on analyzing the surface gene profile, but in the present work, other
197 virulence genes such as toxins and invasins were also studied. The most frequent
198 virulence factors found in our study were *cyl(E)*, *scp(B)*, *rib* and *bca* most of them related
199 to invasion of epithelial cells ¹⁹. These results are in correlation with previous studies
200 carried out in other geographical areas ^{7, 20, 21}. In the present work, the *bca* (related to
201 invasion and antophagocytosis) and *scp(B)* (related to invasion) genes were most
202 prevalent among neonatal **isolates**, probably favoring the colonization and invasion of
203 neonatal mucosa and blood/brain barrier ²¹. **In contrast to our findings, Dutra et al.⁷**
204 **found the *bca* gene more prevalent among colonizing strains and the *scp(B)* gene in**
205 **the 100% of the strains. The virulence factor *rib* related to invasiveness was more**
206 **prevalent among serotype III that is associated with invasive infection and, in our**
207 **study, in agreement with their higher presence in pathogenic isolates. One isolate**
208 **belonging to serotype III and presenting the *spb(1)* and *rib* virulence genes formed**
209 **part of the ST17 associated with neonatal invasive infections ²³.**

210 In the present study, we observed a significant percentage of **isolates** resistant to second
211 choice antibiotics, such as azithromycin, erythromycin and/or clindamycin. Remarkably,
212 we found that the percentage of **isolates** resistant to these antibiotics was significantly
213 higher among pathogenic than colonizing and neonatal **isolates** suggesting that, in
214 contrast with other bacteria such *E. coli*, pathogenic **GBS** strains are more **prone** to
215 acquire antimicrobial resistance ²⁴.

216 Moreover, the rates of resistance to macrolides and clindamycin have also been on the
217 rise worldwide. Domelier *et al.* ²⁵ and Campelo *et al.* ⁹ reported 19% of resistance to
218 erythromycin among GBS strains causing materno-fetal and neonatal disease in France
219 and Las Palmas (Spain), respectively. On the other hand, a study carried out in
220 Switzerland reported 30% of erythromycin resistance and 28% of clindamycin resistance
221 ²⁶, with a significant increase in the number of multidrug-resistant strains. **However,**
222 **other studies have found lower rates of isolates resistant to macrolides among**
223 **pathogenic strains, being lower or equal of 4% ^{7,27}. These differences could be due**
224 **to the selective pressure caused by the antimicrobial treatments that these pregnant**
225 **women could be received during gestation.**

226 Interestingly, the resistance to macrolides and lincosamides in GBS in our study was
227 associated with the presence of the *erm(B)* gene, being consistent with epidemiological
228 studies about invasive GBS isolates collected from neonates in China and South Korea.
229 On the other hand, the coexistence of several resistant genes such as *erm(A)*, *erm(B)*, and
230 *mef(A/E)* (12 isolates) is remarkable in our work. These results differ from those
231 published by other authors, who described combinations of these genes, the most common
232 being the *erm(A)/erm(B)* and *erm(A)/mef(A/E)* genes ^{15,16,28}.

233 **In contrast with other studies that found a significant relationship between**
234 **erythromycin resistance and serotype III ^{29,30,31}, this association was not present**
235 **among our isolates.**

236 In conclusion, although penicillin G and ampicillin continue to be a good treatment for
237 GBS infections, a significant increase in the percentage of **isolates** resistant to macrolides
238 and lincosamines has been observed. The fact that this increase was observed mainly
239 among GBS causing symptomatic disease implies a serious problem for treating this type
240 of infections. Surveillance of the epidemiology, antimicrobial resistance and virulence of

241 GBS could provide great knowledge about these microorganisms and help to improve the
242 treatment of infections caused by this opportunistic pathogen.

243

244

245 **REFERENCES**

- 246 1. Otaguiri ES, Morguette AEB, Tavares ER, dos Santos PMC, Morey AT, Cardoso JD,
247 et al. Commensal *Streptococcus agalactiae* isolated from patients seen at University
248 Hospital of Londrina, Paraná, Brazil: capsular types, genotyping, antimicrobial
249 susceptibility and virulence determinants. *BMC Microbiol.* 2013; 13:297.
- 250 2. Andreu A, Sanfeliu I, Viñas L, Barranco M, Bosch J, Dopico E, et al. Decreasing
251 incidence of perinatal group B streptococcal disease (Barcelona 1994-2002). Relation
252 with hospital prevention policies. *Enferm Infect Microbiol Clin.* 2003; 21:174–9.
- 253 3. Raymond J, Lopez E, Bonacorsi S, Poyart C, Moriette G, Jarreau PH, et al. Evidence
254 for transmission of *Escherichia coli* from mother to child in late-onset neonatal infection.
255 *Ped Infect Dis J.* 2008; 17:186–8.
- 256 4. Poyart C, Tazi A, Réglie-Poupet H, Billoët A, Tavares N, Raymond J, et al. Multiple
257 PCR assay for rapid and accurate capsular typing of group B streptococci. *J Clin*
258 *Microbiol.* 2007; 45:1985–8.
- 259 5. Centers for Disease Control and Prevention (CDC). Hospital-based policies for
260 prevention perinatal Group B streptococcal disease United States, 1999. *MMWR*
261 *Morbidity and Mortality Weekly Report* 2000; 49:936–40.
- 262 6. Bergseng H, Bevanger L, Rygg M, Bergh K. Real-time PCR targeting the sip gene for
263 detection of group B streptococcus colonization in pregnant women at delivery. *J Med*
264 *Microbiol.* 2007; 56:223–8.
- 265 7. Dutra VG, Alves VMN, Olendzki AN, et al. *Streptococcus agalactiae* in Brazil:
266 serotype distribution, virulence determinants and antimicrobial susceptibility. *BMC*
267 *Infect Dis.* 2014; 14:323.

- 268 8. Schrag SJ, Arnold KE, Mohle-Boetani JC, Lynfield R, Zell ER, Stefonek K, et al.
269 Prenatal screening for infectious diseases and opportunities for prevention. *Obst Gynecol.*
270 2003; 102:753–60.
- 271 9. Campelo FA, Pedrosa AC, Antúnez IÁ, Capuz BL. Fenotipos y mecanismos de
272 resistencia a macrólidos y lincosamidas en aislados de *streptococcus agalactiae* con
273 significación clínica en un período de ocho años (2002-2010). *Rev Esp Quimioter.* 2012;
274 25:42–6.
- 275 10. Rubens CE, Wessels MR, Heggen LM, Kasper DL. Transposon mutagenesis of type
276 III group B Streptococcus: correlation of capsule expression with virulence. *Proc Nat*
277 *Acad Sci USA.* 1987; 84:7208–12.
- 278 11. Fluegge K, Supper S, Siedler A, Berner R. Serotype distribution of invasive group B
279 streptococcal isolates in infants: results from a nationwide active laboratory surveillance
280 study over 2 years in Germany. *Clin Infect Dis.* 2005; 40:760–3.
- 281 12. Clinical and Laboratory Standards Institute (CLSI). Performance Standards for
282 Antimicrobial Susceptibility Testing; 24th Informational Supplement, M100-S24. 2014.
- 283 13. Martins ER, Andreu A, Correia P, Juncosa T, Bosch J, Ramirez M, et al. Group B
284 streptococci causing neonatal infections in Barcelona are a stable clonal population: 18-
285 Year surveillance. *J Clin Microbiol.* 2011; 49:2911–8.
- 286 14. Lin FY, Azimi PH, Weisman LE, Philips JB, Regan J, Clark P, et al. Antibiotic
287 susceptibility profiles for group B streptococci isolated from neonates, 1995-1998. *Clin*
288 *Infect Dis.* 2000; 31:76–9.
- 289 15. Wang P, Tong JJ, Ma XH, Song FL, Fan L, Guo CM, et al. Serotypes, antibiotic
290 susceptibilities, and multi-locus sequence type profiles of *Streptococcus agalactiae*

291 isolates circulating in Beijing, China. PLoS One. 2015; 10:1–13.

292 16. Bergal A, Loucif L, Benouareth DE, Bentorki AA, Abat C, Rolain JM. Molecular
293 epidemiology and distribution of serotypes, genotypes, and antibiotic resistance genes of
294 *Streptococcus agalactiae* clinical isolates from Guelma, Algeria and Marseille, France.
295 Eur J Clin Microbiol Infect Dis. 2015; 34:2339–48.

296 17. Ippolito DL, James WA, Tinnemore D, Huang RR, Dehart MJ, Williams J, et al.
297 Group B streptococcus serotype prevalence in reproductive-age women at a tertiary care
298 military medical center relative to global serotype distribution. BMC Infect Dis. 2010;
299 10:336.

300 18. Sadowy E, Matynia B, Hryniewicz W. Population structure, virulence factors and
301 resistance determinants of invasive, non-invasive and colonizing *Streptococcus*
302 *agalactiae* in Poland. J Antimicrob Chemother. 2010; 65:1907–14.

303 19. Lindahl G, Stålhammar-Carlemalm M, Areschoug T. Surface proteins of
304 *Streptococcus agalactiae* and related proteins in other bacterial pathogens. Clin Microbiol
305 Rev. 2005; 18:102-27.

306 20. Beigverdi R, Jabalameli F, Mirsalehian A, Hantoushzadeh S, Boroumandi S,
307 Taherikalani M, et al. Virulence factors, antimicrobial susceptibility and molecular
308 characterization of *Streptococcus agalactiae* isolated from pregnant women. Acta
309 Microbio Immunol Hung. 2014; 61:425–34.

310 21. Rojo-Bezares B, Azcona-Gutiérrez JM, Martín C, Jareño MS, Torres C, Sáenz Y.
311 *Streptococcus agalactiae* from pregnant women: antibiotic and heavy-metal resistance
312 mechanisms and molecular typing. Epidemiol Infect. 2016; 144:3205-14.

313 22. Herbert MA, Beveridge CJE, Saunders NJ. Bacterial virulence factors in neonatal
314 sepsis: group B streptococcus. Curr Op Infect Dis. 2004; 17:225–9.

- 315 23. Tazi A, Disson O, Bellais S, Bouaboud A, Dmytruk N, Dramsi S, et al. The surface
316 protein HvgA mediates group B streptococcus hypervirulence and meningeal tropism in
317 neonates. *J Exp Med.* 2010; 207: 2313-22.
- 318 24. Soto SM, Jimenez de Anta MT, Vila J. Quinolones induce partial or total loss of
319 pathogenicity islands in uropathogenic *Escherichia coli* by SOS-dependent or -
320 independent pathways, respectively. *Antimicrob Agents Chemother.* 2006; 50: 649-53.
- 321 25. Domelier AS, van der Mee-Marquet N, Arnault L, Mereghetti L, Lanotte P, Rosenau
322 A, et al. Molecular characterization of erythromycin-resistant *Streptococcus agalactiae*
323 strains. *J Antimicrob Chemother.* 2008; 62:1227–33.
- 324 26. Capanna F, Emonet SP, Cherkaoui A, Irion O, Schrenzel J, De Tejada BM. Antibiotic
325 resistance patterns among group B Streptococcus isolates: implications for antibiotic
326 prophylaxis for early-onset neonatal sepsis. *Swiss Med Week.* 2013; 25:17–20.
- 327 27. Pinto TC, Costa NS, Vianna Souza AR, Silva LG, Corrêa AB, Fernandes FG, et al.
328 Distribution of serotypes and evaluation of antimicrobial susceptibility among human and
329 bovine *Streptococcus agalactiae* strains isolated in Brazil between 1980 and 2006. *Braz*
330 *J Infect Dis.* 2013; 17: 131-6.
- 331 28. Betriu C, Culebras E, Rodríguez-Avial I, Gómez M, Sánchez BA, Picazo JJ. *In Vitro*
332 Activities of Tigecycline against Erythromycin-Resistant *Streptococcus pyogenes* and
333 *Streptococcus agalactiae*: Mechanisms of Macrolide and Tetracycline Resistance.
334 *Antimicrob Agents Chemother.* 2004; 48:323–5.
- 335 29. De Francesco MA, Caracciolo S, Gargiulo F, Manca N. Phenotypes, genotypes,
336 serotypes and molecular epidemiology of erythromycin-resistant *Streptococcus*
337 *agalactiae* in Italy. *Eur J Clin Microbiol Infect Dis.* 2012; 31: 1741-7.

338 30. Schuab RB, Arêas GP, Souza VC, Barros RR. Molecular epidemiology of
339 *Streptococcus agalactiae* recovered from significant bacteriuria. Infect Dis (Lond). 2015;
340 47: 637-42.

341 31. Yan Y, Hu H, Lu T, Fan H, Hu Y, Li G, Zhang X, Shi Y, Xia R. Investigation of
342 serotype distribution and resistance genes profile in group B Streptococcus isolated from
343 pregnant women: a Chinese multicenter cohort study. APMIS. 2016; 124: 794-9.

344

345