

1 **Genetic and epigenetic associations with pre-COPD lung function trajectories.**

2 David J Martino<sup>1</sup>, Dinh S Bui<sup>2</sup>, Shuai Li<sup>2,3,4,5</sup>, Sabrina Idrose<sup>2</sup>, Jennifer Perret<sup>2</sup>, Adrian Lowe<sup>2</sup>,  
3 Caroline Lodge<sup>2</sup>, Gayan Bowatte<sup>2</sup>, Yuben Moodley<sup>6</sup>, Paul S Thomas<sup>7</sup>, Graeme Zosky<sup>8</sup>, Phillip  
4 Hansbro<sup>9</sup>, John W Holloway<sup>10</sup>, Cecile Svanes<sup>11,12</sup>, Rosa Faner<sup>13,14,15</sup>, Eugene H Walters<sup>2,16\*</sup>  
5 and Shaymali C Dharmage<sup>2\*</sup>.

6 *\* Joint Senior authors*

7

8 *1 Wal-yan Respiratory Research Centre, Telethon Kids Institute, University of Western*  
9 *Australia.*

10

11 *2 Centre for Epidemiology and Biostatistics, Melbourne School of Population and Global*  
12 *Health, The University of Melbourne*

13

14 *3 Centre for Cancer Genetic Epidemiology, Department of Public Health and Primary Care,*  
15 *University of Cambridge*

16

17 *4 Precision Medicine, School of Clinical Sciences at Monash Health, Monash University,*  
18 *Clayton, Victoria, Australia*

19

20 *5 Murdoch Children's Research Institute, Royal Children's Hospital, Parkville, Victoria 3051,*  
21 *Australia*

22

23 *6 Department of Respiratory Medicine, Fiona Stanley Hospital, Murdoch, Australia*

24

25 *7 Respiratory Medicine Prince of Wales' Hospital & Prince of Wales' Clinical School, UNSW,*  
26 *Randwick, NSW 2031, Australia*

27

28 *8 Menzies Institute for Medical Research, University of Tasmania*

29

30 *9 Centre for Inflammation, Centenary Institute and University of Technology Sydney, School of*  
31 *Life Sciences, Faculty of Science, Sydney NSW 2007, Australia*

32

33 *10 Human Development and Health, Faculty of Medicine, University of Southampton,*  
34 *Southampton UK.*

35

36 *11 Centre for International Health, Department of Global Public Health and Primary Care,*  
37 *University of Bergen, Norway.*

38

39 *12 Department of Occupational Medicine, Haukeland University Hospital, Bergen, Norway.*

40

41 *13 University of Barcelona, Immunology Unit, Biomedicine Department. Spain.*

42

43 14. *IDIBAPS-FCRB Institut d'Investigacions Biomediques August Pi i Sunyer-Fundació Clinic*  
44 *Recerca, Barcelona, Spain.*

45

46 15. *Centro de Investigación Biomédica en Red de Enfermedades Respiratorias (CIBER), Spain.*

47

48 16 *School of Medicine, University of Tasmania and School of Population and Global Health,*  
49 *University of Melbourne*

50

51 **Running title: Genomics of pre-COPD lung function trajectories.**

52

53 **Corresponding author:** David Martino

54 Telethon Kids Institute

55 PO Box 855, West Perth, Western Australia, 6872

56

57 **Abbreviations**

58 COPD – Chronic Obstructive Pulmonary Disease

59 CpG – Cytosine phosphate Guanine

60 DMR – Differentially Methylated Region

61 GWAS – Genome Wide Association Study

62 mQTL – Methylation Quantitative Trait Loci

63 SNP – Single Nucleotide Polymorphism

64 TAHS – Tasmanian Longitudinal Health Study

65 To the Editor,

66 Understanding the molecular mechanisms of lung function trajectories that progress to  
67 chronic obstructive pulmonary disease (COPD) (pre-COPD trajectories), especially those with  
68 a rapidly declining phenotype, should inform preventive interventions. The Tasmanian  
69 Longitudinal Health Study (TAHS) previously defined life-course lung function trajectories by  
70 serial spirometry in a cohort of all seven-year-old school children in the state of Tasmania  
71 recruited in 1968 and followed up to age 53 years (1). Of the six pre-bronchodilator FEV<sub>1</sub>  
72 lifetime trajectories identified, three collectively accounted for 75% of chronic obstructive  
73 pulmonary disease (COPD) prevalence at age 53 years (2). These high-risk trajectories were:  
74 1) early below average lung function (with usual rate of subsequent decline), 2) persistently  
75 low, and 3) early below average lung function with accelerated decline. The TAHS cohort  
76 provides a unique opportunity to investigate molecular factors associated with disadvantaged  
77 trajectories, and we conducted a pilot study in this cohort to characterize associations with  
78 COPD high-risk trajectories to inform more extensive longitudinal studies in the future.

79 The rationale for our approach was based on previous studies that demonstrate declining  
80 lung function (3-6) and lower lung function associated with COPD (7, 8) are complex  
81 phenotypes involving the interplay between genomic and environmental factors. Genetic (9)  
82 and epigenetic associations (3, 7, 10) have been previously been described for disadvantaged  
83 lung function trajectories, including epigenetic aging related phenotypes (epigenetic age  
84 acceleration) (5, 10). We therefore conducted a screen of both epigenetic (DNA methylation)  
85 and genetic (single nucleotide polymorphisms; SNPs) markers on available whole blood  
86 samples collected at the 45-year follow-up. We used an extremes of phenotype design to  
87 maximise power randomly selecting 80 subjects from across the three high-risk trajectories

88 and matching on age and smoking status to 80 subjects from the persistently high trajectory.  
89 By design, individuals belonging to the different lung function trajectories varied in post  
90 bronchodilator FEV<sub>1</sub>, but also steroid medication use, and sex so these differences were  
91 examined in adjusted models. We quantified 787,111 DNA methylation markers (CpGs) and  
92 4,456,571 SNPs using the InfiniumMethylationEPIC (v1) and Infinium Global Screening Array  
93 (v3) genotyping microarrays. Some of the results of this study have been previously reported  
94 in the form of an abstract (11).

95 We detected DNA methylation differences at 55 differentially methylated regions (DMRs)  
96 containing 73 unique genes and 6 non-coding regions (FDR adjusted P < 0.05; Figure 1A).  
97 Notable genes in DMRs included *LY6G5C* and *HLA-DQB1* within the major histocompatibility  
98 complex, HOX cluster transcription factors (*HOXB-AS3*, *HOXB3*, *HOXB6*) which have been  
99 implicated in the pathogenesis of pulmonary diseases (12, 13), and transmembrane  
100 glycoproteins (*LGALS3BP*, *OCA2*, *KCNE1*, *PTPRN2*, *TNXB*, *PCDHGA5*, *CDSN*, *PCDHGA4*,  
101 *PCDHGA3*, *PCDHGB3*, *PCDHGA2*, *PCDHGB2*, *PCDHGA1*, *EGFR*, *DPP6*, *FOLH1*, *SGCD*, *CRTAC1*,  
102 *PCDHGB1*, *FIBIN*, *CHST1*, *MUCA4*, *DPEP3*) that play a role in epithelial biology and when  
103 disrupted may lead to EMT (14).

104 This DMR signature was only partially consistent across the high-risk trajectory sub-groups  
105 (24% of DMRs shared across all sub-groups), whereas sub-group specific regions  
106 predominated (Figure 1B) consistent with the notion of COPD risk factor etiologies that likely  
107 exhibit different molecular drivers (15). Current COPD or current asthma explained 17-30% of  
108 methylation differences across the DMRs respectively, but sex and blood cell counts were not  
109 mediators or confounders of these associations. Integrating the genetic and epigenetic data  
110 sets we performed methylation quantitative trait (mQTL) mapping and found that genetic  
111 variation at 381 nearby SNPs (+/- 500kb of DMRs) in 17 genomic loci were associated with

112 23% of CpGs within DMR regions (Figure 1C). The strongest mQTL region was on chromosome  
113 6 at the major histocompatibility locus. Using publicly available tissue specific gene expression  
114 signatures (GTEx catalogue v8) we determined these mQTL SNPs were statistically enriched  
115 among transcripts primarily expressed in the lung (ATP13A4, MUC4, PSORS1C1) (Figure 1D).  
116 Several mQTLs have previously been associated with Lung function phenotypes (HAPLN1,  
117 HLA-DRB1, HLA-DQA1, HLA-DQB1), COPD (HLA-DQB1, HLA-DQA1) and Asthma risk (HLA-  
118 DQB1, HLA-DQA1, HLA-DRB1, HLA-DRB6, PSORS1C1) in the genome-wide association study  
119 (GWAS) catalogue. Consistent with previous studies (5, 10) we also found that epigenetic age  
120 predictions were significantly higher in the high-risk group when measured using the  
121 phenoAge algorithm (16) (Table 1). On average, individuals in the high-risk category had  
122 increased mean predicted chronological age of 1.5 years relative to controls (40.2 v 38.7,  
123  $P=0.03$ , t-test). Stratified analysis suggested age-acceleration was strongest in the early below  
124 average, accelerated decline group (Beta = 2.1,  $P=0.06$  v 1.4,  $P=0.11$ , *below average*; v 1.1,  
125  $P=0.19$ , *persistently low*) although sample size was a limiting factor.

126 To our knowledge this was the first epigenome-wide association analysis in individuals from  
127 COPD-risk lung function trajectories, providing a strong foundation for further delineation of  
128 phenotypes and risk factors to enable precision molecular profiling. We determined blood to  
129 be a phenotypically relevant tissue to explore molecular associations with life-time lung  
130 function trajectories in this cohort. Although causality of the epigenetic associations cannot  
131 be established in this pilot, a subset of epigenetic changes in the high-risk trajectory were  
132 mQTLs whereby genetic variants affected the methylation patterns at these genes. Since the  
133 causality of genetic variation on DNA methylation levels is uni-directional, these analyses aid  
134 in prioritization of methylation-trait associations from epigenome-wide scans. The mQTL  
135 associations are compelling candidates for gene-environment interactions, and might be

136 linked to early life events, as well as processes related to disease progression. Confirmatory  
137 longitudinal are now planned to dissect these environmental and host genomic risk factors  
138 that are reflected in the epigenome. Our analysis of DMR sharing across sub-groups suggests  
139 molecular risk factors will be unique across different life-time lung function trajectories  
140 warranting follow-up studies at cohort-wide scale. We also determined that comorbidities  
141 including current asthma and COPD explained a proportion of variation in the blood  
142 epigenetic markers. Consistent with previous reports for declining lung function and  
143 epigenetic ageing phenotypes (5, 10), we also found that epigenetic age acceleration was  
144 detectable in the high-risk trajectory group, the biology of which is still poorly understood. In  
145 summary this pilot study confirms the utility of our approach and paves the way for future  
146 profiling studies in this unique cohort. An enhanced understanding of molecular risk factors  
147 associated with disadvantaged trajectories will enable more precise biomarker-driven  
148 interventions in the future with potential to redirect the course of respiratory health in  
149 vulnerable individuals.

150

151

152

153

154

155

156

157

158

159

160

161

162

163 **References**

- 164 1. Matheson MC, Abramson MJ, Allen K, Benke G, Burgess JA, Dowty JG, Erbas B, Feather  
165 IH, Frith PA, Giles GG, Gurrin LC, Hamilton GS, Hopper JL, James AL, Jenkins MA,  
166 Johns DP, Lodge CJ, Lowe AJ, Markos J, Morrison SC, Perret JL, Southey MC,  
167 Thomas PS, Thompson BR, Wood-Baker R, Haydn Walters E, Dharmage SC, group  
168 Ti. Cohort Profile: The Tasmanian Longitudinal Health STUDY (TAHS). *Int J*  
169 *Epidemiol* 2017; 46: 407-408i.
- 170 2. Bui DS, Lodge CJ, Burgess JA, Lowe AJ, Perret J, Bui MQ, Bowatte G, Gurrin L, Johns  
171 DP, Thompson BR, Hamilton GS, Frith PA, James AL, Thomas PS, Jarvis D, Svanes  
172 C, Russell M, Morrison SC, Feather I, Allen KJ, Wood-Baker R, Hopper J, Giles GG,  
173 Abramson MJ, Walters EH, Matheson MC, Dharmage SC. Childhood predictors of  
174 lung function trajectories and future COPD risk: a prospective cohort study from the  
175 first to the sixth decade of life. *Lancet Respir Medicine* 2018; 6: 535-544.
- 176 3. Ngo D, Pratte KA, Flexeder C, Petersen H, Dang H, Ma Y, Keyes MJ, Gao Y, Deng S,  
177 Peterson BD, Farrell LA, Bhambhani VM, Palacios C, Quadir J, Gillenwater L, Xu H,  
178 Emson C, Gieger C, Suhre K, Graumann J, Jain D, Conomos MP, Tracy RP, Guo X,  
179 Liu Y, Johnson WC, Cornell E, Durda P, Taylor KD, Papanicolaou GJ, Rich SS, Rotter  
180 JI, Rennard SI, Curtis JL, Woodruff P, Comellas AP, Silverman EK, Crapo JD, Larson  
181 MG, Vasan RS, Wang TJ, Correa A, Sims M, Wilson JG, Gerszten RE, O'Connor GT,  
182 Barr RG, Couper D, Dupuis J, Manichaikul A, O'Neal W, Tesfaigzi Y, Schulz H,  
183 Bowler R. Systemic Markers of Lung Function and FEV(1) Decline across Diverse  
184 Cohorts. *Ann Am Thorac Soc* 2023.
- 185 4. Lee M, Huan T, McCartney DL, Chittoor G, de Vries M, Lahousse L, Nguyen JN, Brody  
186 JA, Castillo-Fernandez J, Terzikhan N, Qi C, Joehanes R, Min JL, Smilnak GJ, Shaw  
187 JR, Yang CX, Colicino E, Hoang TT, Bermingham ML, Xu H, Justice AE, Xu CJ, Rich  
188 SS, Cox SR, Vonk JM, Prokic I, Sotoodehnia N, Tsai PC, Schwartz JD, Leung JM,  
189 Sikdar S, Walker RM, Harris SE, van der Plaats DA, Van Den Berg DJ, Bartz TM,  
190 Spector TD, Vokonas PS, Marioni RE, Taylor AM, Liu Y, Barr RG, Lange LA,  
191 Baccarelli AA, Obeidat M, Fornage M, Wang T, Ward JM, Motsinger-Reif AA,  
192 Hemani G, Koppelman GH, Bell JT, Gharib SA, Brusselle G, Boezen HM, North KE,  
193 Levy D, Evans KL, Dupuis J, Breeze CE, Manichaikul A, London SJ. Pulmonary  
194 Function and Blood DNA Methylation: A Multiancestry Epigenome-Wide Association  
195 Meta-analysis. *Am J Respir Crit Care Med* 2022; 206: 321-336.
- 196 5. Carmona JJ, Barfield RT, Panni T, Nwanaji-Enwerem JC, Just AC, Hutchinson JN, Colicino  
197 E, Karrasch S, Wahl S, Kunze S, Jafari N, Zheng Y, Hou L, DeMeo DL, Litonjua AA,  
198 Vokonas PS, Peters A, Lin X, Schwartz J, Schulz H, Baccarelli AA. Metastable DNA  
199 methylation sites associated with longitudinal lung function decline and aging in  
200 humans: an epigenome-wide study in the NAS and KORA cohorts. *Epigenetics* 2018;  
201 13: 1039-1055.
- 202 6. Fang J, Gao Y, Zhang M, Jiang Q, Chen C, Gao X, Liu Y, Dong H, Tang S, Li T, Shi X.  
203 Personal PM(2.5) Elemental Components, Decline of Lung Function, and the Role of  
204 DNA Methylation on Inflammation-Related Genes in Older Adults: Results and  
205 Implications of the BAPE Study. *Environ Sci Technol* 2022; 56: 15990-16000.

- 206 7. Qiu W, Baccarelli A, Carey VJ, Boutaoui N, Bacherman H, Klanderman B, Rennard S,  
207 Agusti A, Anderson W, Lomas DA, DeMeo DL. Variable DNA methylation is  
208 associated with chronic obstructive pulmonary disease and lung function. *Am J Respir*  
209 *Crit Care Med* 2012; 185: 373-381.
- 210 8. Casas-Recasens S, Noell G, Mendoza N, Lopez-Giraldo A, Garcia T, Guirao A, Agusti A,  
211 Faner R. Lung DNA Methylation in Chronic Obstructive Pulmonary Disease:  
212 Relationship with Smoking Status and Airflow Limitation Severity. *Am J Respir Crit*  
213 *Care Med* 2021; 203: 129-134.
- 214 9. Busch R, Cho MH, Silverman EK. Progress in disease progression genetics: dissecting the  
215 genetic origins of lung function decline in COPD. *Thorax* 2017; 72: 389.
- 216 10. Rezwani FI, Imboden M, Amaral AFS, Wielscher M, Jeong A, Triebner K, Real FG,  
217 Jarvelin M-R, Jarvis D, Probst-Hensch NM, Holloway JW. Association of adult lung  
218 function with accelerated biological aging. *Aging Albany Ny* 2020; 12: 518-542.
- 219 11. Martino D, Bui D, Lodge C, Perret J, Boweatte G, Lowe A, Faner R, Walters E, Thomas  
220 P, Moodley Y, Zosky G, Hamilton G, Holloway J, Svanes C, Abramson M, Dharmage  
221 S. Epigenetic study of COPD-risk lung function trajectories. *Respirology* 2022; 27: 3-  
222 4.
- 223 12. Golpon HA, Geraci MW, Moore MD, Miller HL, Miller GJ, Tuder RM, Voelkel NF. HOX  
224 genes in human lung: altered expression in primary pulmonary hypertension and  
225 emphysema. *Am J Pathol* 2001; 158: 955-966.
- 226 13. Portas L, Pereira M, Shaheen SO, Wyss AB, London SJ, Burney PGJ, Hind M, Dean CH,  
227 Minelli C. Lung Development Genes and Adult Lung Function. *Am J Respir Crit Care*  
228 *Med* 2020; 202: 853-865.
- 229 14. Sohal SS, Mahmood MQ, Walters EH. Clinical significance of epithelial mesenchymal  
230 transition (EMT) in chronic obstructive pulmonary disease (COPD): potential target for  
231 prevention of airway fibrosis and lung cancer. *Clin Transl Med* 2014; 3: 33.
- 232 15. Stolz D, Mkorombindo T, Schumann DM, Agusti A, Ash SY, Bafadhel M, Bai C, Chalmers  
233 JD, Criner GJ, Dharmage SC, Franssen FME, Frey U, Han M, Hansel NN, Hawkins  
234 NM, Kalhan R, Konigshoff M, Ko FW, Parekh TM, Powell P, Mólken MR-v, Simpson  
235 J, Sin DD, Song Y, Suki B, Troosters T, Washko GR, Welte T, Dransfield MT. Towards  
236 the elimination of chronic obstructive pulmonary disease: a Lancet Commission.  
237 *Lancet* 2022; 400: 921-972.
- 238 16. Levine ME, Lu AT, Quach A, Chen BH, Assimes TL, Bandinelli S, Hou L, Baccarelli AA,  
239 Stewart JD, Li Y, Whitsel EA, Wilson JG, Reiner AP, Aviv A, Lohman K, Liu Y,  
240 Ferrucci L, Horvath S. An epigenetic biomarker of aging for lifespan and healthspan.  
241 *Aging Albany Ny* 2018; 10: 573-591.



244 **Figure Legends**

245 **Figure 1** – (A) Volcanoplot of differentially methylated regions. Each point represents a  
246 genomic region, and the number of individual CpGs in the region is shown on the y-axis as a  
247 function of the effect size (x-axis) interpreted as the percent change in methylation ratios ( $10^{-2}$ ). (B) Upset plot showing the number of overlapping DMRs per trajectory sub-group. *ph*=  
248 *persistently high*; *acc.dec* = *accelerated decline*; *bl.ave* = *below average*. (C) Boxplot of the  
249 Mucin 4 mQTL showing methylation ratios expressed as a percentage ( $10^{-2}$ ) stratified by  
250 genotype. Means comparisons by t-test, exact P-values shown. (D) Summary statistics of  
251 tissue-specific enrichment testing for mQTLs and sets of differentially expressed genes for 30  
252 general tissue types in the GTExv8 catalogue.  
253

254

**Table 1 - Logistic regression of PhenoAge clock with case - control group**

	Coefficient	Std. Error	z value	P value
AA	0.09	0.05	1.85	0.06
Male sex	1.11	0.38	2.90	0.06
Asthma	2.23	0.48	4.67	<0.01*
EAA	0.09	0.05	1.85	0.06
Male sex	1.11	0.38	2.90	<0.01*
Asthma	2.23	0.48	4.66	<0.01*
IEAA	0.10	0.05	1.94	0.05*
Male sex	1.09	0.38	2.87	<0.01*
Asthma	2.26	0.48	4.70	<0.01*

256 *Outcome variable = high-risk/persistently low, predictors: AA = Age acceleration residual,*  
 257 *EAA = Extrinsic age acceleration residual, IEAA = Intrinsic age acceleration residual. \* = P*  
 258 *<0.05.*

259

260

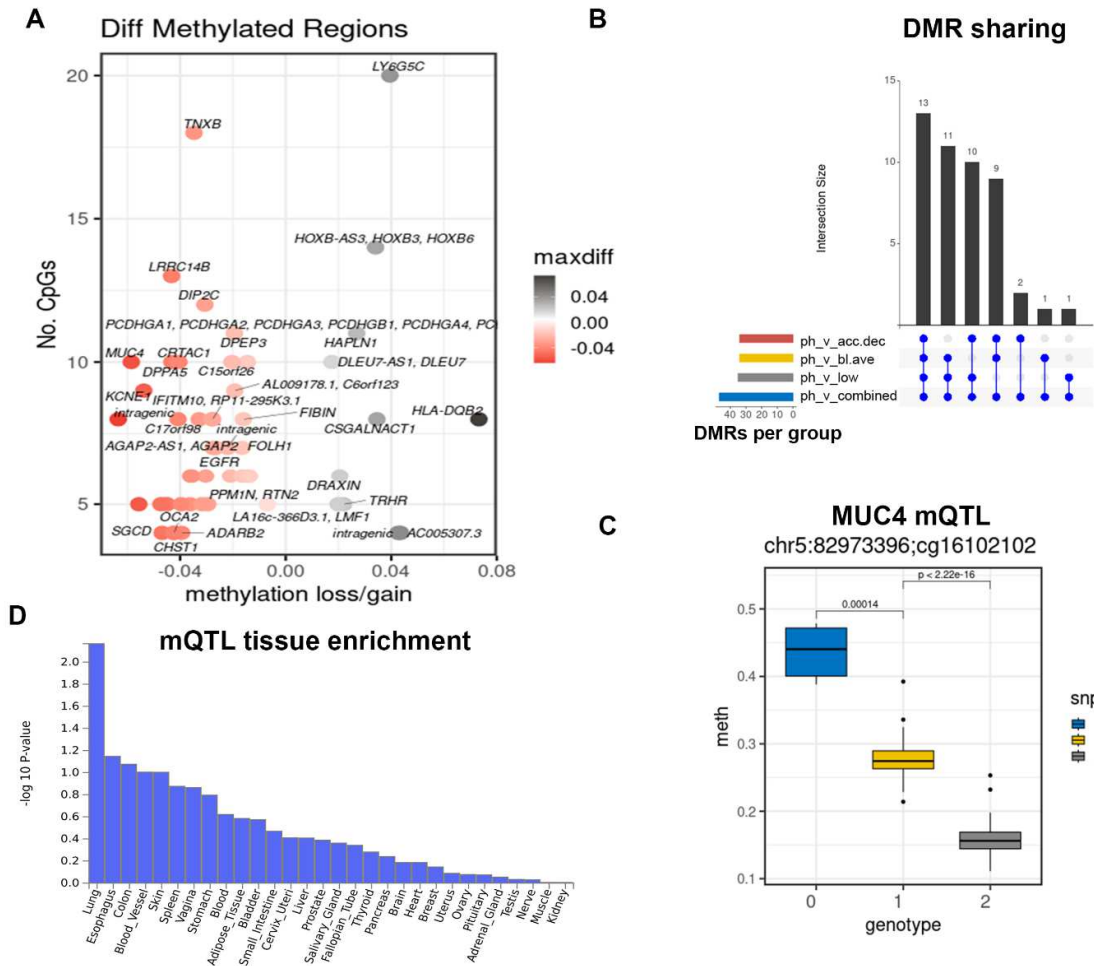
261

262

263

264

265



266

267

268 *Figure 1*

269

270 Supported by Fondo de Investigacion en Salud–Instituto de Salud Carlos III grant PI21/00735,

271 Western Australian Future Health Research and Innovation Fund grant WANMA/2021,

272 National Health and Medical Research Council grants 1175134 and 2010287, and European

273 Research Council grant 101044387..

274 Author Contributions : S.C.D., D.J.M., and D.S.B. participated in the conception and design of

275 the study. R.F., S.L., N.S. 8 , J.P., E.H.W., and S.I. participated in intellectual development,

276 interpretation of data, and drafting of the manuscript. Y.M., G.Z., P.M.H., J.H., C.S., A.L., and

277 C.L. contributed to the development and editing of the manuscript.

278