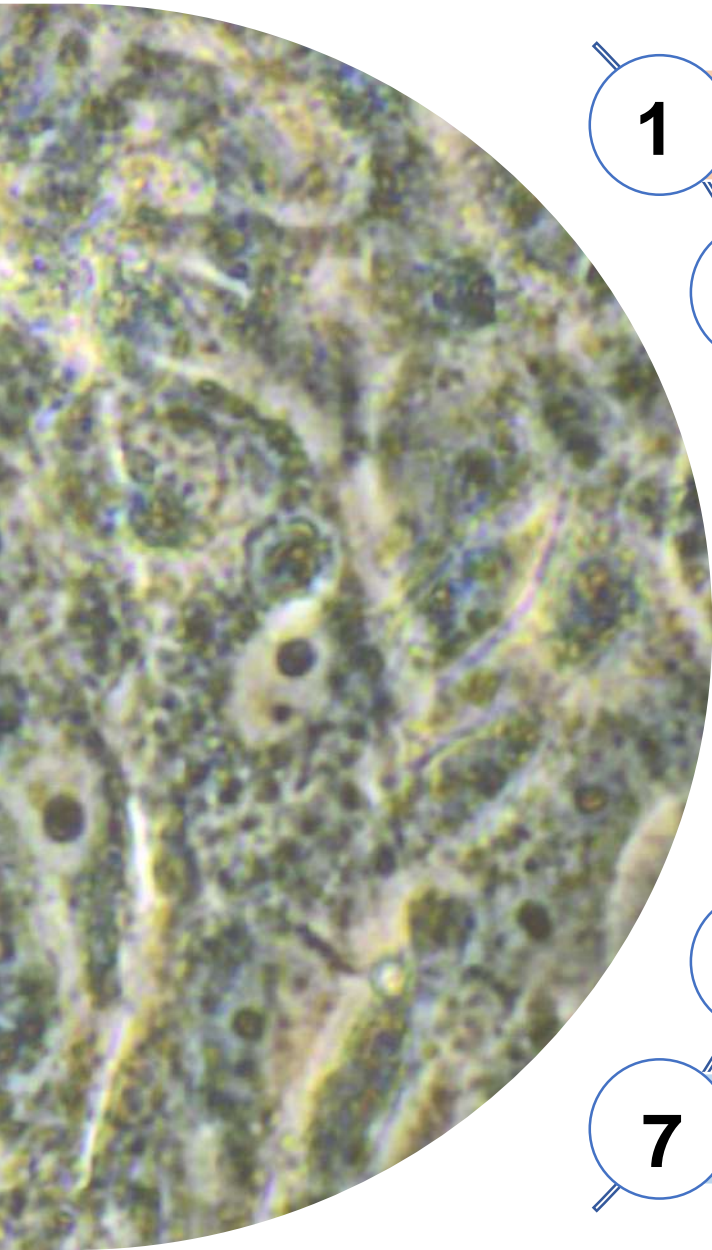


**Identification of anti-parasitic
compounds against *Trypanosoma cruzi*,
the causal agent of Chagas disease,
through the evaluation of diverse
chemical collections**

Nieves Martínez Peinado

Thesis directors: Dr. Julio Alonso Padilla and Prof. Joaquim Gascón

Tutor: Jordi Vila Estapé



- 1** Introduction
- 2** Hypothesis
- 3** Objectives
- 4** Methods
- 5** Results and discussion
- 6** Limitations
- 7** Conclusions

1. INTRODUCTION

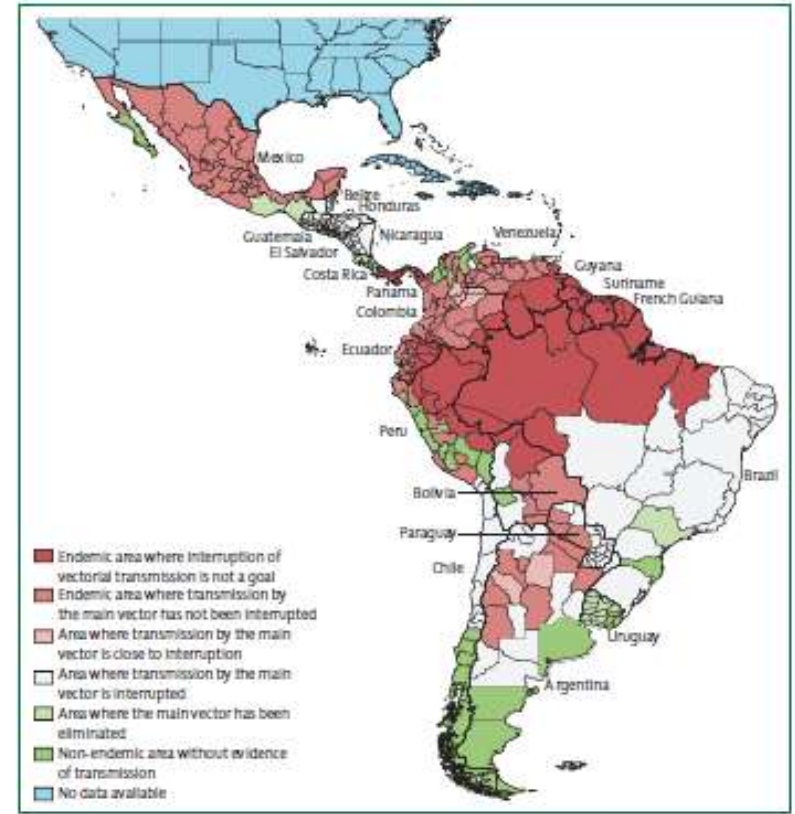
- Is a neglected tropical disease (NTD) caused by the Kinetoplastid protozoan parasite *Trypanosoma cruzi* (*T.cruzi*).

EPIDEMIOLOGY

- ~7 million people are affected by the disease, mainly in Latin America where is endemic in 21 countries.
- Spread to non-endemic areas → global health problem.

TRANSMISION

- Main route: triatomine insects (*family Reduviidae*).
- Blood transfusion, organ transplants, vertical transmission.



(Perez-Molina J.A. et al, Lancet 2018)

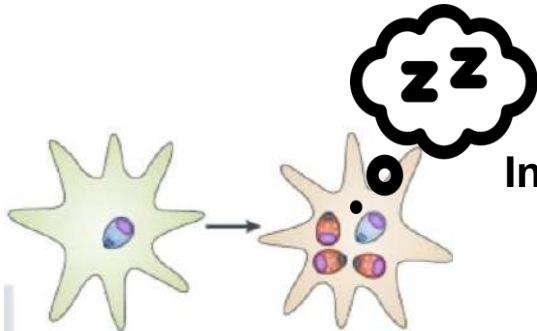
1. INTRODUCTION

TRYPANOSOMA CRUZI LIFE CYCLE



Mammalian host stage

Multiple hosts
(>100 species)
Multiple tissues

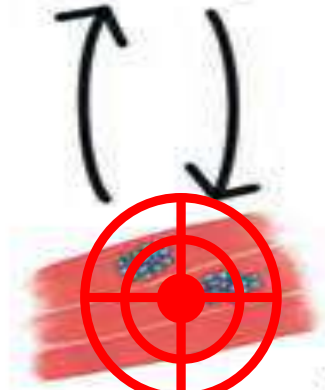


Intracellular replicative in multiple tissues

Motile bloodstream form



Trypomastigote



Amastigote



Epimastigote



Metacyclic Trypomastigote



Replicative in insect gut

Triatomine insect vector stage

Multiple vectors

Mammalian first infective motile form

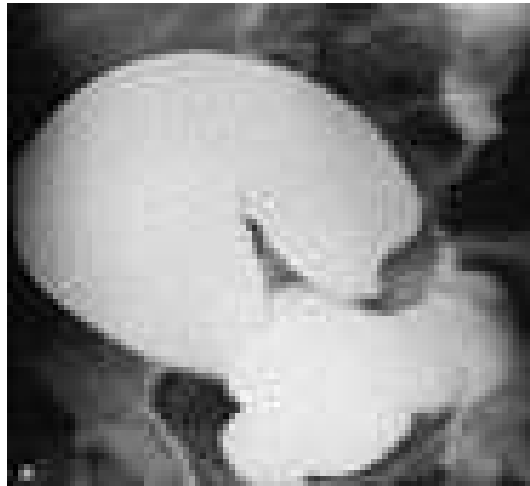
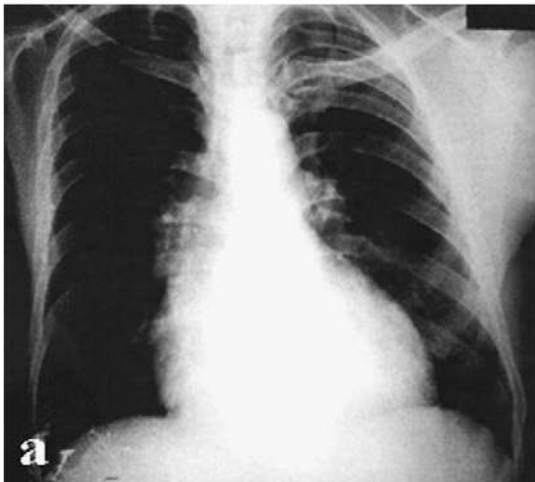
(Atwood et al., Science 2005)

1. INTRODUCTION

CLINICAL MANIFESTATIONS

Chagas disease may progress in two clinical phases:

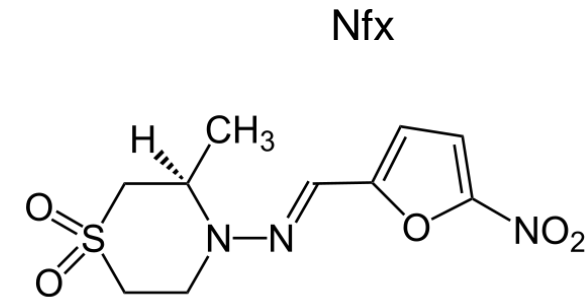
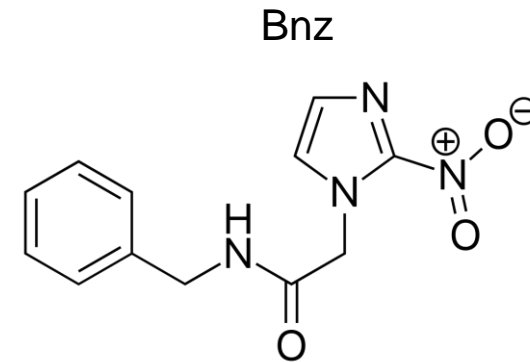
- Acute
- Chronic
 - Indeterminate
 - 30-40% infected: cardiac and/or digestive damage



(Coura J.R. et al, Acta Tropica 2010)

TREATMENT

- Only two drugs available: Benznidazole (Bnz) and Nifurtimox (Nfx).
- Good efficacy in acute phase but diminished as the disease progress.
- High toxicity and frequent adverse events.



URGENT NEED OF NEW ANTI CHAGASIC DRUGS FOR CRHONIC PHASE!

DRUG DISCOVERY



New Compound Sets Identified from High Throughput Phenotypic Screening Against Three Kinetoplastid Parasites: An Open Resource

Imanol Peña¹, M. Pilar Manzano², Juan Cantizani², Albane Kessler², Julio Alonso-Padilla³, Ana I. Bardera¹, Emilio Alvarez¹, Gonzalo Colmenarejo¹, Ignacio Cotillo², Irene Roquero¹, Francisco de Dios-Anton¹, Vanessa Barroso¹, Ana Rodriguez³, David W. Gray⁴, Miguel Navarro⁵, Vinod Kumar⁶, Alexander Sherstnev⁷, David H. Drewry⁸, James R. Brown⁶, Jose M. Fiandor² & J. Julio Martin¹

(Peña I et al., Scientific Reports 2015)

The NEW ENGLAND JOURNAL of MEDICINE

ORIGINAL ARTICLE

Randomized Trial of Posaconazole and Benznidazole for Chronic Chagas' Disease

Israel Molina, M.D., Jordi Gómez i Prat, M.D., Fernando Salvador, M.D., Begoña Treviño, M.D., Elena Sulleiro, M.D., Núria Serre, M.D., Diana Pou, M.D., Sílvia Roure, M.D., Juan Cabezos, M.D., Lluís Valerio, Ph.D., Albert Blanco-Grau, M.D., Adrián Sánchez-Montalvá, M.D., Xavier Vidal, Ph.D., and Albert Pahissa, Ph.D.

Proteasome inhibition for treatment of leishmaniasis, Chagas disease and sleeping sickness

Shilpi Khare^{1*}, Advait S. Nagle^{1*}, Agnes Biggart¹, Yin H. Lai¹, Fang Liang¹, Lauren C. Davis¹, S. Whitney Barnes¹, Casey J. N. Mathison¹, Elmarie Myburgh^{2,3}, Mu-Yun Gao¹, J. Robert Gillespie⁴, Xianzhong Liu¹, Jocelyn L. Tan¹, Monique Stinson¹, Tanne C. Rivera¹, Jaime Ballard¹, Vince Yeh¹, Todd Groessl¹, Glenn Federe¹, Hazel X. Y. Koh⁵, John D. Venable¹, Badry Bursulaya¹, Michael Shapiro¹, Pranab K. Mishra¹, Glen Spraggon¹, Ansgar Brock¹, Jeremy C. Mottram^{2,3}, Frederick S. Buckner⁴, Srinivasa P. S. Rao⁵, Ben G. Wen¹, John R. Walker¹, Tove Tuntland¹, Valentina Molteni¹, Richard J. Glynn¹ & Frantisek Supek¹



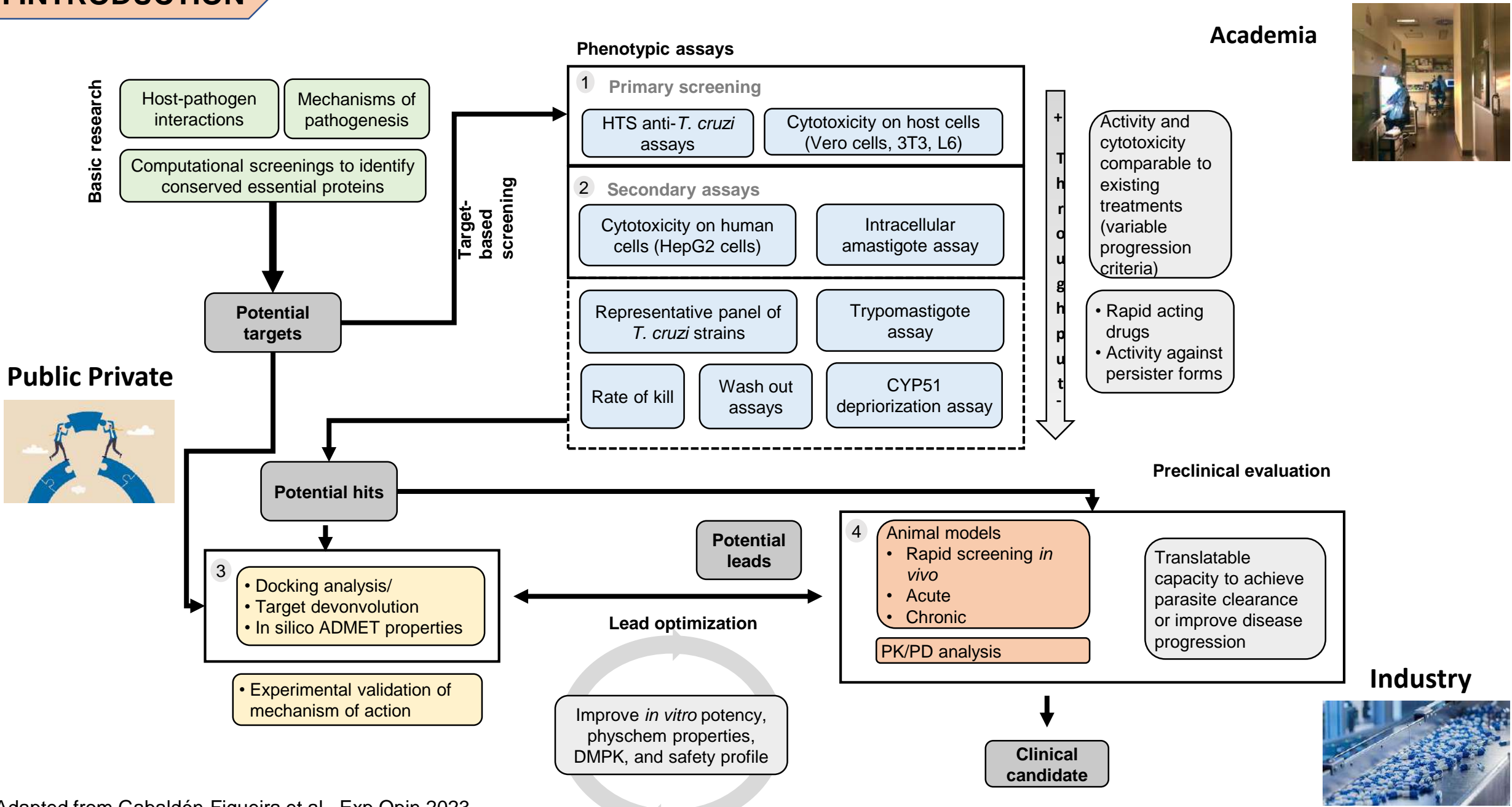
(Khare S, Nature 2016)

Spontaneous dormancy protects *Trypanosoma cruzi* during extended drug exposure

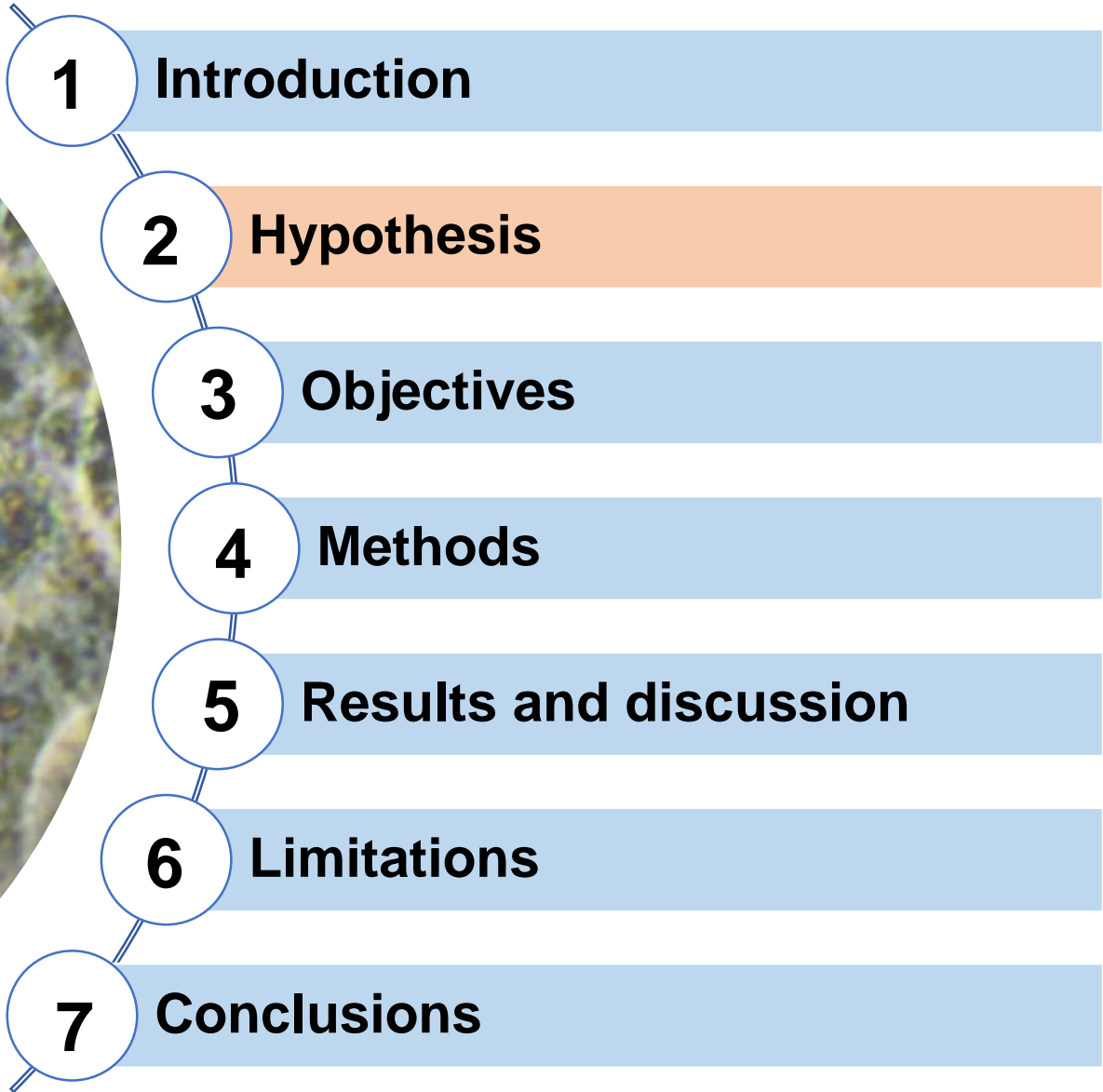
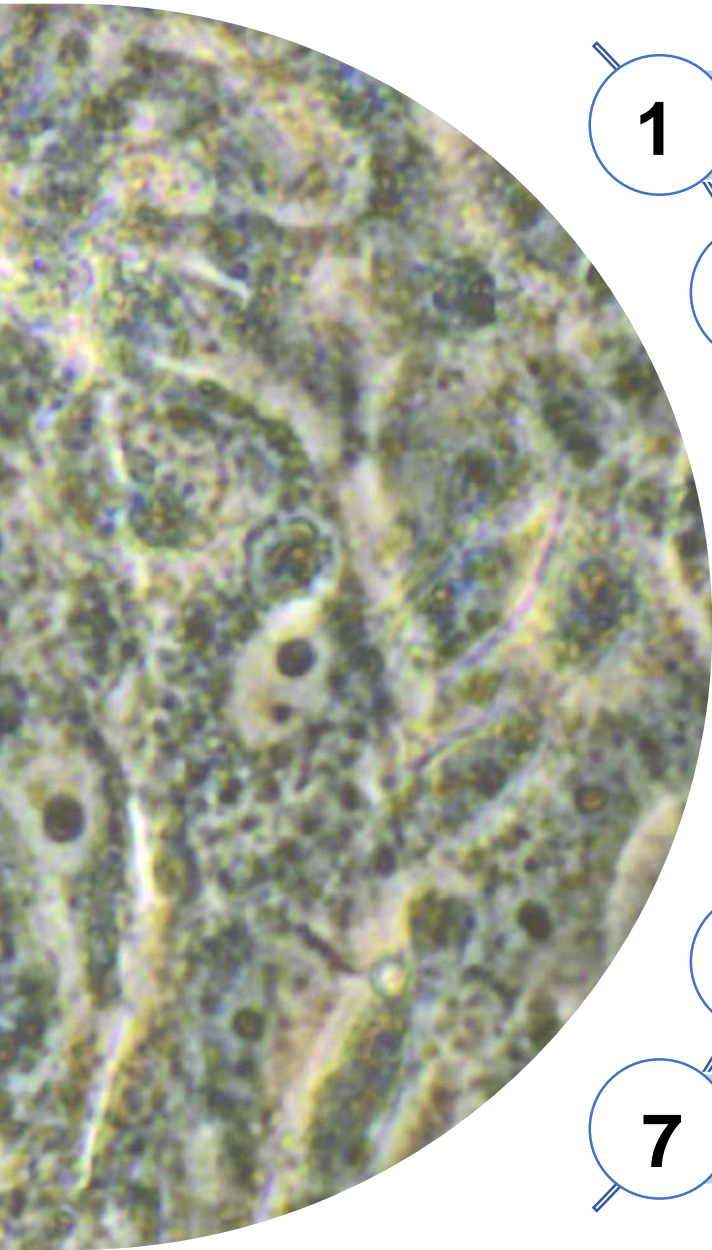
Fernando J Sánchez-Valdéz^{1†}, Angel Padilla^{1,2†}, Wei Wang¹, Dylan Orr¹, Rick L Tarleton^{1,2*}

¹Center for Tropical and Emerging Global Diseases, University of Georgia, Athens, United States; ²Department of Cellular Biology, University of Georgia, Athens, United States

1. INTRODUCTION



Adapted from Gabaldón-Figueira et al., Exp Opin 2023



2. HYPOTHESIS

The exploration of the structural diversity and biological properties from different chemical collections obtained through collaborations will allow to preclinically prioritize chemical entities for the treatment of Chagas disease.



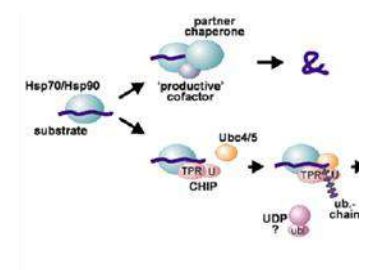
Amaryllidaceae plants

- Natural compounds are a valuable source of active biological substances
- Unique alkaloid constituents



Licensed drugs

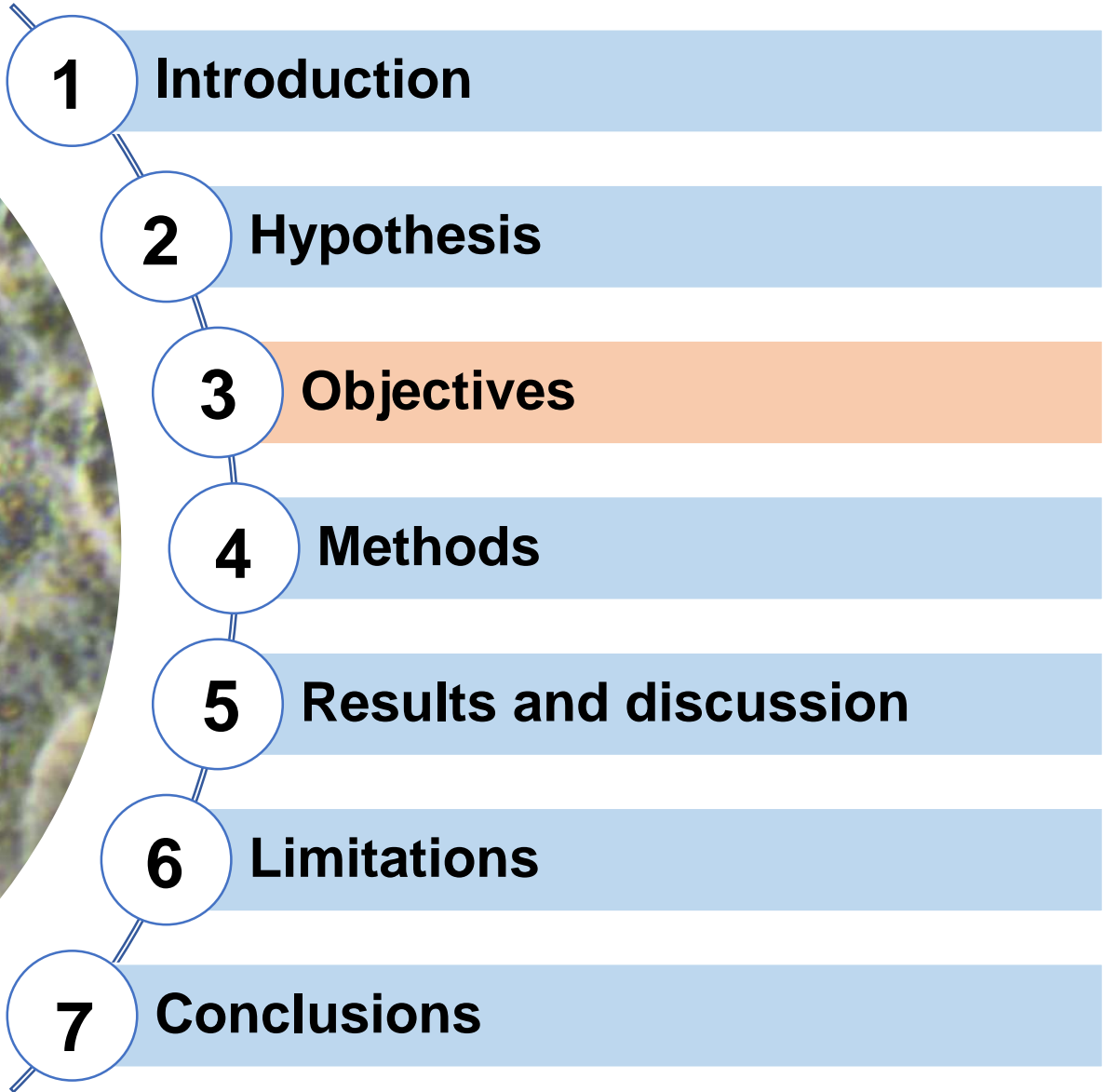
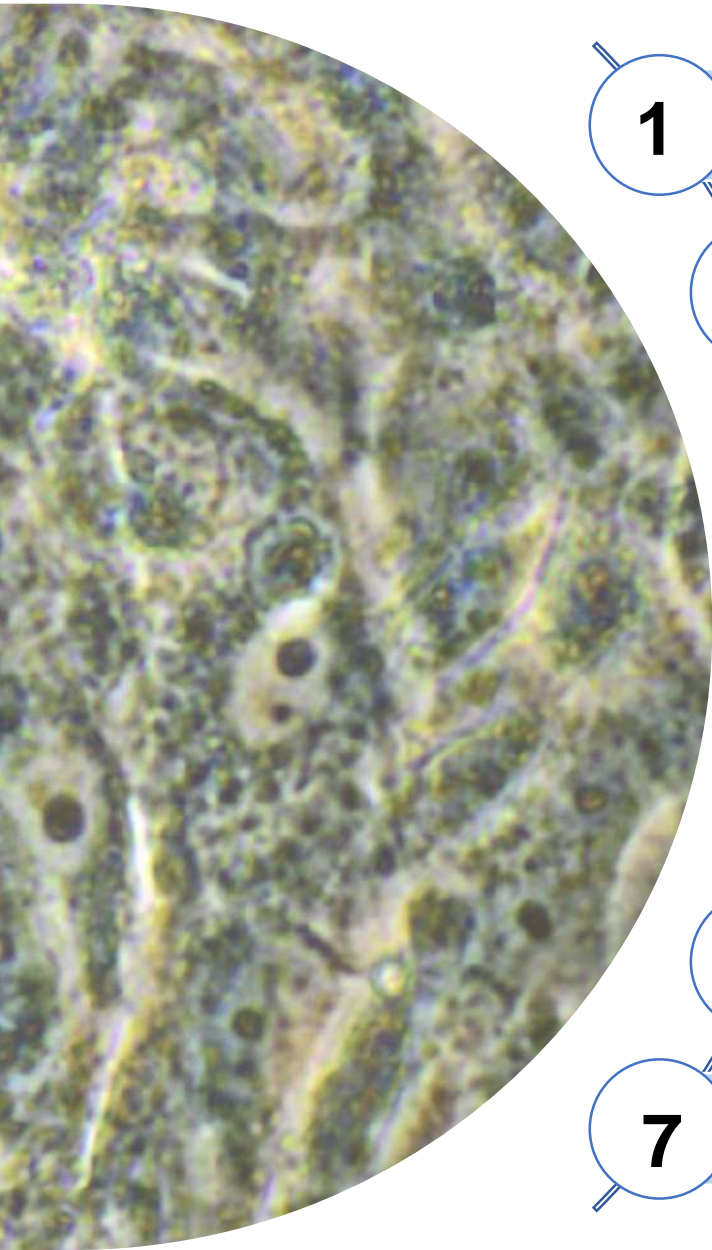
- A fast-track and low-cost strategy
- Pharmacological characteristics and safety profiles
- Posaconazole and E1224



Metabolism modifier compounds

- Metabolic coupling of intracellular pathogens with host cells is essential for successful colonization of the host
- Potential anti-parasitic treatments

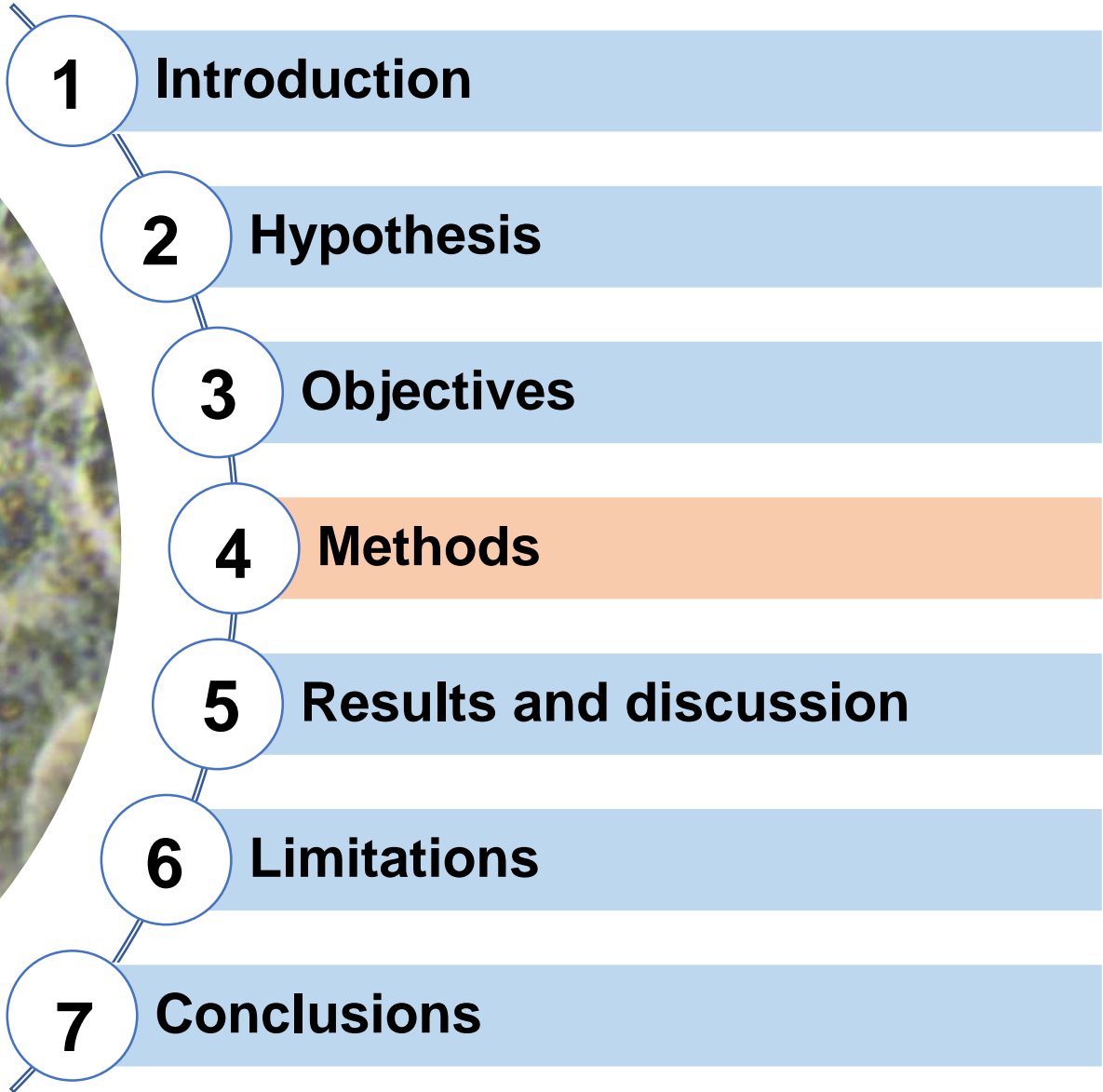
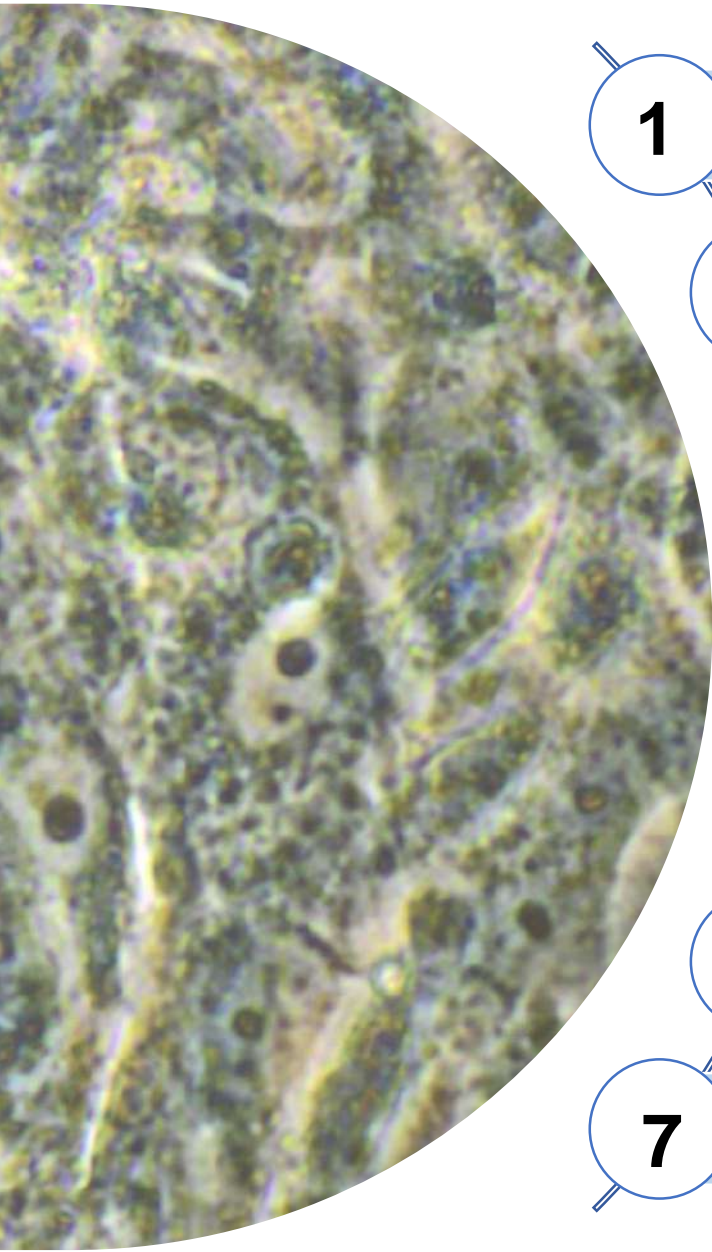
In silico target identification



3. OBJECTIVES

The main objective of this work is the identification of compounds or drugs with potent and specific activity against the parasite *T. cruzi* among different chemical collections.

- Specific Objective 1: development of a statistically robust and reproducible *in vitro* screening cascade to identify compounds specifically acting against *T. cruzi*.
- Specific Objective 2: identification of Amaryllidaceae plant extracts or alkaloids isolated from them with specific anti-*T. cruzi* activity.
- Specific Objective 3: evaluation of the anti-*T. cruzi* activity of a collection of licensed drugs through *in vitro* and *in vivo* experiments.
- Specific Objective 4: exploration of the capacity to modulate or inhibit *T. cruzi* growth of a collection of metabolism modifier compounds.
- Specific Objective 5: deciphering *T. cruzi* molecular targets and mechanisms of action of hit compounds using *in silico* molecular docking studies and the AlphaFold protein database.



4. METHODS

Amaryllidaceae plants



UNIVERSITAT DE BARCELONA

Prof. Jaume Bastida



Dra. Gabriela Feresin

Drugs for repurposing



CLÍNIC BARCELONA Hospital Universitari

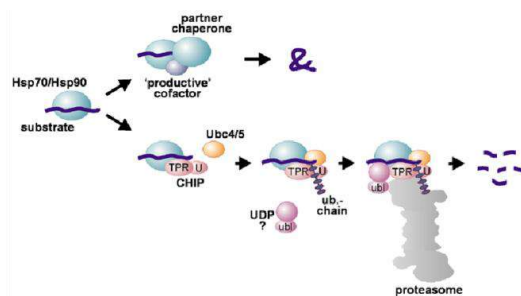
Prof. Joaquim Gascón



UNIVERSITY OF GEORGIA

Dr. Juan Bustamante

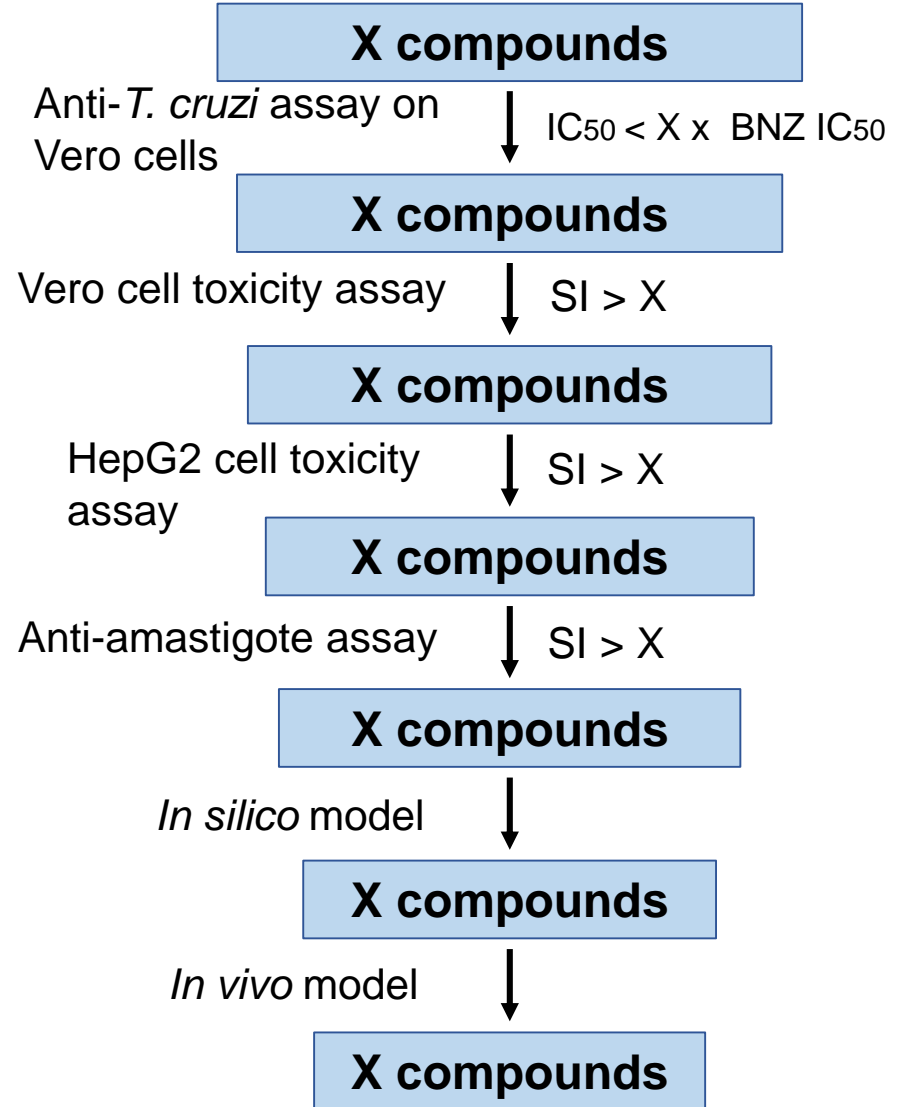
Metabolism modifier compounds



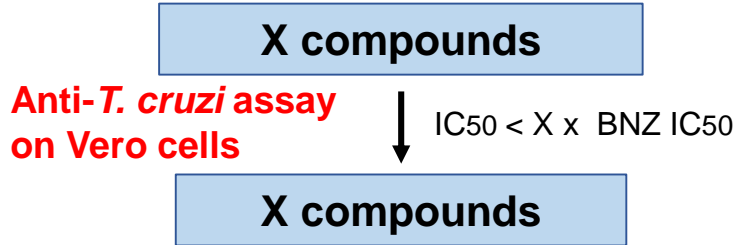
UAB

Universitat Autònoma de Barcelona

Dra. Alhelí Rodríguez-Cortes

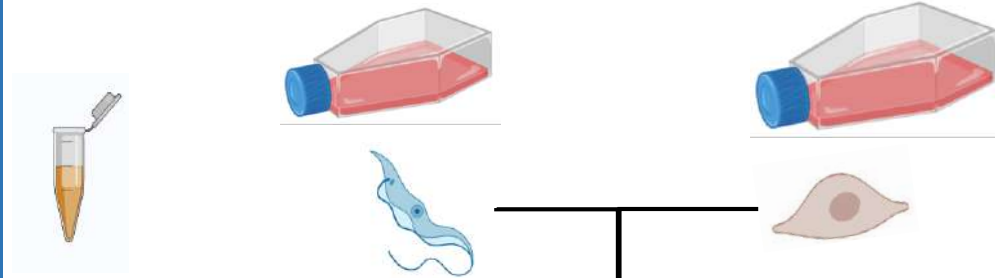


4. METHODS

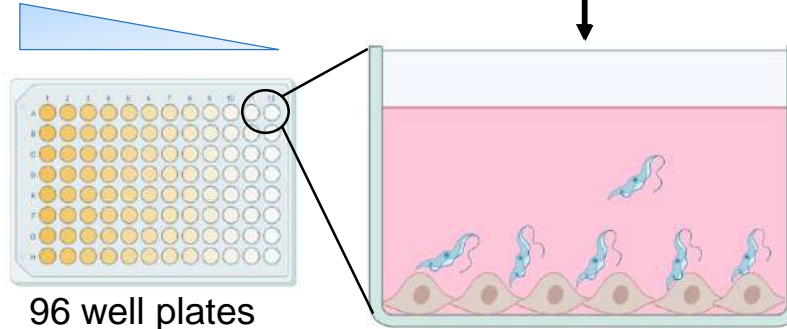


Anti-*T. cruzi* assay on Vero cells

T. cruzi Tulahuen expressing β -galactosidase reporter gene
(Fred Buckner, University of Washington, US).



MOI=1
50,000 cells per well



96 well plates

Day 0-1

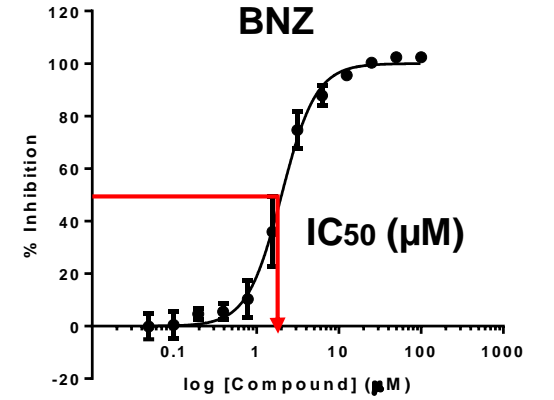
Day 1

Stop assay
by freezing

Day 5

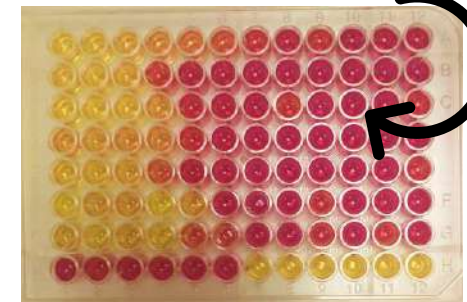
Day 6

Incubation 37° C
5% CO₂

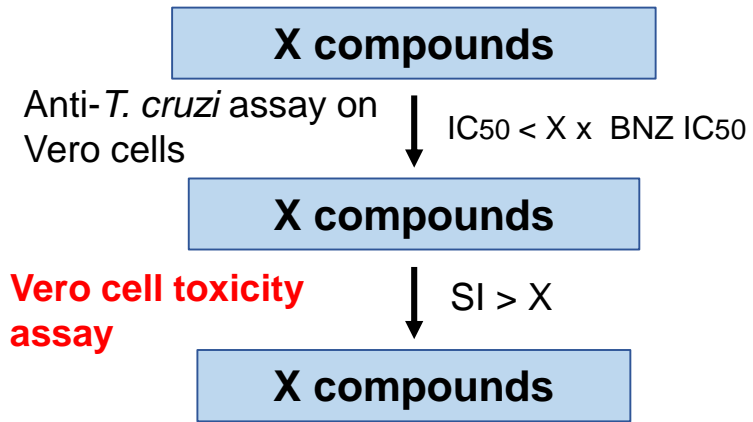


Colored substrate (CPRG)

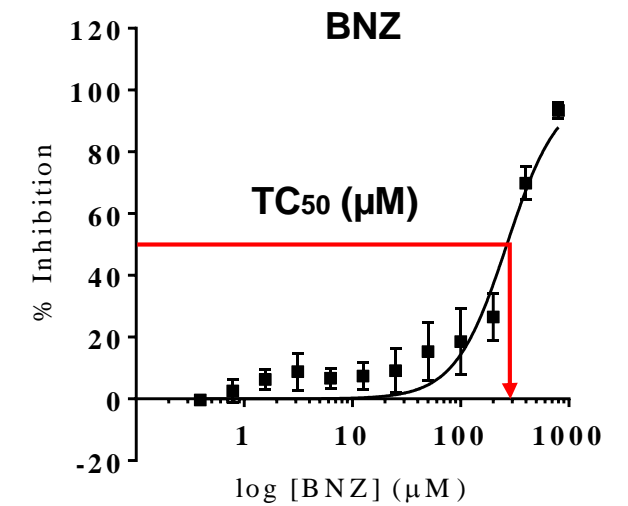
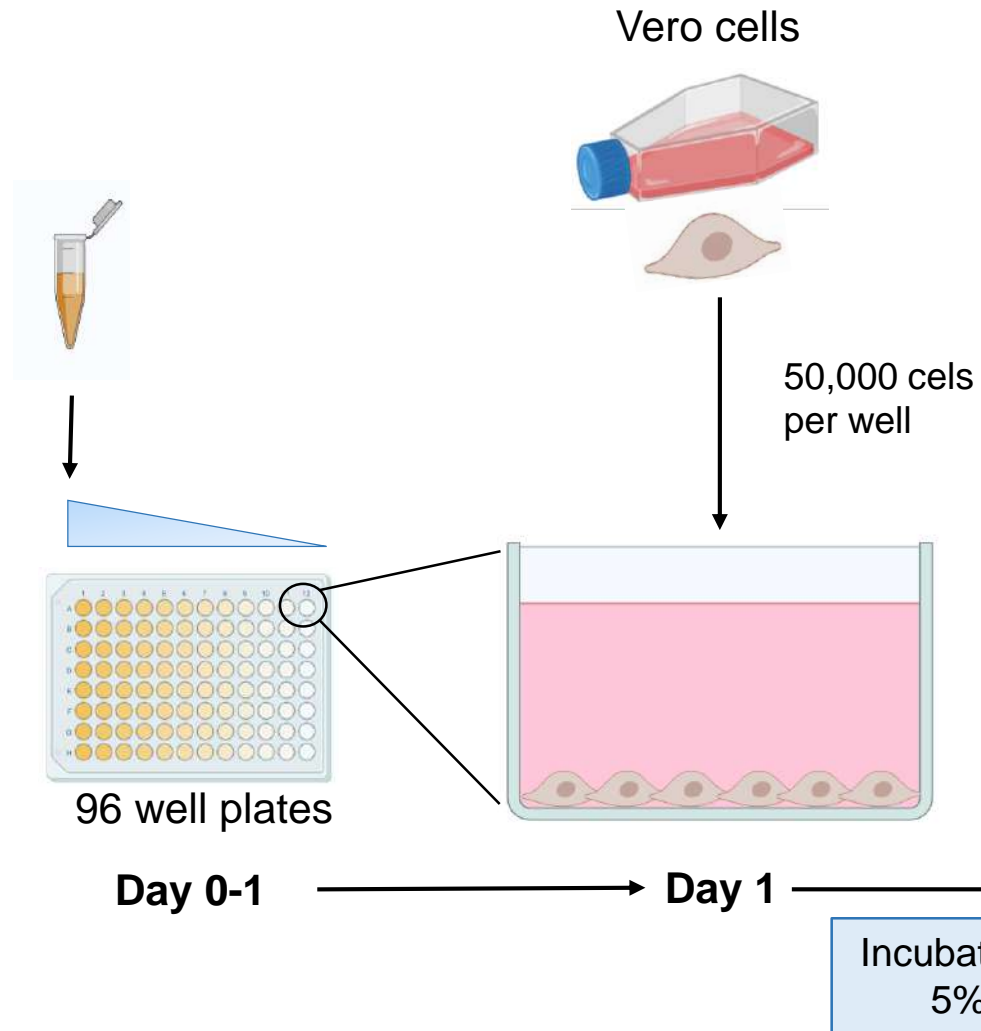
Principle of action: enzymatic activity increase indicates *T. cruzi* replication.



4. METHODS

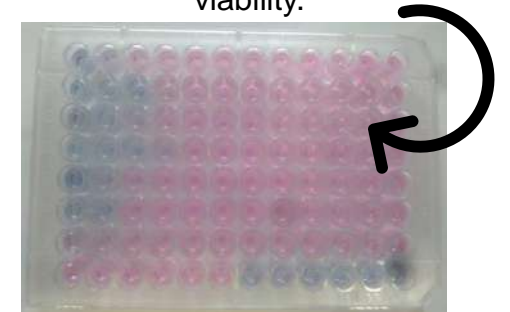


Vero cell toxicity assay

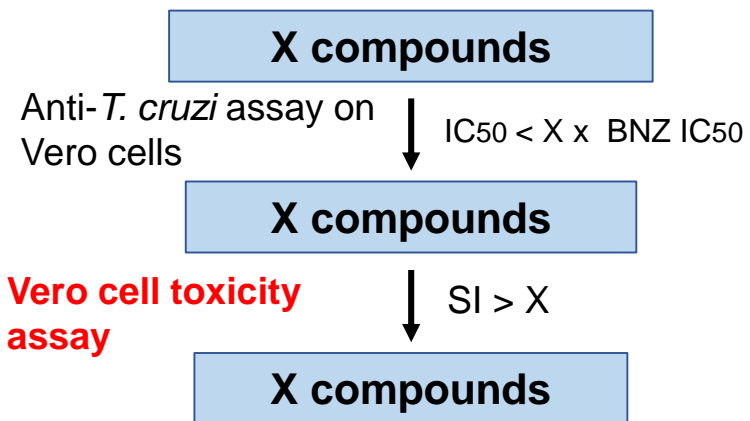


Fluorescent substrate (AlamarBlue)

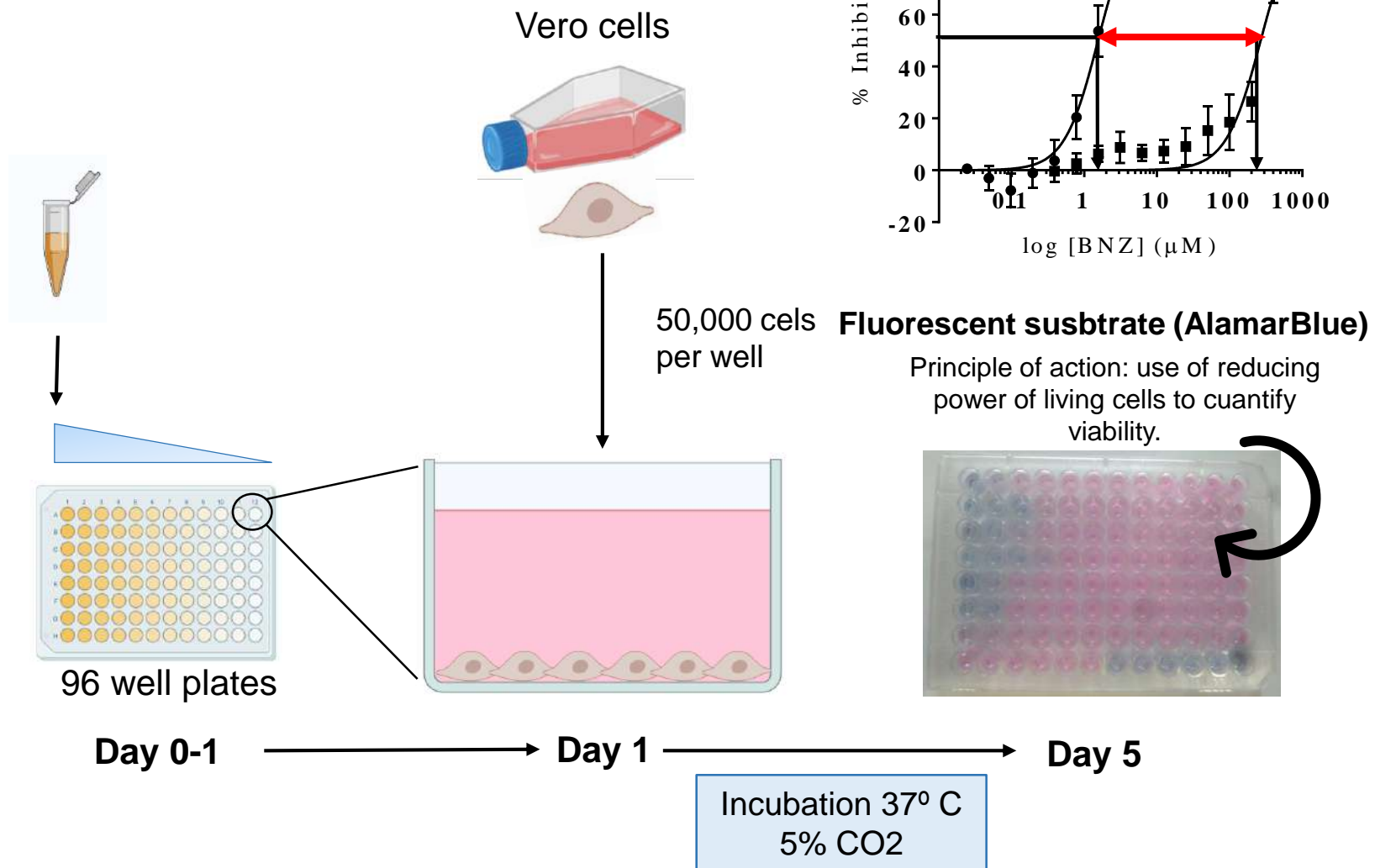
Principle of action: use of reducing power of living cells to quantify viability.



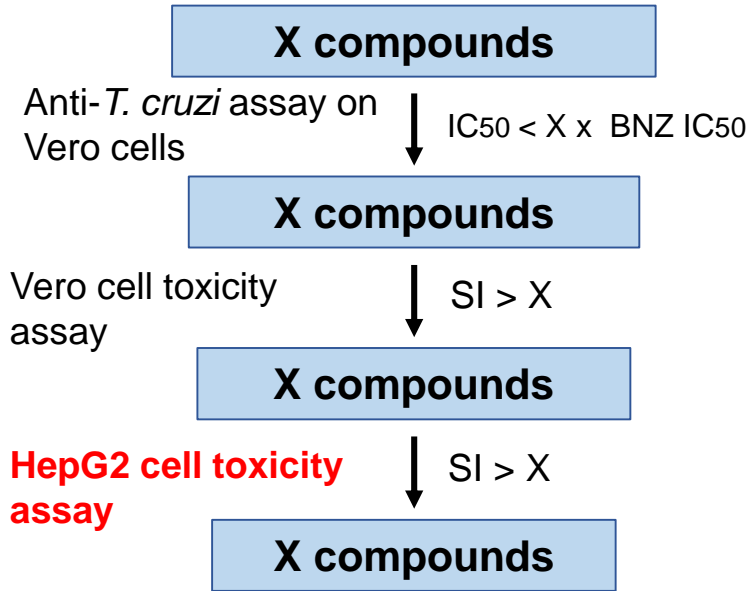
4. METHODS



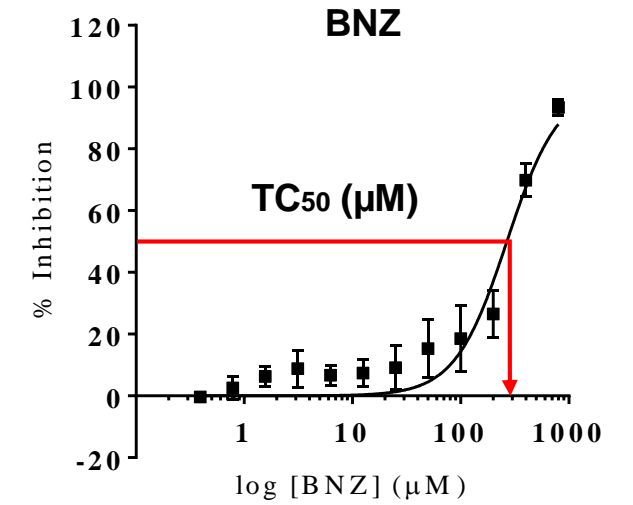
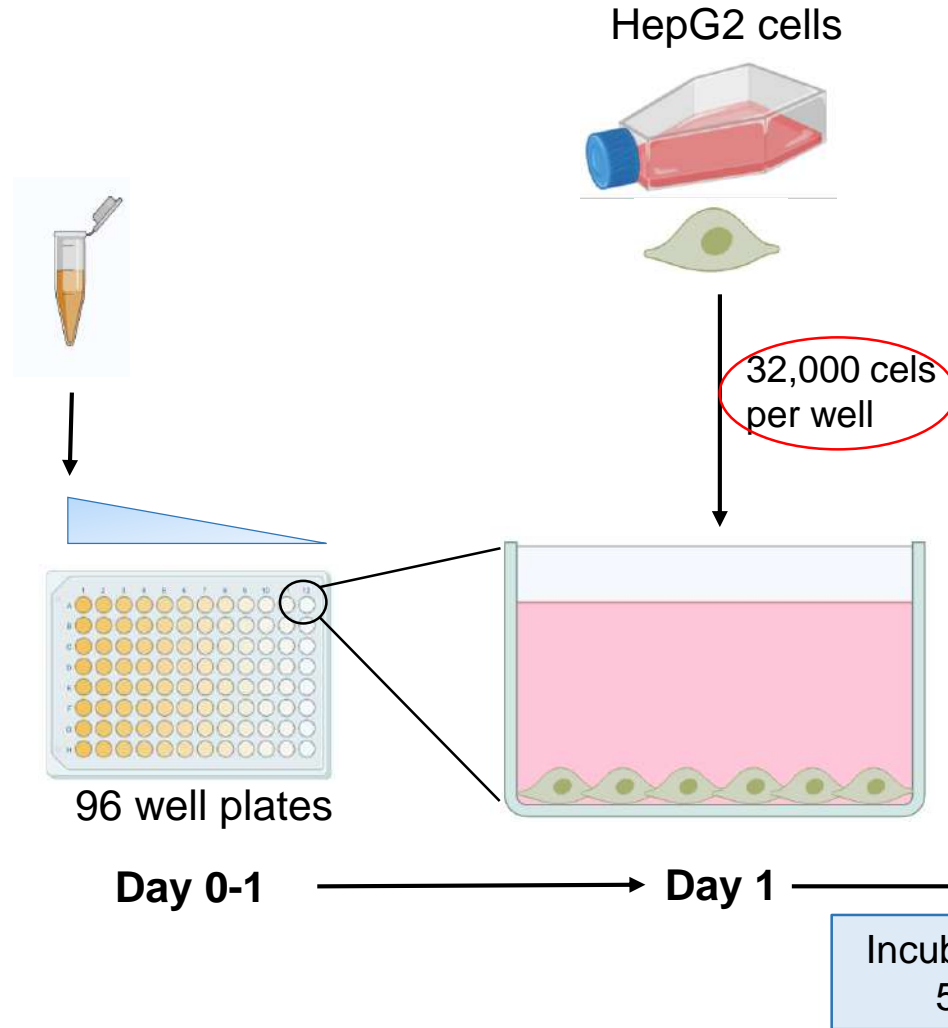
Vero cell toxicity assay



4. METHODS



HepG2 cell toxicity assay



Fluorescent substrate (AlamarBlue)

Principle of action: use of reducing power of living cells to quantify viability.



4. METHODS

X compounds

Anti-*T. cruzi* assay on Vero cells
↓ $IC_{50} < X \times BNZ IC_{50}$

X compounds

Vero cell toxicity assay
↓ $SI > X$

X compounds

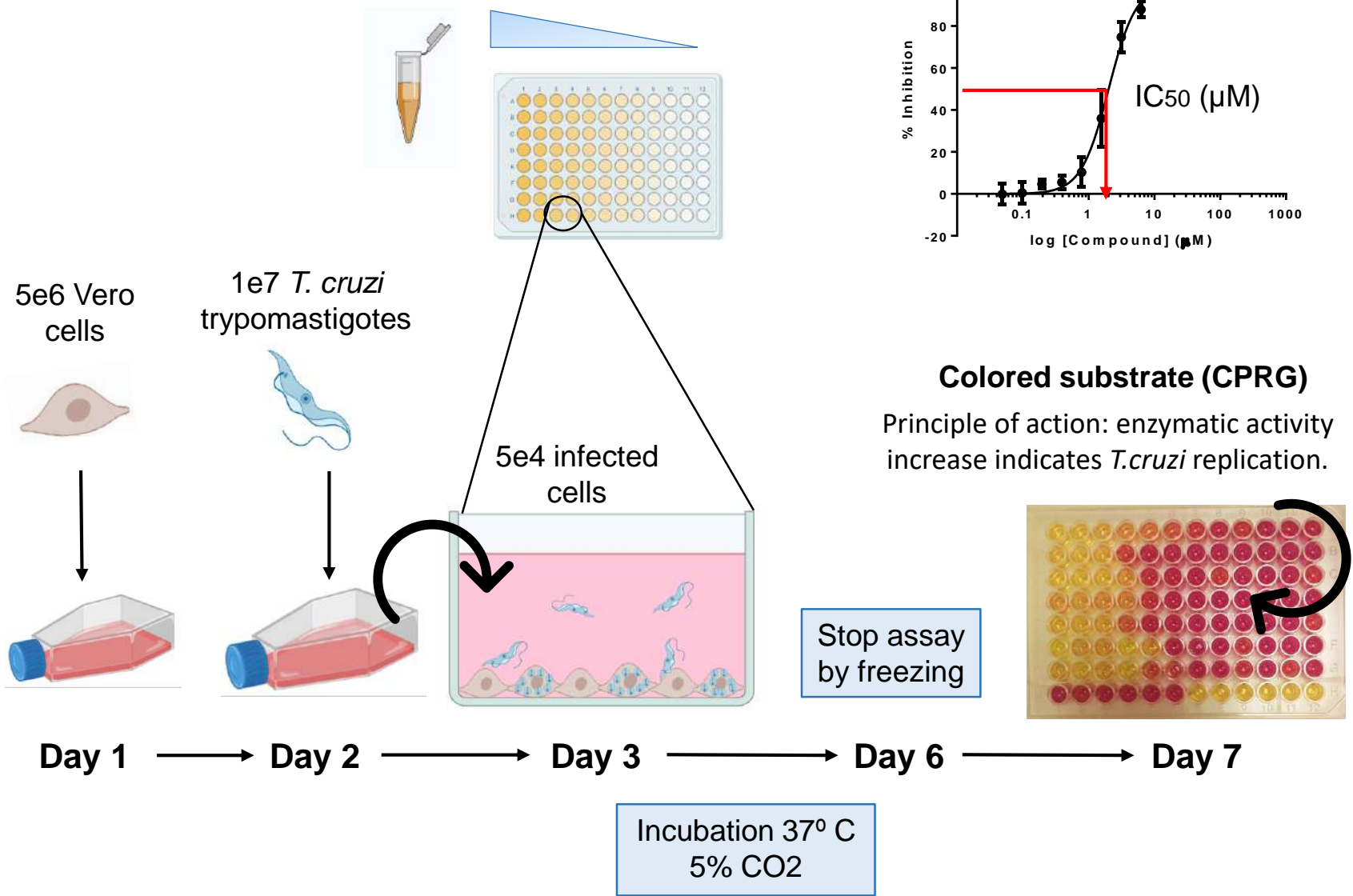
HepG2 cell toxicity assay
↓ $SI > X$

X compounds

Anti-amastigote assay ↓ $SI > X$

X compounds

Anti-amastigote assay



4. METHODS

X compounds

Anti-*T. cruzi* assay on Vero cells
↓ $IC_{50} < X \times BNZ IC_{50}$

X compounds

Vero cell toxicity assay
↓ $SI > X$

X compounds

HepG2 cell toxicity assay
↓ $SI > X$

X compounds

Anti-amastigote assay
↓ $SI > X$

X compounds

NYU *In vivo* model

X compounds

In vivo assays

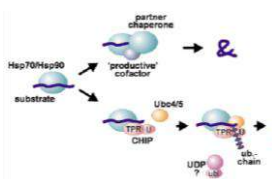
T. cruzi Brazil strain (DTU I) expressing firefly luciferase



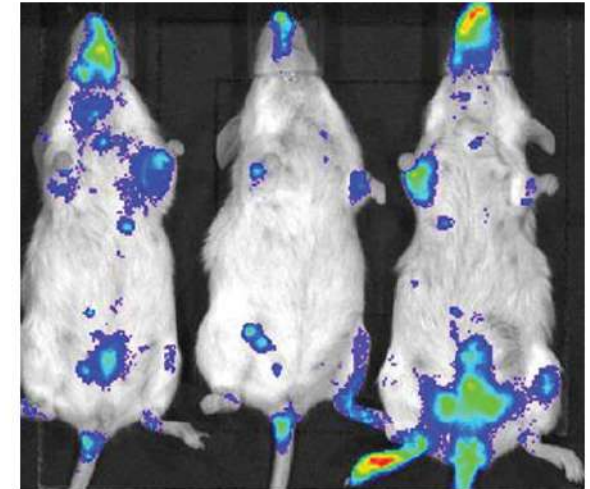
Acute *in vivo* model



Balb/c mice

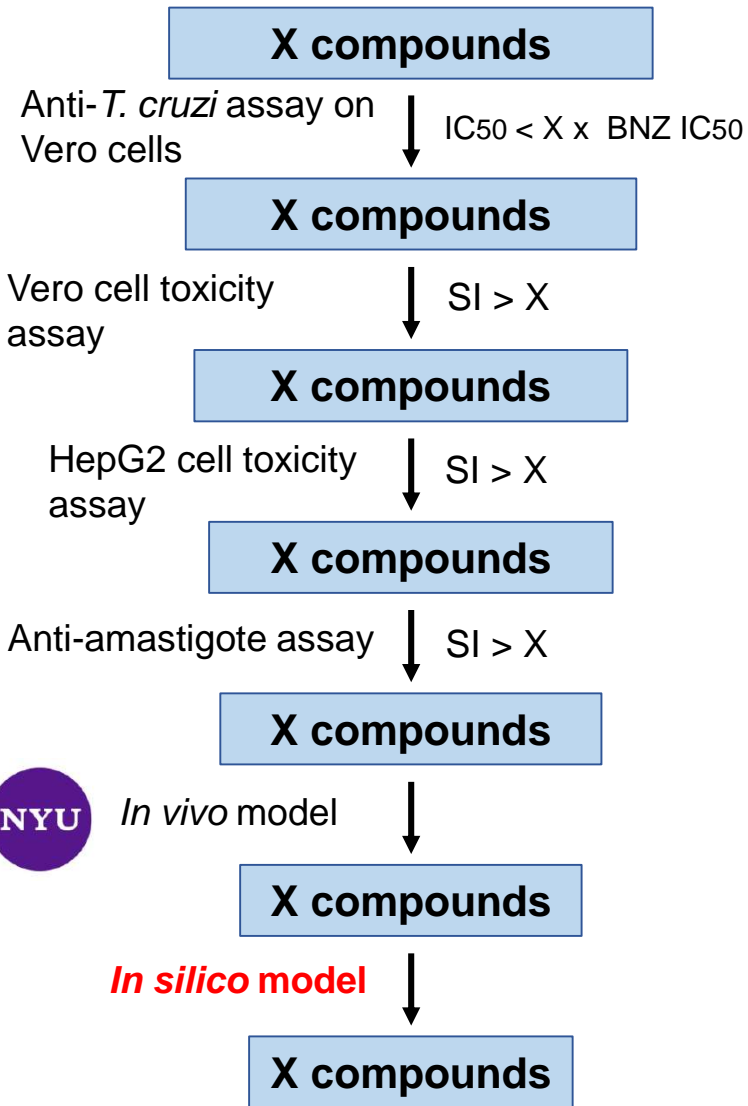


Chronic *in vivo* model



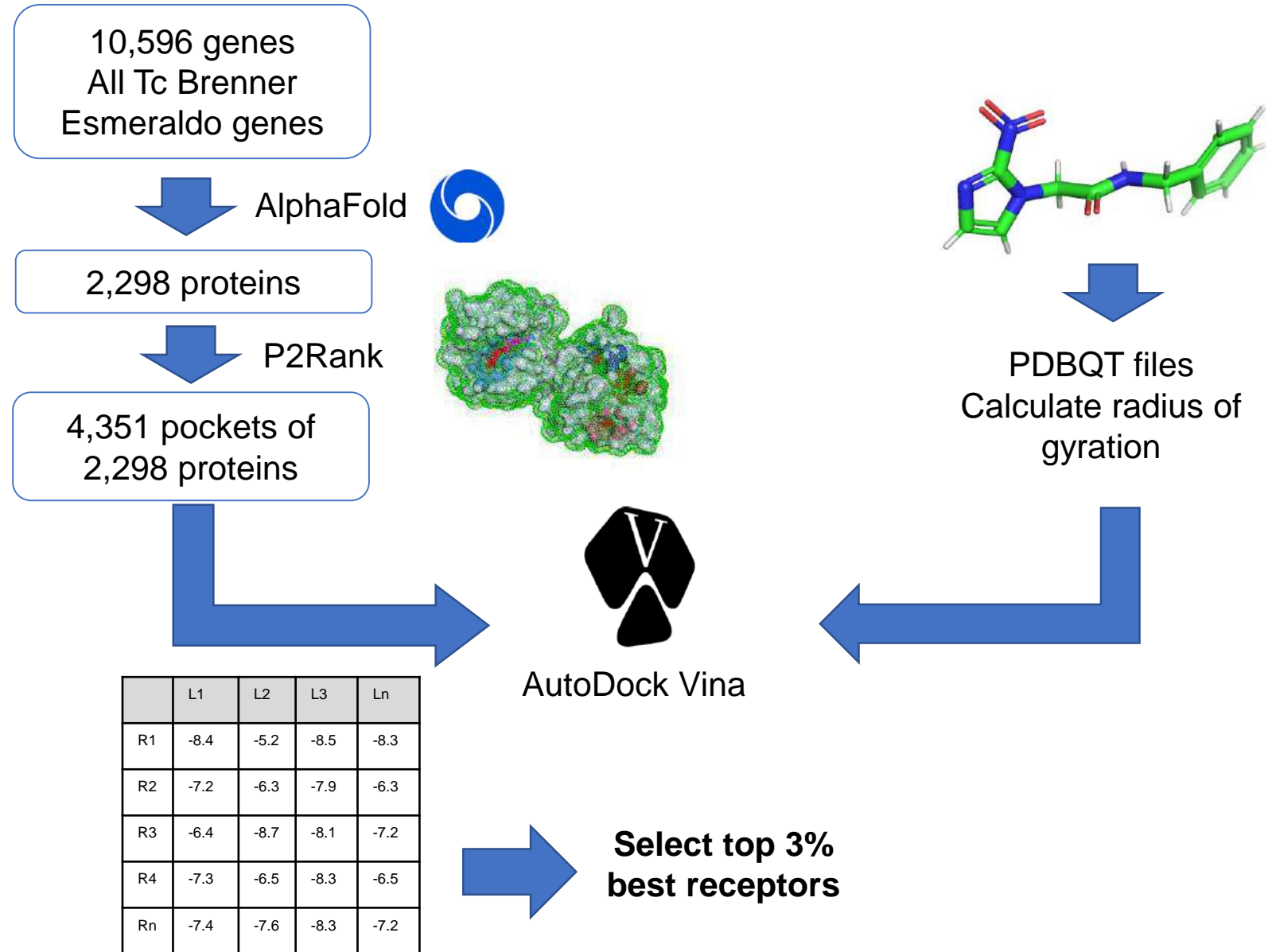
Dra. Ana Rodriguez

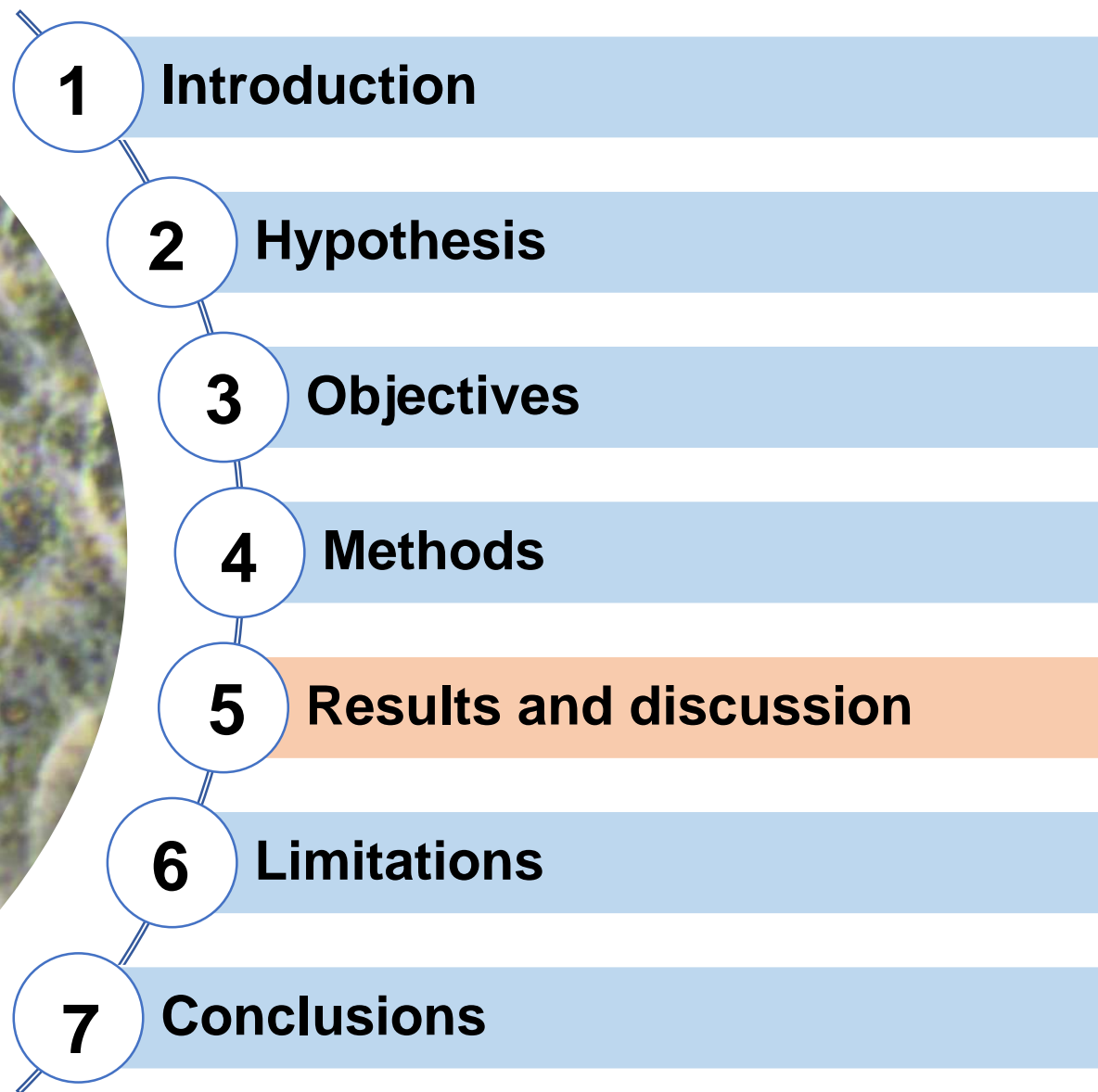
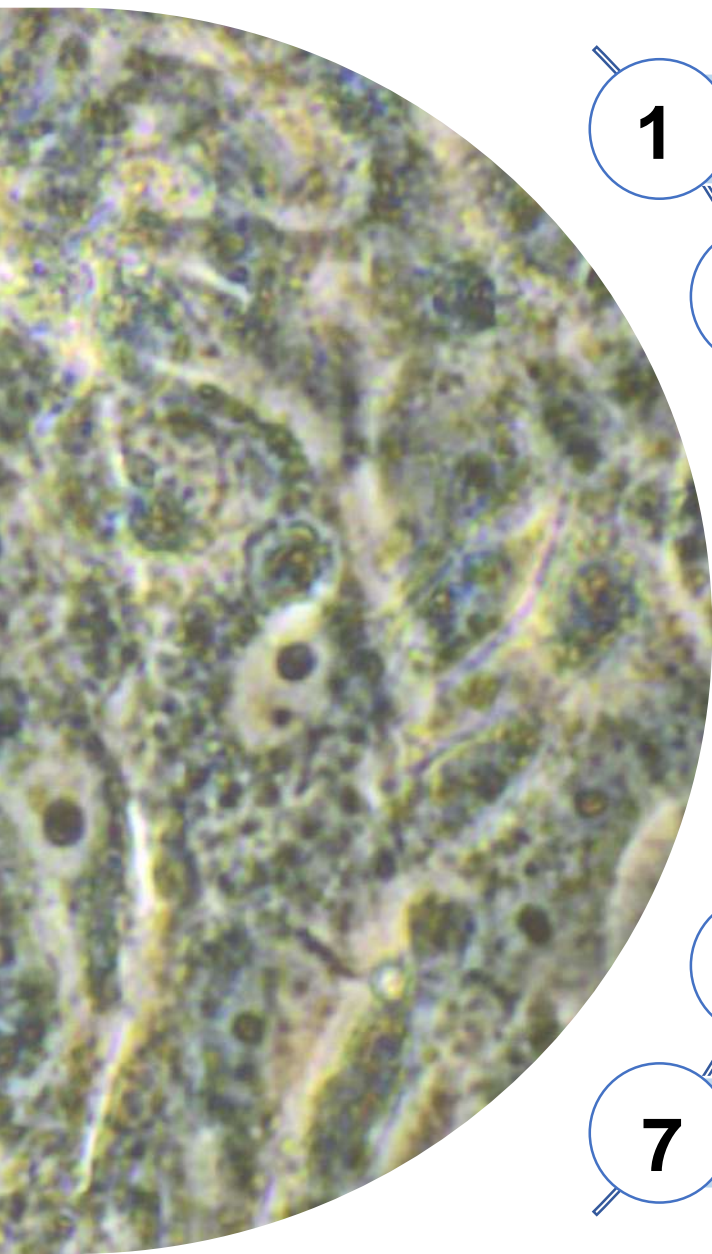
4. METHODS



In silico target identification

Ros-Lucas et al., 2022





- Chapter I. Amaryllidaceae plants
- Chapter II. Drug repurposing
- Chapter III. Metabolism modifier compounds
- Chapter IV. *In silico* target identification

Martínez-Peinado et al. *Parasites Vectors* (2020) 13:299
<https://doi.org/10.1186/s13071-020-04171-6>

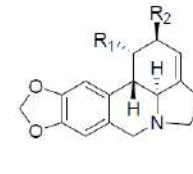
Parasites & Vectors

RESEARCH

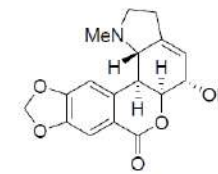
Open Access

Amaryllidaceae alkaloids with anti-*Trypanosoma cruzi* activity

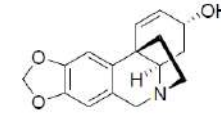
Nieves Martínez-Peinado¹, Nuria Cortes-Serra¹, Laura Torras-Claveria², María-Jesús Pinazo¹, Joaquim Gascon¹, Jaume Bastida² and Julio Alonso-Padilla^{1*}



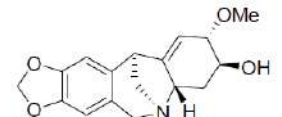
1: Lycorine R₁=R₂=OH
 9: 1-O-acetylcarranine R₁=OAc, R₂=H



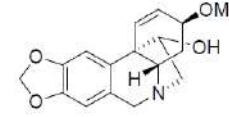
2: Hippeastrine



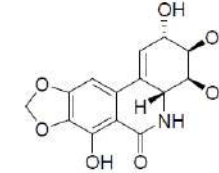
3: Crinine



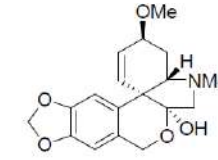
7: Montanine



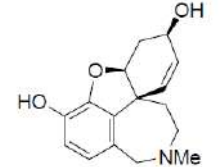
4: Haemanthamine



5: Narciclasine

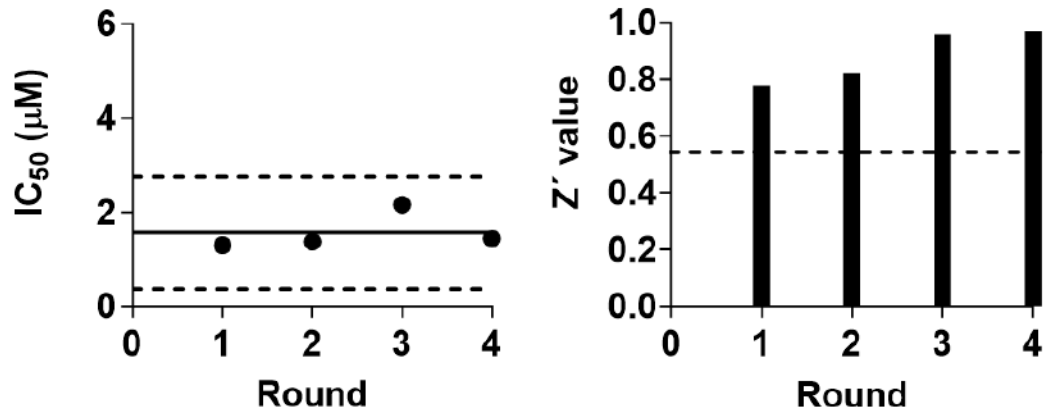


6: Tazettine

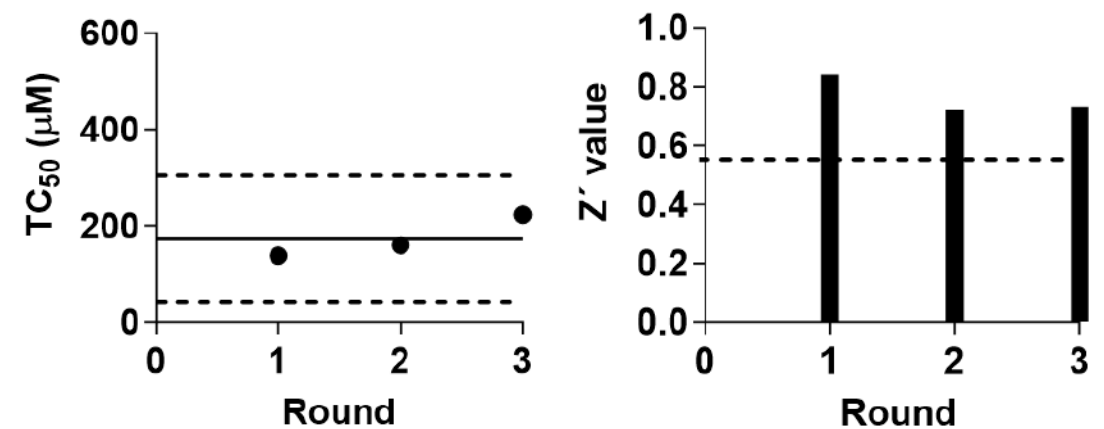


8: Sanguinine

Anti-*T. cruzi* assay



Vero cell toxicity assay



Z' parameter to assess the reproducibility and quality (0.5-1)

Martinez-Peinado et al. *Parasites Vectors* (2020) 13:299
<https://doi.org/10.1186/s13071-020-04171-6>

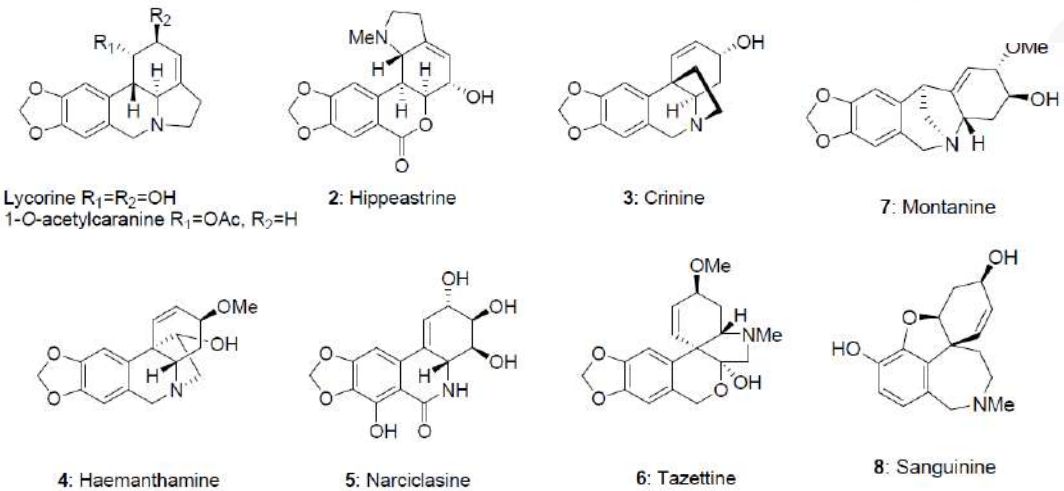
Parasites & Vectors

RESEARCH

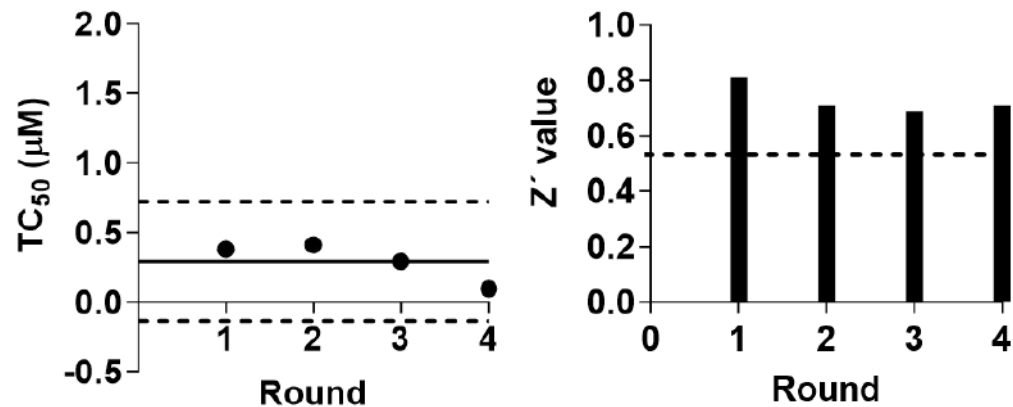
Open Access

Amaryllidaceae alkaloids with anti-*Trypanosoma cruzi* activity

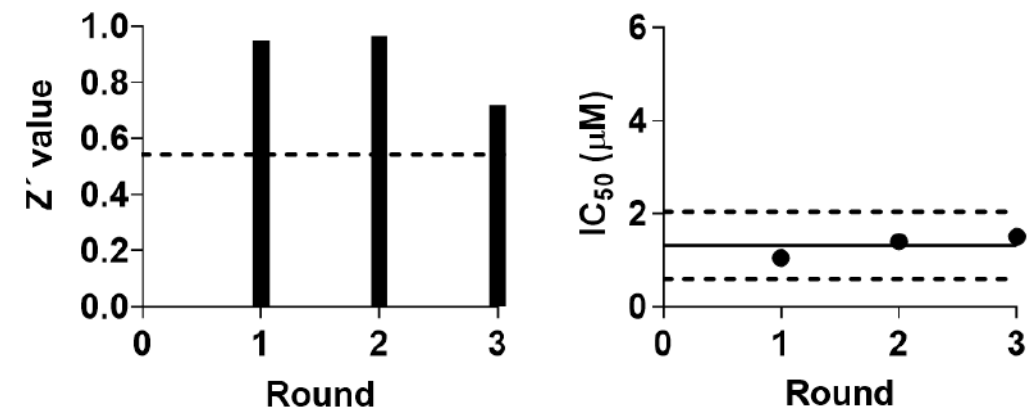
Nieves Martínez-Peinado¹, Nuria Cortes-Serra¹, Laura Torras-Claveria², María-Jesús Pinazo¹, Joaquim Gascon¹, Jaume Bastida² and Julio Alonso-Padilla^{1*}



HepG2 cell toxicity assay

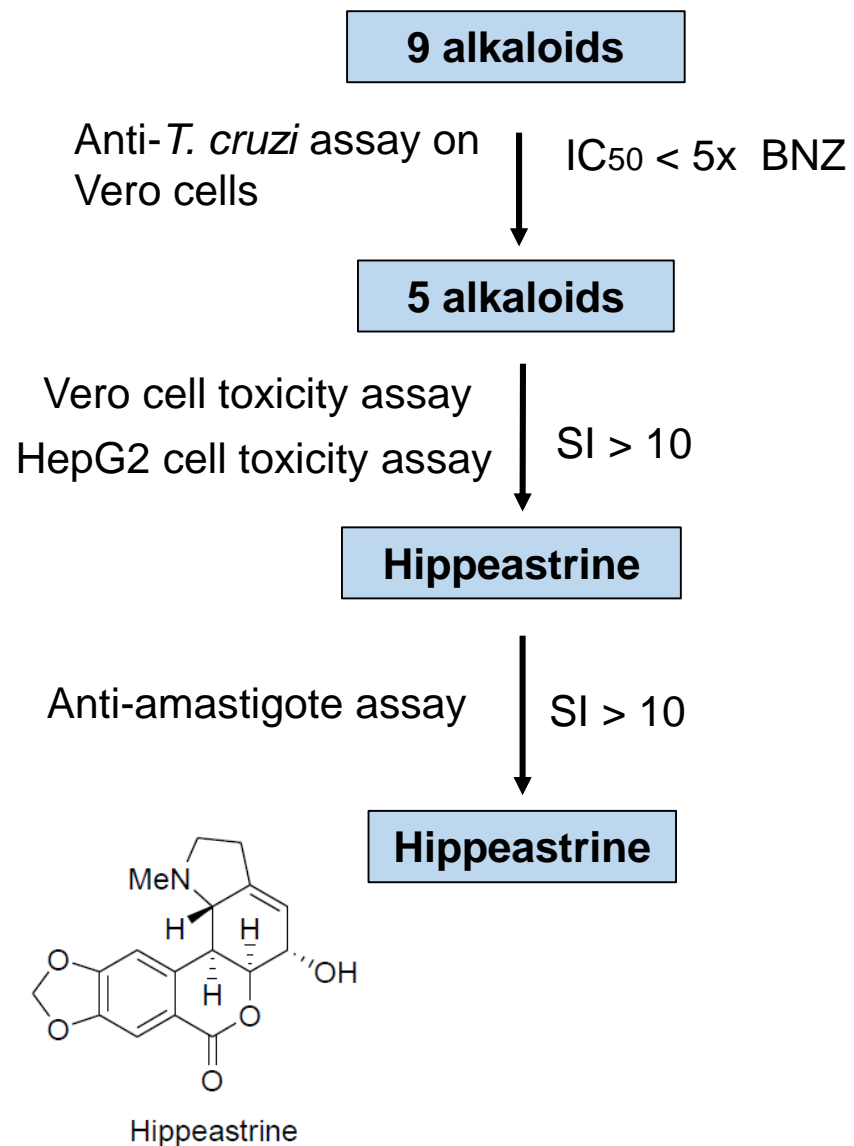


Anti-amastigote assay



Z' parameter to assess the reproducibility and quality (0.5-1)

5. RESULTS AND DISCUSSION. CHAPTER I



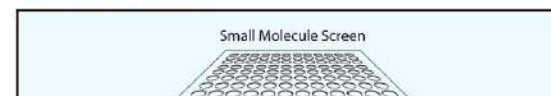
Alkaloid	IC_{50} (μ M)	TC_{50}^a (μ M)	SI^a	TC_{50}^b (μ M)	SI^b	IC_{50} (μ M)	SI^a	SI^b
BNZ	1.56	173.4	111.2	168.76	108.2	1.20	144.5	140.6
Lycorine	0.70	5.21	7.5	21.87	31.2			
Hippeastrine[#]	3.63	45.99	12.7	128.10	35.2	3.31	13.8	38.7
Haemanthamine	1.59	11.52	7.3	42.48	26.7			
Narciclasine	0.49	0.66	1.3	2.73	5.5			
Montanine	1.99	5.04	2.5	46.10	23.1			

Short Article

Cell Stem Cell

High-Content Screening in hPSC-Neural Progenitors Identifies Drug Candidates that Inhibit Zika Virus Infection in Fetal-like Organoids and Adult Brain

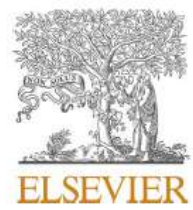
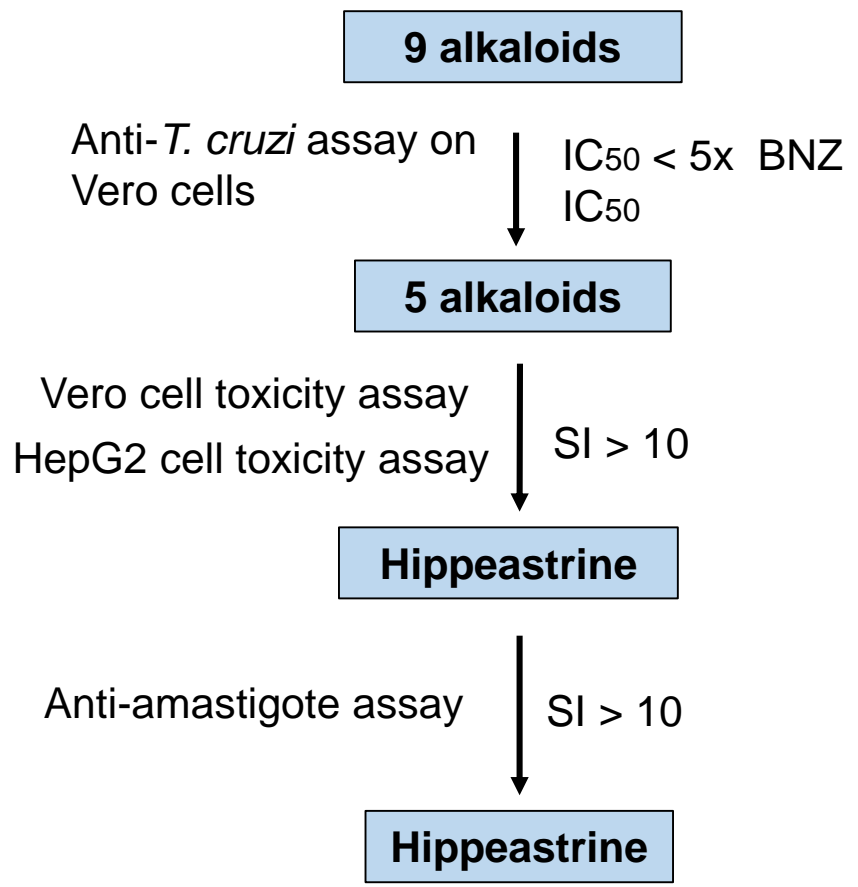
Graphical Abstract



Authors

Ting Zhou, Lei Tan, Gustav Y. Cederquist, ..., Todd Evans, Lorenz Studer, Shuibing Chen

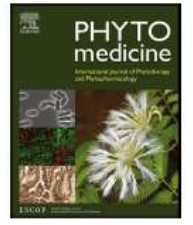
5. RESULTS AND DISCUSSION. CHAPTER I



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Phytomedicine

journal homepage: www.elsevier.com/locate/phymed

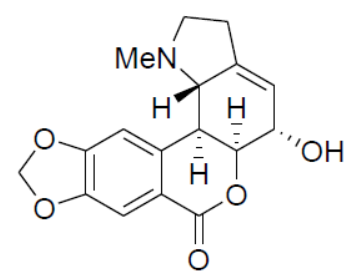


Original Article

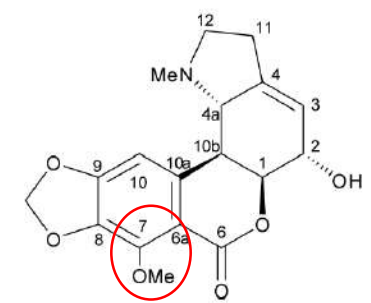
Candimine from *Hippeastrum escoipense* (Amaryllidaceae): Anti-*Trypanosoma cruzi* activity and synergistic effect with benznidazole

Javier E. Ortiz ^{a,b,1}, Mauricio Piñeiro ^{a,b,1}, Nieves Martinez-Peinado ^{c,d}, Patricia Barrera ^e, Miguel Sosa ^e, Jaume Bastida ^d, Julio Alonso-Padilla ^{c,f,§}, Gabriela E. Feresin ^{a,b,§,*}

^a Instituto de Biotecnología, Facultad de Ingeniería, Universidad Nacional de San Juan, Av. Libertador General San Martín, 1109 O San Juan, Argentina
^b Consejo Nacional de Investigaciones Científicas y Técnicas (CONICET), CCT CONICET San Juan, Argentina
^c Barcelona Institute for Global Health (ISGlobal), Hospital Clinic-University of Barcelona, 08036 Barcelona, Spain
^d Departament de Biologia, Sanitat i Medi Ambient, Facultat de Farmàcia i Ciències de l'Alimentació, Universitat de Barcelona, 08028 Barcelona, Spain
^e Facultad de Ciencias Médicas, Instituto de Histología y Embriología "Dr. Mario H. Burgos", Universidad Nacional de Cuyo-CONICET, CC 56 (5500) Mendoza, Argentina
^f CIBER de Enfermedades Infecciosas, Instituto de Salud Carlos III (CIBERINFEC, ISCIII), Madrid, Spain



Hippeastrine
 IC₅₀=3.63, SI=12.7
 IC₅₀(amastigote)=3.31, SI=13.8



Candimine
 IC₅₀=2.49, SI=102.57
 IC₅₀(amastigote)=1.60, SI=159.63

5. RESULTS AND DISCUSSION. CHAPTER I

Martínez-Peinado et al. *Parasites Vectors* (2021) 14:337
<https://doi.org/10.1186/s13071-021-04837-9>

Parasites & Vectors

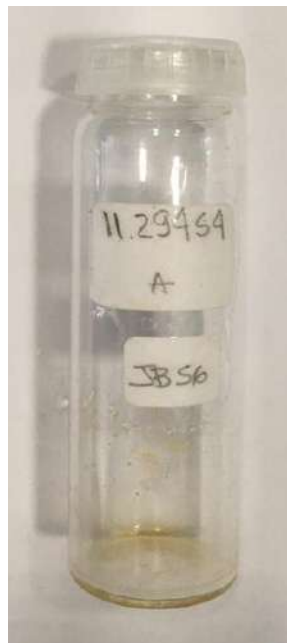
RESEARCH

Open Access

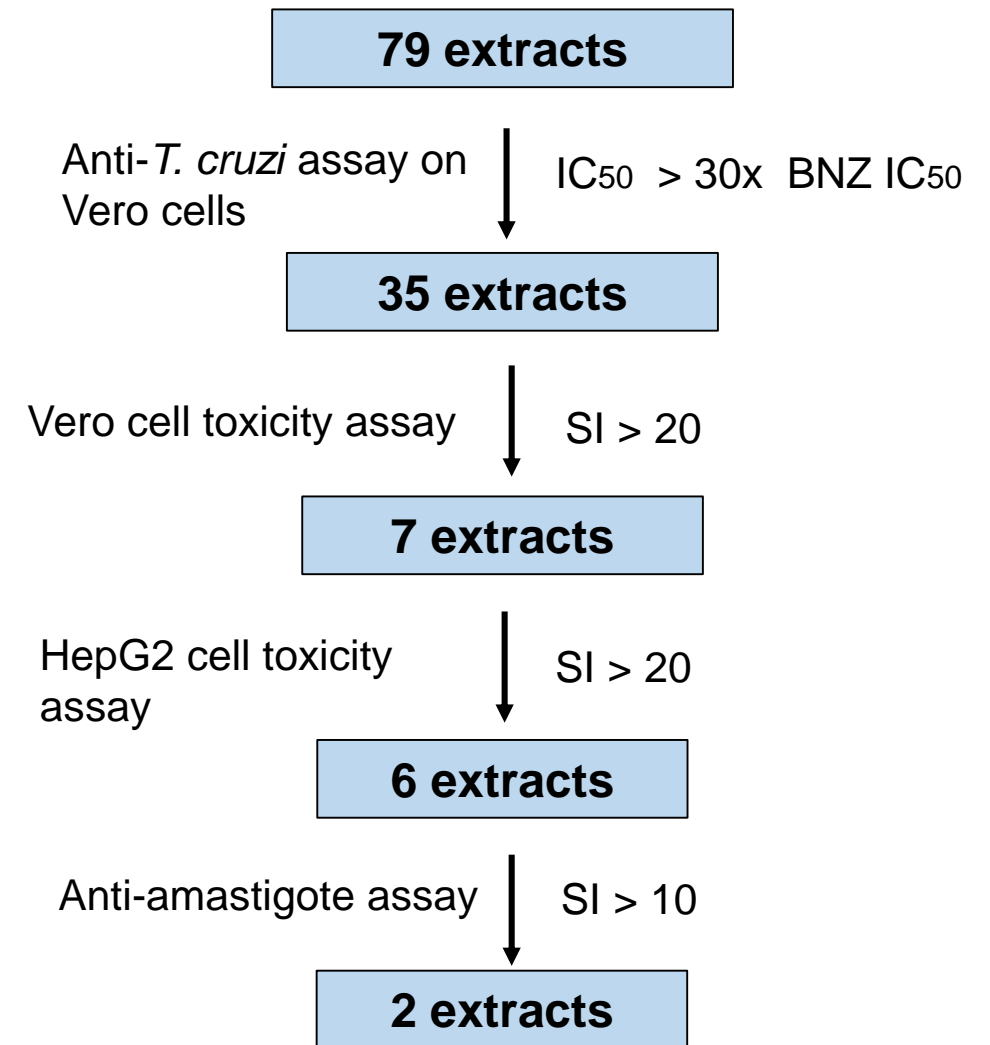
Amaryllidaceae plants: a potential natural resource for the treatment of Chagas disease



Nieves Martínez-Peinado¹, Nuria Cortes-Serra¹, Luciana R. Tallini^{2,3}, Maria-Jesus Pinazo¹, Joaquim Gascon¹, Jaume Bastida^{2*} and Julio Alonso-Padilla^{1*}



Prof. Jaume Bastida



5. RESULTS AND DISCUSSION. CHAPTER I

Martínez-Peinado et al. *Parasites Vectors* (2021) 14:337
<https://doi.org/10.1186/s13071-021-04837-9>

Parasites & Vectors

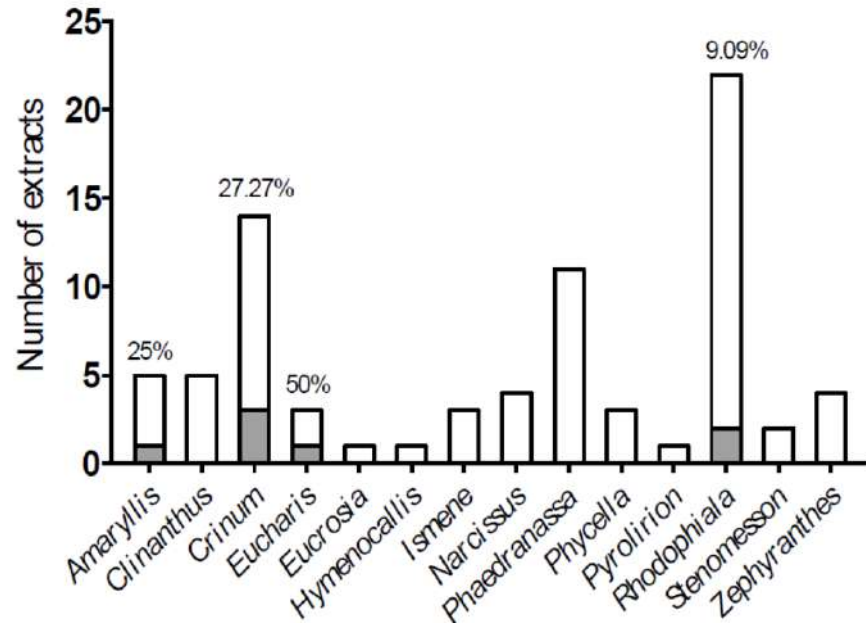
RESEARCH

Open Access

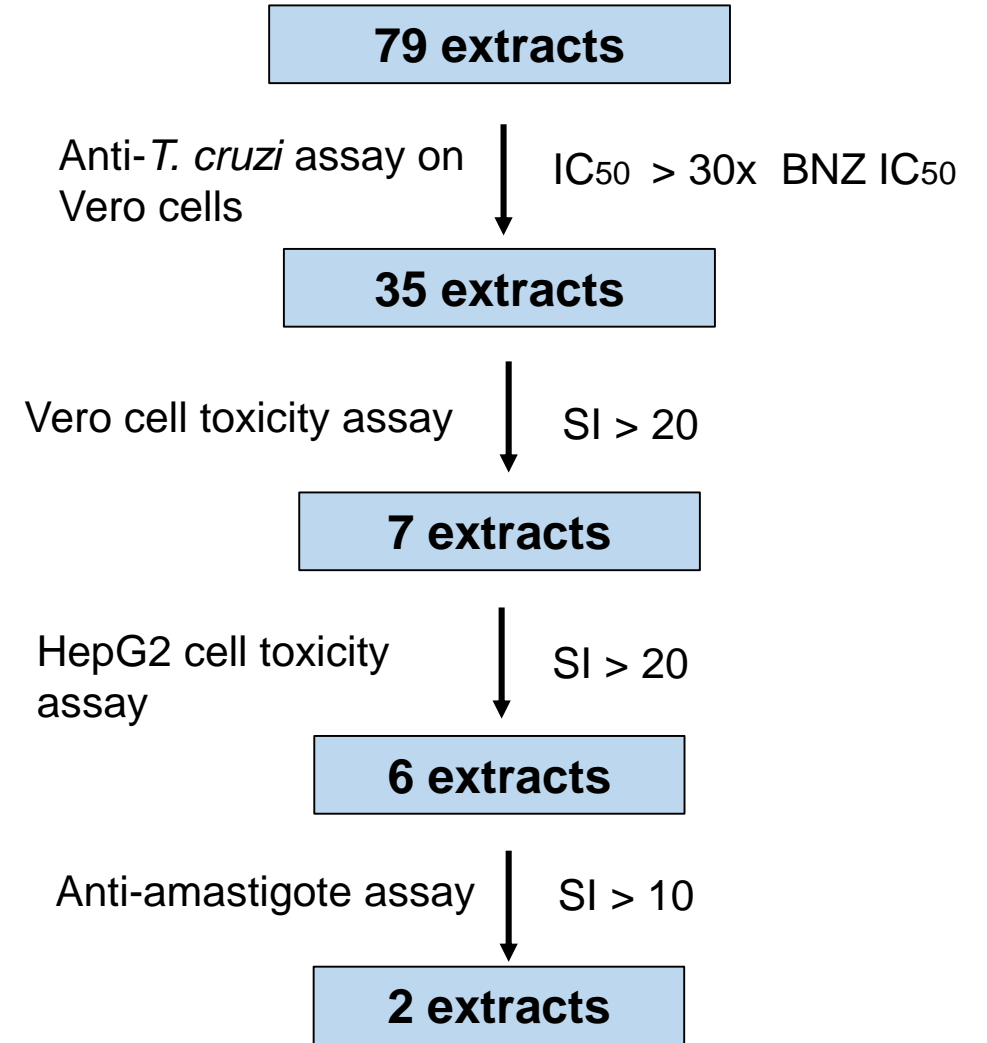


Amaryllidaceae plants: a potential natural resource for the treatment of Chagas disease

Nieves Martínez-Peinado¹, Nuria Cortes-Serra¹, Luciana R. Tallini^{2,3}, Maria-Jesus Pinazo¹, Joaquim Gascon¹, Jaume Bastida^{2*} and Julio Alonso-Padilla^{1*}



Distribution of anti-*T. cruzi* selective extracts per plant genus



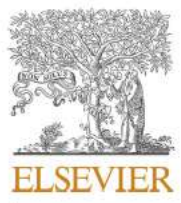
5. RESULTS AND DISCUSSION. CHAPTER I

Extract number	Plant species of origin	Country of collection	Part of the plant*	Vero cells assay			HepG2 cell assay	Anti-amastigote assay	
				IC ₅₀ (ppm)	TC ₅₀ (ppm)	SI	TC ₅₀ (ppm)	IC ₅₀ (ppm)	SI
BNZ	-			0.40	69.60	174	51.47	0.53	131.4
51	<i>Amaryllis belladonna</i>	Chile	B	1.65	41.97	25.4	128.2	37.29	1.12
81	<i>Crinum amabile</i>	Venezuela	B	5.42	211.5	38.9	266.9	25.86	8.2
93	<i>Crinum amabile</i>	Ecuador	B	2.21	60.69	27.5	111.3	20.57	2.9
56	<i>Crinum erubescens</i>	Bolivia	B	9.50	234.7	24.7	678.3	11.10	21.1
101	<i>Eucharis formosa</i>	Ecuador	B	9.71	346.7	35.7	778.9	26.93	12.9
23	<i>Rhodophiala andicola</i>	Chile	B	6.20	134.9	21.8	77.37	-	-
24	<i>Rhodophiala andicola</i>	Chile	AP	6.13	228.4	37.3	188.1	10.18	22.4

5. RESULTS AND DISCUSSION. CHAPTER I

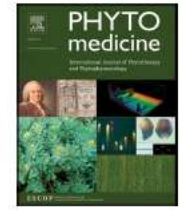
Phytomedicine 101 (2022) 154126

Contents lists available at [ScienceDirect](https://www.sciencedirect.com)



Phytomedicine

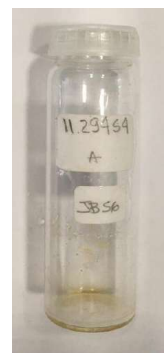
journal homepage: www.elsevier.com/locate/phymed



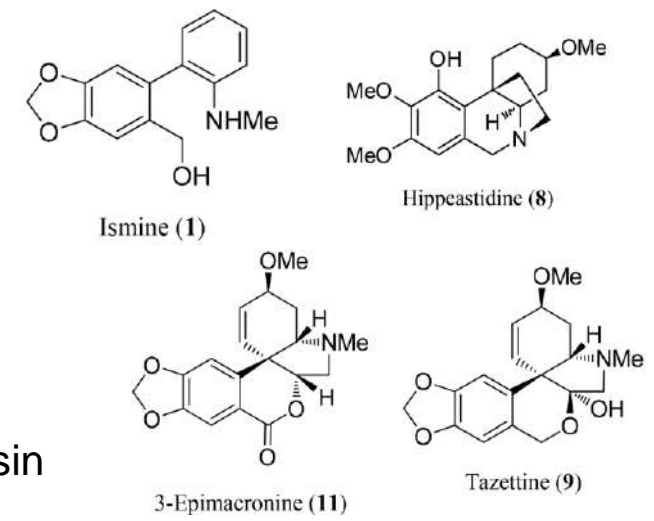
Original Article

Anti-*Trypanosoma cruzi* activity of alkaloids isolated from *Habranthus brachyandrus* (Amaryllidaceae) from Argentina

Nieves Martinez-Peinado ^{a,b,1}, Javier E. Ortiz ^{c,d,1}, Nuria Cortes-Serra ^{a,b}, Maria Jesus Pinazo ^{a,b}, Joaquim Gascon ^{a,b}, Alejandro Tapia ^c, German Roitman ^e, Jaume Bastida ^f, Gabriela E. Feresin ^{c,d,*}, Julio Alonso-Padilla ^{a,b,**}

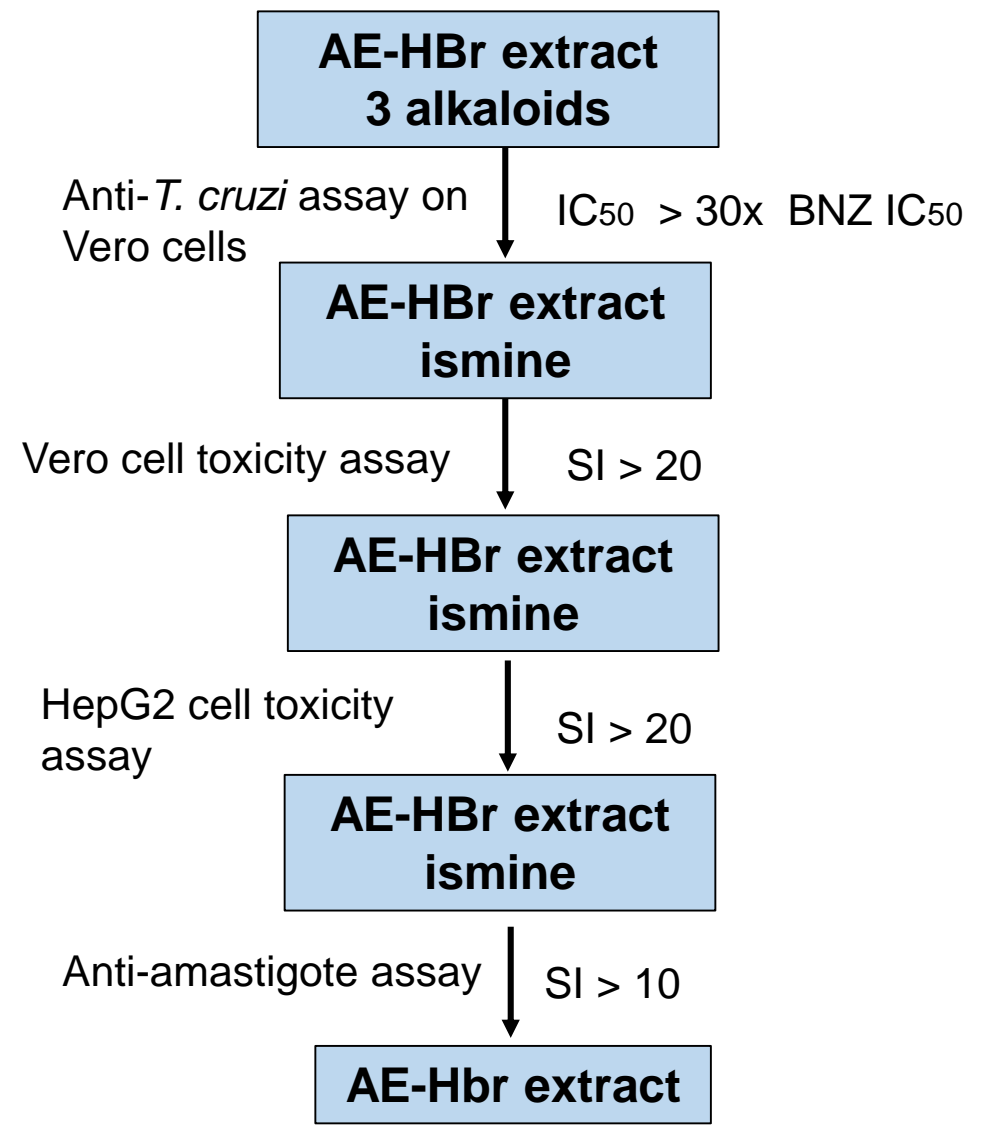


GC-MS spectra, NMR
Chromatographic
techniques

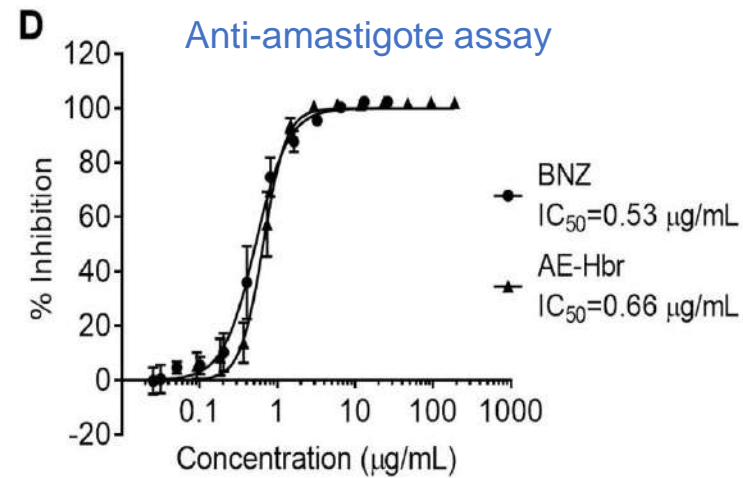
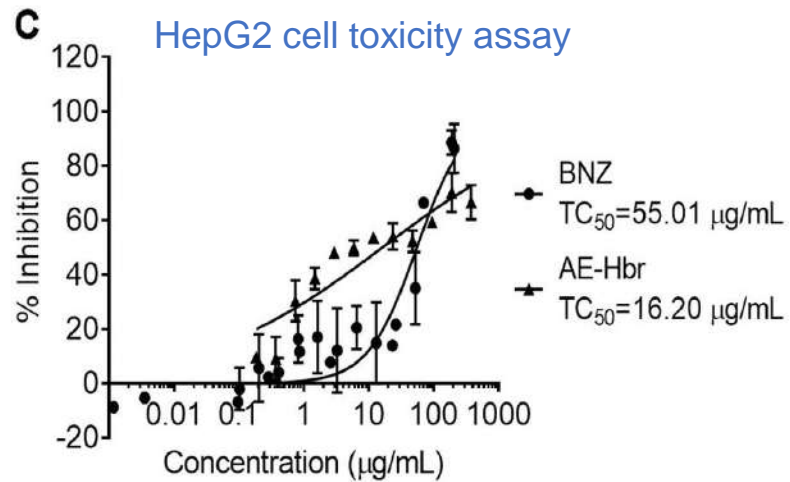
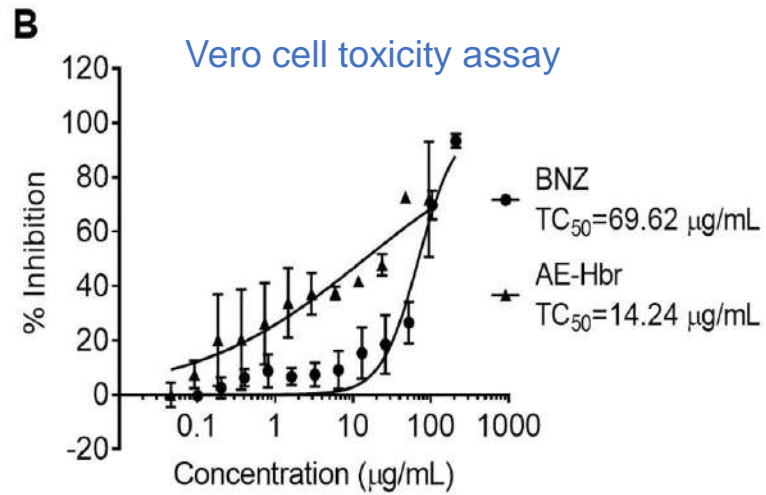
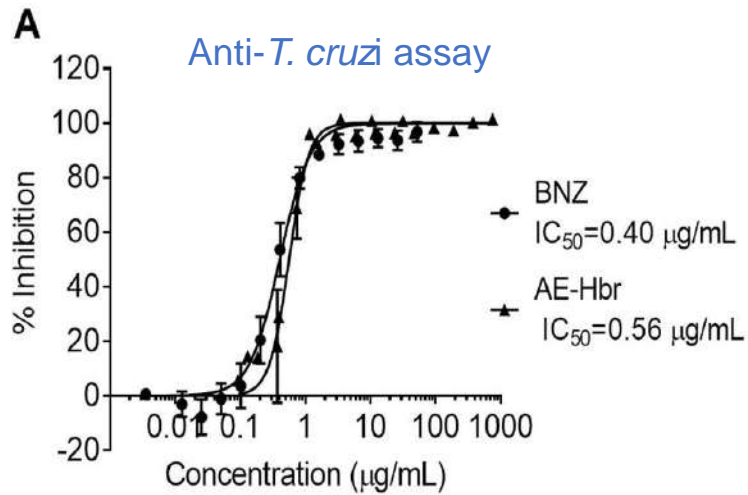


Prof. Jaume Bastida

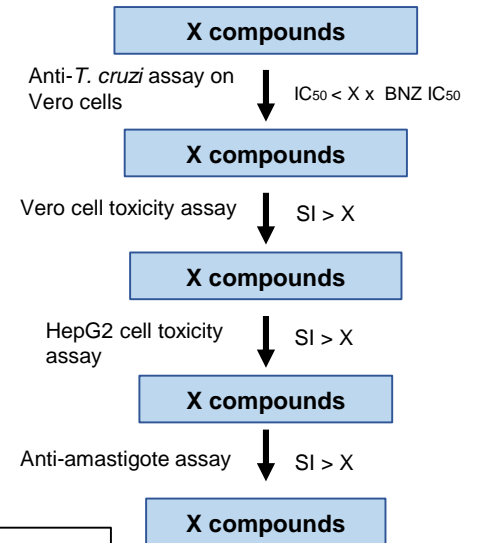
Dr. Gabriela Feresin



5. RESULTS AND DISCUSSION. CHAPTER I



GC-MS spectra, NMR
Chromatographic
techniques



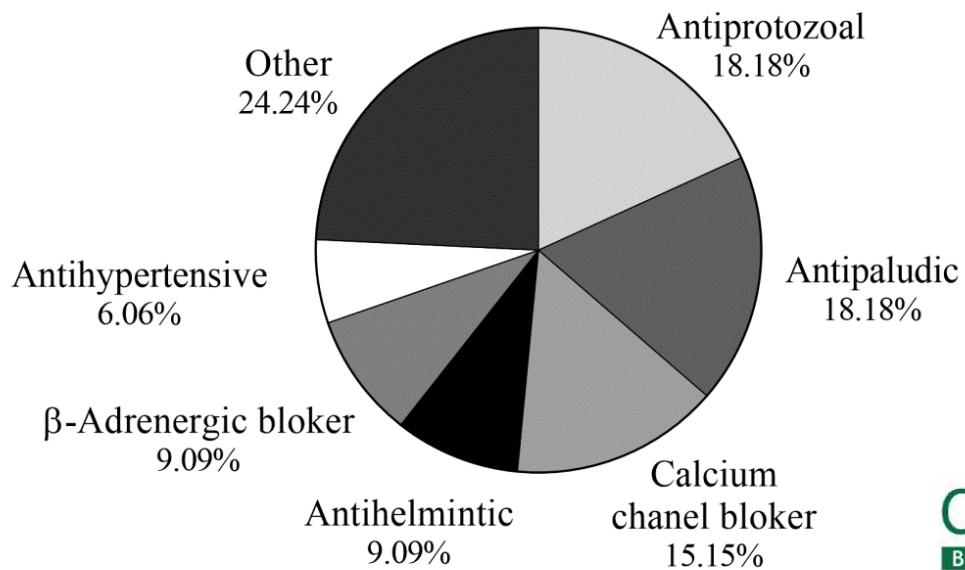
Alkaloids
responsible of
AE-Hbr extract
activity?

5. RESULTS AND DISCUSSION. CHAPTER II

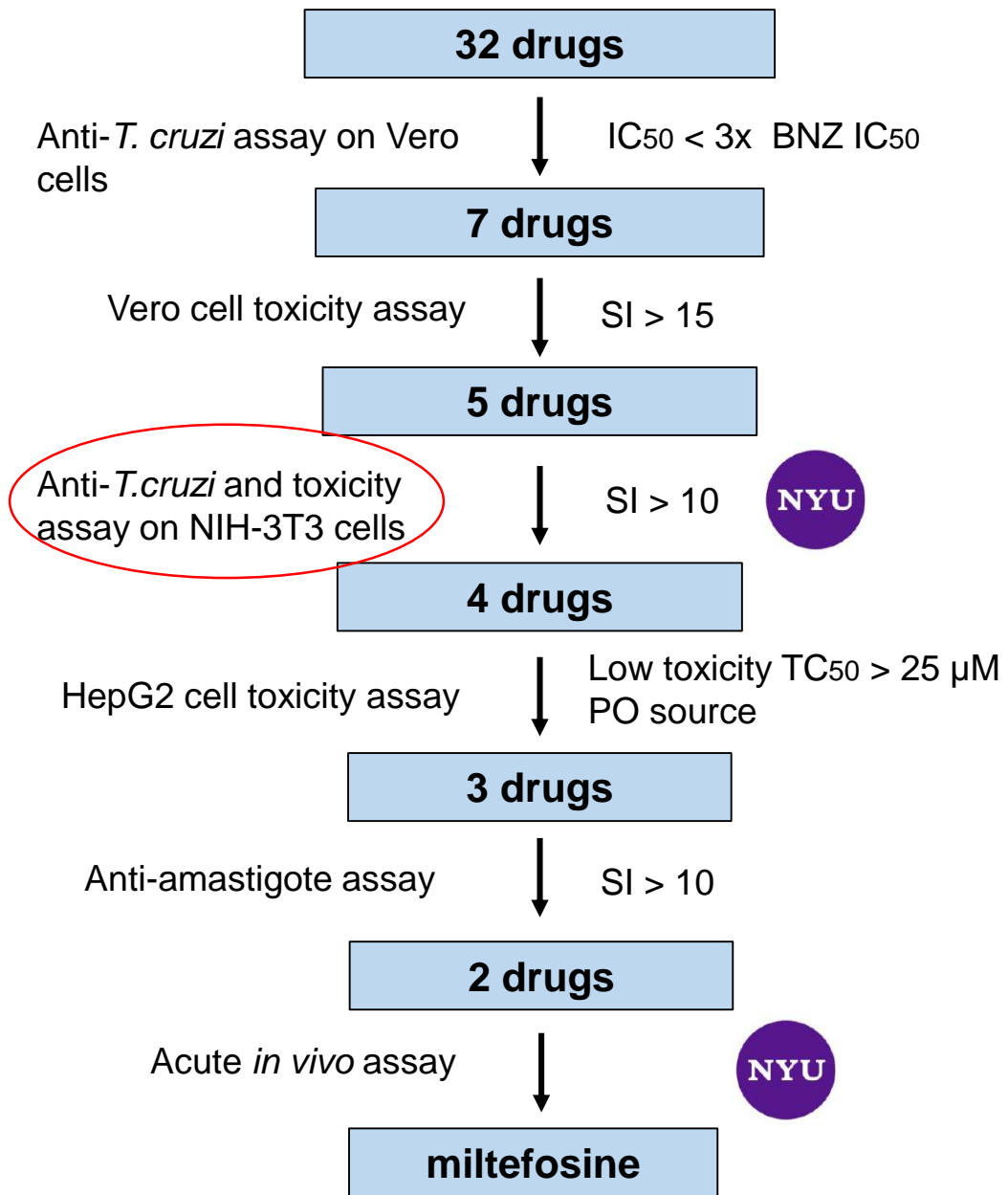


Article Identification of *Trypanosoma cruzi* Growth Inhibitors with Activity In Vivo within a Collection of Licensed Drugs

Nieves Martinez-Peinado ^{1,†}, Nuria Cortes-Serra ^{1,†}, Julian Sherman ², Ana Rodriguez ², Juan M. Bustamante ³, Joaquim Gascon ¹, Maria-Jesus Pinazo ^{1,*} and Julio Alonso-Padilla ^{1,*}



Dr. Juan Bustamante
Prof. Joaquim Gascón



5. RESULTS AND DISCUSSION. CHAPTER II



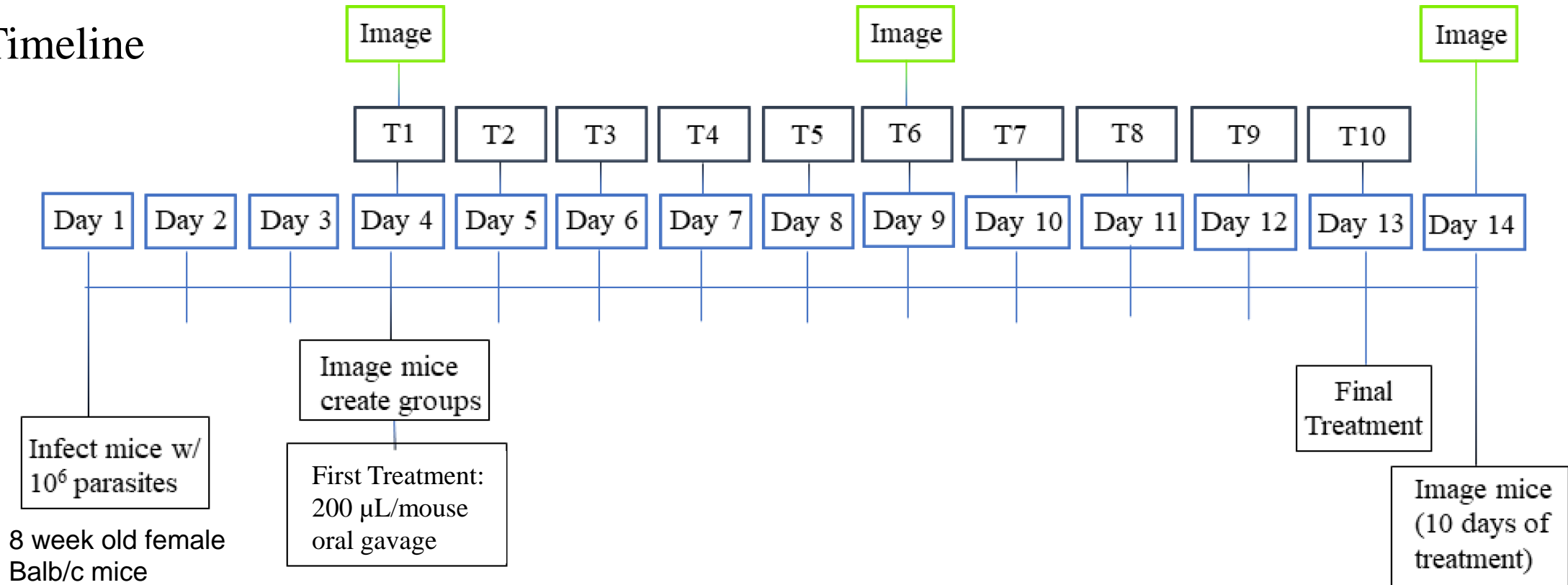
Drug	Vero cells assays			NIH-3T3 cells assays*			HepG2 cells assays	Anti-amastigote assay	
	IC ₅₀ (μM)	TC ₅₀ (μM)	SI	IC ₅₀ (μM)	TC ₅₀ (μM)	SI	TC ₅₀ (μM)	IC ₅₀ (μM)	SI
BNZ	1.93	242.2	125.5	-	-	-	229.8	2.66	91.1
Atovaquone – proguanil	1.26	27.13	21.5	1.32	50	>50	34.36	1.85	14.7
Miltefosine	0.018	78.99	4,388.3	0.037	1.95	52.7	51.28	1.25	63.2
Lidocaine[#]	0.016	0.23	14.4						
Nifedipine	0.19	1.967	10.4						
Pentamidine	1.01	78.96	78.2	0.13	5.9	45.4	39.4		
Piperaquine tetrphosphate - dihydroartemisinin	3.95	75.27	19.1	4.05	27.33	6.8			
Verapamil	3.44	197.4	57.4	0.21	5.72	27.2	170.5	122.5	1.6

values expressed as drug % (v/v).

5. RESULTS AND DISCUSSION. CHAPTER II

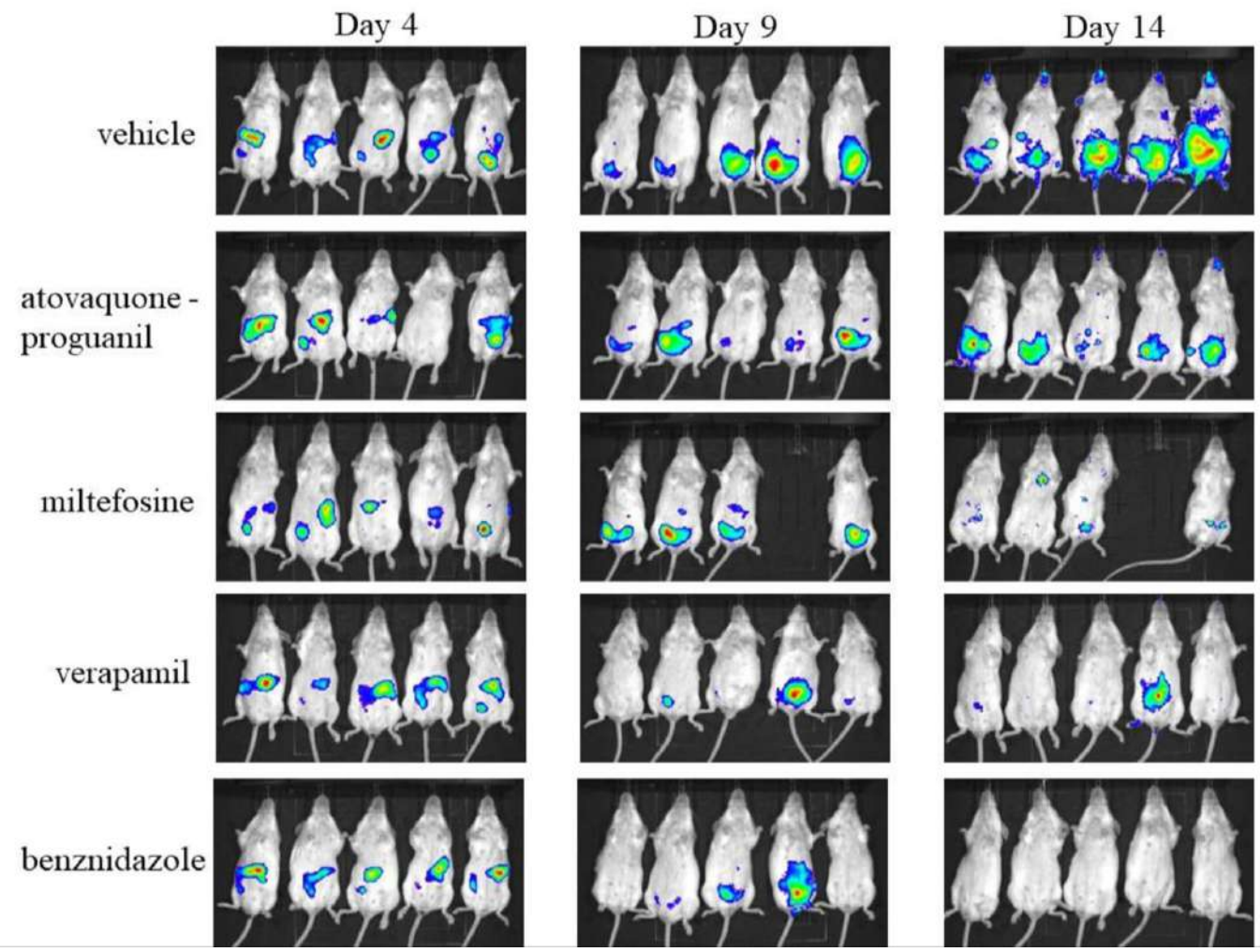
Acute *in vivo* model

Timeline

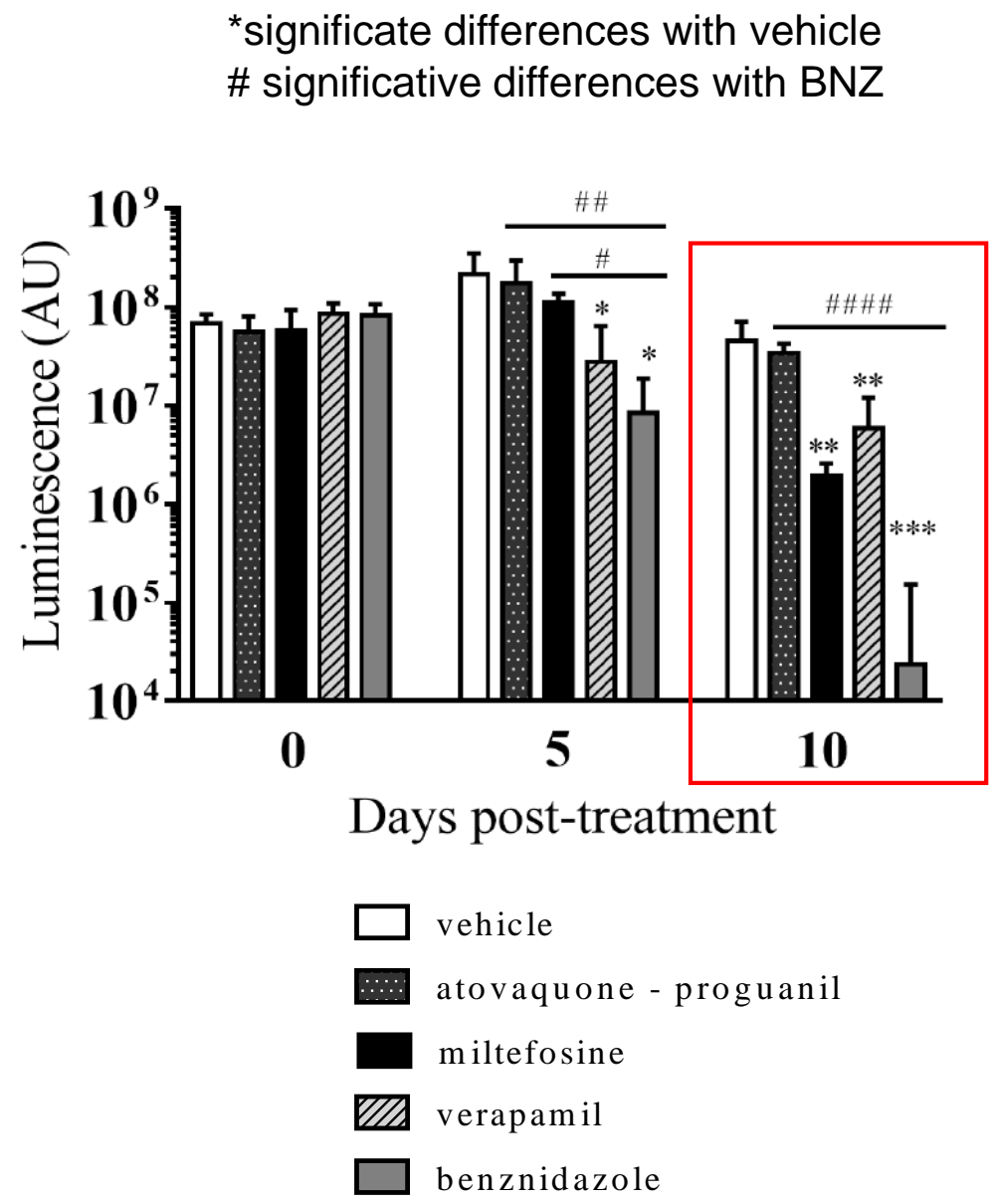


BNZ, miltefosine and autovaquone-proguanil: 30 mg/kg/day
verapamil: 5 mg/kg/day

Acute *in vivo* model



T. cruzi Luc Brazil (DTU I)



5. RESULTS AND DISCUSSION. CHAPTER II



Miltefosine and Benznidazole Combination Improve Anti-Trypanosoma cruzi In Vitro and In Vivo Efficacy

Julián Ernesto Nicolás Gulín^{1,2}, Margarita María Catalina Bisio^{1,3}, Daniela Rocco¹, Jaime Altcheh¹, María Elisa Solana^{4,5} and Facundo García-Bournissen^{1,6*}

OPEN ACCESS

Analysis

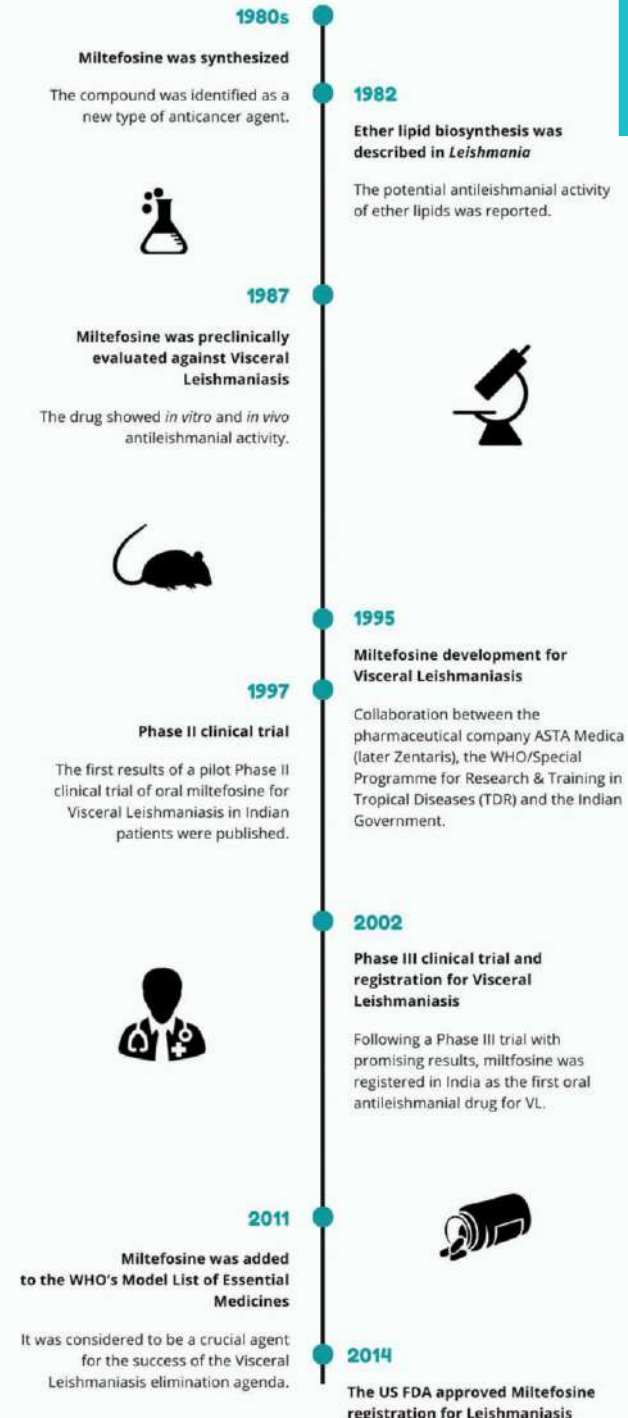
BMJ Global Health

Why miltefosine – a life-saving drug for leishmaniasis – is unavailable to people who need it the most

REVIEW

Miltefosine in the treatment of leishmaniasis: Clinical evidence for informed clinical risk management

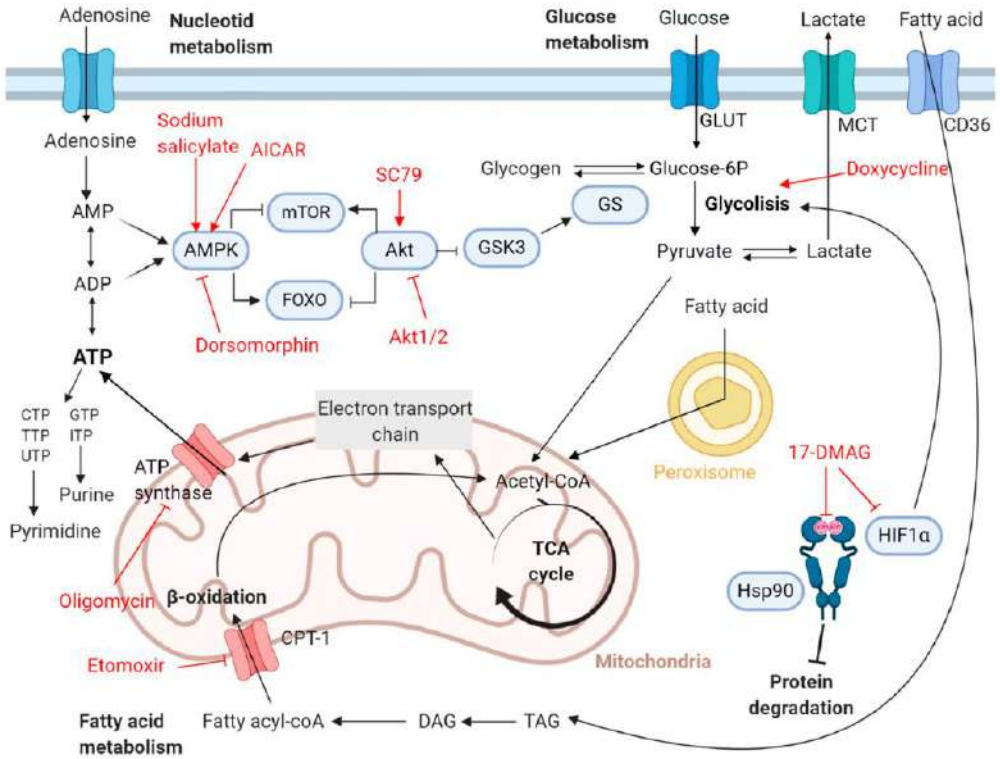
MILTEFOSINE DISCOVERY & DEVELOPMENT



5. RESULTS AND DISCUSSION. CHAPTER III

Article Anti-*Trypanosoma cruzi* Activity of Metabolism Modifier Compounds

Nieves Martinez-Peinado ¹, Clara Martori ², Nuria Cortes-Serra ¹, Julian Sherman ³, Ana Rodriguez ³, Joaquim Gascon ¹, Jordi Alberola ², Maria-Jesus Pinazo ¹, Alheli Rodriguez-Cortes ^{2,*} and Julio Alonso-Padilla ^{1,*}



UAB
Universitat Autònoma
de Barcelona
Dra. Alhelí
Rodríguez-Cortes

9 compounds

Anti-*T. cruzi* assay on Vero cells \downarrow $IC_{50} < 5 \times BNZ IC_{50}$

3 compounds

AlamarBlue
Crystal violet } Vero cell toxicity assay \downarrow $SI > 10$

17-DMAG

Anti-amastigote assay \downarrow $SI > 10$

17-DMAG

In silico model \downarrow

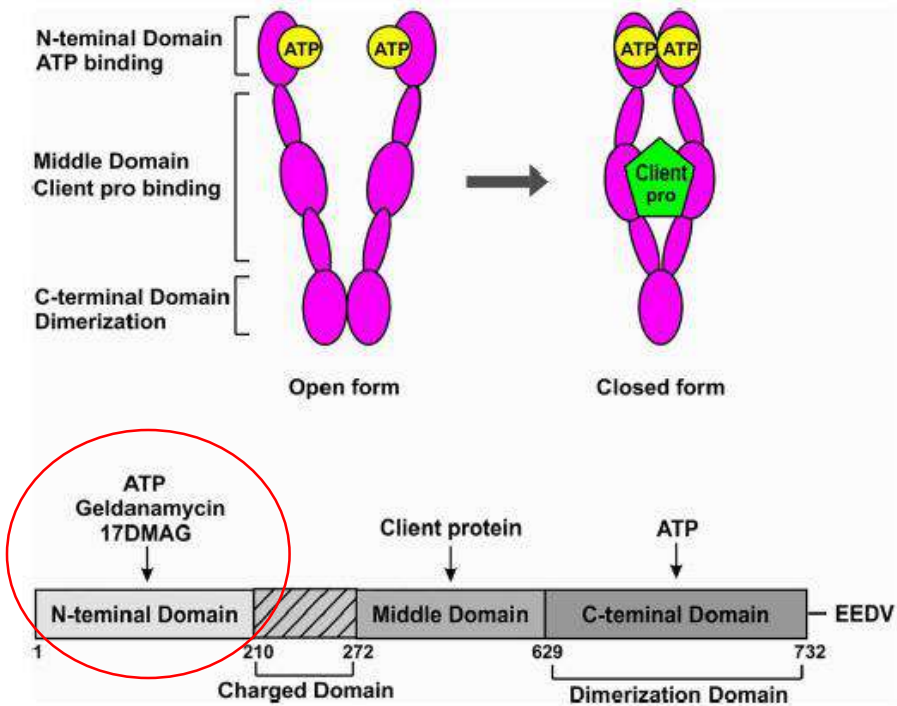
17-DMAG

Chronic *in vivo* model \downarrow

-

5. RESULTS AND DISCUSSION. CHAPTER III

Compound	Vero cell assays Alamar Blue			Crystal violet		Anti-amastigote assay	
	IC ₅₀ (μM)	TC ₅₀ (μM)	SI	TC ₅₀ (μM)	SI	IC ₅₀ (μM)	SI
BNZ	1.63	243.8	149.6	140.2	86	2.02	120.69
17-DMAG	0.017	6.2	366.5	2.97	174.7	0.17	36.5



(Mellyart et al., BiomedPharma 2018)

SCIENTIFIC REPORTS
nature research

OPEN A docking-based structural analysis of geldanamycin-derived inhibitor binding to human or *Leishmania* Hsp90

Received: 20 June 2018
Accepted: 13 September 2019
Published online: 14 October 2019

Luana Carneiro Palma¹, Luiz Felipe Gomes Rebello Ferreira², Antonio Luis de Oliveira Almeida Petersen¹, Beatriz Rocha Simões Dias¹, Juliana Perrone Bezerra de Menezes¹, Diogo Rodrigo de Magalhães Moreira³, Marcelo Zaldini Hermandes² & Patricia Sampaio Tavares Veras¹

OPEN ACCESS Freely available online

PLOS | NEGLECTED TROPICAL DISEASES

Exploring the *Trypanosoma brucei* Hsp83 Potential as a Target for Structure Guided Drug Design

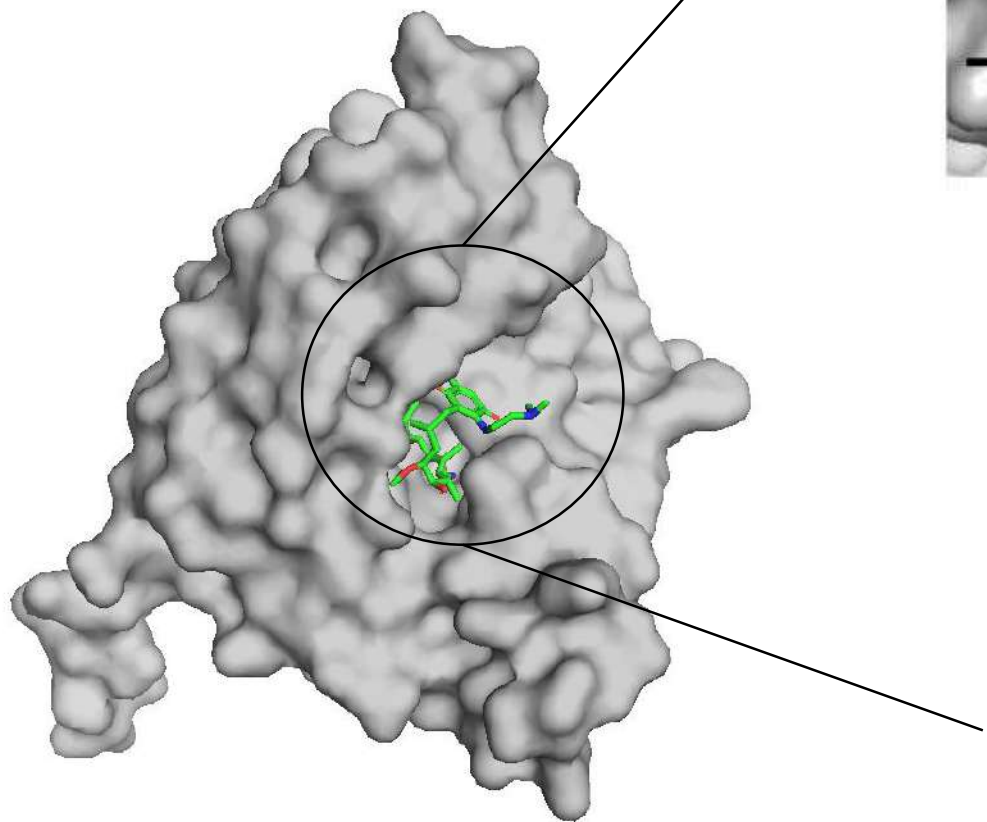
Juan Carlos Pizarro^{1,2*}, Tanya Hills¹, Guillermo Senisterra¹, Amy K. Wernimont¹, Claire Mackenzie³, Neil R. Norcross³, Michael A. J. Ferguson³, Paul G. Wyatt³, Ian H. Gilbert³, Raymond Hui¹

¹ The Structural Genomics Consortium (SGC), University of Toronto, Toronto, Ontario, Canada, ² Department of Tropical Medicine, School of Public Health and Tropical Medicine, Tulane University, New Orleans, Louisiana, United States of America, ³ Division of Biological Chemistry and Drug Discovery, College of Life Sciences, University of Dundee, Dundee, Scotland, United Kingdom

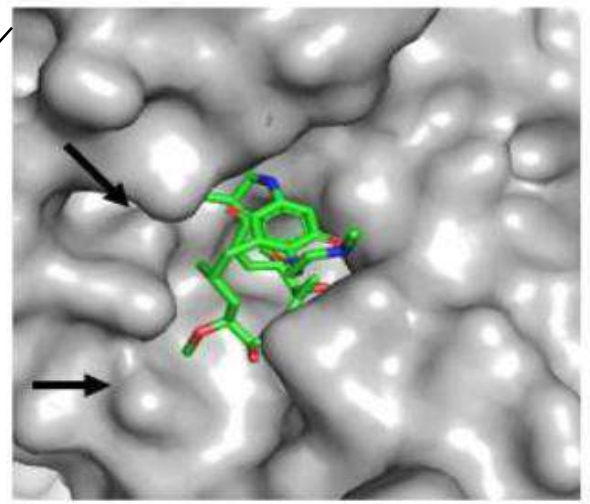
***In silico* study:**

Site-directed mutagenesis

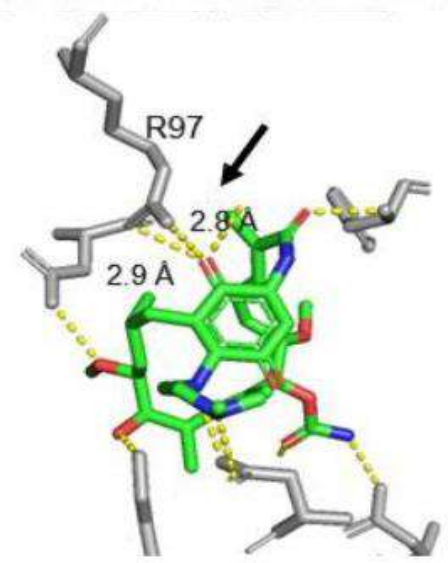
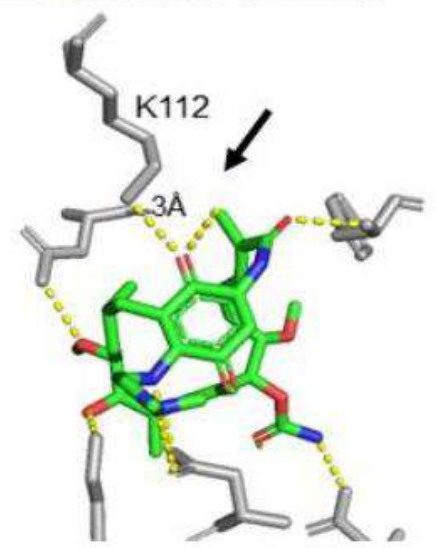
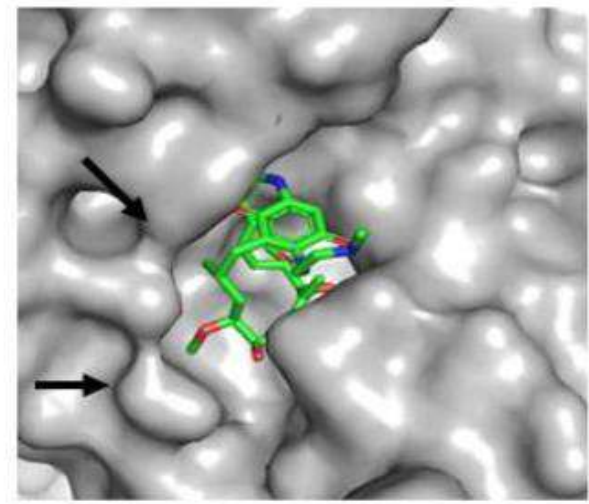
73% sequence identity between human Hsp90 N-terminal and *T. cruzi* Hsp83 N-terminal



(PDB: 1OSF)
Human Hsp90



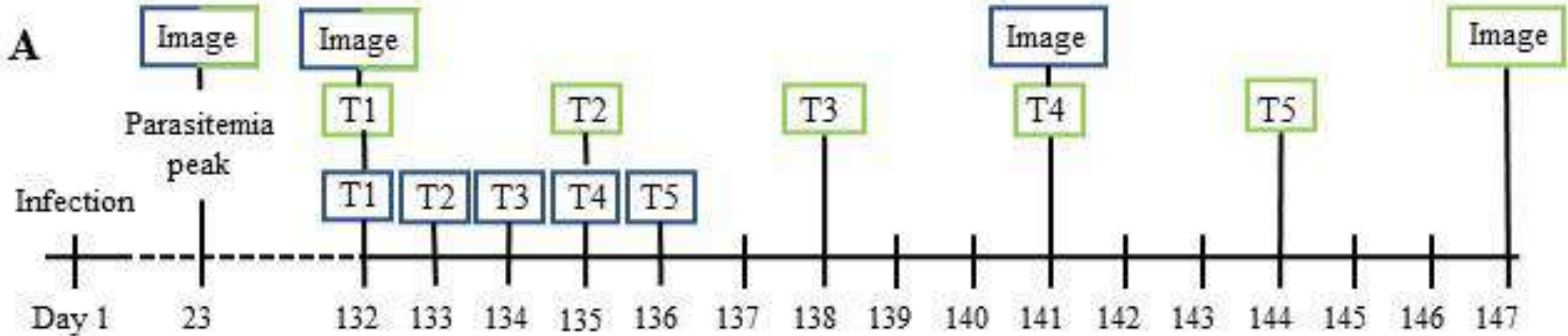
***T. cruzi* Hsp83**



(PyMOL Molecular Graphics System)

Chronic *in vivo* model

Timeline



BNZ intraperitoneal administration: 30 mg/kg/day
 17-DMAG intraperitoneal administration: 30 mg/kg/day

MAJOR ARTICLE

OPEN ACCESS Freely available online

PLOS NEGLECTED TROPICAL DISEASES

Potent Antitrypanosomal Activities of Heat Shock Protein 90 Inhibitors In Vitro and In Vivo

Kirsten J. Meyer¹ and Theresa A. Shapiro^{1,2}
¹Department of Pharmacology and Molecular Sciences and ²Division of Clinical Pharmacology, Department of Medicine, The Johns Hopkins University School of Medicine, Baltimore, Maryland

Chemotherapeutic Potential of 17-AAG against Cutaneous Leishmaniasis Caused by *Leishmania (Viannia) braziliensis*

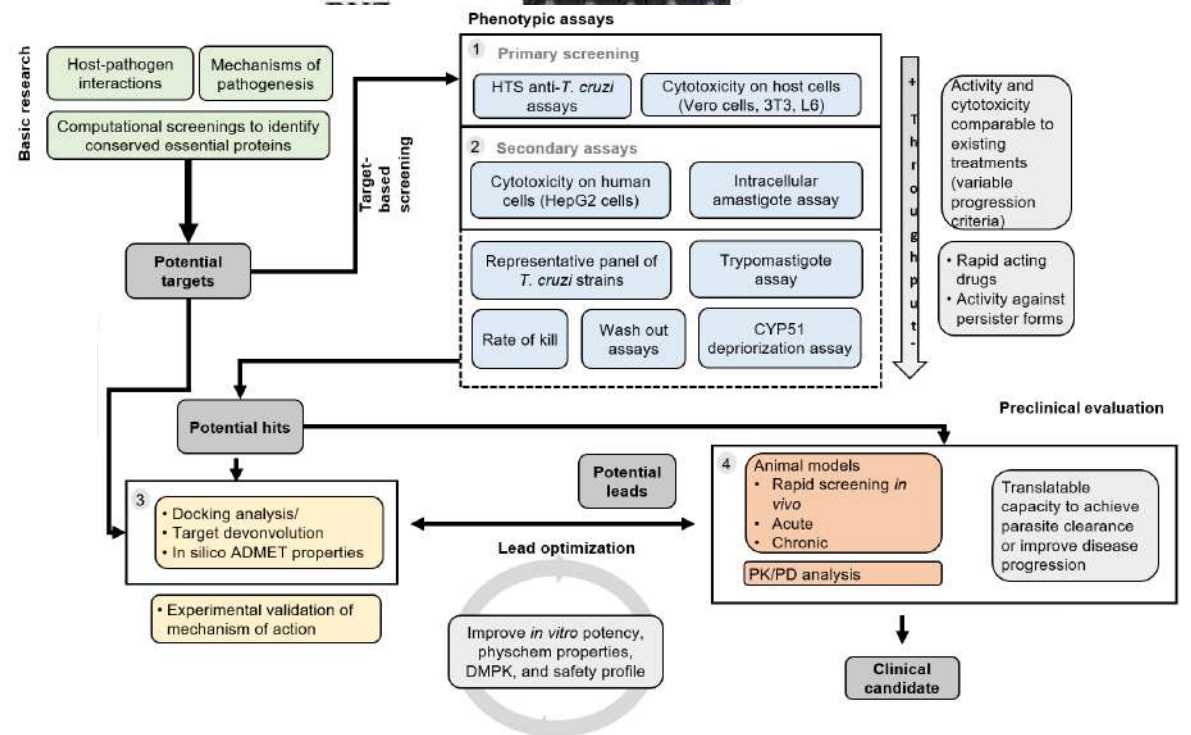
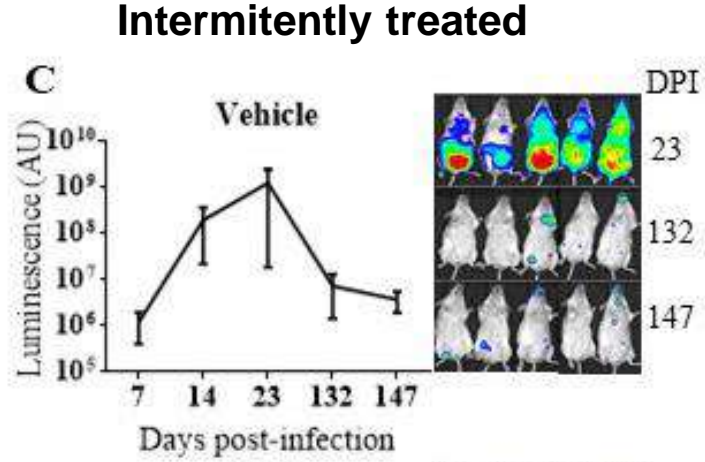
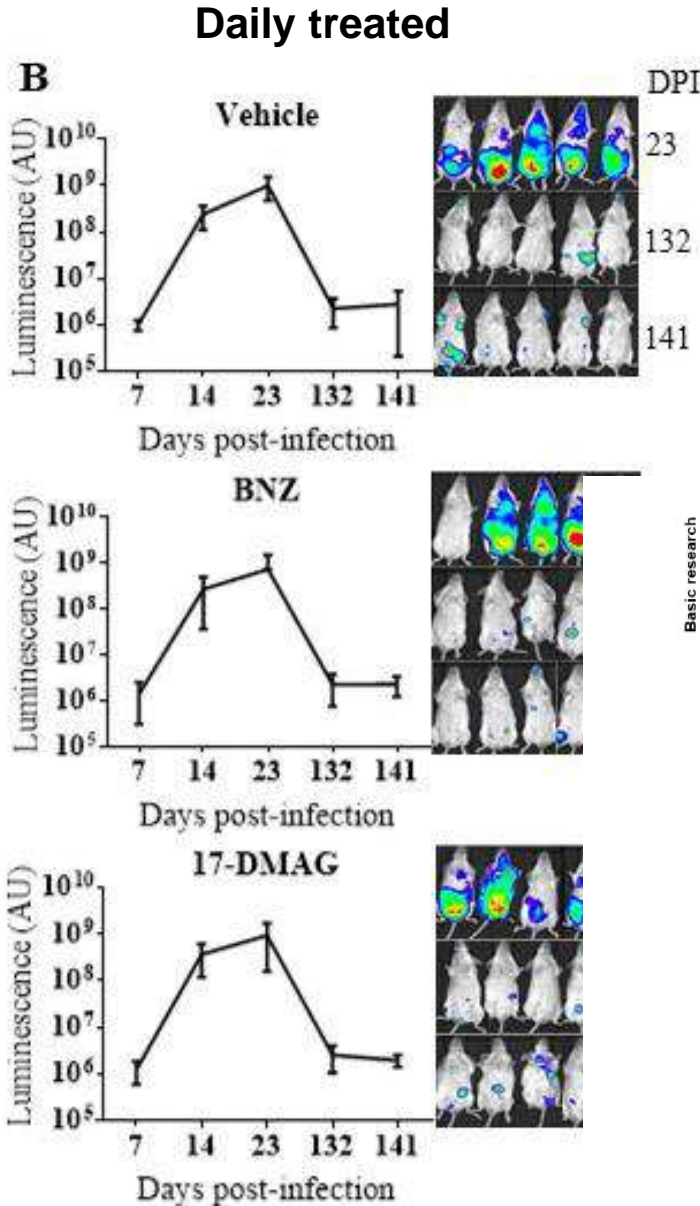
Diego M. Santos^{1*}, Antonio L. O. A. Petersen¹, Fabiana S. Celes¹, Valeria M. Borges^{1,2}, Patricia S. T. Veras¹, Camila I. de Oliveira^{1,2*}

5. RESULTS AND DISCUSSION. CHAPTER III

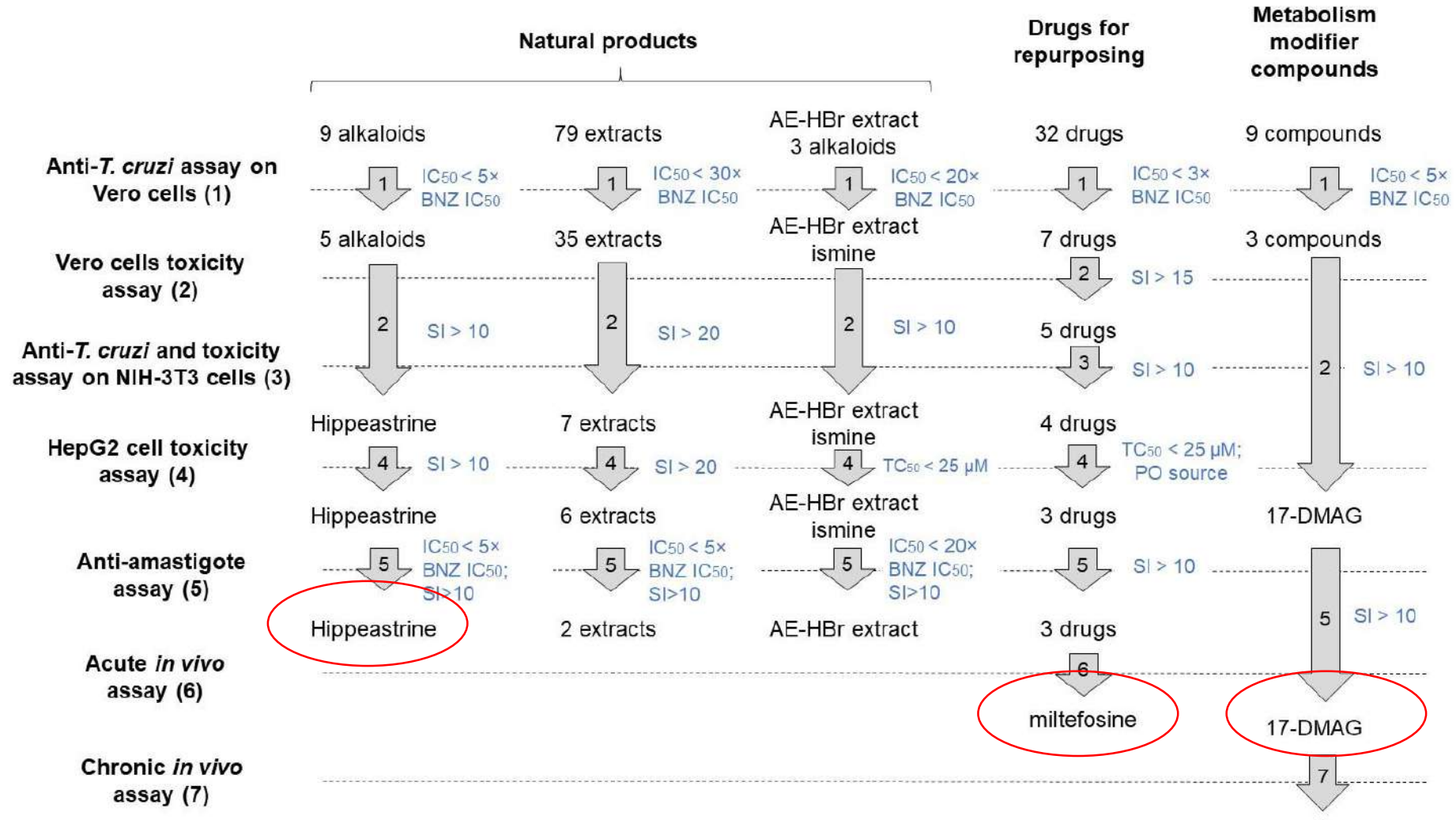
Chronic *in vivo* model

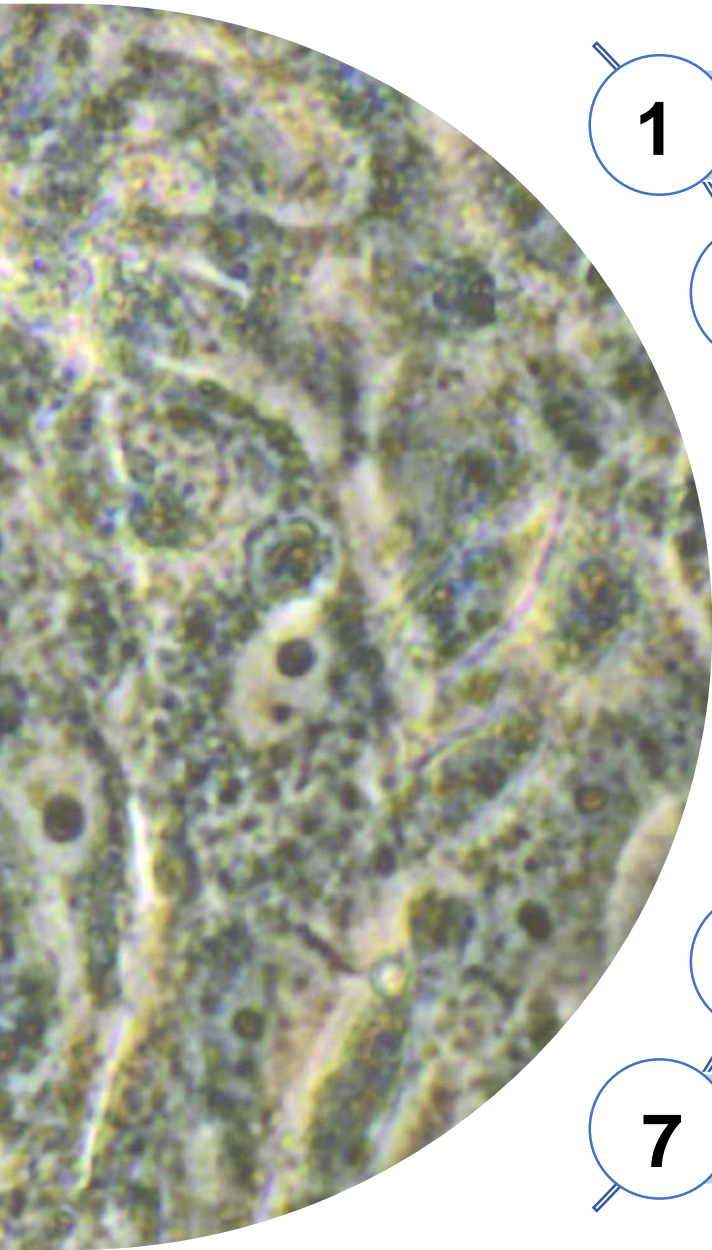


BNZ and 17-DMAG:
30 mg/kg/day



5. RESULTS AND DISCUSSION

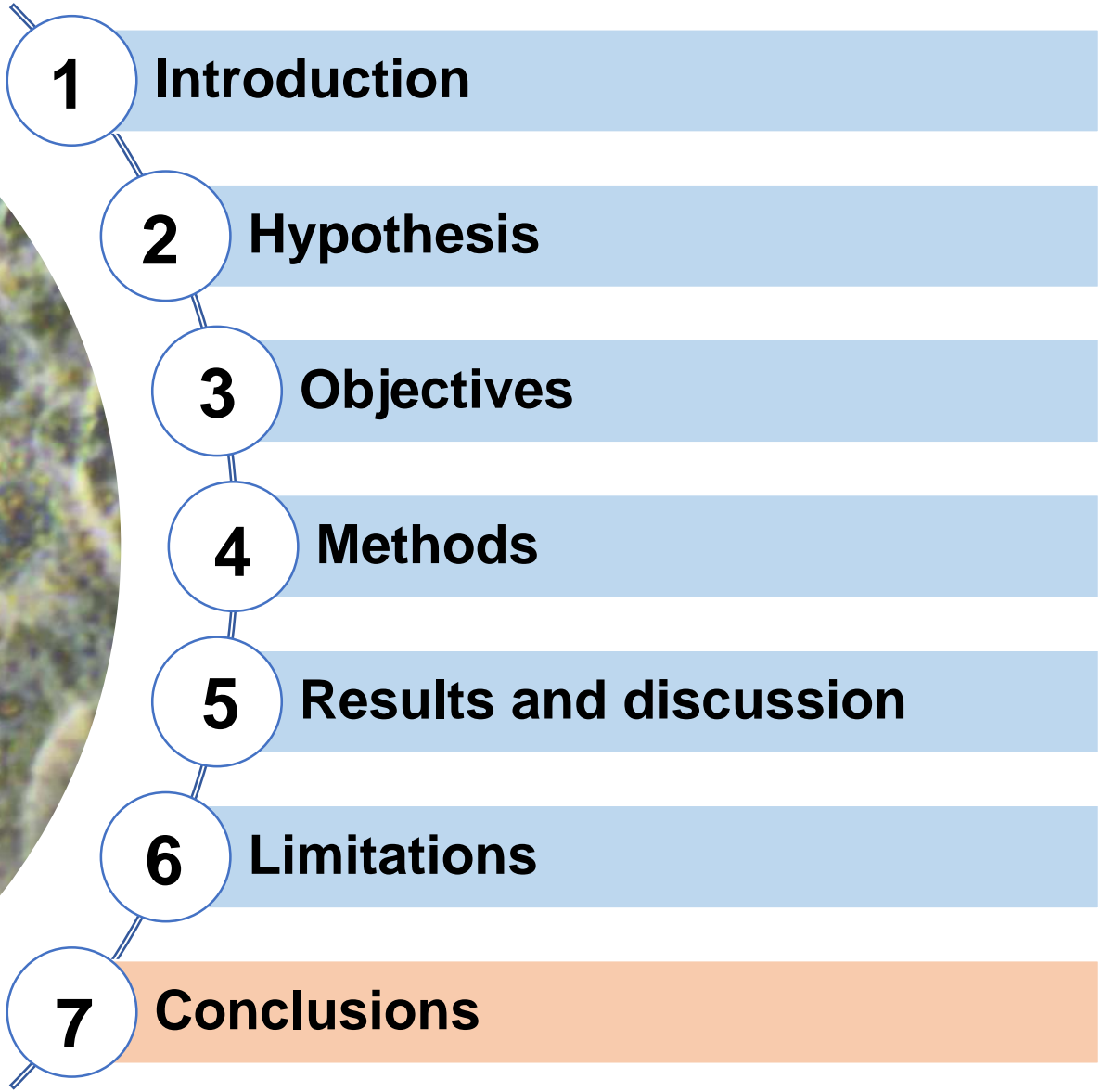
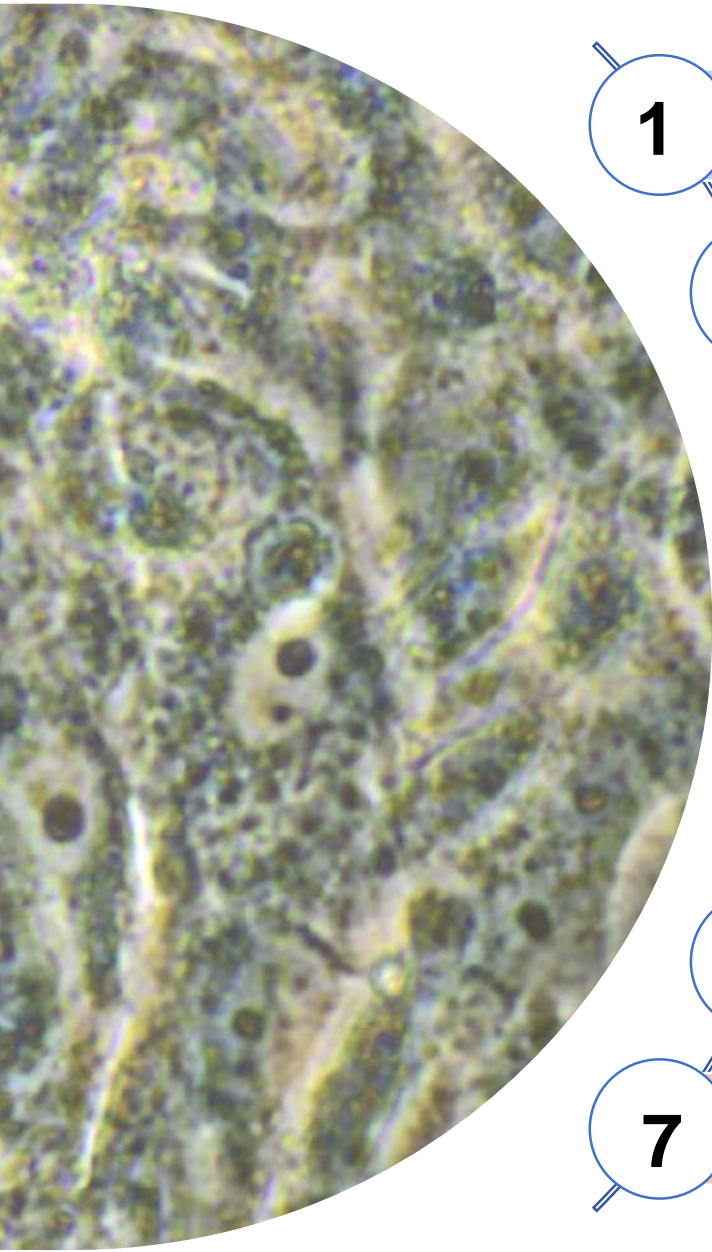




- 1** Introduction
- 2** Hypothesis
- 3** Objectives
- 4** Methods
- 5** Results and discussion
- 6** Limitations
- 7** Conclusions

6. LIMITATIONS

- Chemical collections:
 - a. Unknown alkaloid profile of some of the extracts.
- Our *in vitro* screening cascade would benefit from extra secondary assays:
 - a. Evaluation against a panel of diverse *T. cruzi* strains and host cells.
 - b. Wash-out.
 - c. Rate of kill.
 - d. CYP51 depriorization.
- *In vivo* assays:
 - a. Lack of resources to continue studying chronically infected mice for a longer period of time.
- *In silico* study:
 - a. AlphaFold models vs crystalized proteins.
 - b. Receptor rigid molecular docking.




7. CONCLUSIONS


1. The screening cascade established as part of this thesis encompasses *in vitro*, *in silico* and *in vivo* assays that allow the identification of compounds/drugs with specific activity against *T. cruzi*.
2. Amaryllidaceae plants are a source of biological active alkaloids with anti-*T. cruzi* properties.
3. *C. erubescens*, *R. andicola* and *H. brachyandrus* extracts were active against *T. cruzi* and deserve further exploration to elucidate the alkaloid or alkaloids responsible of such anti-parasitic activity.
4. The alkaloids hippeastrine and ismine were found to be active against the parasite forms infecting mammalian cells and showed low toxicity to Vero and HepG2 cells. However, ismine lacks activity against the replicative amastigote forms.
5. Miltefosine performance *in vitro* and *in vivo* would encourage further investigating its use against *T. cruzi*.
6. The metabolism modifier compound 17-DMAG showed the highest *in vitro* potency against the parasite among all tested compounds, but failed to work in a mouse model of chronic *T. cruzi* infection.
7. Our *in silico* target identification pipeline has allowed us to identify potential molecular targets and hypothesize on the compounds' MOA, although experimental validation would be needed.
8. In summary, we have found compounds with selective anti-*T. cruzi* activity. Although some of them deserve further attention, none has worked *in vivo* as good as the current anti-*T. cruzi* standard drug: benznidazole.

Muchas gracias!


Chagas Initiative

Initiative Co-Directors


 **Quim Gascon**
RESEARCH PROFESSOR, HEAD OF THE CHAGAS PARASITO AND SUPPORT DISEASE PROGRAMME AND DIRECTOR OF THE CHAGAS INITIATIVE


 **Julio Alonso Padilla**
ASSISTANT RESEARCH PROFESSOR AND DIRECTOR OF THE CHAGAS INITIATIVE


Initiative Coordinator


 **Irene Losada**
MEDICAL RESEARCH FELLOW & CHAGAS INITIATIVE COORDINATOR


ISGlobal Team


 **Cristina Alonso-Vega**
CLINICAL TRIAL MONITOR


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 **Elizabeth Posada**
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 **Mirko Rojas**
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Prof. Jaume Bastida



Dra. Gabriela Feresin



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GEORGIA

Dr. Juan Bustamante

UAB

Universitat Autònoma
de Barcelona

Dra. Alhelí Rodríguez-Cortes

