

Acute effects of a session of electroconvulsive therapy on brain-derived neurotrophic factor plasma levels

Miquel Bioque MD, PhD ^{a,*,**}, Karina S. Mac-Dowell MSc, PhD ^{b,*}, Cristina Font MSc ^b, Ana Meseguer RN ^c, Elisabet Macau RN ^d, Marta Garcia-Orellana MD ^e, Marc Valentí MD, PhD ^f, Juan C. Leza MD, PhD ^{b,**}, Miquel Bernardo MD, PhD ^a.

^a Barcelona Clínic Schizophrenia Unit, Neuroscience Institute, Hospital Clínic de Barcelona, Barcelona; Institut d'Investigacions Biomèdiques August Pi i Sunyer (IDIBAPS), Barcelona; Centro de Investigación Biomédica en red en salud Mental (CIBERSAM); Departament de Medicina, Universitat de Barcelona.

^b Department of Pharmacology & Toxicology, Faculty of Medicine, Universidad Complutense de Madrid University, Instituto de Investigación Hospital 12 de Octubre (i+12), IUIIN; CIBERSAM.

^c Barcelona Clínic Schizophrenia Unit, Neuroscience Institute, Hospital Clínic de Barcelona, Barcelona; CIBERSAM.

^d Psychiatry Department, Neuroscience Institute, Hospital Clínic de Barcelona, Barcelona.

^e Anesthesiology Department, Hospital Clínic de Barcelona, Barcelona; Universitat de Barcelona, Barcelona.

^f Barcelona Bipolar Disorder Program, Psychiatry Department, Neuroscience Institute, Hospital Clínic de Barcelona, Barcelona.

*The first and the second author (MBi and KSMD) contributed equally to this work.

** Co-corresponding authors:

Dr. Miquel Bioque
Barcelona Clínic Schizophrenia Unit
Hospital Clínic de Barcelona
Barcelona, Spain
Carrer Villarroel 170 Esc 9 Pta 6
ZIP: 08036
Tel.: +34 932275400 (X 5547)
E-mail address: mbioque@clinic.ub.es

or

Dr. Juan C. Leza
Dept. of Pharmacology & Toxicology
Fac. Medicine Universidad Complutense
ZIP: 28040
Madrid. Spain.
Tel: +34 913941478
jcleza@med.ucm.es

Running title: Acute effects of ECT in BDNF plasma levels

Conflicts of interest

Dr. Bernardo has been a consultant for, received grant/research support and honoraria from, and been on the speakers/advisory board of ABBiotics, Adamed, Angelini, Casen Recordati, Eli Lilly, Janssen-Cilag, Lundbeck, Otsuka, Takeda, Somatics and has obtained research funding from the Ministry of Education, Culture and Sport, the Spanish Ministry of Economy, Industry and Competitiveness (CIBERSAM), by the Government of Catalonia, Secretaria d'Universitats i Recerca del Departament d'Economia i Coneixement (2017SGR1355), Foundation European Group for Research In Schizophrenia (EGRIS), and the 7th Framework Program of the European Union. Dr. Bioque has been a consultant for, received grant/research support and honoraria from, and been on the speakers/advisory board of has received honoraria from talks and/or consultancy of Adamed, Ferrer, Janssen-Cilag, Lundbeck, Otsuka, Pfizer and Sanofi. Dr. Valentí has received research grants from Eli Lilly & Company and has served as a speaker for Abbott, Bristol-Myers Squibb, GlaxoSmithKline, Janssen-Cilag, and Lundbeck. The rest of authors report no competing interests for this study.

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^d Psychiatry Department, Neuroscience Institute, Hospital Clínic de Barcelona, Barcelona.

^e Anesthesiology Department, Hospital Clínic de Barcelona, Barcelona; Universitat de Barcelona, Barcelona.

^f Barcelona Bipolar Disorder Program, Psychiatry Department, Neuroscience Institute, Hospital Clínic de Barcelona, Barcelona.

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** Co-corresponding authors:

Dr. Miquel Bioque
Barcelona Clínic Schizophrenia Unit
Hospital Clínic de Barcelona
Barcelona, Spain
Carrer Villarroel 170 Esc 9 Pta 6
ZIP: 08036
Tel.: +34 932275400 (X 5547)
E-mail address: mbioque@clinic.cat

or
Dr. Juan C. Leza
Dept. of Pharmacology & Toxicology
Fac. Medicine Universidad Complutense
ZIP: 28040
Madrid. Spain.
Tel: +34 913941478
jcleza@med.ucm.es

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Keywords: Biomarkers; BDNF; Brain-derived neurotrophic factor; ECT; Electroconvulsive therapy.

Abstract

Brain-derived neurotrophic factor (BDNF) and nerve growth factor (NGF) are neurotrophins that play critical roles in brain neuronal function. Previous studies have established the association between BDNF and NGF signalling and severe mental disorders, but changes in BDNF plasma levels and electroconvulsive therapy (ECT) response are controversial. The aim of his study was to explore the acute effects of a single session of ECT on these neurotrophins signalling.

Plasma levels of BDNF and NGF and their tyrosine kinase-type receptors expression in peripheral blood mononuclear cells (PBMCs) were determined before and two hours after a single ECT session in 30 subjects with a severe mental disorder.

Two hours after an ECT session we found a statistically significant decrease of BDNF plasma levels ($p=0.007$). We did not find significant acute effects on NGF plasma levels or receptors expression in PBMCs. We found a significant inverse correlation between the time of convulsion and BDNF plasma levels decrease ($r=-0.041$, $p= 0.024$).

We have identified a decrease in BDNF plasma levels after 2 hours of a single ECT session. These results indicate the interest for future research in the role of neurotrophins in the response and safety of ECT.

Introduction

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3 Electroconvulsive therapy (ECT) is recognized as one of the most effective therapies for certain
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5 severe mental disorders resistant to other treatments or in clinical scenarios that need a rapid
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7 response ¹⁻⁶. Despite its efficacy, it is necessary to expand the available knowledge about its
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9 mechanisms of action and to identify useful biological markers linked either to clinical response
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11 or to the appearance of side effects.
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15 ECT affects a wide range of molecules, including neurotransmitters, inflammatory pathways and
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17 neurotrophic factors ^{7, 8}. Brain-derived neurotrophic factor (BDNF) and nerve growth factor
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19 (NGF) are neurotrophins that play critical roles in neurodevelopment and a wide range of adult
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21 neuronal functions ^{9, 10}. Both BDNF and NGF are activated by their own tyrosine kinase-type
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23 receptors, named TrkB and TrkA respectively ¹¹. In the case of TrkB, two types of receptors
24
25 have been described: an active full-length form of the receptor (TrkB-F) and a truncated form
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27 (TrkB-T). The last one lacks kinase activity and inhibits the TrkB-F function by competing in
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29 binding to BDNF ¹². For its part, TrkA mediates on multiple effects of NGF, including neuronal
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31 differentiation and programmed death inhibition ¹¹.
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38 Several studies and meta-analyses have established the association between BDNF and NGF
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40 signalling and severe mental disorders, including major depressive disorder (MDD), bipolar
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42 disorder (BD) and schizophrenia (SZ) ¹³⁻¹⁶. A meta-analysis conducted by Molendijk et al.
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44 concluded that serum BDNF levels were reduced in patients suffering from major depressive
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46 disorder (MDD) and in patients receiving a course of antidepressants ¹⁷.
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51 Despite a growing number of studies, findings on changes in BDNF levels and the response to
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53 ECT are controversial ¹⁸. A recent meta-analysis of 22 studies of BDNF blood levels after ECT in
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55 patients with major depressive disorder concluded that BDNF levels may increase after ECT,
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57 being a candidate to be used as an indicator of treatment response after one or more weeks of
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59 ECT ¹⁹. However, some studies reported that serum or plasma BDNF levels increase at different
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time points after ECT ²⁰⁻²⁹, while other research groups did not find any influence of ECT on BDNF levels ³⁰⁻³⁵. Recently, Sorri and collaborators reported a decrease of BDNF levels during the fifth ECT session, between the baseline and the 2-hr samples, with no association between serum or plasma BDNF levels and remission ¹⁰. Finally, baseline serum BDNF levels and the BDNF Val66Met polymorphism did not show any clinical utility in predicting ECT response in treatment-resistant MDD patients ³⁶. Taken together, these results indicate a need for further research of the effects of ECT on the neurotrophins signalling and of the predictive value of BDNF plasma determination in patients who will receive this treatment.

Considering this background, the main objective of the present study is to explore the acute effects of a single session of ECT on the neurotrophins signalling in a selected group of patients with a severe mental disorder. Besides, we pretend to analyze if there are different effects considering clinical features of the studied sample and ECT parameters.

Subjects and Methods

Subjects

Thirty subjects who attended the Hospital Clínic de Barcelona ECT facilities during the recruitment period were recruited. The study inclusion criteria were: (1) Having a severe mental disorder (Major Depressive Disorder, Bipolar Disorder, Schizophrenia or Schizoaffective Disorder) according to the DSM-5 criteria; and (2) age between 18 and 70 years. The exclusion criteria were the following: (1) being pregnant; (2) presenting acute signs of infection in the previous day, such as fever (>38^o) or leukocytosis (> 10,000); (3) being following a treatment of anti-inflammatory drugs, antioxidants, antibiotics or immunological therapies; (4) have been vaccinated in the previous month; and (5) subjects with neurological diseases, traumatic

1 brain injury with loss of consciousness, mental retardation or generalized developmental
2 disorders history.
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5 All the potential candidates identified during the period of recruitment were discussed in a
6 specific ECT committee and were informed about the study by members of the research group.
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9 All participants signed the study informed consent ⁸. The study had been approved by the
10 Ethics Committee of the Hospital Clínic de Barcelona (record CB/2017/0369).
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13 Members of the research group collected the study variables from medical records,
14 sociodemographic and clinical data from an interview prior to the ECT session. Participants
15 could be in any ECT regimen (acute: first 6-12 ECT sessions; continuation: first six months of
16 treatment after acute regimen; maintenance: after first six months).
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19 All the ECT stimuli were administered with a MECTA SPECTRUM 5000Q[®] device (MECTA Corp,
20 Lake Oswego, USA). Adequate ictal response (> 20 seconds, < 60 seconds) were monitored
21 through Electroencephalographic (EEG) and motor seizure registers. Patients underwent the
22 ECT session under a general anesthesia, using Succinylcholine (25–110 mg), atropine (0–1 mg),
23 and sodium tiopental (75–400 mg). Before and after the stimulus ventilation was assisted a
24 face mask with high oxygen concentration. Mild hyperventilation was maintained for at least
25 one minute before the brain stimulation. All through the procedure a PHILIPS MP-20 (PHILIPS[®],
26 Boeblingen, Germany) Anesthesia monitor was used to monitor non-invasive arterial blood
27 pressure, EKG and heart rate and pulse oximetry. The MECTA EMR[®] software was used to
28 automatically collect certain characteristics of the stimulation.
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31 Study variables were recorded by the research group members, including age, gender,
32 diagnosis, duration of the illness, information regarding work disability, ECT regimen, ECT
33 electrode placement, EEG time of convulsion and total number of ECT sessions. Prescribed
34 psychopharmacological treatment was also recorded. The prescribed daily doses of
35 antipsychotics were converted to an estimated equivalent amount of chlorpromazine
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1 following the international consensus ³⁷, while the prescribed daily doses of antidepressants
2 were converted to an estimated equivalent amount of fluoxetine following published
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4 guidelines ³⁸.
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10 **Sample collection and biochemical measurements**

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14 10 mL of venous blood samples were collected after overnight fasting before the ECT session
15 (PRE-ECT), and two hours after (Post-ECT) from all participants ⁸. In order to be able to
16 compare our results to previous comparable studies that had taken this same measure ¹⁰ and
17 to ensure that both measures (pre and post ECT session) were being collected under fasting
18 conditions, we decided to sample at two hours post-ECT. Blood samples were kept in a fridge
19 (at 4°C) until further manipulation after approximately 1 h. Blood tubes were centrifuged (641
20 g × 10 min). The separated resultant plasma samples were stored at -80°C. The rest of the
21 sample was 1:2 diluted in RPMI 1640 (LifeTech) culture medium. A gradient with Ficoll-Paque
22 (GE Healthcare) was used to isolate Peripheral blood mononuclear cells (PBMC) by
23 centrifugation at room temperature (800 g × 40 min). PBMC layer was aspirated and
24 resuspended in the culture medium, then centrifuged at room temperature (1116 g × 10 min).
25
26 Once the supernatant layer was removed, the PBMC-enriched pellet was stored at -80°C at
27 the Hospital Clinic de Barcelona facilities. Once the whole recruitment was over, all samples
28 were sent to the Neuropsychopharmacology laboratory of the Medicine faculty of the
29 Universidad Complutense de Madrid for subsequent determinations.
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50 Plasma levels of BDNF and NGF were determined using enzymatic assays (ELH-BDNF-1,
51 RayBiotech®; ab99986, abcam), according to the manufacturer's instructions, and measured
52 using a Synergy 2 multi-mode reader (BioTek®). The sensitivity of the assay for BDNF was 80
53 pg/mL and <14 pg/ml for NGF; for both kits the manufacturer's intra- and interassay
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1 coefficients of variation (CV) were respectively <10% and <12%, while own lab's CVs were <7%
2 and <8%.

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5 Protein levels of TrkA, TrkB-F, and TrkB-T receptors in PBMCs were quantified by Western Blot
6 analysis. In brief, 15 µg of cytosolic extracts were loaded onto electrophoresis gels ³⁹. Protein
7 samples were separated and transferred onto nitrocellulose membrane (Transfer Pack,
8 Biorad®). After blocking, membranes were incubated with specific antibodies: (1) TrkA, rabbit
9 polyclonal antibody dilution of 1:750 in BSA 1% (sc118, SCB); (2) TrkB-F, rabbit polyclonal
10 antibody dilution of 1:750 in BSA 1% (sc12, SCB); (3) TrkB-T, rabbit polyclonal antibody dilution
11 of 1:1000 in BSA 1% (ab1987, abcam); (4) β-actin mouse monoclonal in a dilution 1:10000
12 (A5441, Sigma, Spain). Proteins were recognized by the respective horseradish peroxidase-
13 linked secondary antibodies and visualized using an Odyssey® Fc System (Li-COR Biosciences®)
14 and quantified by densitometry (NIH ImageJ® software). Values were normalized to the
15 loading control (β-actin, the blots are shown in figure 1, c-e). All western blots were performed
16 at least three times in separate assays.

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19 Given the counterbalancing effect of TrkB-T and TrkB-F, we decided to choose the ratio of
20 TrkB-F to TrkB-T expression (hereafter F/T ratio) as our index variable for describing BDNF
21 receptor expression.

22 **Statistical analysis**

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25 A paired t-test was used to assess the differences of biomarkers expression before and after
26 the ECT session on those variables (TrkA, TrkB-F, TrkB-T and F/T ratio) which distribution met
27 the assumption of normality in the Kolmogorov-Smirnov (with Lilliefors correction). For those
28 variables which distribution did not met the assumption of normality (BDNF and NGF plasma
29 levels), a nonparametric Wilcoxon Signed Rank test was used.

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In order to explore whether certain changes in the neurotrophins expression were associated with some of the demographic, clinical, pharmacological or ECT variables, several sub-analyses were performed. To avoid a large number of post-hoc analyses, only those biomarkers with significant pre/post-ECT activity changes were included in these sub-analyses. Being these exploratory analysis, corrections were not made for multiple comparisons ⁴⁰. A mixed between-within subjects analysis of variance was conducted to assess the impact categorical variables on neurotrophins expression. The relationship between the biomarkers variations and continuous variables were investigated using Pearson correlation coefficient. Preliminary analyses were performed to ensure no violation of the assumptions of normality, linearity and homoscedasticity. The strength of these correlations were evaluated according to Cohen's guidelines (small $r=0.10$ to 0.29 ; medium $r=0.30$ to 0.49 ; large $r=0.50$ to 1 ,) ⁴¹.

All analysis with a p -value <0.05 were considered statistically significant. Data were managed and analyzed with the IBM SPSS Statistics (v.23).

Results

Table 1 shows general demographic, clinical, pharmacological and ECT data of all the study participants. There wasn't any recorded incidence during these ECT procedures. Figure 1 summarizes the main results.

Table 1 around here

Figure 1 around here

Table 2 summarizes the changes in neurotrophins plasma levels and in the expression of the receptors in PBMCs before and two 2 hours after the ECT session, when there was a significant decreased of the BDNF plasma levels (from 82.16 ± 6.79 to 63.53 ± 6.75 ; $Z=-2.7$, $p= 0.007$).

Table 2 around here

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3 No baseline (pre-ECT) differences between genders, diagnosis or ECT regimen were found. We
4
5 found a significant inverse correlation for the EEG time of convulsion and BDNF plasma levels
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7 at the endpoint ($p= 0.024$), with a medium strength of the relationship ($r=-0.041$).
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10 We conducted group of analyses to explore if the observed changes in BDNF plasma levels
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12 after the ECT session were modified by concurrent pharmacological treatment studied. A
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14 mixed between-within subjects analysis of variance was conducted to assess the impact of
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16 concurrent psychopharmacologic treatment on BDNF levels changes after the ECT session. We
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18 did not find significant differences in BDNF levels changes in patients with or without
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20 prescriptions of antipsychotic, antidepressant, benzodiazepines, anticonvulsants, lithium or
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22 anticholinergics, meaning that changes of BDNF levels were not related to being or not under
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24 those treatments. We neither found significant correlation between BDNF plasma levels
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26 changes and antipsychotic ($p=0.17$) or antidepressant ($p=0.76$) doses.
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37 **Discussion**

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39 The results of this study point to the acute effects that a single ECT session produces on the
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41 neurotrophins pathway. Specifically, two hours after an ECT session we found a statistically
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43 significant decrease of the levels of BDNF in plasma (figure 1 a). We also found a significant
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45 inverse correlation for the EEG duration of convulsion and BDNF plasma levels decrease ($r=-$
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47 0.041 , $p= 0.024$). We did not find a significant acute effect over NGF plasma levels or receptors
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49 expression in PBMCs (figure 1 b-e).
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54 These results agree with previous reports from Sorri and collaborators ¹⁰, who recently
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56 reported a decrease of BDNF levels between the baseline and the 2-hour samples during the
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58 fifth ECT session. Stelzhammer and collaborators also reported decreased serum BDNF levels
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1 after 4 weeks of acute ECT ⁴². By contrast, some studies have reported plasma BDNF levels
2 increases at different time points after ECT ²⁰⁻²⁸. A recent meta-analysis of 22 studies, revealed
3 no significant changes in the serum BDNF levels between patients who responded and those
4 that did not respond to ECT, but reported a significant increase in the plasma BDNF levels after
5 ECT treatment in the first week and month in patients with MDD ¹⁹. Other research groups did
6 not report any influence of ECT on BDNF plasma levels ³⁰⁻³⁵.

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14 In our opinion, a major difference between these studies and ours is the timing of blood
15 sampling ^{10, 42}. In some of studies mentioned above BDNF plasma levels were measured during
16 a course of ECT, while in others blood samples were taken in a variable period of one day to
17 one month after finishing the whole ECT course (generally 9-12 sessions). Besides, previous
18 studies have shown that BDNF levels stored in plasma and serum might differ significantly,
19 which could be a confusion factor that might explain part of the controversial findings so far ¹⁹.

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30 Overall, these apparently controversial results sstrengthens the hypothesis that BDNF may
31 undergo differential regulation after different time periods following an ECT course ⁴². In that
32 vein, a gene profiling study found increased BDNF expression 2 hours after both acute and
33 chronic ECT, with decreased levels after 6 hours ⁴³. The mechanisms of action of ECT and
34 psychotropic drugs may be different, so their effects on specific biomarkers of therapeutic
35 response may be different too.

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45 Some limitations should be considered when analyzing these results. First, the relatively small
46 group of participants, which could limit the statistical power of the findings. Secondly, there
47 was heterogeneity of the participants in certain clinical variables such as diagnostics,
48 psychopharmacological treatment or ECT regimen. Thirdly, we studied the effects of a single
49 ECT session over the neurotrophins pathway after two hours, while it would very informative
50 to have re-tested the patients at different moment after the ECT session (i.e. 1 hour, 3 hours,
51 24 hours, 48 hours, or even one week later) and at the end of the whole acute treatment

1 (which generally takes between nine and twelve sessions). Fourthly, to our knowledge, the
2 effects of anesthesia, apnea or seizures over the pathway studied are unknown. Finally, we did
3 not have a group of healthy controls paired with the study sample with which to compare the
4 expression of biomarkers at baseline.
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10 Despite these limitations, we believe that our study has identified significant acute effects of
11 ECT treatment over BDNF plasma levels in a group of patients with severe mental disorders.
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13 Future studies could identify the optimal time point to determine the levels of BDNF in plasma,
14 serum or even directly from the brain, comparing ECT responders to partial/non-responders
15 levels across different timepoints during and after ECT treatment ¹⁹. Besides, large genetic
16 consortium such as the International Consortium on the Genetics of Electroconvulsive Therapy
17 and Severe Depressive Disorders (Gen-ECT-ic) may be able to determine the role of the
18 different BDNF genetic polymorphisms in ECT response ⁴⁴.
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30 In conclusion, and regarding to implications for clinical practice, in this study we have
31 identified a decrease of BDNF plasma levels after 2 hours of an ECT session. Nevertheless,
32 predictive value of BDNF for effects of ECT remains uncertain. The results indicate that this is
33 an area of interest for future research, in which the role of neurotrophins in the response and
34 safety of ECT should be clarified.
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46 **Figure 1:** Mean differences (SD; univariate analysis) on biomarkers at baseline (Pre) and two
47 hours after (Post) a single electroconvulsive session. BDNF and NGF plasma levels (a, b);
48 Protein expression of TrkA, TrkB-F and TrkB-T in PBMC (c-e); Ratio of TrkB-F/TrkB-T expression
49 (f). BDNF: Brain-derived neurotrophic factor; NGF: Nerve growth factor; PBMC: Peripheral
50 blood mononuclear cells; TrkA: Tyrosine kinase-type A receptor; TrkB-F: Full-length form of
51 the tyrosine kinase-type B receptor; TrkB-T: Truncated-length form of the tyrosine kinase-type
52 B receptor.
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3 **Conflicts of interest**
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6 Dr. Bernardo has been a consultant for, received grant/research support and honoraria from,
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8 and been on the speakers/advisory board of ABBiotics, Adamed, Angelini, Casen Recordati, Eli
9
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21
22 from, and been on the speakers/advisory board of has received honoraria from talks and/or
23
24 consultancy of Adamed, Ferrer, Janssen-Cilag, Lundbeck, Otsuka, Pfizer and Sanofi. Dr. Valentí
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30 report no competing interests for this study.
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Contributors

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2
3 MBI conducted the literature review, recruited the participants, applied the ECT session,
4
5 collected data, conducted the main statistical analysis, wrote the first draft of the manuscript
6
7 and handled subsequent drafts after receiving coauthors feedback. KSMD & CF, performed all
8
9 biochemical determinations in plasma and in cells and prepared sub-cellular samples, assisted
10
11 with the analysis and wrote the first draft of the manuscript. AM collected the biological
12
13 samples and separated plasma and PBMC. EM recruited the participants, collected the blood
14
15 samples and applied the ECT session. The rest of coauthors participated in the ECT procedures,
16
17 collected data and commented on drafts. All of the authors contributed to the final version of
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19 the paper.
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Table 1. Demographic, clinical characteristics and electroconvulsive therapy parameters

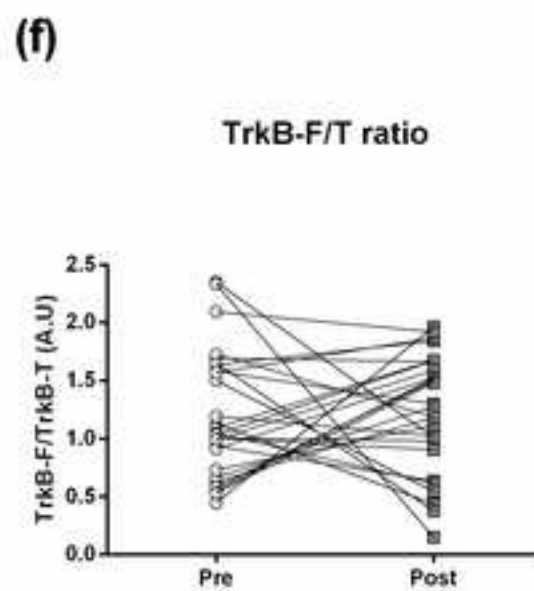
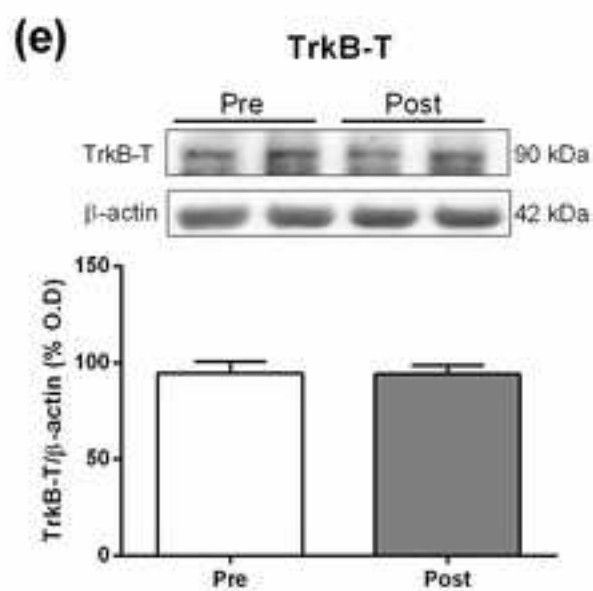
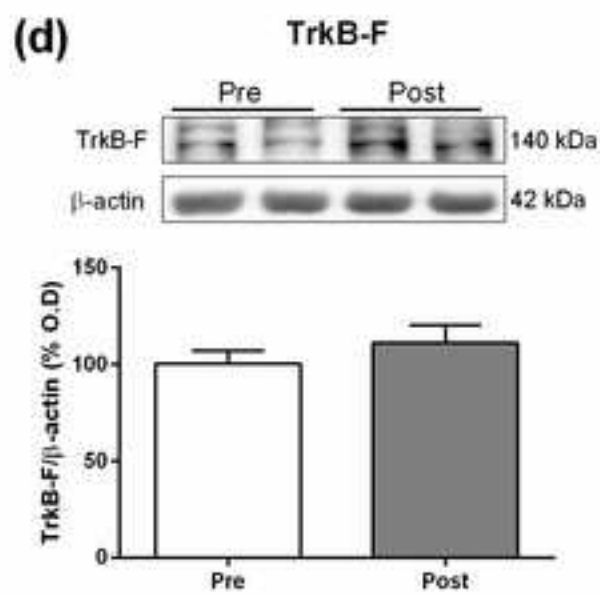
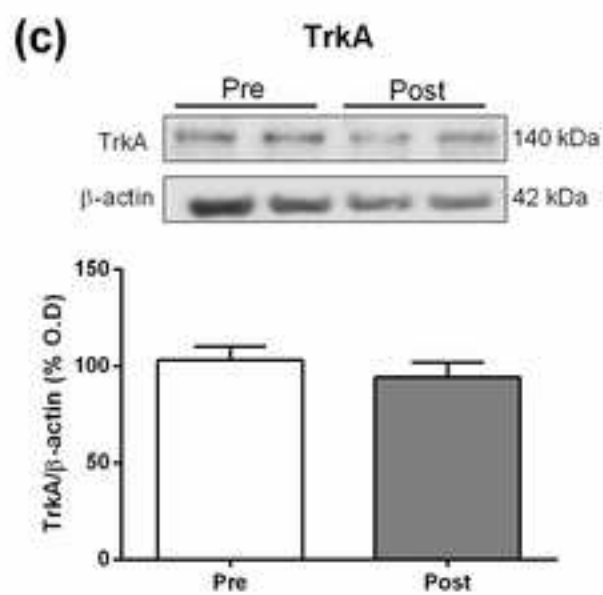
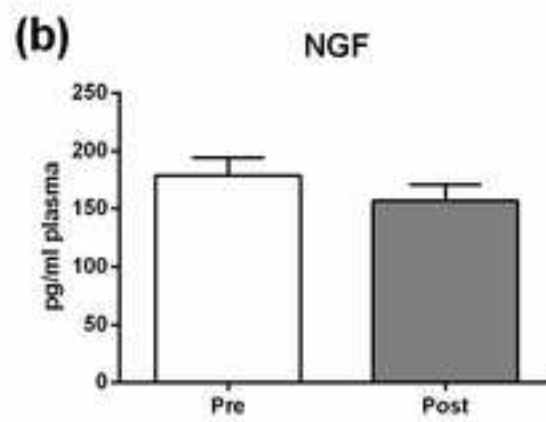
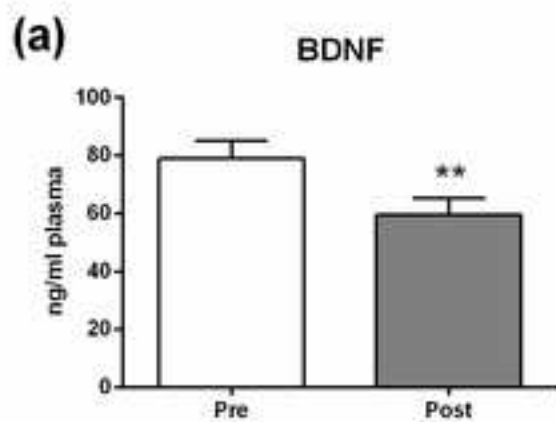
	n=30
Age – years [mean (sd)]	52,97 (16,81)
Gender – no. (%)	
Female	19 (57,7%)
Male	11 (33,3%)
Diagnosis – no. (%)	
Affective Psychosis	24 (80%)
Non-affective Psychosis Diagnosis	6 (20%)
Duration of illness – years [mean (sd)]	19,03 (12,11)
Subjects with an official work disability - no. (%)	26 (76,5%)
ECT regimen – no. (%)	
Acute	6 (20,0%)
Continuation/Maintenance	24 (80%)
ECT application – no. (%)	
Right unilateral	3 (10%)
Bitemporal	27 (90%)
Lifetime ECT number of sessions	60,77 (66,27)
Duration (seconds) of EEG convulsion	40,27 (12,49)
Subjects with psychopharmacological treatments – no. (%)	
Antidepressants	21 (61.8)
Antipsychotics	25 (73.5)
Benzodiazepines	20 (50.8)
Anticonvulsants	7 (20.6)
Lithium	7 (20.6)
Anticholinergics	3 (8.8)
Fluoxetine equivalent mean daily dose of antidepressants - mg/d (sd)	64.39 (± 36.64)
Chlorpromazine equivalent mean daily dose of antipsychotics - mg/d (sd)	668.21 (± 508.84)

In ECT regimen: “Acute” refers to the first 6-12 ECT sessions, “Continuation” for the following sessions during the first six months and “Maintenance” for the sessions received after this first six-month period. ECT: electroconvulsive therapy; EEG: electroencephalographic.

Table 2. Mean differences (\pm SD) in neurotrophins plasma levels and expression of receptors in PBMCs between pre-ECT and post-ECT session (2 hours after stimulus) in 30 patients with severe mental disorders.

Marker	PRE-ECT	POST-ECT	Statistics	p-value
BDNF plasma, ng/ml	82,16 \pm 37,22	63,53 \pm 39,99	Z=-2,70	0,007
NGF plasma, pg/ml	178,78 \pm 82,39	156,96 \pm 76,75	Z=-1,59	0,112
TrkBF WB, % OD	101,74 \pm 36,11	111,86 \pm 50,04	t=-0,91	0.368
TrkBT WB, % OD	93,98 \pm 34,27	94,12 \pm 24,65	t=-0,02	0.986
TrkBF/TrkBT	1,209 \pm 0,54	1,22 \pm 0,51	t=-0,08	0.938
TrkA WB, % OD	103,07 \pm 38,14	95,26 \pm 41,49	t=0,82	0.416

Data expressed as mean \pm standard deviation. BDNF: Brain-derived neurotrophic factor; NGF: Nerve growth factor; OD: Optical density; Plasma: plasma levels; TrkA: Tyrosine kinase-type A receptor; TrkBF: Full-length form of the tyrosine kinase-type B receptor; TrkBT: Truncated-length form of the tyrosine kinase-type B receptor; WB: Western Blot. See Methods section for detail.



Conflicts of interest

Dr. Bernardo has been a consultant for, received grant/research support and honoraria from, and been on the speakers/advisory board of ABBiotics, Adamed, Angelini, Casen Recordati, Eli Lilly, Janssen-Cilag, Lundbeck, Otsuka, Takeda, Somatics and has obtained research funding from the Ministry of Education, Culture and Sport, the Spanish Ministry of Economy, Industry and Competitiveness (CIBERSAM), by the Government of Catalonia, Secretaria d'Universitats i Recerca del Departament d'Economia i Coneixement (2017SGR1355), Foundation European Group for Research In Schizophrenia (EGRIS), and the 7th Framework Program of the European Union. Dr. Bioque has been a consultant for, received grant/research support and honoraria from, and been on the speakers/advisory board of has received honoraria from talks and/or consultancy of Adamed, Ferrer, Janssen-Cilag, Lundbeck, Otsuka, Pfizer and Sanofi. Dr. Valentí has received research grants from Eli Lilly & Company and has served as a speaker for Abbott, Bristol-Myers Squibb, GlaxoSmithKline, Janssen-Cilag, and Lundbeck. The rest of authors report no competing interests for this study.