

NH₂ as a directing group: from the cyclopalladation of amino esters to the preparation of benzolactams by Pd(II)-catalyzed carbonylation of *N*-unprotected arylethylamines

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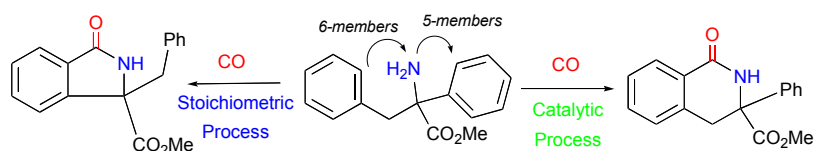
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TOC



Abstract

An unusual NH_2 -directed Pd(II)-catalytic carbonylation of quaternary aromatic α -amino esters to yield benzolactams has been developed. The steric hindrance around the amino group is pivotal for the success of the process. The stoichiometric cyclometallation of a variety of amino esters has been studied in order to evaluate the influence of the different variables (size of the metallacycle, aromatic ring substituents, and steric bulk) in the process, and a complete kinetic-mechanistic study of the cyclopalladation process has been carried out. The experimental results indicate that the full substitution of the carbon in the α position of the amino esters plays an important role in their cyclopalladation reaction. The reaction shows a strong bias to 6-membered lactams over the 5-membered analogues, which can be explained by a greater reactivity of the six-membered palladacycles.

Introduction

The development of selective methods for the direct conversion of carbon-hydrogen bonds into carbon-heteroatom and carbon-carbon bonds remains a critical challenge in organic chemistry. An interesting approach to address this issue involves the use of substrates that contain coordinating atoms (or directing groups)¹ which bind to the metal center in a first step; a further rearrangement of some atoms allows the C–H bond activation. The latter process, at stoichiometric scale, is the well-known cyclometallation reaction.² The first cyclometallated compounds were reported in the mid 1960's.³ Since then, this reaction has been extensively studied and has acquired a great interest given the application of metallacycles in many areas which include organic synthesis, catalysis, design of metalomesogens and antitumoral drugs, asymmetric synthesis, resolution of racemic ligands, intermolecular aromatic C-H bond activation, or in the synthesis and reactivity of organometallic complexes with biologically relevant ligands.⁴ In this respect, the development of ligand-directed reactions has led to a renewed interest in the cyclometallation reactions. Palladium complexes are particularly attractive catalysts for such transformations because ligand-directed C-H functionalization at palladium centers can be used to obtain different types of C-Y bonds (Y being carbon, oxygen, nitrogen, sulfur, or halogen). Furthermore, palladium can activate C-H bonds both at sp^2 and sp^3 sites and a wide range of catalytic processes has been described with different nitrogen-based directing groups. These include imines, oxime ethers, azobenzenes, amides, *N*-alkylanilines, benzodiazepines, pyridines, pyrazoles and isoxazolines. Oxygen-based ligands, such as carboxylic acids and aldehydes, have also been used as directing groups in some cases.^{1,5} In contrast, to the best of our knowledge, the use of primary amines as directing groups has not been described so far.

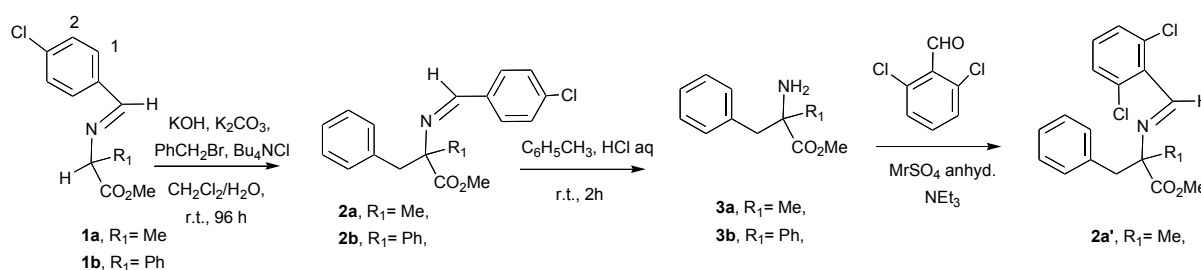
The transition-metal-catalyzed carbonylation of arenes with gaseous CO is a significant chemical transformation, since it extends the carbon chain length and also introduces a synthetically versatile carbonyl group. Arenes were first carbonylated to obtain carboxylic acids by Fujiwara *et al.* in 1980 using $Pd(AcO)_2$ under 15 atm of CO, and the arene substrates as solvent.⁶ However, no control over regioselectivity was observed for substituted arenes. This problem has been overcome by different research groups using the directing group approach.^{1,7} Thus, Yu *et al.* has very recently described the palladium acetate-catalyzed carbonylation of anilides to obtain *N*-acyl anthranilic acids.^{7d} Similarly, Orito *et al.* have reported the direct carbonylation of aromatic C-H bonds with CO in *N*-alkyl- ω -

arylalkylamines to obtain benzolactams using a Pd(AcO)₂/Cu(AcO)₂/air system in toluene solution at 120 °C.^{7b,8} However, the authors stated: “*carbonylation of primary amines, including benzylic amines or phenylethylamines, under the same conditions, produced no benzolactams but produced ureas in good yields*”. It should be noted that Pd(II) catalysts are readily reduced by CO, in a reaction that also produces Ac₂O, which could cause secondary reactions with primary amines.⁹ Thus, a method for catalyzed C-H activation/carbonylation of primary amines under a CO environment has not been established.

Here we describe the preparation of benzolactams *via* palladium acetate-catalyzed aromatic carbonylation of quaternary α -amino α -alkyl esters, by an unusual process that uses NH₂ as a directing group.¹⁰

Results and Discussion

As part of an ongoing research project on bioorganometallic chemistry,¹¹ we attempted the cyclometallation of imines RCH=NC(Me)(CH₂Ph)(COOMe) (**2a**: R = 4-ClC₆H₄; **2a'**: R = 2,6-Cl₂C₆H₃), derived from a quaternary α -amino ester, with Pd(AcO)₂ in toluene or acetic acid solution. Ligand **2a** was obtained by alkylation of imine **1a** with a mixture of KOH, K₂CO₃ and benzyl bromide and ligand **2a'** was prepared from amino ester **3a** (obtained by hydrolysis of the imine **2a**) *via* a condensation reaction with 2,6-dichlorobenzaldehyde. The direct alkylation of the imine **1a'**, 2,6-Cl₂C₆H₃CH=NCH(Me)(COOMe), gave low yield of **2a'**, probably the steric hindrance of the di-*ortho* chloro substituted fragment (Scheme 1) **hampers** the process.



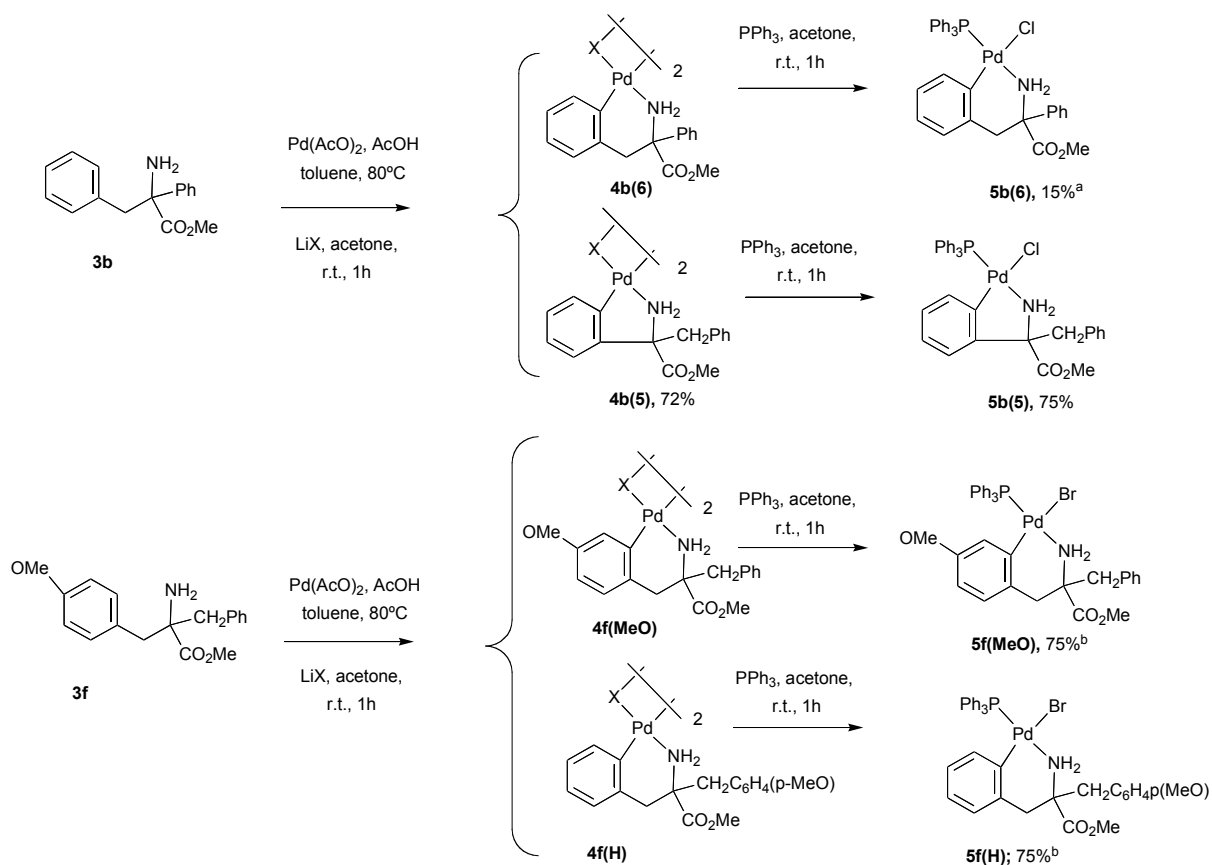
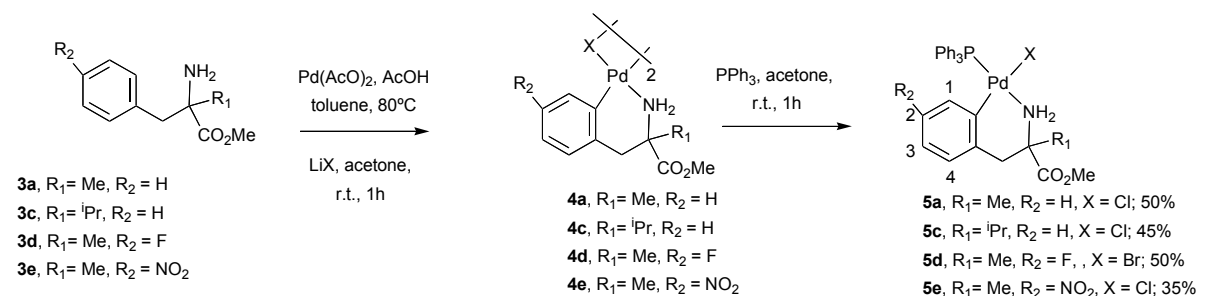
Scheme 1. Synthesis of imines **2** and aminoesters **3**.

Treatment of imines **2a** and **2a'** with Pd(AcO)₂ in toluene or acetic acid solution at 80 °C did

not produce the expected imine palladacycles. Only compound **5a**, [PdCl(CN)(PPh₃)] (being CN the metallated amino ester) was isolated in low yield, after subsequent reaction with LiCl and PPh₃. Reaction of the free amino ester **3a** under the same conditions produced palladacycle **5a** in a much better yield, 50% (Scheme 2). Nevertheless, proton NMR batch monitoring of the reaction between **2a** and palladium acetate, under milder conditions, allowed the detection of the metallated imine (see below).

The preferential metallation of the aminoesters with respect to the corresponding imine derivatives is an unexpected result because closely related imines, derived from methyl glycinate, alaninate, valinate and tyrosinate, have been reported to metallate in good yields;^{11c} furthermore, the cyclometallation of primary amines has always been considered problematic.¹²

Bearing these facts in mind we tried to extend the above cyclopalladation process to amino esters **3b-f** (Scheme 2), readily obtained by alkylation of the imines arising from commercial α -amino acids or α -amino esters hydrochlorides by standard procedures.¹³ These were selected in order to study the influence of the different variables (size of the metallacycle formed, substituents in the aromatic ring, and steric bulk) in the process. Thus, ligand **3f** seems especially interesting due to the fact that it can afford two different metallacycles: one by activation of an unsubstituted benzene ring and the other by metallation of a MeO substituted aromatic ring. Ligand **3b** is also remarkable as it can afford either a five- or a six-membered metallacycle, depending on which aromatic ring undergoes the metallation reaction.



Scheme 2. Synthesis of cyclometallated compounds **4** and **5**.
 Compounds **4a**, **4b(6)**, **4c**, **4d**, **4e** and **4f** could not be purified.
^a Detected by proton NMR.
^b Mixture 1: 1 of regioisomers

The reaction between amino esters **3a-f** and palladium acetate in toluene at 80 °C afforded, as expected, the dinuclear acetato bridged compounds **4a-f** (X = AcO). Dinuclear halide bridged compounds **4a-f** (X = Cl or Br) can also be obtained by reaction of the dinuclear acetato

bridged compounds with lithium halide in acetone at room temperature (30 minutes). Unfortunately, all attempts to purify the dinuclear six-membered metallacycles, either by column chromatography or by recrystallization, were unsuccessful, the complexes partially decomposing during the purification processes. In contrast **4b** (X = Br) can be purified by flash chromatography (hexane/EtAcO 8:2) to afford the five-membered palladacycle **4b(5)** in 72% yield (see experimental). Consequently, the preparation of the more stable mononuclear compounds **5** was attempted (see Scheme 2). The reaction was carried out from the acetato derivatives **4** (X = AcO) by reaction with PPh₃ and the corresponding lithium halide. Compounds **5** could be purified by column chromatography and isolated in good yields, as expected.

The yields of the metallation reactions of amino esters **5a**, **5d** and **5e** to afford the corresponding six-membered metallacycles do not vary significantly, showing that the substituents on the aromatic ring that experience metallation do not play a significant role in process. This fact has already been observed in the kinetic-mechanistic studies carried out on a family of carefully tuned imine derivatives.¹⁴ Furthermore, metallation of both aromatic rings, in a equimolar ratio, is observed with ligand **3f**, which agrees with the non-electrophilic substitution behavior of the observed process. The sequence is better explained through a mechanism involving a concerted proton abstraction from the metallating C-H unit by an ancillary ligand. The trends observed in the thermal and pressure activation parameters obtained in the above-mentioned studies suggest that once the C-H bond being activated reaches its correct positioning, the process is fairly independent of its electronic nature.¹⁴ Recent computational studies of C-H bond activation at late transition metal systems indicate that assistance *via* co-ligands (especially carboxylates) is a good way of cleaving these bonds; the term ‘ambiphilic metal ligand activation’ has been proposed to describe such reactions.¹⁵ In contrast, the size of the metallacycle seems to play an important role in the cyclometallation reaction; metallation of ligand **3b** affords the five-membered metallacycle as the major isomer with respect to the six-membered analogue (6:1 ratio, by proton NMR).

Good XRD-quality crystals of compounds **5a**, **5c** and the six-membered ring isomer of **5b**, **5b(6)**, were obtained by vapor diffusion at 298 K of mixtures CH₂Cl₂/MeOH; CH₂Cl₂/C₆H₁₄ and MeC₆H₅/C₆H₁₄ respectively (Figures 1-3). It should be noted that the six-membered metallacycle **5b(6)**, crystallizes from the mixture of five- and six-membered **5b** compounds, despite **5b(6)** being the minor component of the mixture. The strong intermolecular

interactions present in the structure of this compound (see below) easily explain this fact.

For all compounds the distances between palladium and the coordinated atoms are similar to those reported and the smallest angle in the coordination sphere of palladium corresponds to the C-Pd-N bite angle (Table 1).¹⁶ The phosphorus and nitrogen atoms adopt a *trans* arrangement, the six-membered metallacycle has a boat conformation in all cases, and the coordination plane shows a slight tetrahedral distortion in both **5a** and **5c**.

The structures of compounds **5c** and **5b(6)** reveal diverse intermolecular interactions: conventional NH \cdots O hydrogen bonds and non-conventional CH \cdots O and CH \cdots X hydrogen bonds. In contrast the crystal structure of **5a** reveals only non-conventional C-H \cdots O hydrogen bonds.

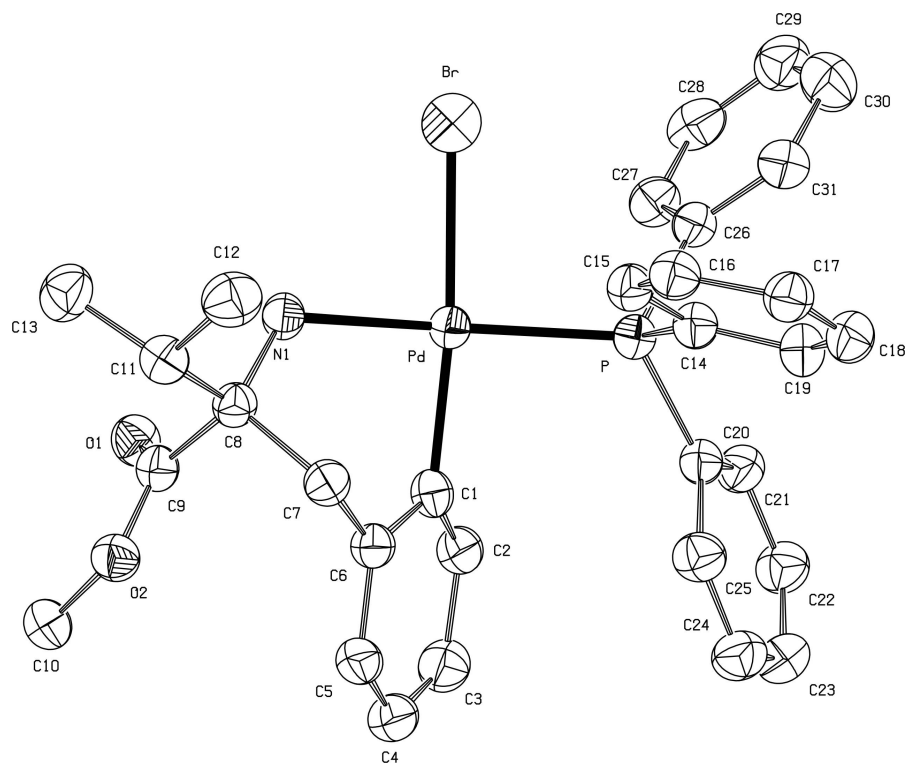


Figure 3. ORTEP plot of **5c**. Hydrogen atoms have been omitted for clarity.

Table 1. Selected bond lengths (in Å) and bond angles (in deg.) of **5a**, **5c**, and **5b(6)**.

	5a	5c	5b(6)
<i>Bond lengths</i>			
Pd(1)-C(1)	1.986(3)	2.037(5)	2.003(2)
Pd(1)-N(1)	2.136(3)	2.134(4)	2.131(2)
Pd(1)-P(1)	2.2488(15)	2.255(2)	2.2507(11)
Pd(1)-X(1)*	2.4198(13)	2.4874(14)	2.4056(9)
N(1)-C(8)	1.480(4)	1.480(6)	1.502(3)
O(2)-C(9)	1.320(4)	1.349(6)	1.333(3)
O(2)-C(10)	1.459(5)	1.445(6)	1.453(3)
O(1)-C(9)	1.203(4)	1.180(6)	1.210(3)
<i>Bond angles</i>			
C(1)-Pd(1)-N(1)	84.61(11)	89.14(17)	88.29(9)
C(1)-Pd(1)-P(1)	93.52(9)	92.76(14)	93.75(7)
N(1)-Pd(1)-X(1)*	86.08(9)	85.96(12)	85.41(6)
P(1)-Pd(1)-X(1)*	96.12(5)	92.78(6)	92.58(3)
C(7)-C(8)-N(1)	109.6(3)	109.1(4)	109.28(18)
C(8)-C(9)-O(1)	124.6(3)	123.9(4)	123.8(2)
C(8)-C(9)-O(2)	111.4(3)	111.0(4)	112.0(2)

- X = Cl for **5a** and **5c(6)**, and X = Br for **5c**.

The unexpected results found in the cyclopalladation of some of these ligands prompted us to carry a complete kinetic-mechanistic study on the reaction between palladium acetate, amino esters **3a**, **3b** and **3f**, and imine **2a** in toluene solution. The experiments were performed at [Pd]:[N-donor ligand] ratios within the 0.9-1.1 margin to avoid the formation of the dead-end *trans*-[Pd(AcO)₂(N-donor)₂] species, while having practically all the reactants as {Pd(AcO)(N-donor)} metallating units.¹⁷ The reactions were monitored by UV-Vis spectroscopy in the full 300-800 nm range, with *absorbance versus time* traces derived where larger differences were detected. The kinetic and thermal and pressure activation parameters thus obtained are collected in Table 2 (Figure 4). The parameters corresponding to the cyclopalladation of the *N*-benzylideneamine and benzylamine, previously reported^{12b,17b} have also been included in the same table for comparative purposes. The values obtained fall with the range of the values determined for similar systems, involving a large organization on going to the transition state which is accompanied by an important contraction in volume.

Table 2. Kinetic and activation parameters for the cyclometallation reactions studied.

Metallating ligand	Solvent	$10^4 \times {}^{350}k$ /s ⁻¹	ΔH^\ddagger /kJ mol ⁻¹	ΔS^\ddagger /J K ⁻¹ mol ⁻¹	ΔV^\ddagger /cm ³ mol ⁻¹
<i>N</i> -benzylamine ^{12b}	Toluene	22	73	-91	-16
	Acetic acid	100	100	3	-11
3a	Toluene	170	81±2	-50±6	-13±1
	Acetic acid	53	49±2	-152±7	-16±2
3b	Toluene	460	71±4	-71±14	not measured ^a
5-membered metallation contribution ^b		580	70±1	-72±1	
6-membered metallation contribution ^b		350	76±2	-59±7	
3f	Toluene	520	50±2	-130±5	not measured ^a
<i>N</i> -benzylidenebenzylamine ^{17b}	Toluene	24	52±3	-150±10	-15±2
2a	Toluene	4.8	62±3	-135±10	-14±2

^a Not measured given the impossibility of separation of the 5- and 6-membered cyclometallation contribution under the conditions needed.

^b Proton NMR monitoring at different temperatures of the final 5- to 6-membered cyclometallated compound ratio allows the estimation of $k_{5\text{-membered}}/k_{6\text{-membered}}$ value and thus the contribution of each reaction to the overall process.

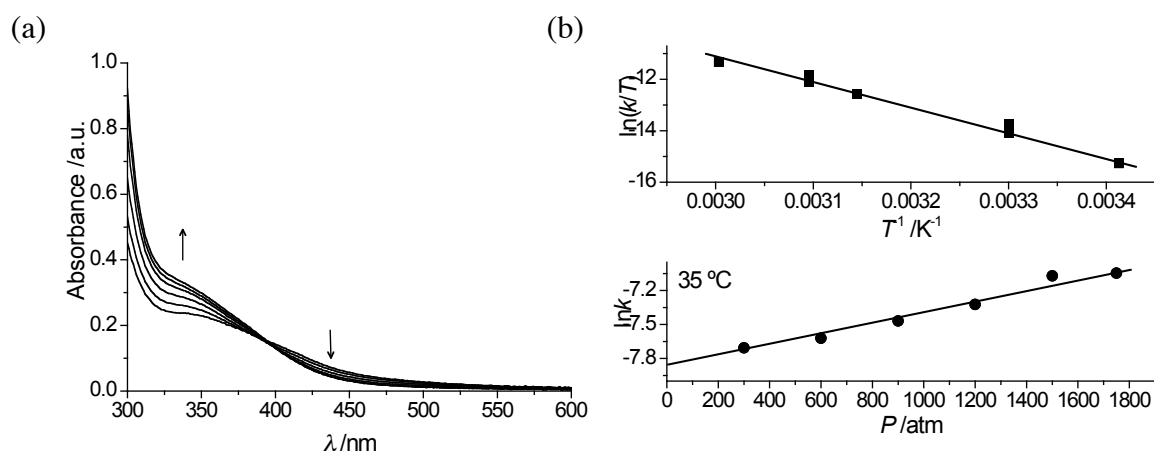


Figure 4.- (a) UV-Vis spectral changes observed for the reaction of an equimolar mixture of palladium acetate and amino ester **3b** in toluene at 30 °C, total time 150 minutes. (b) Eyring (top) and $\ln k$ versus P (bottom) plots for the reaction of amino ester **3a**.

At a first glance it is evident that the cyclometallation of amino esters **3a**, **3b** and **3f** in toluene are definitively faster than for other N-donor ligands previously studied.^{14,17} For amino esters **3a** and **3b** it is also clear that this effect is related with a noticeable decrease in ΔS^\ddagger requirements, while for amino ester **3f** the difference in ΔH^\ddagger is responsible for this fact. In this respect, the feasibility of the separation of the five- and six-membered contributions to the metallation of amino ester **3b**, by NMR measurements on the final reaction mixture, allows a deeper view of the differences. While the differences in ΔH^\ddagger favor the formation of the five-membered metallacycles, the entropic terms indicate a less demanding process for the formation of the six-membered derivative, probably due to the greater flexibility of the starting material arrangement.

A further comparison is also possible between the cyclometallation of amino ester **3a** and its corresponding imino derivative, **2a**; for this system the cyclometallation of the imine derivative is *ca.* 30-fold slower than that of the amino ester. The effect is, nevertheless, not originated in the separate values of ΔH^\ddagger or ΔS^\ddagger , and the acceleration decreases to 10-fold at 300 K due to the much larger temperature dependence of the reaction rate of the amino ester.¹⁸ Probably the higher rigidity of the starting imine material induces the need of a more demanding organization on going to the transition state providing lesser enthalpic

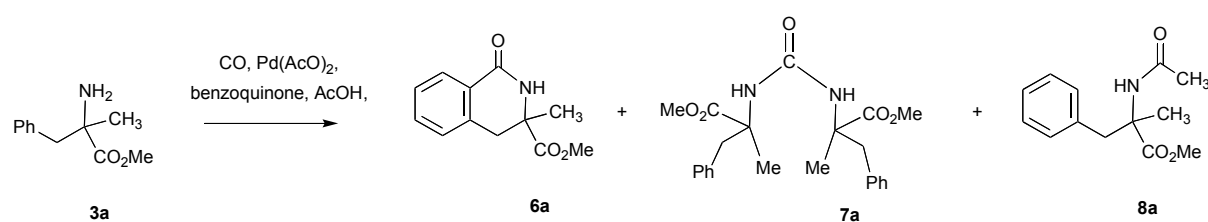
requirements. This is especially relevant as far as the initial preparative observation (see before) of compound **4a** as the solely metallated compound on reaction of **2a** with Pd(AcO)₂ at 80 °C. It is evident that the metallated Pd(II) center promotes the hydrolysis of the cyclometallated imine derivative under the preparative conditions and the consequent faster metallation of the amino ester occurs within the reaction mixture. Some preliminary kinetic runs indicate that the cyclometallated **2a** derivative undergoes the C=N bond hydrolysis process at a rate of $3.5 \times 10^{-5} \text{ s}^{-1}$ at 350 K in toluene solution, producing compound **4a**.

Given the fact that an acceleration of the cyclopalladation reactions is observed in protic media, due to the formation of a highly ordered ambiphilic transition state very sensitive to the presence of any protons, the use of acetic acid as solvent was also tried in the present study.^{14,19} Surprisingly, the monitoring of the cyclopalladation reaction in acetic acid of amino ester **3a** does not produce the expected rate enhancement. In fact, the values measured for ΔH^\ddagger , ΔS^\ddagger and ΔV^\ddagger follow the opposite trend from what have been observed previously: *i.e.* increase in the enthalpic demands and less negative entropy, accompanied by practically the same compression. Obviously, the existence of some interactions between the polar groups of the amino ester and the acetic acid used as a solvent have to be considered responsible for this observed difference. In this respect, the X-ray determined structure of **5a**, **5c** and **5b(6)** (see above) clearly shows the tendency of these amino esters to form hydrogen bond interactions. This lack of enhancement of the metallation process in acetic acid solution is rather relevant for the catalytic results indicated below. It is clear from the results obtained from the kinetic experiments that the effect of the acetic acid solvent in the catalysis cannot be related to the formation of the cyclometallated derivative.

Summarizing, results indicate that the full substitution of the carbon in the α position of the amino esters play a pivotal role in their cyclopalladation reaction behavior. The difference between the ΔS^\ddagger values of metallation of aminoester **3a** ($-50 \text{ J K}^{-1} \text{ mol}^{-1}$) and benzylamine ($-91 \text{ J K}^{-1} \text{ mol}^{-1}$) suggests that this effect is related with a noticeable decrease in ΔS^\ddagger requirements. This specificity can also related with the Thorpe–Ingold effect^{2b,20} (or *gem*-dimethyl effect) that improves the outcome of organic cyclization reactions when alkyl substituents are present on the acyclic carbon backbone. Nevertheless the role played by others factors such as ΔH^\ddagger or the hydrogen bond interactions, when acetic acid is used as a solvent, can not be discarded.

Catalytic results

The great tendency showed by the free amino esters studied to undergo cyclopalladation prompted us to study their palladium-catalyzed NH_2 -directed carbonylation at low pressure. Initially palladacycle **4a** was carbonylated to benzolactam **6a** with CO (1 atm) in different solvents at room temperature, thus indicating the feasibility of the process. From then the palladium acetate-catalyzed carbonylation of racemic amino acid **3a**, using $\text{Cu}(\text{AcO})_2/\text{O}_2$ as the oxidant in toluene, was studied. Unfortunately, only urea **7a** (Table 3) was obtained under different experimental conditions, not the expected benzolactam **6a**. Given the strong accelerating effect of acetic acid in $\text{Pd}(\text{AcO})_2$ -catalysed reactions,²¹ we swapped to this solvent. Under these conditions, the desired **6a** lactam was obtained although contaminated with acetamide **8a**. The best ratio being $\text{6a}/\text{8a} = 64/36$ in a 91% yield. Since the formation of acetamide may also be favored by the Cu(II) salts which can enhance the amide bond formation,²² an alternative oxidant was tried; benzoquinone proved to be the better choice (Table 3). The best results were obtained with a 1.5×10^{-2} M solution of **3a** in refluxing AcOH using 5% molar $\text{Pd}(\text{AcO})_2$ and an amino ester/benzoquinone molar ratio of 1:2. Under these conditions the yield was 91% and the benzolactam/acetamide ratio was 90 : 10 (entry 3).

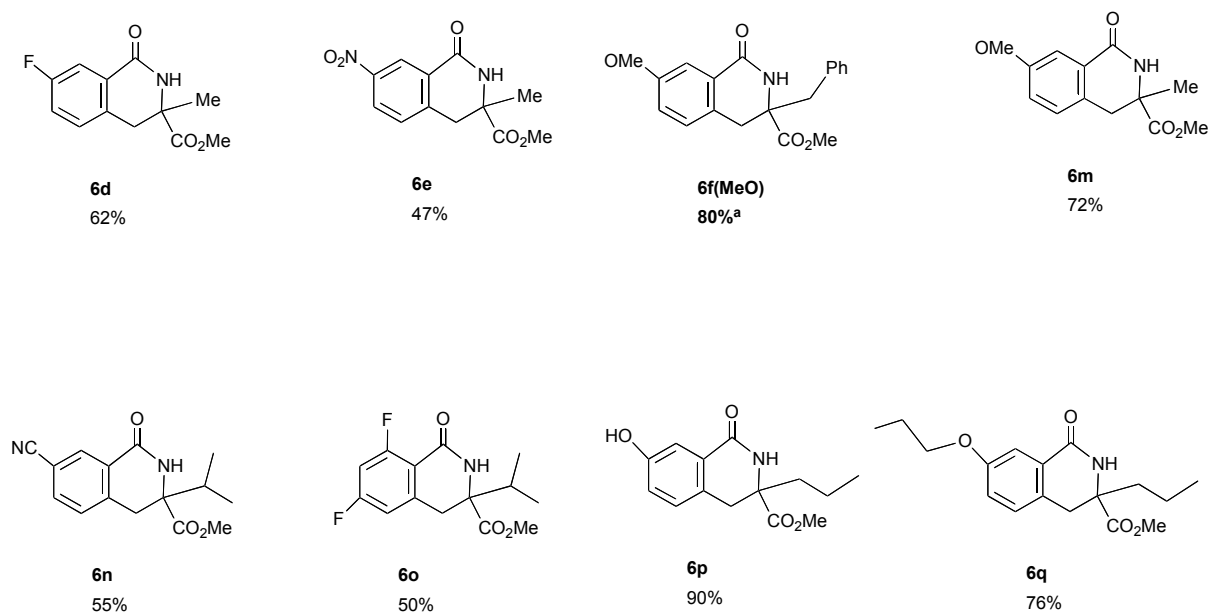
Table 3. Optimization of carbonylation of **3a**

Entry	t/h	T/°C	Benzoquinone	Overall Yield	6a/7a/8a
			(% molar)	(%)	ratio
1 ^a	6	Reflux	100	98	80 : — : 20
2	6	65	100	95	58 : — : 42
3	6	Reflux	200	91	90 : — : 10
4 ^b	6	Reflux	100	92	70 : — : 30
5	6	Reflux	135	98	86 : — : 14
6	3	Reflux	200	94	84 : — : 16

^a 2% molar of Pd(OAc)₂. ^b Two-fold **3a** and benzoquinone concentration.

The catalytic process was successfully expanded to other racemic phenylethylamines and to some lactams substituted in the aromatic ring. The results are shown in table 4 and scheme 3, respectively. From the data it is clear that the steric hindrance of the R and R' groups plays a crucial role in the process. Thus, the carbonylation of methylphenylalaninate produced a rather low benzolactam/acetamide ratio: 46/54 (R' = H, entry 4), which improved for compound **3a** (Table 3). Even no acetamides were found in the preparation of **6g**, **6f** and **6h** bearing larger R' groups (entries 1–3, R' = propyl, benzyl, and *para*-methoxybenzyl respectively). An increase in the steric hindrance around the amino group prevents competitive acetylation. Interestingly, the presence of the ester group is not essential for the success of the catalytic carbonylation (entry 6). However, the presence of a neighboring

coordinating hydroxymethyl or allyl group erodes or inhibits completely the formation of benzolactam (entries 7 and 5, respectively). It should be also noted that the presence of MeO, CN or F groups on the aromatic ring (**6f(MeO)** and **6m-o** in Scheme 3) is compatible with the formation of lactam.



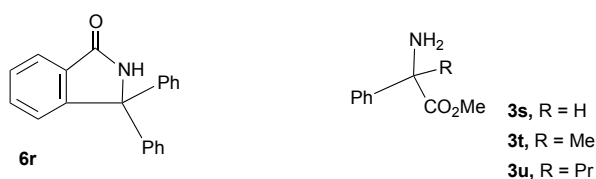
Scheme 3. Benzolactams substituted on the aromatic ring.
^aMixture of regioisomers (see table 4)

The reaction is also found highly sensitive to the size of the benzolactam formed, no five-membered lactams were detected when the reaction was performed with ligands **3s**, **3t** and **3u** (Scheme 4). Nevertheless a 57% yield of the five-membered lactam **6r** was obtained from triphenylmethylamine with a total selectivity. It should be noted that, in this last case, the corresponding amino ester presents a larger steric hindrance around the amino group. These results are in sharp contrast with those reported by Orito *et al.*⁸ for the related carbonylation of secondary amines using Cu(II) as co-oxidant in which the five-membered benzolactams were favored over the six-membered analogues.

Table 4. Carbonylation of phenylethylamines

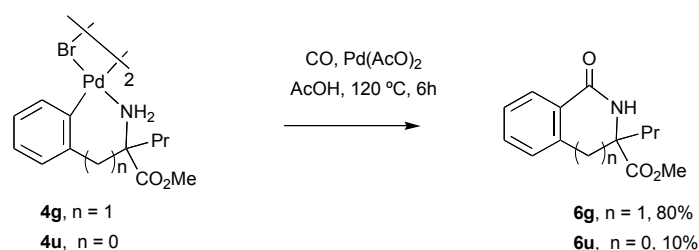
Entry	R	R'	Lactam	Overall Yield (%)	Lactam/acetamide ratio
1	CO ₂ Me	Propyl	6g	98	100 : 0
2	CO ₂ Me	Bn	6h	93	100 : 0
3	CO ₂ Me	p-MeO-Bn	6f	80 ^a	100 : 0
4	CO ₂ Me	H	6i	91	46 : 54
5	CO ₂ Me	Allyl	6j	—	— ^b
6	Me	Me	6k	89	82 : 18
7	CH ₂ OH	Bn	6l	85	50 : 50 ^c

^a Mixture of regioisomers (**6f(H)**/**6f(MeO)**) = 6 : 4). ^b Complex mixture of compounds. ^c Lactam **6l** is not acetylated on the hydroxyl group.

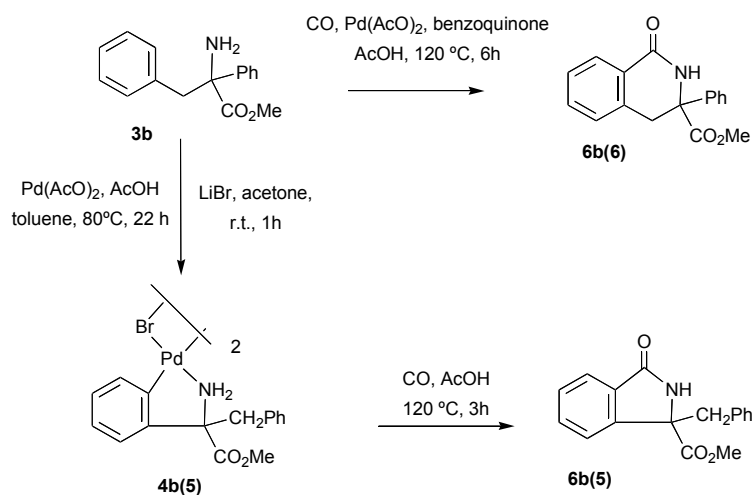
**Scheme 4.** Benzolactam **6r** and methyl phenylglycinates **3s-u**

A greater reactivity of the six-membered palladacycles involved could explain this result. To assess this assumption, the reactivity of the six- and five-membered cyclopalladated derivatives **4g** and **4u** (Scheme 5) with CO was studied. Thus, after 1 hour of reaction at 50 °C, the six-membered benzolactam **6g** was obtained in a 80% yield from **4g** whereas **4u** afforded only a 10% of compound **6u**, in full agreement with the catalytic results (Scheme 5).

In contrast, the catalytic carbonylation of **3b** produced only lactam **6b(6)** which originates from the six-membered metallacycle, despite the fact that the stoichiometric cyclometallation of **3b** favors the five-membered palladacycle **4b(5)** as a major product in the **4b** product mixture (Scheme 2). Nevertheless, carbonylation of a pure sample of **4b(5)** in refluxing AcOH afforded the corresponding five-membered lactam **6b(5)** in 86% yield, thus indicating that both benzolactams sizes (six or five) are attainable depending on the carbonylation method (catalytic or stepwise) (Scheme 6). **The results indicate that, even though both five-membered and six-membered palladacycles are capable of carbonylation to produce the corresponding lactams, the latter reacts more quickly with CO affording **6b(6)** as the only isomer.**



Scheme 5. Carbonylation of **4g** and **4u**



Scheme 6. Catalytic and stepwise carbonylation of **3b**.

Conclusions

An adequate selection of the R groups positioned on the acyclic carbon backbone of phenylethylamines and benzylamines allows an unprecedented NH_2 -directed catalytic carbonylation with high selectivity and yield. Kinetic results of the cyclometallation process indicate that the formation of the metallated palladium intermediate is much faster than for other systems, even though the presence of protic medium does not favor such process as for other N-donor ligands. Furthermore, these studies show that the effect of acetic acid as the solvent in the catalytic process can not be related to the formation of the cyclometallated derivative. The good catalytic results obtained with the system palladium acetate/acetic acid can be explained by the fact that some important steps are assisted by strong hydrogen bonding with AcOH molecules. Finally, the unexpected strong bias to the six-membered lactams over five-membered analogues can also be explained by the greater reactivity of the six-membered palladacycles formed. Studies designed to expand the process to other organic derivatives of interest are currently under way.

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Experimental

Materials and methods.

Amino esters, aldehydes, benzyl bromide, K_2CO_3 , KOH, benzoquinone, $PdCl_2$, $Pd(AcO)_2$, LiCl, LiBr and PPh_3 were obtained from commercial sources and used as received. Solvents were distilled and dried before use.²³

Elemental analyses were carried out at the Serveis de Científico-Tècnics (*Universitat Barcelona*). Mass spectra were performed at the Servei d'Espectrometria de Masses (*Universitat de Barcelona*). Infrared spectra were obtained with a Nicolet 400FTIR instrument using KBr pellets and only the most relevant absorptions of the new products are presented in the following sections. High-resolution 1H -NMR spectra and the two-dimensional $\{^1H-^1H\}$ -NOESY and COSY experiments were registered with a Varian VRX-500 or a Bruker Avance DMX-500MHz instruments. The solvent used for NMR experiments was $CDCl_3$ (99.9%) and the references were $SiMe_4$ [for 1H NMR] and $P(OMe)_3$ [$\delta(^{31}P) = 140.17$ ppm] for ^{31}P -NMR. The chemical shifts (δ) are given in ppm and the coupling constants (J) in Hz. In the characterization section of each product the assignment of signals detected in the 1H -NMR spectra refers to the labelling patterns presented in Schemes 1 and 2.

Procedures for synthesis and characterization of organic compounds **3** are given in the Supporting Information.

Preparation of the palladium(II) complexes

Compounds 4, Typical Procedure:

4b(5): A mixture of **3b** (500 mg, 1.96 mmol) and palladium acetate (439 mg, 1.96 mmol) in 40 mL of toluene was stirred at 80 °C for 22 h. The solvent was removed under vacuum and the residue was treated with lithium bromide (213 mg, 2.45 mmol) in acetone (40 mL) for 1 h at rt. The suspension was filtered to obtain 783 mg (91%) of a 6:1 mixture of 5-membered and 6-membered palladacycles (1H NMR of the crude). The crude was purified by flash chromatography (hexane/EtAcO 8:2) to afford 5-membered palladacycle **4b(5)** (625 mg, 72%). Compound **4b(5)**: Brownish solid; mp 212–214 °C; R_f (hexane/EtAcO 8:2): 0.32; 1H NMR (400 MHz; $CDCl_3$): δ 7.44 (3H, m, ArH), 7.33 (3H, m, ArH), 7.18 (1H, dd, $J = 7.7, 1.2$ Hz, ArH), 7.04 (1H, m, ArH), 6.94 (1H, m, ArH), 4.75 (1H, br d, $J = 10.3$, NHH), 3.86 (3H, s,

OCH₃), 3.72 (1H, d, *J* = 14.1 Hz, CHH), 3.61 (1H, d, *J* = 14.1 Hz, CHH), 3.67 (1H, br d, *J* = 10.3, NHH); ¹³C NMR (CDCl₃, 101 MHz): δ 171.1, 136.2, 133.9, 130.2, 130.0, 129.5, 129.4, 128.3, 127.2, 125.0, 123.0, 74.9 (q), 53.3 (OCH₃), 47.4 (CH₂); IR (KBr): ν_{max} 3296, 3256, 1730; HRMS (Maldi-TOF) calcd for C₃₂H₃₂BrN₂O₄Pd₂ (M-Br)⁺ 798.9609, found 798.9626. Anal. calcd for C₃₂H₃₂Br₂N₂O₄Pd₂: C, 43.61; H, 3.66; N, 3.18. Found: C, 43.7; H, 3.5; N, 3.3.

4g: It was obtained using the same procedure as that described above from 191 mg (0.87 mmol) of amine **3g** as brownish solid. Yield: 305 mg (87%). Mp 110-112 °C; ¹H NMR (300 MHz; CDCl₃): δ 7.38 (1H, d, *J* = 7.8 Hz), 6.90 (1H, m), 6.83 (1H, m), 6.75 (1H, dd, *J* = 7.3, 1.8 Hz), 4.46 (1H, br d, *J* = 11.3 Hz, NHH), 3.68 (3H, s, OCH₃), 3.59 (1H, d, *J* = 13.8 Hz, CHH), 3.25 (1H, d, *J* = 13.8 Hz, CHH), 3.14 (1H, br d, *J* = 11.3 Hz, NHH), 1.98 (1H, m), 1.84 (1H, m), 1.43 (1H, m), 1.24 (1H, m), 0.95 (3H, t, *J* = 7.2 Hz, CH₃); ¹³C NMR (CDCl₃, 101 MHz): δ 173.9 (COO), 136.8, 135.0, 127.5, 127.4, 125.6, 124.6, 59.7 (q), 53.2 (OCH₃), 51.5 (CH₂), 40.0 (CH₂), 17.7 (CH₂), 14.0 (CH₃); IR (KBr): ν_{max} 3302, 3237, 1726, 1571, 1558, 1435, 1230; HRMS (MALDI-TOF) calcd for C₂₆H₃₆BrN₂O₄Pd₂ (M-Br)⁺ 730.9928, found 730.9899; Anal. calcd for C₂₆H₃₆Br₂N₂O₄Pd₂: C, 38.40; H, 4.46; N, 3.44. Found: C, 38.67; H, 4.33; N, 3.60.

4u: It was obtained using the same procedure as that described above from 100 mg (0.48 mmol) of amine **3u** as brownish solid. Yield: 305 mg (87%). Mp 122-124 °C; ¹H NMR (300 MHz; CDCl₃): δ 7.38 (1H, d, *J* = 7.8 Hz), 6.98 (2H, m), 6.88 (1H, m), 5.10 (1H, br d, *J* = 10.3 Hz, NHH), 3.83 (3H, s, OCH₃), 3.63 (1H, br d, *J* = 10.3 Hz, NHH), 2.18 (2H, m), 1.45 (1H, m, CHH), 1.34 (1H, m, CHH), 0.99 (3H, t, *J* = 7.2 Hz, CH₃); ¹³C NMR (CDCl₃, 101 MHz): δ 172.4 (COO), 150.6, 144.9, 135.6, 127.1, 124.8, 123.9, 74.7 (q), 53.5 (OCH₃), 42.7 (CH₂), 17.3 (CH₃), 14.0; IR (KBr): ν_{max} 3296, 3252, 1729, 1571, 1433, 1210; HRMS (MALDI-TOF) calcd for C₂₄H₃₂BrN₂O₄Pd₂ (M-Br)⁺ 702.9615, found 702.9609; Anal. calcd for C₂₄H₃₂Br₂N₂O₄Pd₂: C, 36.71; H, 4.11; N, 3.57. Found: C, 36.65; H, 4.05; N, 3.75.

Compounds **5**, Typical Procedure:

5a: A suspension of **3a** (318 mg, 1.64 mmol) and palladium acetate (358 mg, 1.59 mmol) in toluene (25 mL) was stirred at 80 °C for 22 h. The reaction mixture was cooled and volatiles were removed under vacuum to obtain a solid. This solid is dissolved in acetone, LiCl (180 mg, 4.24 mmol) and PPh₃ (442 mg, 1.69 mmol) were added to the solution, and the resulting mixture was stirred at room temperature for 1 h. The solution was filtered and concentrated to

afford, after addition of ethyl ether, a solid, which was purified by chromatography, using $\text{CHCl}_3/\text{MeOH}$ (98/2) as eluent to afford **5a** (532 mg, 50%). ^1H NMR (250 MHz, CDCl_3): δ 7.59-7.50 (6H, m, PPh_3), 7.41-7.29 (9H, m, PPh_3), 6.70 (2H, d, $J = 4.2$ Hz, H^3 , H^4), 6.46 (1H, dd, $J = 7.5$, 4.9 Hz, H^1), 6.34 (1H, m, H^2), 4.21 (2H, m, NH_2), 3.67 (3H, s, OCH_3), 3.59 (1H, d, $J = 13.3$ Hz, CHH), 3.30 (1H, d, $J = 13.2$ Hz, CHH), 1.76 (3H, s, CH_3); $^{31}\text{P}\{^1\text{H}\}$ NMR (250 MHz, CDCl_3): δ 34.16; $^{13}\text{C}\{^1\text{H}\}$ NMR (101 MHz, CDCl_3): δ 136.93, 134.78 (d, $J = 11.7$ Hz, PPh_3), 130.34 (s, PPh_3), 128.07 (d, $J = 10.8$ Hz, PPh_3), 126.53, 125.63, 123.58, 54.15 (CH_2), 53.07 (CH_3O), 25.68 (CH_3); ESI-MS (+) $\{\text{H}_2\text{O}:\text{CH}_3\text{CN}\}$, m/z : $[\text{M}-\text{Cl}]^+ = 560.10$, $[2\text{M}-\text{Cl}]^+ = 1157.17$; Anal. calcd for $\text{C}_{29}\text{H}_{29}\text{ClNO}_2\text{PPd}$: C, 58.40 %; H, 4.90 %; N, 2.35 %. Found: C, 58.6 %; H, 5.1 %; N, 2.5 %.

5b(5): It was obtained using the same procedure as that described above from 311 mg (1.22 mmol) of amino ester **3b**. Yield: 833 mg (75%). ^1H NMR (400 MHz, CDCl_3): δ 7.75-7.70 (6H, m, PPh_3), 7.50-7.27 (17H, m), 6.89 (1H, m), 6.45 (2H, m), 4.76 (1H, dd, $J = 3.9$, 10.5 Hz, HNH), 4.16 (1H, dd, $J = 3.1$, 3.8 Hz, HNH), 3.82 (3H, s, CH_3O), 3.62 (1H, d, $J = 13.9$ Hz, HCH), 3.79 (1H, d, $J = 14.3$ Hz, HCH); $^{31}\text{P}\{^1\text{H}\}$ NMR (250 MHz, CDCl_3): δ 41.24; $^{13}\text{C}\{^1\text{H}\}$ NMR (101 MHz, CDCl_3): δ 139.00, 135.27 (d, $J = 12.0$ Hz, PPh_3), 130.68 (PPh_3), 128.13 (d, $J = 10.8$ Hz, PPh_3), 126.38, 124.20, 123.00, 53.06 (CH_3O), 48.03 (CH_2); ESI-MS (+) $\{\text{H}_2\text{O}:\text{CH}_3\text{CN}\}$, m/z : $[\text{M}-\text{Cl}]^+ = 622.12$; Anal. calcd for $\text{C}_{34}\text{H}_{31}\text{ClNO}_2\text{PPd}$: C, 62.02 %; H, 4.75 %; N, 2.13%. Found: C, 62.3 %; H, 5.0 %; N, 1.9 %.

5c: It was obtained using the same procedure as that described above from 255 mg (1.15 mmol) of amino ester **3c**. Yield: 344 mg (45%). ^1H NMR (400 MHz, CDCl_3): δ 7.56-7.51 (6H, m, PPh_3), 7.40-7.34 (3H, m, PPh_3), 7.31-7.27 (6H, m, PPh_3), 6.70-6.65 (2H, m, H^3 , H^4), 6.46 (1H, dd, $J = 7.6$, 5.0 Hz, H^1), 6.35-6.25 (1H, m, H^2), 3.75 (1H, d, $J = 12.7$ Hz, HCH), 3.60 (3H, s, CH_3O), 3.13 (1H, d, $J = 12.8$ Hz, HCH), 2.44 (1H, m, CHCH_3), 1.34 (3H, d, $J = 6.9$ Hz, CH_3CH), 1.03 (3H, d, $J = 6.9$ Hz, CH_3CH); $^{31}\text{P}\{^1\text{H}\}$ NMR (162 MHz, CDCl_3): δ 33.85; $^{13}\text{C}\{^1\text{H}\}$ NMR (101 MHz, CDCl_3): δ 137.03, 134.75 (d, $J = 11.6$ Hz, PPh_3), 130.28 (s, PPh_3), 127.96 (d, $J = 10.7$ Hz, PPh_3), 126.31, 125.45, 123.45, 52.79 (CH_3O), 51.36 (CH_2), 34.678 (CHCH_3), 18.26 (CH_3CH), 17.18 (CH_3CH); ESI-MS (+) $\{\text{H}_2\text{O}:\text{CH}_3\text{CN}\}$, m/z : $[\text{M}-\text{Br}]^+ = 588.13$; $[\text{M}-\text{Br}+\text{CH}_3\text{CN}]^+ = 629.15$; Anal. calcd for $\text{C}_{31}\text{H}_{33}\text{BrNO}_2\text{PPd}$: C, 55.66 %; H, 4.97 %; N, 2.09 %. Found: C, 55.7 %; H, 4.8 %; N, 1.9 %.

5d: It was obtained using the same procedure as that described above from 110 mg (0.52 mmol) of amino ester **3d**. Yield: 170 mg (50%). ^1H NMR (400 MHz, CDCl_3): δ 7.58-7.53 (6H, m, PPh_3), 7.43-7.38 (3H, m, PPh_3), 7.34-7.30 (6H, m, PPh_3), 6.64 (1H, dd, $J = 8.1, 5.7$ Hz, H^4), 6.38 (1H, td, $J = 8.3, 2.3$ Hz, H^3), 6.08 (1H, ddd, $J = 2.6, 4.8, 9.2$ Hz, H^1), 3.67 (3H, s, CH_3O), 3.55 (1H, d, $J = 13.3$ Hz, HCH), 3.28 (1H, d, $J = 13.3$ Hz, HCH), 1.74 (3H, s, CH_3C); $^{31}\text{P}\{^1\text{H}\}$ NMR (162 MHz, CDCl_3): δ 33.84 (br). $^{19}\text{F}\{^1\text{H}\}$ NMR (376 MHz, CDCl_3): δ -115.4 (d, $J = 105.9$ Hz); $^{13}\text{C}\{^1\text{H}\}$ NMR (101 MHz, CDCl_3): δ 135.75 (d, $J = 11.5$ Hz, PPh_3), 130.56 (PPh_3), 128.14 (d, $J = 10.9$ Hz, PPh_3), 126.75 (d, $J = 7.33$ Hz, C^4), 123.02, 110.16 (d, $J = 21.9$ Hz, C^3), 53.31 (CH_2), 53.15 (s, CH_3O), 25.51 (s, CH_3C); ESI-MS (+) $\{\text{H}_2\text{O}:\text{CH}_3\text{CN}\}$, m/z : $[\text{M}-\text{Br}]^+ = 578.09$; Anal. calcd for $\text{C}_{29}\text{H}_{28}\text{BrFNO}_2\text{PPd}$: C, 52.87 %; H, 4.28 %; N, 2.13 %. Found: C, 52.9 %; H, 4.2 %; N, 1.9 %.

5e: It was obtained using the same procedure as that described above from 189 mg (0.79 mmol) of amino ester **3e**. Yield: 160 mg (35%). ^1H NMR (400 MHz, CDCl_3): δ 7.62-7.55 (6H, m, PPh_3), 7.41-7.36 (3H, m, PPh_3), 7.34-7.29 (6H, m, PPh_3), 7.53 (1H, dd, $J = 2.3, 8.2$ Hz, H^3), 7.24 (1H, dd, $J = 2.3, 4.2$ Hz, H^1), 6.80 (1H, d, $J = 8.2$ Hz, H^4), 3.66 (3H, s, CH_3O), 3.63 (1H, d, $J = 13.3$ Hz, HCH), 3.37 (1H, d, $J = 13.2$ Hz, HCH), 1.76 (3H, s, CH_3C); $^{31}\text{P}\{^1\text{H}\}$ NMR (250 MHz, CDCl_3): δ 34.37; $^{13}\text{C}\{^1\text{H}\}$ NMR (101 MHz, CDCl_3): δ 155.53, 144.85, 134.80 (d, $J = 11.5$ Hz, PPh_3), 131.75, 130.88 (PPh_3), 130.09 (PPh_3), 128.43 (d, $J = 10.9$ Hz, PPh_3), 126.25, 118.89, 56.72 (CH_3C), 53.87 (CH_2), 53.47 (CH_3O), 25.88 (CH_3C); ESI-MS (+) $\{\text{H}_2\text{O}:\text{CH}_3\text{CN}\}$, m/z : $[\text{M}-\text{Cl}]^+ = 605.07$; Anal. calcd for $\text{C}_{29}\text{H}_{28}\text{ClN}_2\text{O}_4\text{PPd}$: C, 54.31 %; H, 4.40 %; N, 4.37 %. Found: C, 54.4 %; H, 4.5 %; N, 3.9 %.

5f: It was obtained, as a mixture 1/1 of compounds, using the same procedure as that described above from 257 mg (0.86 mmol) of amino ester **3f**. Yield: 477 mg (75%). **5f(MeO)** ^1H NMR (400 MHz, CDCl_3): δ 7.59-7.53 (6H, m, PPh_3), 7.41-7.36 (3H, m, PPh_3), 7.33-7.28 (6H, m, PPh_3), 6.66 (1H, d, $J = 8.1$ Hz, H^4), 6.27 (1H, dd, $J = 2.5, 8.1$ Hz, H^3), 5.98 (1H, dd, $J = 2.5, 5.3$ Hz, H^1), 3.77 (3H, s, CH_3O), 3.65 (3H, s, CH_3OCO); $^{31}\text{P}\{^1\text{H}\}$ NMR (162 MHz, CDCl_3): δ 34.47; $^{13}\text{C}\{^1\text{H}\}$ NMR (101 MHz, CDCl_3): δ 134.90 (d, $J = 11.5$ Hz, PPh_3), 130.35 (PPh_3), 128.06 (d, $J = 10.6$ Hz, PPh_3), 126.60, 120.78, 110.43, 55.21 (CH_3O), 52.81 (CH_3OCO); **5f(H)** ^1H NMR (400 MHz, CDCl_3): δ 7.59-7.53 (6H, m, PPh_3), 7.36-7.41 (3H, m, PPh_3), 7.28-7.33 (6H, m, PPh_3), 6.72-6.33 (4H, m, $\text{H}^1\text{-H}^4$), 3.65 (3H, s, CH_3OCO), 3.18 (s, 3H, CH_3O); $^{31}\text{P}\{^1\text{H}\}$ NMR (162 MHz, CDCl_3): δ 34.13; $^{13}\text{C}\{^1\text{H}\}$ NMR (101 MHz, CDCl_3): δ 139.39, 134.90 (d, $J = 11.5$ Hz, PPh_3), 130.35 (PPh_3), 128.06 (d, $J = 10.6$ Hz, PPh_3), 126.33,

114.39, 54.53 (CH₃O), 52.81 (CH₃OCO); ESI-MS (+) {H₂O:CH₃CN}, m/z: [M-Br]⁺=666.14; [M-Br+CH₃CN]⁺=707.17; Anal. calcd for C₃₆H₃₅BrNO₃PPd: C, 57.89 %; H, 4.72 %; N, 1.88 %. Found: C, 57.8 %; H, 4.9 %; N, 1.7 %.

Synthesis of **6a** by stoichiometric carbonylation of **4a**

A suspension of **4a**(X= Br) (120 mg, 0.16 mmol) in methanol (25 mL) was stirred at room temperature in an atmosphere of nitrogen containing carbon monoxide delivered from a toy balloon (~200 mL) for 24 h. The reaction mixture was filtered, washed with 10% aqueous NaHCO₃ solution and dried over MgSO₄. The solvent was removed in a rotatory evaporator to obtain **6a** in 93% yield (65 mg).

6a: mp 180-182 °C (lit.²⁴ 181-183 °C); *R_f* (CH₂Cl₂/MeOH 98:2): 0.31; ¹H NMR (400 MHz; CDCl₃): δ 8.07 (1H, d, *J* = 7.7 Hz, Ar*H*), 7.46 (1H, m, Ar*H*), 7.36 (1H, m, Ar*H*), 7.22 (1H, d, *J* = 7.5 Hz, Ar*H*), 6.58 (1H, br s, NH), 3.69 (3H, s, OCH₃), 3.40 (1H, d, *J* = 15.7 Hz, CH*H*), 3.11 (1H, d, *J* = 15.7 Hz, CH*H*), 1.56 (3H, s, CCH₃); ¹³C NMR (CDCl₃, 101 MHz): δ 174.6 (COO), 165.5 (CONH), 136.1 (q), 132.8, 128.3, 127.9, 127.8, 127.6, 59.0 (q), 53.1 (OCH₃), 37.9 (CH₂), 25.9 (CH₃); IR (ATR): ν_{max} 3195, 1737, 1665; HRMS (ESI⁺) calcd for C₁₂H₁₄NO₃ (M+H)⁺ 220.0968, found 220.0968.

Stepwise preparation of lactam **6b(5)**:

A stirred solution of palladacycle **4b(5)** (110 mg, 0.125 mmol) in AcOH (25 mL) was gently refluxed in an oil bath at 120 °C in an atmosphere of nitrogen containing carbon monoxide delivered from a toy balloon (~200 mL) for 3 h. The reaction mixture was cooled, a filtered through a thin pad of Celite[®]. The volatiles were removed under vacuum and the solid obtained was purified by flash chromatography to afford **6b(5)** (61 mg, 86%).

Methyl 1-benzyl-3-oxoisindoline-1-carboxylate, **6b(5)**: White solid; mp 140-142 °C; *R_f* (hexane/EtAcO 8:2): 0.32; ¹H NMR (400 MHz; CDCl₃): δ 7.82 (1H, d, *J* = 7.7 Hz, Ar*H*), 7.79 (1H, d, *J* = 7.5 Hz, Ar*H*), 7.64 (1H, m, Ar*H*), 7.52 (1H, m, Ar*H*), 7.27 (3H, m, Ar*H*), 7.12 (2H, m, Ar*H*), 6.53 (1H, br s, NH), 3.77 (1H, d, *J* = 13.4 Hz, CH*H*), 3.71 (3H, s, OCH₃), 2.96 (1H, d, *J* = 13.4 Hz, CH*H*); ¹³C NMR (CDCl₃, 101 MHz): δ 170.6 (CO), 169.5 (CO), 144.9

(q), 134.5, 132.5, 130.8, 129.8, 129.5, 128.7, 127.7, 124.0, 123.3, 68.8 (q), 53.0 (OCH₃), 45.0 (CH₂); IR (ATR): ν_{\max} 3219, 1733, 1695, 1611, 1250; HRMS (ESI⁺) calcd for C₁₇H₁₆NO₃ (M+H)⁺ 282.1130, found 282.1128.

Catalytic synthesis of benzolactames

Typical Procedure: A stirred suspension of methyl 2-amino-2-benzyl-3-phenylpropanoate **3h** (100 mg, 0.38 mmol), benzoquinone (83 mg, 0.76 mmol) and palladium acetate (4.5 mg, 0.02 mmol) in AcOH (25 mL) was gently refluxed in an oil bath at 120 °C in an atmosphere of nitrogen containing carbon monoxide delivered from a toy balloon (~200 mL) for 6 h. The reaction mixture was cooled, a filtered through a thin pad of Celite[®]. The volatiles were removed under vacuum to obtain a solid corresponding to almost pure benzolactam **6h**. The residue was purified by flash chromatography to afford **6h** (105 mg, 93%).

Methyl 1-oxo-3-phenyl-1,2,3,4-tetrahydroisoquinoline-3-carboxylate, **6b(6)**: *R_f* (hexane/EtAcO 7:3): 0.43; ¹H NMR (300 MHz; CDCl₃): δ 8.04 (1H, d, *J* = 7.7 Hz, *ArH*), 7.48-7.25 (7H, m, *ArH*), 7.20 (1H, d, *J* = 7.6 Hz, *ArH*), 6.84 (1H, br s, NH), 3.75 (3H, s, OCH₃), 3.71 (1H, d, *J* = 15.6, *CHH*), 3.63 (1H, d, *J* = 15.6, *CHH*); ¹³C NMR (CDCl₃, 101 MHz): δ 171.5 (COO), 165.5 (CONH), 139.1, 135.6, 132.9, 128.9, 128.5, 128.2, 128.0, 127.8, 127.7, 127.5, 125.3, 64.3 (CNH), 53.3 (OCH₃), 38.0 (CH₂); IR (ATR): ν_{\max} 3174, 1733, 1661, 1602, 1446, 1377; HRMS (ESI⁺) calcd for C₁₇H₁₆NO₃ (M+H)⁺ 282.1130, found 282.1144.

Methyl 7-fluoro-3-methyl-1-oxo-1,2,3,4-tetrahydroisoquinoline-3-carboxylate, **6d**: White solid; mp 123-127 °C; ¹H NMR (400 MHz; CDCl₃): δ 7.75 (1H, dd, *J* = 8.8, 2.4 Hz, *ArH*), 7.22-7.13 (2H, m, *ArH*), 6.26 (1H, br s, NH), 3.71 (3H, s, OCH₃), 3.37 (1H, d, *J* = 14.8 Hz, *CHH*), 3.07 (1H, d, *J* = 15.9 Hz, *CHH*), 1.54 (3H, s, CH₃); ¹³C NMR (CDCl₃, 101 MHz): δ 174.5, 173.9, 162.7 (d, *J*_{CF} = 245 Hz), 131.4 (d, *J*_{CF} = 8 Hz), 120.9 (d, *J*_{CF} = 22 Hz), 115.3 (d, *J*_{CF} = 21 Hz), 114.9 (d, *J*_{CF} = 23 Hz), 59.1, 53.3, 40.2, 37.3, 25.8, 23.5; IR (ATR): ν_{\max} 3201, 1734, 1668, 1443, 1199; HRMS (ESI⁺) calcd for C₁₂H₁₃FNO₃ (M+H)⁺ 238.0874, found 238.0870.

Methyl 3-methyl-7-nitro-1-oxo-1,2,3,4-tetrahydroisoquinoline-3-carboxylate, **6e**: reddish solid; mp 104-108 °C; ¹H NMR (400 MHz; CDCl₃): δ 8.91 (1H, d, *J* = 2.4 Hz, *ArH*), 8.31 (1H, dd, *J* = 8.4, 2.4 Hz, *ArH*), 7.44 (1H, d, *J* = 8.4 Hz, *ArH*), 6.51 (1H, br s, NH), 3.85 (1H, d, *J* = 16.4 Hz, *CHH*), 3.71 (3H, s, OCH₃), 3.11 (1H, d, *J* = 16.4 Hz, *CHH*), 1.59 (3H, s,

CH₃); ¹³C NMR (CDCl₃, 101 MHz): δ 171.9, 164.5, 164.4, 145.4, 144.2, 135.4, 129.1, 127.4, 121.1, 54.7, 52.3, 41.2, 22.8; IR (ATR): ν_{max} 3338, 1737, 1671, 1516, 1343, 1206; HRMS (ESI⁺) calcd for C₁₂H₁₃N₂O₅ (M+H)⁺ 265.0819, found 265.0798.

Mixture of methyl 3-(4-methoxybenzyl)-1-oxo-1,2,3,4-tetrahydroisoquinoline-3-carboxylate **6f(OMe)** and methyl 3-benzyl-6-methoxy-1-oxo-1,2,3,4-tetrahydroisoquinoline-3-carboxylate **6f(H)**: White solid; ¹H NMR (400 MHz, CDCl₃): δ 8.09 (1H, d, *J* = 7.5 Hz, *ArH*), 7.48 (1H, td, *J* = 7.5, 1.5 Hz, *ArH*), 7.37 (1H, t, *J* = 7.5 Hz, *ArH*), 7.24 (1H, d, *J* = 7.5 Hz, *ArH*), 7.33-7.02 (4H, m, *ArH*), 6.25 (1H, br s, *NH*), 3.79 (3H, s, OCH₃), 3.64 (3H, s, OCH₃), 3.43 (1H, d, *J* = 13.6 Hz, *CHH*), 3.23 (1H, d, *J* = 13.6 Hz, *CHH*) 3.17 (1H, d, *J* = 13.6 Hz, *CHH*), 2.96 (1H, d, *J* = 13.6 Hz, *CHH*); ¹³C NMR (101 MHz, CDCl₃): δ 172.7, 165.1, 135.6, 111.3, 62.9, 55.2, 52.7, 43.8, 36.8; IR (ATR): ν_{max} 3189, 1734, 1666, 1610, 1513; HRMS (ESI⁺) calcd for C₁₉H₂₀NO₄ (M+H)⁺ 326.1387, found 326.1412.

Methyl 1-oxo-3-propyl-1,2,3,4-tetrahydroisoquinoline-3-carboxylate, **6g**: White solid; mp 120-121 °C; *R_f*(hexane/EtAcO 1:1): 0.48; ¹H NMR (400 MHz; CDCl₃): δ 8.06 (1H, d, *J* = 7.6 Hz, *ArH*), 7.46 (1H, m, *ArH*), 7.36 (1H, m, *ArH*), 7.22 (1H, d, *J* = 7.5 Hz, *ArH*), 6.43 (1H, br s, *NH*), 3.71 (3H, s, OCH₃), 3.37 (1H, d, *J* = 15.7 Hz, *CHH*), 3.13 (1H, d, *J* = 15.7 Hz, *CHH*), 1.79 (2H, m), 1.40 (1H, m), 1.25 (1H, m), 0.90 (3H, t, *J* = 7.3 Hz); ¹³C NMR (CDCl₃, 101 MHz): δ 173.3 (COO), 165.1 (CONH), 135.9, 132.6, 128.1, 127.9, 127.8, 127.4, 61.9 (CNH), 52.8 (OCH₃), 40.7, 36.6 (CH₂), 17.0 (CH₂), 13.9 (CH₃); IR (ATR): ν_{max} 3209, 1727, 1664, 1606, 1578, 1390; HRMS (ESI⁺) calcd for C₁₄H₁₈NO₃ (M+H)⁺ 248.1287, found 248.1294.

Methyl 3-benzyl-1-oxo-1,2,3,4-tetrahydroisoquinoline-3-carboxylate, **6h**: White solid; mp 128-129 °C; *R_f*(hexane/EtAcO 7:3): 0.30; ¹H NMR (400 MHz; CDCl₃): δ 8.09 (1H, d, *J* = 7.6 Hz, *ArH*), 7.49 (1H, m, *ArH*), 7.38 (1H, m, *ArH*), 7.31-7.23 (4H, m, *ArH*), 7.06 (2H, m, *ArH*), 6.26 (1H, br s, *NH*), 3.63 (3H, s, OCH₃), 3.44 (1H, d, *J* = 15.8 Hz, CH₂), 3.24 (1H, d, *J* = 13.4 Hz, CH₂), 3.23 (1H, d, *J* = 15.8 Hz, CH₂), 3.03 (1H, d, *J* = 13.4 Hz, CH₂); ¹³C NMR (CDCl₃, 101 MHz): δ 172.6 (COO), 165.0 (CONH), 135.5 (q), 134.0 (q), 132.7, 129.7, 128.7, 128.2, 127.8, 127.6, 127.5, 62.8 (CNH), 52.7 (OCH₃), 44.6, 36.8; IR (ATR): ν_{max} 3191, 1731, 1663, 1603, 1384; HRMS (ESI⁺) calcd for C₁₈H₁₈NO₃ (M+H)⁺ 296.1281, found 296.1283.

Methyl 1-oxo-1,2,3,4-tetrahydroisoquinoline-3-carboxylate, **6i**: Oil; lit²⁵ ¹H NMR (300 MHz; CDCl₃): δ 8.04 (1H, d, *J* = 7.5 Hz, *ArH*), 7.47 (1H, m, *ArH*), 7.37 (1H, m, *ArH*), 7.25 (1H, m,

ArH), 6.48 (1H, br s, NH), 4.41 (1H, ddd, $J = 9.9, 5.4, 2.1$ Hz, NHCH), 3.79 (3H, s, OCH₃), 3.33 (1H, dd, $J = 15.7, 5.2$ Hz, CHH), 3.22 (1H, dd, $J = 15.7, 9.9$ Hz, CHH); ¹³C NMR (CDCl₃, 101 MHz): δ 170.8 (COO), 165.1 (CONH), 136.1, 132.5, 129.3, 128.1, 127.5, 127.4, 53.0 (OCH₃), 52.8 (NHCH), 31.1 (CH₂); IR (ATR): ν_{\max} 3271, 1737, 1656, 1546, 1536, 1215, 1176; HRMS (ESI⁺) calcd for C₁₁H₁₂NO₃ (M+H)⁺ 206.0817, found 206.0803.

3,3-Dimethyl-3,4-dihydroisoquinolin-1(2*H*)-one, **6k**: White solid; mp 146-147 °C (lit.²⁶ 146-147); ¹H NMR (300 MHz; CDCl₃): δ 8.06 (1H, d, $J = 7.6$ Hz, *ArH*), 7.45 (1H, m), 7.36-7.13 (2H, m, *ArH*), 6.37 (1H, br s, NH), 2.92 (2H, s, CH₂), 1.32 (6H, s, CH₃); ¹³C NMR (CDCl₃, 101 MHz): δ 165.5 (CONH), 137.5, 132.2, 130.4, 127.8, 126.9, 126.2, 52.0 (CNH), 41.6 (CH₂), 28.8 (CH₃); IR (KBr): ν_{\max} 3395, 1660; HRMS (ESI⁺) calcd for C₁₁H₁₄NO (M+H)⁺ 176.1075, found 176.1069.

3-Benzyl-3-(hydroxymethyl)-3,4-dihydroisoquinolin-1(2*H*)-one, **6l**: Brownish oil; *Rf* (hexane/EtAcO 1:1): 0.20; ¹H NMR (300 MHz; CDCl₃): δ 8.05 (1H, d, $J = 7.6$ Hz, *ArH*), 7.50 (1H, m, *ArH*), 7.36 (1H, m, *ArH*), 7.32-7.15 (6H, m, *ArH*), 6.91 (1H, br s, NH, *ArH*), 3.60 (1H, d, $J = 11.2$ Hz, CHH), 3.52 (1H, d, $J = 11.2$ Hz, CHH), 3.06 (1H, $J = 13.6$ Hz), 2.95-2.80 (4H, m); ¹³C NMR (CDCl₃, 101 MHz): δ 165.8 (CONH), 136.7, 135.9, 132.8, 130.4, 128.5, 128.1, 128.0, 127.1, 126.9, 65.7 (CNH), 58.4, 33.7, 30.9; IR (ATR): ν_{\max} 3383, 2927, 1651, 1387, 1253, 1094; HRMS (ESI⁺) calcd for C₁₇H₁₈NO₂ (M+H)⁺ 268.1338, found 268.1345.

Methyl 7-methoxy-3-methyl-1-oxo-1,2,3,4-tetrahydroisoquinoline-3-carboxylate, **6m**: White solid; mp 171-174 °C; ¹H NMR (300 MHz; CDCl₃): δ 7.58 (1H, d, $J = 2.8$ Hz, *ArH*), 7.11 (1H, d, $J = 8.4$ Hz, *ArH*), 7.01 (1H, dd, $J = 8.4, 2.8$ Hz, *ArH*), 6.27 (1H, br s, NH), 3.84 (3H, s, OCH₃), 3.70 (3H, s, OCH₃), 3.33 (1H, d, $J = 15.6$ Hz, CHH), 3.03 (1H, d, $J = 15.6$ Hz, CHH), 1.53 (3H, s, CCH₃); ¹³C NMR (CDCl₃, 101 MHz): δ 174.0 (COO), 165.4 (CONH), 158.9, 130.7, 128.8, 127.9, 120.1, 111.2, 58.9 (CNH), 55.4 (OCH₃), 52.7 (OCH₃), 37.0 (CH₂), 25.5 (CH₃); IR (KBr): ν_{\max} 3195, 3075, 2951, 1736, 1668, 1493, 1451, 1437, 1382; HRMS (ESI⁺) calcd for C₁₃H₁₆NO₄ (M+H)⁺ 250.1079, found 250.1068.

Methyl 7-cyano-3-isopropyl-1-oxo-1,2,3,4-tetrahydroisoquinoline-3-carboxylate, **6n**: White solid; mp 176-178 °C; *Rf* (hexane/EtAcO 1:1): 0.43; ¹H NMR (400 MHz; CDCl₃): δ 8.44 (1H, d, $J = 1.6$ Hz, *ArH*), 7.72 (1H, dd, $J = 8.0, 1.6$ Hz, *ArH*), 7.36 (1H, d, $J = 8.0$ Hz, *ArH*), 6.29 (1H, br s, NH), 3.68 (3H, s, OCH₃), 3.38 (1H, d, $J = 16.4$ Hz, CHH), 3.26 (1H, d, $J = 16.4$ Hz, CHH), 2.18 (1H, m, CHCH₃), 1.00 (3H, d, $J = 6.8$ Hz, CHCH₃), 0.98 (3H, d, $J = 6.8$ Hz,

CHCH₃); ¹³C NMR (CDCl₃, 101 MHz): δ 172.7 (COO), 163.7 (CONH), 141.5, 135.4, 132.8, 128.9, 128.7, 117.9 (CN), 111.5, 65.3 (CNH), 52.8 (OCH₃), 35.0, 33.5, 17.1 (CH₃), 16.8 (CH₃); IR (ATR): ν_{max} 3205, 2963, 2229, 1670, 1610, 1433, 1330, 1277, 1189; HRMS (ESI⁺) calcd for C₁₅H₁₇N₂O₃ (M+H)⁺ 273.1239, found 273.1230.

Methyl 6,8-difluoro-3-isopropyl-1-oxo-1,2,3,4-tetrahydroisoquinoline-3-carboxylate, **6o**: White solid; mp 138-140 °C; *R_f* (hexane/EtAcO 1:1): 0.45; ¹H NMR (300 MHz; CDCl₃): δ 6.77 (2H, m, ArH), 6.29 (1H, br s, NH), 3.68 (3H, s, OCH₃), 3.27 (1H, d, *J* = 15.8 Hz, CHH), 3.17 (1H, d, *J* = 15.8 Hz, CHH), 2.12 (1H, hept, *J* = 6.9 Hz, CHCH₃), 0.99 (3H, d, *J* = 6.9 Hz, CHCH₃), 0.96 (3H, d, *J* = 6.9 Hz, CHCH₃); ¹³C NMR (CDCl₃, 101 MHz): δ 172.5 (COO), 166.0 (CONH), 164.7, 165.0 (dd, *J_{CF}* = 251.2, 13.3 Hz), 163.3 (dd, *J_{CF}* = 274.2, 8.0 Hz), 141.3 (d, *J_{CF}* = 10.4 Hz), 110.9 (dd, *J_{CF}* = 21.9, 4.0 Hz), 104.4 (t, *J_{CF}* = 25.5 Hz), 64.9 (CNH), 52.7 (OCH₃), 34.9, 29.7, 17.2 (CH₃), 16.9 (CH₃); IR (ATR): ν_{max} 3226, 3090, 2960, 1724, 1667, 1615, 1307, 1228, 1122; HRMS (ESI⁺) calcd for C₁₄H₁₆F₂NO₃ (M+H)⁺ 284.1098, found 284.1085.

Methyl 7-hydroxy-1-oxo-3-propyl-1,2,3,4-tetrahydroisoquinoline-3-carboxylate, **6p**: White solid; mp 169-173 °C, ¹H NMR (400 MHz; CDCl₃): δ 7.73 (1H, d, *J* = 2.8 Hz, ArH), 7.08 (1H, d, *J* = 7.6 Hz, ArH), 7.00 (1H, dd, *J* = 8.4, 2.8 Hz, ArH), 6.32 (1H, br s, NH), 3.71 (3H, s, OCH₃), 3.28 (1H, d, *J* = 15.6 Hz, CHH), 3.04 (1H, d, *J* = 15.9 Hz, CHH), 1.78 (2H, t, *J* = 9.2 Hz, CH₂CH₂CH₃), 1.42-1.16 (2H, m, CH₂CH₂CH₃), 1.02 (3H, t, *J* = 7.2 Hz, CH₂CH₂CH₃); ¹³C NMR (CDCl₃, 101 MHz): δ 173.3, 165.5, 156.0, 129.1, 127.2, 120.5, 114.8, 62.3, 52.8, 40.5, 35.8, 17.1, 13.9; IR (ATR): ν_{max} 3124, 1723, 1658, 1383, 1260; HRMS (ESI⁺) calcd for C₂₈H₃₅N₂O₈ (2M+H)⁺ 527.2388, found 527.2381.

Methyl 1-oxo-7-propoxy-3-propyl-1,2,3,4-tetrahydroisoquinoline-3-carboxylate, **6q**: White solid; mp 119-123 °C; ¹H NMR (400 MHz; CDCl₃): δ 7.56 (1H, d, *J* = 2.8 Hz, ArH), 7.10 (1H, d, *J* = 8.4 Hz, ArH), 7.00 (1H, dd, *J* = 8.4, 2.8 Hz, ArH), 6.32 (1H, br s, NH), 3.95 (2H, t, *J* = 6.8 Hz, OCH₂), 3.71 (3H, s, OCH₃), 3.29 (1H, d, *J* = 15.6 Hz, CHH), 3.04 (1H, d, *J* = 15.9 Hz, CHH), 1.84-1.74 (2H, m, OCH₂CH₂CH₃), 1.42-1.17 (2H, m, CH₂CH₂CH₃), 1.02 (3H, t, *J* = 7.6 Hz, CH₂CH₂CH₃), 0.89 (3H, t, *J* = 7.6 Hz, CH₂CH₂CH₃); ¹³C NMR (CDCl₃, 101 MHz): δ 171.9, 164.7, 154.7, 135.2, 130.3, 128.7, 117.2, 115.4, 71.5, 55.7, 53.1, 39.0, 29.6, 23.1, 14.8, 14.1, 10.4; IR (ATR): ν_{max} 3195, 1738, 1663, 1451, 1378, 1066; HRMS (ESI⁺) calcd for C₁₄H₁₈NO₄ (M+H)⁺ 306.1700, found 306.1695.

3,3-Diphenylisoindolin-1-one, **6r**: White solid; mp 208-210 °C (lit.²⁷ 210-211 °C); ¹H NMR (300 MHz; CDCl₃): δ 7.88 (1H, d, *J* = 7.5 Hz, *ArH*), 7.56 (1H, m, *ArH*), 7.31-7.51 (2H, m, *ArH*), 7.35-7.20 (9H, m, *ArH*), 7.15 (1H, m, *ArH*), 6.65 (1H, br s, *NH*); ¹³C NMR (CDCl₃, 101 MHz): δ 169.7 (CONH), 150.1, 142.7, 138.1, 132.4, 130.5, 128.7, 128.5, 128.0, 127.9, 127.0, 126.3, 124.5, 124.3, 71.1 (CNH); IR (ATR): ν_{max} 291, 1692, 1651, 1446, 1258, HRMS (ESI⁺) calcd for C₂₀H₁₆NO (M+H)⁺ 286.1232, found 286.1256.

Crystallography

A prismatic crystal of **5a**, **5c** or **5b(6)** (Table S1) was selected and mounted on a MAR345 diffractometer an image plate detector. Unit-cell parameters were determined from 332 (for **5a**), 1574 (for **5c**), and 719 (for **5b(6)**) reflections, in the range, 3 < θ < 31°, and refined by least-squares methods. Intensities were collected with graphite monochromatized Mo K_α radiation. The number of reflections collected were 11514 (for **5a**), 15530 (for **5c**), and 13074 (for **5b(6)**), in the ranges 1.65° ≤ θ ≤ 30.65°, 1.65° < θ < 32.32°, and 1.57° ≤ θ ≤ 30.67°, for **5a**, **5c** and **5b(6)**, respectively}, of which 6314 (for **5a**), 8579 (for **5c** and 7050 (for **5b(6)**) were non-equivalent by symmetry. The number of reflections assumed as observed applying the condition *I* > 2σ(*I*) were 5492, 6203 and 6874 (for **5a**, **5c** and **5b(6)**, respectively). Lorentz-polarization corrections were made.

The structures were solved by Direct methods, using SHELXS computer program²⁸ and refined by full-matrix least-squares method with SHELX97 computer program²⁹ using 11514, 15530, and 13074 reflections for **5a**, **5c** and **5b(6)**, respectively, (very negative intensities were not assumed). The function minimized was Σw | |F_o|² - |F_c|² |², where w = [σ²(*I*) + (0.0494P)² + 0.2769P]⁻¹ (for **5a**), w = [σ²(*I*) + (0.1182P)² + 0.5077P]⁻¹ (for **5c**) and w = [σ²(*I*) + (0.0607P)² + 1.4868P]⁻¹ for (**5b(6)**) and P = (|F_o|² + 2|F_c|²)/3; *f*, *f'*, and *f''* were taken from the bibliography.³⁰ The final R(on F) factor was 0,0357, 0.0660 and 0.0313 for **5a**, **5c** and **5b(6)**, respectively and the goodness of fit values equal to 1.127 (for **5a**), 1.050 (for **5c**), and 1.195 (for **5b(6)**). Further details concerning the resolution and refinement of these crystal structures are given in Table S2.

Kinetics

The kinetic profiles for the reactions were followed by UV-Vis spectroscopy in the 700-300 nm range. Atmospheric pressure runs were recorded on HP8452A or Cary50 instruments equipped with thermostated multicell transports. Observed rate constants were derived from absorbance *versus* time traces at the wavelengths where a maximum increase and/or decrease of absorbance was observed. For runs at variable pressure, a previously described pressurizing system and pill-box cell was used,³¹ the system was connected to a J&M TIDAS spectrophotometer which was used for the absorbance measurements. The calculation of the observed rate constants from the absorbance *versus* time monitoring of reactions, studied under second or first order concentration conditions, were carried out using the SPECFIT software.³² The general kinetic technique is that previously described.³² The solutions for the kinetic runs were prepared by mixing the calculated amounts of stock solutions of the palladium compounds and the metallating ligands in the desired solvent. In all cases no dependence on the concentration of palladium was detected and it was kept in the $(2-5)\times 10^{-4}$ M margin. Table S2 collects all the obtained k_{obs} values for all the systems studied as a function of the metallating ligand, solvent, temperature and pressure. All post-run fittings were carried out by the standard available commercial programs.

CCDC n.- 905213-905215 contain the supplementary crystallographic data for this paper. These data can be obtained free of charge from the *Cambridge Crystallographic Data Centre* via: www.ccdc.cam.ac.uk/data_request/cif

Notes and references

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