

## Draft Genome Sequence of JVAP01<sup>⊤</sup>, the Type Strain of the Novel Species Acinetobacter dijkshoorniae

AMERICAN SOCIETY FOR MICROBIOLOGY gen@meAnnouncements™

Dietmar Fernández-Orth,<sup>a</sup> Clara Cosgaya,<sup>a</sup> Murat Telli,<sup>b</sup> Noraida Mosqueda,<sup>a</sup> Marta Marí-Almirall,<sup>a</sup> <sup>(b</sup>Ignasi Roca,<sup>a</sup> <sup>(b</sup>Jordi Vila<sup>a</sup>

Department of Clinical Microbiology and ISGlobal, Barcelona Ctr. Int, Health Res. CRESIB, Hospital Clínic, Universitat de Barcelona, Barcelona, Spaina; Department of Clinical Microbiology, School of Medicine, Adnan Menderes University, Aydin, Turkey<sup>b</sup>

**ABSTRACT** Here, we report the draft genome sequence of the type strain of *Acinetobacter dijkshoorniae*, a novel human pathogen within the *Acinetobacter calcoaceticus–Acinetobacter baumannii* (ACB) complex. Strain JVAP01<sup>T</sup> has an estimated genome size of 3.9 Mb, exhibits a 38.8% G+C content, and carries a plasmid with the  $bla_{NDM-1}$  carbapenemase gene.

The Acinetobacter calcoaceticus–Acinetobacter baumannii (ACB) complex currently comprises six different Acinetobacter species, the environmental A. calcoaceticus and five Acinetobacter species that are potential human pathogens, that is, A. baumannii, A. pittii, A. nosocomialis, A. seifertii, and A. dijkshoorniae, the latter two only recently discovered (1). Members of the ACB complex are virtually undistinguishable from a biochemical standpoint and can only be differentiated by means of molecular methods (2, 3). Hospital outbreaks are mostly attributed to A. baumannii, whose innate ability to accumulate multiple antibiotic resistance mechanisms is greatly feared (4). The advent of more reliable identification methodologies, however, has shown an alarming abundance of all other species in the clinical setting, as well as their potential to bear resistance mechanisms to last resort antibiotics (5, 6). Here we report the draft genome sequence of strain JVAP01<sup>T</sup> (CECT 9134<sup>T</sup>, LMG 29605<sup>T</sup>), the type strain of Acinetobacter dijkshoorniae that was recovered in 2009 from a urine sample in Turkey. JVAP01<sup>T</sup> produces the NDM-1 metallo- $\beta$ -lactamase and is resistant to  $\beta$ -lactam antibiotics and kanamycin (7).

Genomic DNA was extracted from cultured bacteria and an Illumina library was generated following Nextera XT (Illumina, Inc., San Diego, CA, USA) manufacturer's protocol with paired-end libraries ( $2 \times 150$ ). Sequencing was performed in an Illumina MiSeq system. *De novo* assembly was performed using Velvet version 1.2.10 in conjunction with the Velvet optimizer (http://bioinformatics.net.au/software.velvetoptimiser.shtml), ABySS v1.5.2 and Spades v3.5.0 (8–10). Contigs for all assemblers were joined using CISA v1.3 (11). CISA contigs below 200 nucleotides were discarded to yield a total of 92 contigs with a 90-fold coverage. The draft genome comprised a total assembly length of 3,858,459 bp and the G+C content was in accordance with that of *Acinetobacter* spp., at 38.8%.

The sequence of the 47 kilobase plasmid (pNDM-JVAP01) containing the  $bla_{NDM-1}$  gene and a type VI secretion system was previously published (7).

All 92 contigs and the plasmid were further annotated using the RAST server (12), which predicted 3,599 coding sequences (CDS), 26 rRNAs, and 134 tRNAs in the genome. In order to classify the antibiotic resistance gene pools, Resfinder v2.1 with a threshold of 85% identity and a minimum length of 40% was used (13). Results showed the presence of the *bla*<sub>NDM-1</sub> and *aphA6* genes, described previously and conferring

Received 7 November 2016 Accepted 8 November 2016 Published 12 January 2017

**Citation** Fernández-Orth D, Cosgaya C, Telli M, Mosqueda N, Marí-Almirall M, Roca I, Vila J. 2017. Draft genome sequence of JVAP01<sup>T</sup>, the type strain of the novel species *Acinetobacter dijkshoorniae*. Genome Announc 5:e01480-16. 10.1128/genomeA.01480-16.

**Copyright** © 2017 Fernández-Orth et al. This is an open-access article distributed under the terms of the Creative Commons Attribution 4.0 International license.

Address correspondence to Ignasi Roca, Ignasi.Roca@isglobal.org.

D.F.-O. and C.C. contributed equally to this article.

resistance to most  $\beta$ -lactams and aminoglycosides, respectively (7), but also of the  $bla_{ADC}$ - and  $bla_{OXA-213}$ -family genes, respectively encoding an *Acinetobacter*-derived cephalosporinase and a class D oxacillinase, both of chromosomal location and conferring resistance to  $\beta$ -lactams. PathogenFinder v1.1 was used for the prediction of bacterial pathogenicity (14). Results revealed this species as human pathogen (83.8% probability) matching common sequences with 24 pathogenic families. Those, among others, were from *A. baumannii* ATCC 17978, AB0057, ACICU, and AYE strains.

**Accession number(s).** This whole-genome shotgun project has been deposited in DDBJ/ENA/GenBank under the accession no. LJPG00000000 and KM923969 for its associated plasmid. The version described in this paper is LJPG00000000.1.

## ACKNOWLEDGMENTS

This study was supported by the Ministerio de Economía y Competitividad, Instituto de Salud Carlos III, cofinanced by European Regional Development Fund (ERDF) "A Way to Achieve Europe," by the Government of Catalonia, the Adnan Menderes University and the European commission under the MagicBullet project. The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

## REFERENCES

- Cosgaya C, Marí-Almirall M, Van Assche A, Fernández-Orth D, Mosqueda N, Telli M, Huys G, Higgins PG, Seifert H, Lievens B, Roca I, Vila J. 2016. *Acinetobacter dijkshoorniae* sp. nov., a new member of the *Acinetobacter calcoaceticus–Acinetobacter baumannii* complex mainly recovered from clinical samples in different countries. Int J Syst Evol Microbiol. https:// doi.org/10.1099/ijsem.0.001318.
- Higgins PG, Lehmann M, Wisplinghoff H, Seifert H. 2010. gyrB multiplex PCR to differentiate between Acinetobacter calcoaceticus and Acinetobacter genomic species 3. J Clin Microbiol 48:4592–4594. https://doi.org/ 10.1128/JCM.01765-10.
- La Scola B, Gundi VA, Khamis A, Raoult D. 2006. Sequencing of the *rpoB* gene and flanking spacers for molecular identification of *Acinetobacter* species. J Clin Microbiol 44:827–832. https://doi.org/10.1128/ JCM.44.3.827-832.2006.
- Roca I, Espinal P, Vila-Farrés X, Vila J. 2012. The Acinetobacter baumannii oxymoron: commensal hospital dweller turned pan-drug-resistant menace. Front Microbiol 3:148. https://doi.org/10.3389/fmicb.2012.00148.
- Wisplinghoff H, Paulus T, Lugenheim M, Stefanik D, Higgins PG, Edmond MB, Wenzel RP, Seifert H. 2012. Nosocomial bloodstream infections due to Acinetobacter baumannii, Acinetobacter pittii, and Acinetobacter nosocomialis in the United States. J Infect 64:282–290. https://doi.org/ 10.1016/j.jinf.2011.12.008.
- Zander E, Fernández-González A, Schleicher X, Dammhayn C, Kamolvit W, Seifert H, Higgins PG. 2014. Worldwide dissemination of acquired carbapenem-hydrolysing class D beta-lactamases in *Acinetobacter* spp. other than *Acinetobacter baumannii*. Int J Antimicrob Agents 43: 375–377. https://doi.org/10.1016/j.ijantimicag.2014.01.012.
- Espinal P, Mosqueda N, Telli M, van der Reijden T, Rolo D, Fernández-Orth D, Dijkshoorn L, Roca I, Vila J. 2015. Identification of NDM-1 in a putatively novel Acinetobacter species ("NB14") closely related to Acin-

etobacter pittii. Antimicrob Agents Chemother 59:6657-6660. https://doi.org/10.1128/AAC.01455-15.

- Zerbino DR, Birney E. 2008. Velvet: algorithms for *de novo* short read assembly using de Bruijn graphs. Genome Res 18:821–829. https:// doi.org/10.1101/gr.074492.107.
- Bankevich A, Nurk S, Antipov D, Gurevich AA, Dvorkin M, Kulikov AS, Lesin VM, Nikolenko SI, Pham S, Prjibelski AD, Pyshkin AV, Sirotkin AV, Vyahhi N, Tesler G, Alekseyev MA, Pevzner PA. 2012. SPAdes: a new genome assembly algorithm and its applications to single-cell sequencing. J Comput Biol 19:455–477. https://doi.org/10.1089/cmb.2012.0021.
- Simpson JT, Wong K, Jackman SD, Schein JE, Jones SJ, Birol I. 2009. ABySS: a parallel assembler for short read sequence data. Genome Res 19:1117–1123. https://doi.org/10.1101/gr.089532.108.
- Lin SH, Liao YC. 2013. CISA: contig integrator for sequence assembly of bacterial genomes. PLoS One 8:e60843. https://doi.org/10.1371/ journal.pone.0060843.
- Aziz RK, Bartels D, Best AA, DeJongh M, Disz T, Edwards RA, Formsma K, Gerdes S, Glass EM, Kubal M, Meyer F, Olsen GJ, Olson R, Osterman AL, Overbeek RA, McNeil LK, Paarmann D, Paczian T, Parrello B, Pusch GD, Reich C, Stevens R, Vassieva O, Vonstein V, Wilke A, Zagnitko O. 2008. The RAST Server: Rapid Annotations using Subsystems Technology. BMC Genomics 9:75. https://doi.org/10.1186/1471-2164-9-75.
- Zankari E, Hasman H, Cosentino S, Vestergaard M, Rasmussen S, Lund O, Aarestrup FM, Larsen MV. 2012. Identification of acquired antimicrobial resistance genes. J Antimicrob Chemother 67:2640–2644. https:// doi.org/10.1093/jac/dks261.
- Cosentino S, Voldby Larsen M, Møller Aarestrup F, Lund O. 2013. PathogenFinder—distinguishing friend from foe using bacterial whole genome sequence data. PLoS One 8:e77302. https://doi.org/10.1371/ journal.pone.0077302.