

PDR-1/hParkin negatively regulates the phagocytosis of apoptotic cell corpses in *Caenorhabditis elegans*

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Apoptotic cell death is an integral part of cell turnover in many tissues, and proper corpse clearance is vital to maintaining tissue homeostasis in all multicellular organisms. Even in tissues with high cellular turnover, apoptotic cells are rarely seen because of efficient clearance mechanisms in healthy individuals. In *Caenorhabditis elegans*, two parallel and partly redundant conserved pathways act in cell corpse engulfment. The pathway for cytoskeletal rearrangement requires the small GTPase CED-10 Rac1 acting for an efficient surround of the dead cell. The CED-10 Rac pathway is also required for the proper migration of the distal tip cells (DTCs) during the development of the *C. elegans* gonad. Parkin, the mammalian homolog of the *C. elegans* PDR-1, interacts with Rac1 in aged human brain and it is also implicated with actin dynamics and cytoskeletal rearrangements in Parkinson's disease, suggesting that it might act on engulfment. Our genetic and biochemical studies indicate that PDR-1 inhibits apoptotic cell engulfment and DTC migration by ubiquitylating CED-10 for degradation.

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Multicellular organisms execute the majority of unwanted cell populations in a regulated manner via the process of apoptosis.^{1,2} Remarkably, even in tissues with high cellular turnover, apoptotic cells are rarely seen *in situ*, which is thought to be owing to efficient clearance mechanisms.

Early studies in the nematode *Caenorhabditis elegans* (*C. elegans*) suggested that apoptotic cells are recognized and cleared before they are 'fully dead'.^{3,4}

Two evolutionary conserved and partially redundant signaling pathways act together in the clearance of cell corpses during development and in the germline in *C. elegans*: (a) CED-2, CED-5 and CED-12 and (b) CED-1, CED-6 and CED-7, both converging in the activation of the small GTPase CED-10 (Rac1 in mammals), which, eventually, promote cytoskeletal reorganization and cell corpse engulfment.^{5,6} In addition, two independent signaling pathways have been described very recently: the one acting through the ABI-1 tyrosine kinase and the other being regulated by the SLI-1 ligase.^{7,8} However, the redundancy of these new pathways to the canonical engulfment branches still cannot be discarded.

CED-10/Rac1 is a member of the Rho family GTPases. GTP-bound Rac has an evolutionary conserved positive effect on engulfment, and Rac activation at sites of apoptotic cell recognition subsequently leads to cytoskeletal rearrangement.^{9,10} Once Rac1 is activated, it is ubiquitylated for targeting to proteasomal degradation.^{11,12} The specific E3 ubiquitin ligase regulating Rac1 degradation in the scenario of the engulfment remains elusive.

E3 ubiquitin ligases constitute the third step of the ubiquitination process and have an obligatory role in catalyzing the conjugation of ubiquitin to a lysine residue in the target proteins. Rac1 activity is regulated by the proteasomal machinery in other scenarios, and the E3 ligases HACE1, c-IAP and XIAP are some examples.^{13–15} HACE-1 has been involved in the mechanism of pathogen infection and c-IAP and XIAP regulate carcinogenesis.^{16,17} In the IAPs context, the downregulation of the E3 ligases leads to an increase of Rac1 levels and activity in cells, promoting cell elongation and migration.

Parkin encodes a protein with E3 ligase activity whose loss of function is considered causative of autosomal

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Abbreviations: ced, cell-death abnormality; DTC, distal tip cell; PD, Parkinson's disease; UBL, ubiquitin-like domain; DIC, differential interference contrast

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recessive-juvenile parkinsonism. Human Parkin (hParkin) consists of four domains: an N-terminal ubiquitin-like domain (UBL), and two RING-finger domains flanking a cysteine-rich in-between domain. The UBL domain has been implicated in proteasome binding,¹⁸ substrate recognition¹⁹ and regulation of Parkin stability.²⁰ A number of substrates of Parkin have been identified in various experimental systems.^{21–24} Some of the identified mutations in the gene encoding for Parkin have been shown to impair its E3 ubiquitin ligase activity for several substrates.^{25,26}

Parkin function has been associated with multiple cellular processes, among them is the stability of cytoskeletal components. Thus, Parkin is associated with actin filaments in neuronal and non-neuronal cells.²⁷

Protein degradation by ubiquitin-mediated targeting to the proteasomal machinery has a crucial role in cell signaling, notably in pathways that control actin cytoskeleton dynamics.^{11,28} We hypothesized that PDR-1/hParkin might act in engulfment pathways. In addition, we asked whether it did so by interacting with the *C. elegans* CED-10.

We now present evidence that PDR-1 inhibits apoptotic cell engulfment. Interestingly, we find that PDR-1 regulates the turnover of CED-10 by polyubiquitylation. This work provides the first link between cellular processes of ubiquitylation/proteasomal degradation and the ability to clear apoptotic cells efficiently in the nematode. Further studies are needed to elucidate the exact mechanism working in humans.

Results

Pdr-1 mutations decreased the number of cell corpses of engulfment mutants. Searching for Parkin interactors involved in cytoskeletal rearrangements, we detected a positive interaction between Parkin and Rac1 in aged human brains (Supplementary Figure S1). The nematode *C. elegans* was then used as the model to study the regulation of *ced-10* (human Rac1, hRac1) by *pdr-1* (hParkin) because of the simplicity of *C. elegans* as an animal model to study genetic interactions, and because mechanisms controlling the engulfment in *C. elegans* are conserved in metazoa.⁴ To test first whether *pdr-1* had a role in engulfment, the number of unengulfed cell corpses was counted in the heads of first larval stage (L1) animals, harboring mutations in *pdr-1* and *ced-10*. The number of unengulfed corpses varies with the strength of the engulfment defect and defines a quantitative assay of engulfment abnormalities.²⁹ In the *ced-10* gene, the presence of the *n1993* lesion is a G-to-T transition that results in a change of valine 190 to glycine.³⁰ The mutation *ced-10(n3246)* is a G-to-A transition resulting in a change of glycine 60 of CED-10 to arginine (G60R).³¹ This mutation results in altered function (gain-of-function or dominant-negative phenotypes described in Reddien and Horvitz³¹ and Shakir *et al.*³⁰). Because of the increased defects in the engulfment, this allele is considered to be strong, in comparison with *ced-10(n1993)* allele that is considered to be weak. Both these alleles are not null and they still maintain some residual activity.^{30,31}

Two presumptive null alleles of *pdr-1* were used in this study, *lg103* and *tm395*. The allele *lg103* carries in-frame deletion encoding the internally truncated PDR-1 protein

Δ aa24–247, resulting in the loss of the UBL and the complete unique Parkin domain and the following RING domain.²² PDR-1 activity is being abolished as a consequence of this loss-of-function mutation.^{22,32} The allele *tm395* is an out-of-frame deletion that results in an earlier stop codon so that its capacity of association with the proteasome is not modified.²² Mutant alleles *lg103* and *tm395* were backcrossed with N2 wild-type 7 at least twice respectively.²²

Pdr-1 effect on engulfment was first tested in a *ced-10*/hRac1 background. Neither wild-type nor *pdr-1* mutation alone had any obvious effect on engulfment. However, mutations of *pdr-1* suppressed both *ced-10* alleles, as the number of unengulfed corpses were decreased in the heads of the two *ced-10*;*pdr-1* double mutants (Table 1 and Figures 1a–c).

The suppression of the *ced-10* stronger allele (such as that generated by the (*n3246*) mutation) by *pdr-1* mutations is consistent with a general inhibition of the engulfment by *pdr-1*. We could not use the *ced-10(t1875)* null allele in these studies owing to its embryonic lethality phenotype, which prevented scoring larval corpses.³³

In addition to *ced-10*, the loss-of-function *ced-5(n1812)* and *ced-1(e1735)* alleles were also partially suppressed by *pdr-1(lg103)* (Table 1). *Dyn-1* mutants died at the embryonic stage and were not analyzed. As the main purpose was to investigate the interaction between Parkin and Rac1, the other engulfment mutant alleles were not tested.

The fact that loss of *pdr-1* function suppresses the engulfment defects caused by *ced-1*, *ced-5* and *ced-10* mutations suggested that PDR-1 may act in parallel or downstream to both engulfment pathways. Because of our previous results obtained in human brains (Supplementary Figure S1), the regulation of *ced-10* by *pdr-1* cannot be excluded.

Like CED-10,³⁴ PDR-1 protein is ubiquitously expressed. Expression starts in embryogenesis and is maintained throughout development until adulthood.²² Both CED-10 and PDR-1 act in the same subcellular compartments.^{22,35} To determine whether *pdr-1* was responsible for the engulfment increase detected in *ced-10*;*pdr-1* worms, we used the rescue strain, *pdr-1(lg103);byEx429[K08E3];sel-12::gfp*,²² as described in the Methodology section. The cosmid K08E3 is a genomic subclone containing the complete *pdr-1* operon.²² PDR-1 wild type in double mutants *ced-10*;*pdr-1* rescued the suppressor activity of *pdr-1* (Table 1 and Figures 1d and e). This indicates a solid rescue of mutant *pdr-1* suppressor activity and identifies PDR-1 as a new inhibitor in the engulfment of apoptotic cells.

Pdr-1 is not involved in the execution of apoptosis. A reduction in persistent cell corpse numbers can be achieved not only through a restoration of engulfment activity but also through reduced apoptosis or delayed initiation of cell death.³⁶ To discard any role of *pdr-1* in the apoptosis execution, *pdr-1* influence in the generation of apoptotic cells was subsequently tested. Whereas 11–13 cells remained alive in the anterior pharynx of *ced-3* loss-of-function (*lf*) mutants (allele *ced-3(n717)*),^{36–38} no surviving extra cells were observed in *pdr-1* mutants alone (Table 2). In each *ced-3(lf)* and *ced-3(lf)*;*pdr-1* mutant animals from our experiment, some undead cells were not found as referenced also by the

Table 1 *pdr-1* mutations suppress the engulfment defects of engulfment *ced-10* gene mutations

Genotype	Number of cell corpses \pm s.d.	n	P-value
N2	0 \pm 1	30	—
<i>pdr-1(tm395)</i>	0	30	—
<i>pdr-1(lg103)</i>	0 \pm 1	30	—
<i>ced-10(n3246)</i>	21.4 \pm 0.6	67	—
<i>ced-10(n1993)</i>	15.41 \pm 1.054	30	—
<i>ced-10(n1993);pdr-1(tm395)</i>	6.1 \pm 0.3	48	<0.0001 to <i>ced-10(n1993)</i>
<i>ced-10(n1993);pdr-1(lg103)</i>	7.4 \pm 0.4	30	<0.0001 to <i>ced-10(n1993)</i>
<i>ced-10(n3246);pdr-1(lg103)</i>	8.9 \pm 0.3	121	<0.0001 to <i>ced-10(n3246)</i>
<i>ced-10(n3246);pdr-1(tm395)</i>	10.5 \pm 0.7	35	<0.0001 to <i>ced-10(n3246)</i>
<i>pdr-1(lg103);byEx[PDR-1 wild type;sel-12::gfp]</i>	0 \pm 1	30	—
<i>ced-10(n3246);pdr-1(lg103);byEx[PDR-1 wild type;sel-12::gfp]</i>	19.48 \pm 0.5	63	0.0117 to <i>ced-10(n3246)</i>
<i>ced-5(n1812)^a</i>	25.07 \pm 0.8	30	—
<i>ced-5(n1812);pdr-1(lg103)</i>	19.05 \pm 0.7	31	<0.0001 to <i>ced-10(n3246)</i>
<i>ced-1(e1375)^b</i>	16.7 \pm 0.5	31	—
<i>ced-1(e1375);pdr-1(lg103)</i>	9.9 \pm 0.6	51	<0.0001 to <i>ced-10(n1993)</i>

pdr-1 accelerates the engulfment process. First larval stage (L1) animals were anesthetized and viewed using DIC microscopy. Cell corpses were counted in the head of L1s. The P-value refers to comparisons between the strain with the engulfment mutation alone, *ced-10(n3246)*, *ced-10(n1993)*, *ced-5(n1812)*, *ced-1(n1375)*, and the strains with the engulfment mutation with either the *pdr-1(tm395)* or *pdr-1(lg103)* alleles or the rescue strain *ced-10(n3246);pdr-1(lg103);byEx[PDR-1 wt::sel-12-gfp]*. All data are means \pm s.d. (statistics: paired t-test; two-tailed, unequal variances)

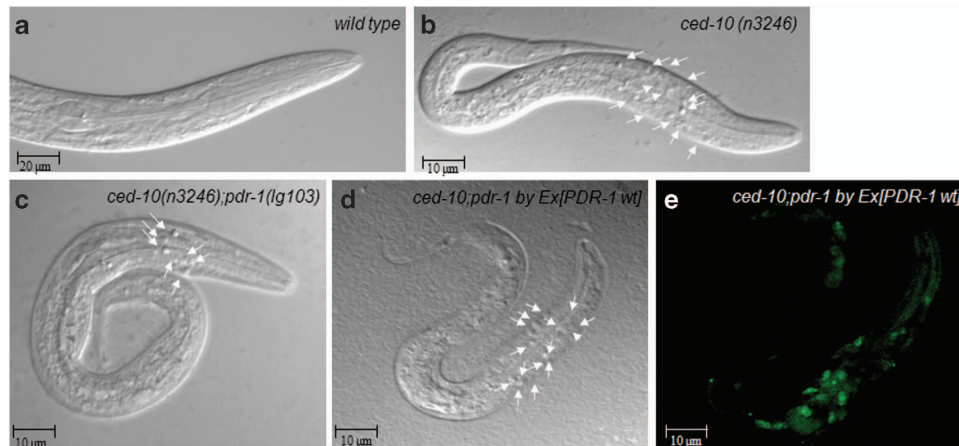


Figure 1 Loss of *pdr-1* accelerates the engulfment machinery in *ced-10* mutants. (a–e) DIC microscopy images of freshly hatched L1 larvae of the indicated genotypes. Arrowheads indicate persistent cell corpses. Scale bar, 20 and 10 μ m as indicated. (d and e) Expression of PDR-1 wild type rescues the *ced-10(n3246);pdr-1(lg103)* phenotype. Alleles used were as follows: *ced-10(n3246)*, *pdr-1(lg103)*, *ced-10(n3246);pdr-1(lg103)* and the rescue strain *ced-10(n3246);pdr-1(lg103); byEx[PDR-1 wt::sel-12-gfp]*

detailed protocol described in Schwartz.³⁸ Even so the number of extra cells was not altered when *ced-3(lf)* worms were crossed with *pdr-1* mutants (Table 3). The same effect was obtained under conditions limiting caspase activity through *ced-3(n2438)*.^{36,37} Therefore, taking into account the genetic interaction of *pdr-1* with the engulfment machinery described above, *pdr-1* selectively affects the engulfment of apoptotic cells.

Loss of *pdr-1* accelerates the engulfment of dying cells. Apoptotic cell corpses are visible from the time they are generated until they are engulfed and degraded by neighboring cells. The kinetics of the engulfment was investigated by four-dimensional (4D) Nomarski time-lapse video microscopy (Leica DM 6000, Madrid, Spain) to follow the first 13 programmed cell deaths in the AB cell lineage³ (Figure 2 and Supplementary Table S2 and Figure S2).

Surprisingly, alterations were found in both the kinetics of the engulfment time and the onset of apoptosis in *pdr-1* allele alone (Figure 2 and Supplementary Figure S2A, respectively). As demonstrated by Supplementary Figure S2B, the first mutant embryo development occurred mostly synchronized with the wild type, but the cell cycle was progressively delayed (Supplementary Figure 2B). Both cell lineages, respectively, are evolving unsynchronized with the increasing embryonic developmental time. Therefore, the delay in the onset of apoptosis cannot be ascribed to a function of *pdr-1* in the initiation of apoptosis *per se*, but may correspond to a general defect in cell cycle progression affecting the developing embryo. Strikingly, the mean engulfing time was shorter in *pdr-1* compared with wild-type worms (Figure 2 and Supplementary Table S2). Given the general delay in the developing embryos, a delay in the engulfment resulting in accumulated cell corpses at L1 stage could then

also be expected. Moreover, it is worth noting the lack of differences in cell corpse persistence between *ced-10* mutants alone in comparison with *ced-10;pdr-1* double mutant animals (Figure 2b and Supplementary Table S2), albeit a clear reduction in cell corpse number (Figure 1 and Table 1). While in the 4D recordings only the first 13 cell deaths are being analyzed (from this developing time the embryo is moving faster and the engulfing time is impossible to follow appropriately³⁹), the counting of cell corpses at L1 stage represents the final process and reflects the global efficiency of the engulfment, including all the unphagocytosed cell corpses.

Table 2 *pdr-1* is not involved in the execution of apoptosis

Genotype	Extra cells ± s.d.	n	P-value
N2	0.1 ± 0.2	15	—
<i>pdr-1(lg103)</i>	0.1 ± 0.2	15	—
<i>ced-3(n717)</i>	8 ± 0.4	15	—
<i>ced-3(n717);pdr-1(lg103)</i>	8 ± 0.6	35	0.711 to <i>ced-3(n717)</i>
<i>ced-3(n2438)</i>	2.3 ± 0.1	13	—
<i>ced-3(n2438);pdr-1(lg103)</i>	1.9 ± 0.2	35	0.463 to <i>ced-3(n2438)</i>

pdr-1 does not affect the execution of apoptosis. Third larval stage (L3) animals were anesthetized and viewed using DIC microscopy. The number of extra cell nuclei in the anterior pharynx (pro- and metacarpus) was counted. Alleles used were as follows: *ced-3(n717)*, *ced-3(n2438)* and *pdr-1(lg103)*. All data are means ± s.d. (statistics: paired t-test; two-tailed, unequal variances)

Table 3 *ced-10* activity in the engulfment of cell corpses depends on the proteasomal activity

Genotype	Treatment	Number of cell corpses ± s.d.	n	P-value
wild type	MG-132	0	15	—
<i>ced-10(n3246)</i>	DMSO	16.2 ± 3.9	20	—
<i>ced-10(n3246)</i>	MG-132	5.7 ± 2.3	25	<i>P</i> < 0.001

Therefore, the increase in the rate of engulfment observed in the 4D recordings reinforces and lends further support to the initial enhancement in the engulfment observed in *pdr-1* animals.

***Pdr-1* has cell-autonomic functions in the engulfing cells.**

To corroborate the hypothesis of *pdr-1* acting in the engulfing cell, PDR-1::GFP was expressed in a ‘tissue-specific’ manner by two different cell-type-specific promoters.^{40,41} Overexpression of PDR-1 controlled by the CED-1 promoter (*Pced-1::pdr-1::gfp*), which drives gene expression specifically in engulfing cells,⁶ fully rescued the reduced cell corpses observed in *ced-10(n3246);pdr-1(lg103)* mutants. This rescuing activity was not observed when PDR-1 expression was controlled by the *egl-1* promoter (*Pegl-1::pdr-1::gfp*), which drives gene expression specifically in dying cells⁴² (Figure 3a and Supplementary Table S1). However, the rescue data on the *ced-10* weak allele (*n1993*) were not fully accomplished when *pdr-1(lg103)* was the allele improving the engulfment (Figure 2b and Supplementary Table S1). As already reported, the use of hypomorphic alleles in the study of epistasis and double mutant analysis is problematic in the nematode.⁴³ Still, a partial rescue was obtained when this weak allele of *ced-10* was in combination with *pdr-1(tm395)*. These results demonstrate that PDR-1 functions in engulfing rather than in dying cells, but a minor function of *pdr-1* in the apoptotic cell cannot be excluded completely.

PDR-1 suppresses the distal tip cell migration defect associated with CED-10.

The two distal tip cells (DTCs) migrate during development from the center of the animal outward and then back again, meeting approximately in the center of the animal. As they move, the gonads form behind them, resulting in two U-shaped gonads.⁴⁴ In *ced-10* pathway mutants, the gonads often have extra turns or arms caused by abnormal DTC migration

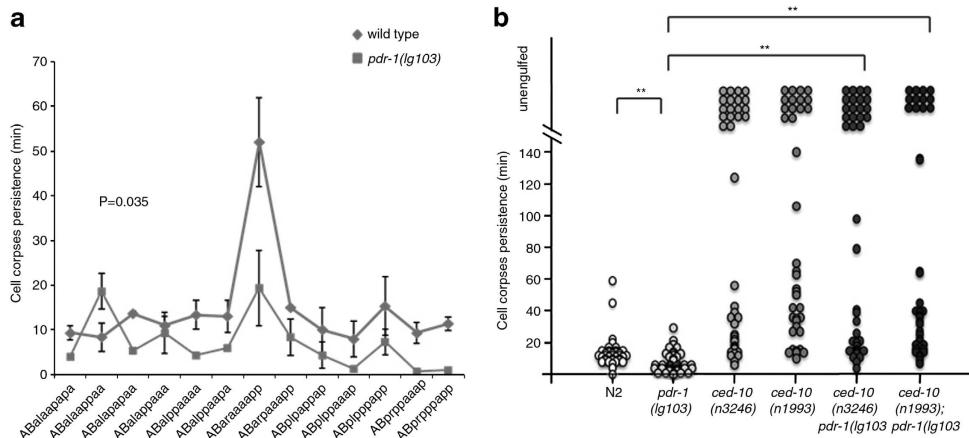


Figure 2 Loss of *pdr-1* increases engulfment kinetics. The first 13 apoptotic cell deaths in the AB lineage were followed by 4D microscopy and the cell corpse persistence was measured. Cells indicated as not engulfed either floated off the tissue into the egg-shell cavity or their lineage could not be followed anymore owing to the beginning of muscle contraction at the 1.5-fold stage (320 min after fertilization or approximately 120 min after the onset of cell death). Analysis was performed in three independent individuals in each genetic background (*n* = 3 × 13 cells). Alleles used were as follows: *ced-10(n3246)*, *ced-10(n1993)*, *pdr-1(lg103)* and *pdr-1(tm395)*. (a) The name of each analyzed cell was represented by the X axis and statistical analysis was carried out using Wilcoxon’s rank-sum test. (b) Each circle represents a single cell and the statistical analysis was carried out using Kolmogorov–Smirnov test (***P* < 0.01)

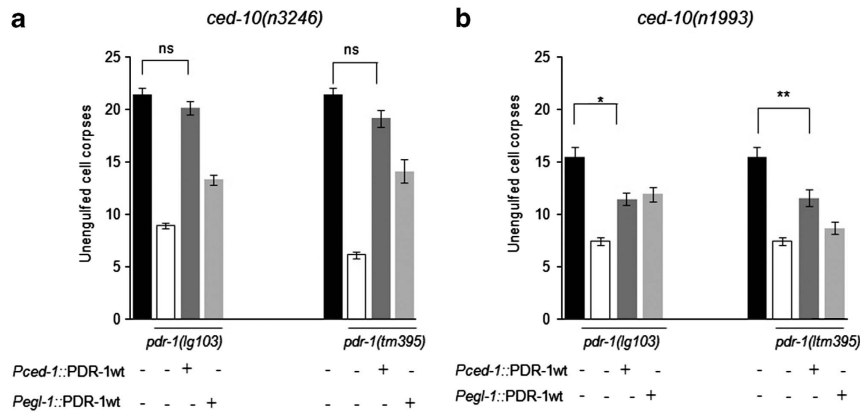


Figure 3 Overexpression of *pdr-1* in the engulfing cell reverses the effect of *pdr-1* mutations. (a) The rescue of *pdr-1::gfp* in the engulfing cell (driven by *Pced-1*) was partially or completely achieved in the double mutant strains *ced-10(n3246);pdr-1(lg103)* and *ced-10(n3246);pdr-1(tm395)*. (b) The rescue of the transgenic strain *pdr-1::gfp* in the engulfing as well as in the apoptotic cells (driven by *Pegl-1*) was partially accomplished in the *ced-10(n1993);pdr-1(lg103)* double mutant. The rescue of *pdr-1::gfp* in the engulfing cell was also partially achieved in the *ced-10(n1993);pdr-1(tm395)* double mutant strain. This *P*-value refers to comparisons between the strains with the wild-type *pdr-1* under the two different promoters (*Pced-1* or *Pegl-1*) with the double mutant strains in which *pdr-1* decreases the number of cell corpses. All data are means \pm s.d. (Statistics: paired *t*-test; two-tailed, unequal variances) (***P* < 0.01 and **P* < 0.05). Between 30 and 50 animals were scored per genotype

(Wu *et al.*⁴⁵ and Figures 4b and c). To elucidate whether this pathway was also affected by *pdr-1*, DTC migration defects were analyzed in *pdr-1* mutants alone and in combination with *ced-10*. Animals containing only any of the mutations of *pdr-1* presented here showed the normal U-shaped gonadal arms. However, mutations of *pdr-1* decreased the percentage of gonadal morphology defects in the two *ced-10* alleles tested (Figure 4). In the context of DTC, the effect exerted by both alleles of *pdr-1* on the most severe mutation *ced-10(n3246)* robustly confirms the genetic role of the modulation of the DTC migration phenotype by *pdr-1*.

The amount of CED-10 is increased in the absence of PDR-1. The results presented above strongly suggest PDR-1 as a negative modulator of the engulfment.

Both *ced-10* alleles analyzed still maintain some remaining activity.^{30,31} An increase in the pool of CED-10 active protein will also accelerate the engulfment machinery, resulting in less cell corpses in L1 animals. The decrease in the cell corpses observed in *ced-10;pdr-1* double mutants might be the consequence of CED-10 accumulation because of PDR-1 miss function. This hypothesis was analyzed biochemically by immunoblot. The genetic strain *ced-10(n3417);CED-10::GFP* (described in the Supplementary Methodology section) was crossed with both *pdr-1* mutants used in the engulfment assays. Worm lysates were prepared as described previously,⁴⁶ and the amount of GFP was detected by chemiluminescence. A band of 46 kDa was observed in all lanes, corresponding to CED-10::GFP (Figure 5a). CED-10::GFP was increased in the worms containing both alleles of *pdr-1* (Figure 5a, second and third lane of the western blot analysis). The 25 kDa band observed in all samples corresponds to the molecular weight of the GFP protein.

These results strongly show a direct contribution of PDR-1/Parkin to the modulation of the CED-10/Rac1 turnover. Moreover, the increased accumulation of CED-10 induced by the strong loss of function allele *lg103* clearly indicates that

the elimination of PDR-1 E3 ubiquitin ligase activity (the deletion removes the UBL) effectively increased the amount of CED-10.

Similar results were obtained when the amount of CED-10::GFP was analyzed from confocal images and quantified (Figures 5b–e and Supplementary Figure S4). The lack of *pdr-1* did not affect the subcellular expression of CED-10::GFP (Supplementary Figures S3 and S4). Total fluorescence was significantly increased in the worms containing the allele *pdr-1(lg103)*, in comparison with the strain *ced-10(n3417);CED-10::GFP*, thus corroborating the results obtained by western blot analysis. Results were statistically significant only for the *lg103* allele of *pdr-1* (*P* < 0.01) (Figure 5e and Supplementary Figure S3).

CED-10 is targeted by PDR-1 for proteasomal degradation. We show for the first the interaction between Rac1 and Parkin, the human orthologs of CED-10 and PDR-1, respectively, in aged human brain (Supplementary Figure S1). The genetic interaction between *pdr-1* and *ced-10* plus the biochemical studies performed in worms led us to test whether the two nematode proteins can also physically interact. The unavailability of specific *C. elegans* antibodies against PDR-1 and CED-10 for immunoprecipitation and for immunohistochemical approaches⁴⁷ hampered analysis of the interaction between the endogenous proteins in the nematode. In consequence, interaction between the nematode proteins was tested in cells. Co-immunoprecipitation experiments were performed in human embryonic kidney 293T (HEK293T) cells transiently transfected with the nematode-tagged proteins FLAG-PDR1 and GST-CED-10. Figure 6a (first panel, lanes 3 and 5, respectively) shows that CED-10 co-precipitate with the two tested forms of PDR-1, the wild-type and the mutant.

As it was previously described for *pdr-1* mutants, they retain the capability of interaction with their protein partners,³² suggesting that the PDR-1-CED-10 interaction was not affected by PDR-1 mutations.

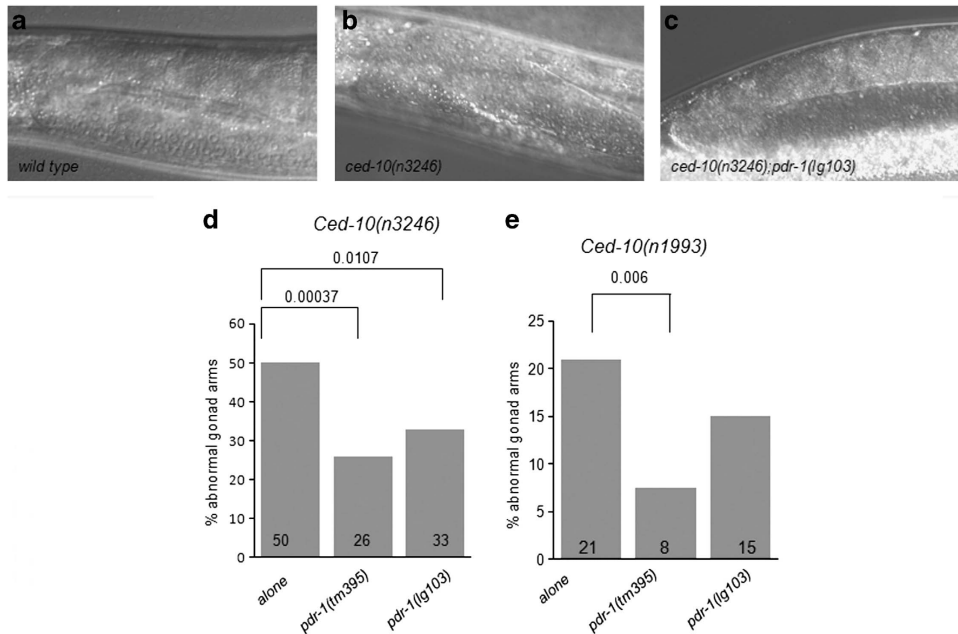


Figure 4 *Pdr-1* mutation suppresses DTC migration defects in both hypomorphic *ced-10* mutants. The gonads of *ced-10(n3246)* or *ced-10(n1993)* animals at the fourth larval stage (L4) with or without mutations in *pdr-1* were scored for morphology using DIC microscopy (d, left and right, respectively). (a) Representative picture showing the normal U-shaped gonad in wild type, *pdr-1* and a percentage of *ced-10;pdr-1* animals. (b and c) Representative DIC pictures showing the abnormalities scored and analyzed as in (d). (d and e) Bar graph representing the scoring of DTC abnormalities in the two *ced-10* backgrounds, *ced-10(n3246)* and *ced-10(n1993)*, respectively. More than 50 gonad arms were scored for all genotypes. *P*-values derived using Fisher's exact test are shown

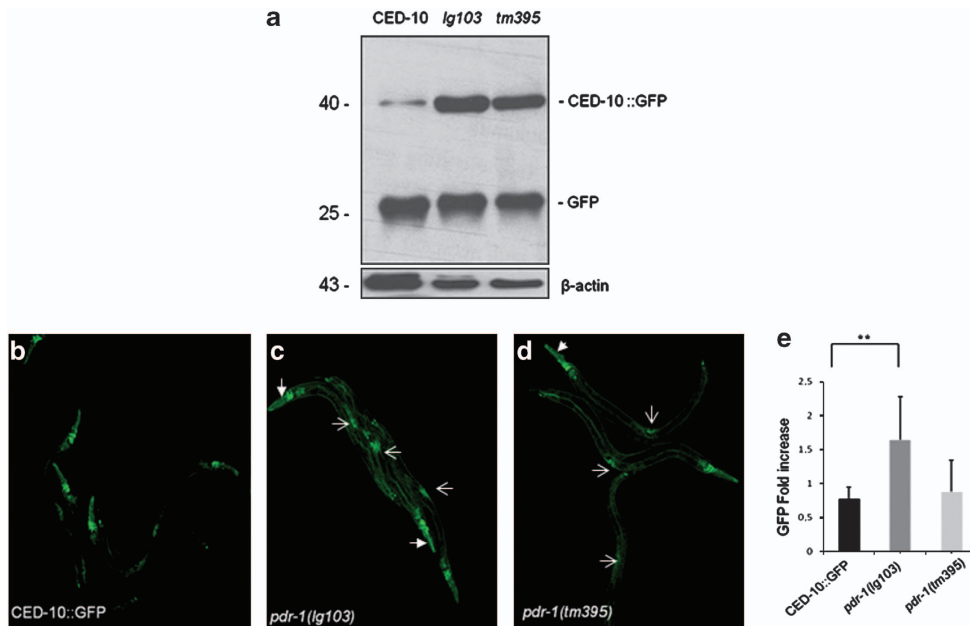


Figure 5 Parkin miss function affects the amount of CED-10. (a) The strain *ced-10(n3417)* harboring the integrated array CED-10::GFP was crossed individually with the *pdr-1* mutant strains, and the amount of GFP correlated with the amount of CED-10 detected by immunoblotting. Twenty micrograms of protein was electrophoresed and blotted against GFP (upper panel). The amount of CED-10::GFP was increased in both *pdr-1* mutant strains analyzed. Antibody anti- β -actin was used as a loading control (bottom panel). The same results were obtained in three independent experiments. (b-d) Confocal fluorescent micrographs of CED-10 translational GFP reporter expression pattern and quantification of total CED-10::GFP in L4 animals. The most evident fluorescent organs are labeled by white arrows, vulva by light arrow and head by thick arrow. Scale bars in (b-d) = 40 μ m. (e) Values are means \pm s.e.m. ($n = 10-15$ worms imaged in three independent experiments). ** $P < 0.01$ in the strain containing the allele *pdr-1(lg103)* compared with the control strain. *P*-value in the strain containing the allele *pdr-1(tm395)* = 0.78948142

Importantly, overexpressed PDR-1 interacted with endogenous human Rac1, the human ortholog of CED-10 (Figure 6b), suggesting a common mechanism of regulation in metazoan.

As PDR-1-CED-10 interaction was not affected by PDR-1 mutations, we next investigated whether the ubiquitylation capacity of PDR-1 was compromised by PDR-1 mutation. First, it was analyzed whether PDR-1 induced the ubiquitylation of CED-10. To test this, HEK293T cells were transiently transfected with the constructs encoding FLAG-PDR-1 and GST-CED-10 and/or His-ubiquitin, and further treated either with or without the proteasome inhibitor MG-132. Levels of ubiquitylation were determined by pulling down His-ubiquitin with Ni²⁺-NTA-agarose beads and immunoblotting for GST-CED-10 protein (Figure 6c). Slowly migrating species of CED-10 corresponding to the ubiquitylated forms were detected only in cells treated with MG-132. No CED-10 immunostaining was observed when ubiquitin-K48R was used in pull-down assays, indicating that PDR-1 ubiquitylated CED-10 through K48 ubiquitin linkages (Figure 6c, lanes 4 and 5, respectively). As expected, the E3 ligase-null mutant of PDR-1 (Ig103) failed to induce CED-10 ubiquitylation in the presence of MG-132 in comparison with the non-mutated PDR-1 (Figure 6d, lanes 4 and 5, respectively).

The proteasomal machinery regulates the engulfment pathway. The modulation of CED-10 turnover by PDR-1 strongly suggested that the proteasomal machinery was a mechanism underlying the regulation of CED-10 activity during the engulfment. To further corroborate this hypothesis *in vivo*, *ced-10* mutant animals were treated by the proteasomal inhibitor MG-132 and the amount of cell corpses were counted as described in the head of L1 larvae (Figure 7). Indeed, the amount of cell corpses was decreased in *ced-10* mutant animals after MG-132 treatment, thus confirming the proteasomal regulation of the CED-10 activity during the engulfment.

Discussion

Our findings support that PDR-1 negatively regulates two mechanisms in which CED-10 has a pivotal role: the engulfment of apoptotic cells and the migration of DTCs during gonadogenesis. Ectopic expression experiments indicate that PDR-1 action in engulfing cells is partly dependent on its UBL-binding domain. PDR-1 ubiquitin ligase function is crucial to modulate the CED-10 ubiquitylation and turnover. Consistent with these findings, the general proteasomal inhibition also ameliorates the engulfment phenotype in

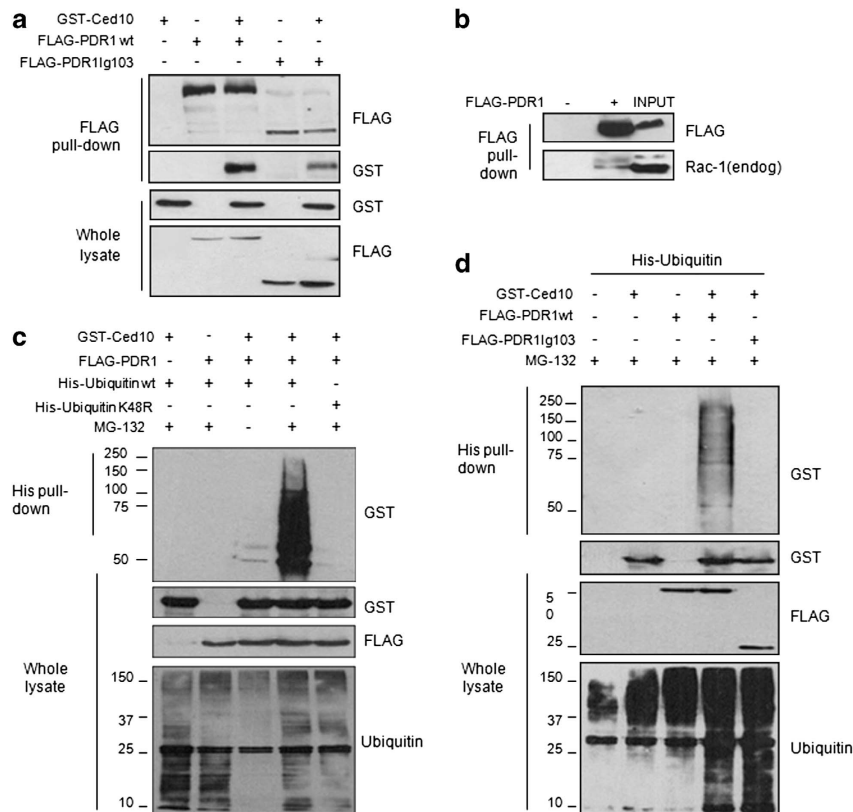


Figure 6 PDR-1 loss-of-function fails to ubiquitylate CED-10. (a) HEK293T cells were transfected with the indicated expression plasmids, and 500 μ g of lysates were used to purify overexpressed FLAG-PDR1 using a FLAG-agarose resin. Immune complexes were immunoblotted for CED-10 and PDR-1 using anti-GST and anti-FLAG antibodies, respectively. Similar results were obtained in three independent experiments. (b) One microgram of HEK293T cells overexpressing FLAG-PDR1 was immunoprecipitated with anti-FLAG-agarose beads. Immune complexes were immunoblotted for human endogenous Rac1 and PDR-1 using anti-Rac1 and anti-FLAG antibodies. (c and d) Cells overexpressing the indicated plasmids were treated with 20 μ M MG-132 for 18 h. His-tagged ubiquitin was purified using Ni²⁺-NTA-agarose beads and the ubiquitylated forms of CED-10 were visualized by immunoblotting using an anti-GST antibody

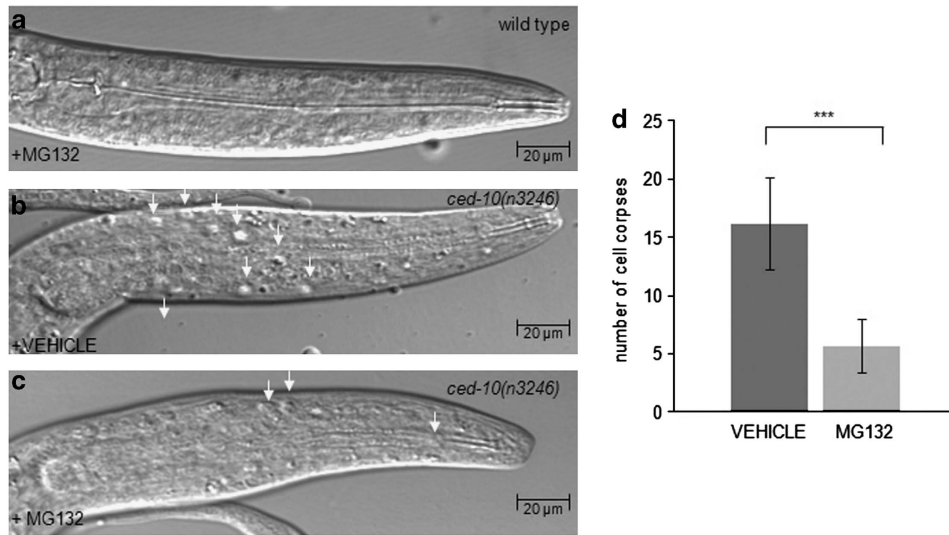


Figure 7 The activity of *ced-10* during engulfment is dependent on proteasomal activity. (a–c) DIC microscopy images of synchronized L1 larvae of the indicated genotypes. Arrowheads indicate persistent cell corpses. Scale bar, 20 μ m. (a) The proteasomal inhibitor MG-132 is not affecting the number of cell corpses in wild-type animals. (b and c) MG-132 treatment decreased the amount of cell corpses in *ced-10(n3246)* animals, (c) compared with (b), in which the worms were incubated only with the solvent DMSO 0.2% as a control. All data are means \pm s.d. (Statistics: paired *t*-test; two-tailed, unequal variances). Between 15 and 25 animals were scored per genotype. (d) Values are means \pm s.d. ****P* < 0.001 in *ced-10* mutant animals treated with MG-132 compared with the *ced-10* animals incubated only with the DMSO 0.2% as a vehicle for the treatment

ced-10 mutants, thus confirming for the first time a proteasomal modulation of the engulfment machinery.

Proteasomal regulation of CED-10/Rac1 in the engulfment. Protein degradation represents the most reliable way to dampen the amplitude and duration of signal-transduction pathways.⁴⁸ It has been previously shown *in vitro* that Rac1, after activation, suffers ubiquitylation and proteasomal degradation.^{11,12} Ubiquitin-mediated degradation of Rac1 (and RhoA) is specifically impaired in different immortalized and cancer cell lines, suggesting that several enzymes specifically target Rho protein members to ubiquitin-mediated proteasomal degradation.⁴⁹ In fact, it has been recently described that Rac1 activity is regulated by the proteasomal machinery in other scenarios, and, as described in the introduction, the E3 ligases HACE1, c-IAP and XIAP are some examples.^{13–15} Our MG-132 experiments clearly demonstrate that the activity of CED-10 during the engulfment is regulated by the proteasomal machinery as well. Concretely from the proteasomal machinery ligases, PDR-1 interacts with human Rac1 and ubiquitylates CED-10. The ameliorated phenotypes of *ced-10;pdr-1* mutants might be the consequence of an increased CED-10 stability owing to the lack of degradation by loss-of-function PDR-1. The use of hypomorphic but severe alleles with remaining activity, such as *ced-10(n3246)*, is crucial to investigate activity modulators. Obviously, this aim cannot be achieved by using null alleles such as *ced-10(n3417)* or *ced-10(t1875)* in which the whole activity is absent.

In conclusion, our results add Parkin/PDR-1 to the ligases regulating Rac1/CED10, and propose Parkin/PDR-1 as the one regulating cellular engulfment. Both proteins Parkin/PDR-1 and Rac1/CED10 act in the same cellular compartment,^{22,35} and according to our scenario, PDR-1 miss function would

increase CED-10 stability during the development of the worm, thus improving the GTPase activity, and lastly resulting in an increase in the rate of engulfment.

Although the high degree of homology between PDR-1 and CED-10 and their human counterparts has been well established so far, further work must be performed to extrapolate these results to humans.

PDR-1/Parkin as a suppressor of the engulfment machinery. Negative regulators of the engulfment machinery in *C. elegans* are *swan-1*,⁵⁰ *mtm-1*,⁴⁰ *abl-1*,⁷ *pgrn-1*,⁵¹ *srpg-1*³⁹ and the recently reported E3 ligase *sli-1*,⁸ in which the E3 ligase activity is dispensable to modulate engulfment, contrary to that detected by *pdr-1*. Among them, only SRGP-1 (Slit-Robo Gap-1)³⁹ and PGRN-1⁵¹ show effects on the kinetics of engulfment as PDR-1 does.

The multitude of regulatory molecules acting on CED-10 (Rac1) emphasizes the key position of this GTPases in the engulfment. The CED-5–CED-12 complex acting as a GEF (by promoting the active, GTP-bound state)⁵² and SRGP-1 acting as a GAP (by promoting the GDP-bound state and inactivation)³⁹ are the main regulators described so far acting in engulfment machinery. Our results add a further complexity in this process. We propose PDR-1 as the E3 ligase responsible for the maintenance of the right ubiquitylation state, to guarantee the stabilization of CED-10 and its efficient regulation by the corresponding GEFs and GAPs. However, it is likely that additional regulators exist that act at other points within these pathways.

All living tissues have some mechanism(s) in place to handle corpse clearance, and most cell types possess the ability to phagocytose apoptotic cells, underlining the relevance of this process in metazoan cells. Engulfment of apoptotic debris induces anti-inflammatory responses that result in a quiet

death, free from inflammation and cytotoxicity to neighboring cells.⁵³ In contrast, materials associated with Alzheimer's disease or Parkinson's disease appear to induce a proinflammatory glial response that is neurotoxic (summarized in Sokolowski and Mandell⁵⁴). In this scenario, engulfment might be associated with the clearance of neuronal debris and increased generation of reactive oxygen species as a consequence. Besides, a correct and efficient clearance will avoid an exaggerated immune response after cell death. Further work is required to study which are the consequences of the Rac1/Parkin interaction in human disease and whether this interaction is relevant for the development of the juvenile variant Parkinson's disease, characterized by Parkin mutations. Finally, future therapeutic drugs designed to modulate efficiently the activity of Rac1 might contribute to the amelioration of diseases in which Rac1 activity and cell clearance are clearly involved, such as cancer and autoimmunity.

Materials and Methods

The complete methodology is extensively described in the Supplementary Methodology section.

General methods and strains. *C. elegans* strains were maintained at 20 °C on nematode growth medium plates seeded with *Escherichia coli* strain OP50 as described previously by Brenner.⁵⁵ N2 (Bristol) was used as the wild-type strain. The mutations used in this study are listed by chromosomes as follows: linkage group LGI – *ced-1(e1375)*; LGIII – *pdr-1(lg103)*, *pdr-1(tm395)*; LGIV – *ced-3(n717)*, *(n2438)*, *ced-5(n1812)*, *ced-10(n1993)*, *(n3246)*. Extrachromosomal array – *pdr-1(lg103);byEx429[K08E3];sel-12::gfp*.²² Integrated array – *ced-10(n3417)::CED-10::GFP* (gift from Dr. Erik Lundquist, University of Kansas, Lawrence, KS, USA; strain LE999).

Molecular biology. Recombinant DNA, polymerase chain reaction and cloning procedures were performed using standard protocols.⁵⁶

Quantification of engulfment defects. Unengulfed apoptotic cell corpses were visualized directly in the heads of young larvae as refractile discs using Nomarski differential interference contrast (DIC) microscopy (Leica DM 6000 microscope).^{57,58} Statistical analysis was performed using GraphPadPrism Version 4 software (GraphPAD Software, La Jolla, CA, USA). *P*-values for pairwise comparisons were calculated using the Student's *t*-test.

Quantification of cell-death defects. For quantification of cell-death defects in the anterior pharynx, animals in the third larval stage (L3) were anesthetized and viewed with DIC microscopy as described.³⁸

Rescue experiments. *Ced-10(n3246);pdr-1(lg103)* hermaphrodites were crossed with males of the already generated rescue strain *pdr-1(lg103);byEx434[pBY1500]*, in which pBY1500 contains the complete operon of *pdr-1* named K08E3 and *sel-12::gfp* has been used as a comarker.²² To facilitate the understanding of the rescue experiments, the cosmid name has been substituted by 'PDR-1 wild type' and the comarker *sel-12::gfp* has been omitted. L1 green worms from the F3 generation were scored for cell corpses as described previously.

Cell autonomous rescue. To address the question of the role of *pdr-1* in the engulfing and/or the dying cell, we used two already well-characterized promoters of *ced-1* and *egl-1*, respectively.^{6,42} CED-1 is a cell surface phagocytic receptor that recognizes cell corpses,⁶ and EGL-1 localizes specifically in apoptotic cells.⁴² The corresponding translational fusions were generated as described previously.²² *Pced-1::gfp* and *Pegl-1::gfp* were kindly provided by Dr. Z Zhou (Baylor College of Medicine, Houston, TX, USA) and Dr. B Conrath (Ludwig-Maximilians-Universität München (LMU), Munich, Germany), respectively.

Video recording. 4D microscopy was carried out using standard live-animal mounting techniques on a Leica DM 6000 microscope fitted with Nomarski optics.

The use of Nomarski optics allows identification of key time points during apoptosis without using any dye or fluorescent marker that might alter the process. Embryonic cell death was determined as described.⁵⁹

***C. elegans* western blot analysis.** To elucidate the dependence on the amount of CED-10 by PDR-1, we took advantage of the strain LE999 in which *ced-10(n3417)* IV harbors the transgene CED-10::GFP in an array (strain kindly provided by Dr. E Lundquist).³⁵ *Ced-10(n3417)* lacks exons 2 and 3 (from 666 bp 3' of the *ced-10* ATG, within intron 2, to 1637 bp 3' of the *ced-10* ATG, within intron 3) of the *ced-10* locus, which is predicted to result in the expression of only the first 35 amino acids of CED-10.³⁵ The *pdr-1* mutant strains *pdr-1(lg103)* and *pdr-1(tm395)* were crossed individually with the strain LE999. Worm pellets from the corresponding genotypes were processed for western blot analysis as described.⁴⁶ The expanded method is detailed in the Supplementary Methodology section.

Confocal analysis of the CED-10::GFP reporter. *In vivo* analysis of the CED-10 reporter was investigated by analyzing the strain LE999 described above. To quantify GFP expression, L4 stage animals were mounted on 5%, 20 mM sodium azide agar pads and confocal images were collected from 10–15 animals. We then measured the pixels produced by the fluorescence in the whole body of animals and determined the fluorescence level relative to worm area, using the ImageJ software (<http://imagej.nih.gov/ij/plugins/index.html>). The final value, by means of internal density, was obtained by subtracting the background of individual images by using the pluggin 'background subtraction from ROI' included in the software. *P*-values for pairwise comparisons were calculated using the Student's *t*-test.

Cell culture, transient transfection, immunoprecipitation and immunoblotting. HEK293T cells were cultured in 10-cm diameter dishes at 37 °C in Dulbecco's modified Eagle's medium (Sigma, Barcelona, Spain), containing 10% fetal bovine serum and antibiotics. Cells were lysed in ice-cold lysis buffer (50 mM Tris-HCl (pH 7.5), 1 mM EDTA, 150 mM NaCl, 0.5% Nonidet P-40 and protease inhibitor cocktail), centrifuged at 12 000 × *g* for 15 min at 4 °C and supernatants were then stored at –20 °C until use. Immunoprecipitation experiments were performed as described previously.⁶⁰

Ubiquitination *in vivo* assay. The purification of His-ubiquitinated conjugates was performed following the method described by Erazo *et al*.⁶¹

Proteasome inhibitor treatment. To test *in vivo* whether the CED-10 activity during the engulfment was regulated by the proteasomal machinery, the worms were treated with the commonly used proteasome inhibitor MG-132 (Calbiochem, Barcelona, Spain). The MG-132 experiments were performed as described previously, with some modifications.^{62,63} Briefly, L4 stage larvae of the corresponding genotypes were grown on *E. coli* (strain OP50)-seeded NGM plates containing fresh MG-132 50 μM with 0.2% DMSO as a solvent control both in the agar media and in the OP50-like bacteria. After 2 days, the mothers were washed off the plates and bleached, and the hatched L1 progeny was incubated in M9 + MG-132 50 μM with 0.2% DMSO as a solvent control overnight at 20 °C. To see the difference between MG-132-treated and -non-treated worms, animals were mounted on agar pads and the quantification of the engulfment defects, by means of the amount of cell corpses, was performed as described above. *Ced-10(n3246)* worms treated with 0.2% DMSO were used as a control, to discard the effects of the solvent *per se* in the quantification of the cell corpses. *P*-values for pairwise comparisons were calculated using the Student's *t*-test.

Conflict of Interest

The authors declare no conflict of interest.

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The constructs PDR-1-FLAG and PDR-1(LG103)-FLAG were kindly provided by Dr. Wolf Dieter Springer. This work was supported in part by The Spanish Ministry of Science and Competitiveness grants PC0009/003 and PI1100968 (E Dafo and I Ferrer respectively), Riojasalud Foundation and The Spanish Ministry of Science and Innovation Grant BFU2010-21794 (J Cabello and E Gómez), Grant DFG (Deutsche Forschungsgemeinschaft) SFB850 and SFB746 (R Baumeister) and PIF fellowship from Universitat Autònoma de Barcelona and Spanish Ministerio de Educación y Ciencia grant BFU2007-60268 (T Erazo and J Lizcano respectively).

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