

1 **Photoelectrocatalytic inactivation of *Pseudomonas***
2 ***aeruginosa* using an Ag-decorated TiO₂ photoanode**

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17

18 **Abstract**

19 Fast and total inactivation of Gram-negative *Pseudomonas aeruginosa* in suspensions at
20 natural pH 5.9 has been achieved by photoelectrocatalysis (PEC), using Ag-decorated
21 TiO₂ photoanodes onto transparent [conducting indium tin oxide \(ITO\)](#) under UVA
22 irradiation. The assays were made with 100 mL of bacterial suspensions in an undivided
23 cell equipped with a photoanode, a stainless steel cathode and Ag|AgCl (3 M KCl) as
24 reference electrode. Total inactivation was obtained in only 5 min using coatings with 4
25 wt.% Ag, 25 mM Na₂SO₄ as the electrolyte and 1.70 V as applied bias potential.
26 Comparative photocatalytic treatments reached total inactivation at much longer time,
27 suggesting the crucial role of hydroxyl radicals in PEC. These oxidants, which were
28 detected by electron spin resonance, attacked the outer cell wall very effectively, since
29 the recombination of the electron/hole pairs photoinduced under UVA irradiation was
30 reduced. As characterized by [high-resolution transmission electron microscopy](#)
31 [\(HRTEM\)](#), the best synthesized Ag-TiO₂ thin-film photoanode mainly contains anatase
32 TiO₂ nanopowder decorated with Ag nanoparticles of ca. 45 nm. Analyses by X-ray
33 powder diffraction and UV/Vis spectroscopy were also performed. The potential use of
34 PEC for bacterial disinfection was confirmed for the rod-shaped Gram-positive *Bacillus*
35 *atrophaeus*, which was more slowly inactivated due to its different cell wall structure.
36 Scanning electron micrographs of both bacteria showed that PEC induced a high
37 roughness, cell lysis and accumulation of cellular debris.

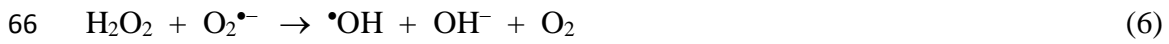
38 *Keywords:* *Bacillus atrophaeus*; Bacterial inactivation; Photoelectrocatalysis;
39 *Pseudomonas aeruginosa*; Water disinfection

40 1. Introduction

41 Recently developed electrochemical advanced oxidation processes (EAOPs) like
42 electrochemical oxidation (EO), electro-Fenton, UV- or solar-assisted photoelectro-
43 Fenton and photoelectrocatalysis (PEC) have shown their great potential for water
44 decontamination and disinfection [1-8]. The destruction of suspended or attached
45 biofilms of pathogenic bacteria is a crucial need in drinking water production,
46 swimming pools, washing in food processing, water distribution or ballast water
47 management [9]. Ozone, chlorine dioxide and chlorine are typically utilized for water
48 disinfection. However, they can yield hazardous products, including haloacetic acids
49 and trihalomethanes [1]. To overcome these drawbacks, non-toxic antibacterial
50 nanostructures have been proposed [10,11]. EAOPs also represent an alternative for
51 bacterial inactivation, since they are eco-friendly methods based on the generation of
52 reactive oxygen species (ROS) like the hydroxyl radical ($\bullet\text{OH}$), which is able to react
53 with cell membrane components causing lysis and the death of bacteria [1,12,13].

54 PEC is electrochemically-assisted photocatalysis (PC) [14]. PC consists in the
55 photoexcitation of an electron from the valence band (VB) of a semiconductor to the
56 conduction band (CB, e^-_{CB}), with production of a positively charged vacancy or hole
57 (h^+_{VB}) from reaction (1) [14-18]. The photogenerated holes can oxidize organic
58 molecules or react with water to form the $\bullet\text{OH}$ via reaction (2). The photoinduced
59 electrons can produce other ROS like superoxide radical anion ($\text{O}_2^{\bullet-}$), hydroperoxyl
60 radical ($\text{HO}_2\bullet$) and H_2O_2 from reactions (3)-(6).





67 The main drawback of PC is the fast recombination of the e^-_{CB}/h^+_{VB} pairs, which
68 causes a great efficiency loss. This is solved in PEC by applying either a constant
69 current density (j) or a constant bias anodic potential (E_{bias}) to the illuminated
70 semiconductor. This allows the continuous extraction of the e^-_{CB} from the anode
71 through the external electrical circuit, thus inhibiting reactions (3)-(6) and
72 recombination. Hence, a higher effective accumulation of holes and $\bullet OH$ from reactions
73 (1) and (2) is feasible, leading to much higher overall efficiency than PC [14,17].

74 The best semiconductor for PC and PEC is TiO_2 because it is non-toxic, highly
75 stable against photocorrosion and inexpensive. It is typically utilized as a mixture of
76 two nanocrystalline forms, namely anatase and rutile, needing UV light to photoexcite
77 the electrons from their VB since their bandgap (E_{bg}) is 3.0-3.2 eV [17]. TiO_2 thin
78 layers from powders can be prepared by several procedures including sol-gel [14,19],
79 microwave-assisted dip-coating [20], chemical vapor deposition [14] or electrophoretic
80 deposition [21-23]. Ti, indium tin oxide (ITO) and fluorinated tin oxide (FTO) are
81 widely used as substrates.

82 The photocatalytic efficiency of TiO_2 thin films in practice is limited by the small
83 UV fraction ($\sim 5\%$) of the solar spectrum [24]. Lately, three main strategies have been
84 followed to improve the oxidation power of this semiconductor in PEC [14,17]: (i) use
85 of nanostructured TiO_2 materials like nanotube bundles or arrays, nanobelts, nanowires
86 or nanorods, showing large effective area that allows greater light absorption; (ii) band-
87 gap engineering to absorb visible light, based on TiO_2 doping to substitute O atoms
88 either with transition metal ions like Cr, Co, W, Zr and Fe or with non-metals like N, F,

89 S, B and C; and (iii) surface decoration with noble metal nanoparticles like Ag, Pd or
90 Au to act as electron trap of photoinduced electrons. In the case of Ag-TiO₂, Ag
91 promotes the charge transfer at the interface with TiO₂, minimizing the e⁻_{CB}/h⁺_{VB}
92 recombination [17,18]. Furthermore, the surface plasmon resonance of Ag nanoparticles
93 enhances the visible light response of the photocatalyst [25]. The use of Ag-TiO₂ for
94 PEC disinfection is particularly interesting because of the high antibacterial activity and
95 biocompatibility of Ag [26-28].

96 The decoration of semiconductors with Ag particles is carried out by dipping the
97 substrate into an Ag⁺ solution, followed by exposure to UV radiation to induce Ag⁺
98 photoreduction [28-33]. The efficient photocatalytic power of Ag-TiO₂ films for the
99 degradation of 10 mg L⁻¹ phenol solutions at pH 3.0 has been reported using UV light
100 [34] and sunlight [35]. Recently, several authors have described the inactivation of
101 different bacteria by PC under UV and solar irradiation using Ag-TiO₂ [36-38]. Ag-
102 TiO₂ photoanodes have also been studied for PEC inactivation of *Escherichia coli* and
103 *Staphylococcus aureus* [39], *Mycobacterium smegmatis* [28] and *Mycobacterium*
104 *kansasii* and *Mycobacterium avium* [40], showing the superiority of the decorated
105 photoanodes.

106 *Pseudomonas aeruginosa* is a rod-shaped Gram-negative bacterium, ubiquitous in
107 soil and water as well as in animals and in plants [12]. This aerobic pathogen can cause
108 mild to severe infections such as pneumonia, dermatitis, keratitis, otitis and nosocomial
109 infections [41]. The increasing antibiotic resistance of such bacteria makes necessary
110 the use of efficient methods to ensure a fast inactivation in water. Nouri et al. [11]
111 reported the mortality of 99% of 5.6×10¹³ CFU mL⁻¹ of *P. aeruginosa* using chitosan
112 films loaded with 3% mormollite/CuO nanocomposite. Hussain et al. [42] showed
113 complete disinfection of water with 1.8×10⁹ CFU mL⁻¹ in 0.3% NaCl at pH 7 after 20

114 min of adsorption onto a graphite intercalation compound. Inactivation of about 99%
115 was also found by electrolyzing 10^8 CFU mL⁻¹ of *P. aeruginosa* with a stainless steel
116 anode at $j = 70$ $\mu\text{A cm}^{-2}$ for 1 h [43]. Additionally, the effectiveness of $\bullet\text{OH}$ has been
117 well proven by means of Fenton process with ferrocene-loaded polymeric micellar
118 nanoplateforms [10], EO with boron-doped diamond (BDD) and dimensionally stable
119 anodes [44], and PC with Ag-TiO₂ microspheres under UVC illumination [36].

120 The aim of this work is to study the inactivation of *P. aeruginosa* by UVA-assisted
121 PEC with Ag-decorated TiO₂ deposited onto ITO. The influence of the percentage of
122 Ag loading, supporting electrolyte content and E_{bias} on the disinfection process was
123 examined. Comparative treatment by PC was made to show the advantages of PEC. The
124 production of $\bullet\text{OH}$ was confirmed by electron paramagnetic resonance (EPR). The
125 characteristics of the best Ag-TiO₂ photoelectrode were ascertained by high-resolution
126 transmission electron microscopy (HRTEM), energy dispersive X-ray analysis (EDX),
127 X-ray powder diffraction (XRD) and UV/Vis spectroscopy. The study was extended to
128 the Gram-positive bacterium *Bacillus atropheus* to show the performance of the
129 photoanode against other kinds of cell walls. The changes in bacterial morphology were
130 analyzed by scanning electron microscopy (SEM).

131 2. Materials and methods

132 2.1. Chemicals

133 Sodium sulfate used as supporting electrolyte was of analytical grade purchased
134 from Panreac Química. Reagents used for the preparation of the photoanodes, bacterial
135 culture and all the analysis were of analytical grade purchased from Panreac Química,
136 Merck, Fluka and Sigma-Aldrich. Solutions were prepared with ultrapure water from a
137 Millipore Milli-Q system (resistivity > 18.2 M Ω cm).

138 2.2. Preparation of the Ag-TiO₂ photoanodes

139 First, TiO₂ nanoparticles were deposited onto ITO glass substrates purchased from
140 MSE Supplies (25 mm × 50 mm, 1.1 mm thickness, 10 Ω sq⁻¹), which were
141 electrochemically cleaned and then introduced as the cathode in a conventional
142 electrophoretic cell equipped with a boron-doped diamond (BDD) anode placed in
143 parallel at an interelectrode distance of 1.0 cm. The back side of the ITO was
144 conveniently isolated to prevent any deposition. The cell was filled with 30 mL of a
145 suspension of 1.5 g TiO₂ P-25 (80% anatase, 20% rutile, average particle diameter of 20
146 nm) purchased from Degussa in 5% (v/v) isopropanol/water, after 15 min of sonication.
147 TiO₂ deposition was performed by applying a cell voltage of 4 V for only 60 s at
148 ambient temperature. The resulting ITO/TiO₂ thin film was subsequently annealed at
149 450 °C for 30 min. The deposition / annealing cycle was made four times in order to
150 ensure the formation of a compact layer of TiO₂ with good adherence and crystallinity.
151 [A sketch of the setup use for the preparation of ITO/TiO₂ is shown in Fig. S1a.](#)

152 The decoration of the above ITO/TiO₂ material with Ag was performed using a
153 solution containing 27.5 mL isopropanol, 0.8 mL Milli-Q water, 0.4 mL HNO₃ and
154 either 0.0153 or 0.0612 g AgNO₃ (Sigma-Aldrich) to obtain photoanodes with 1% or
155 4% (w/w) Ag, respectively. The ITO/TiO₂ was immersed into the corresponding
156 solution for 16 h at ambient temperature in the dark, dried at ambient temperature for 15
157 min and then irradiated with a 125 W UVC light bulb ($\lambda_{\max} = 254$ nm) for 90 min to
158 photoreduce the adsorbed Ag⁺ ion to Ag ([see Fig. S1b](#)). Homogeneous and stable Ag-
159 TiO₂ thin-film photoelectrodes of purple color were obtained after annealing at 450 °C
160 for 30 min.

161 2.3. Bacteria and culture

162 The inactivation of the rod-shaped Gram-negative bacterium *P. aeruginosa* ATCC
163 15442 and Gram-positive bacterium *B. atrophaeus* ATCC 9372 (previously named *B.*
164 *subtilis*) was studied. The strains of both bacteria were cultured in Trypticasein Soy
165 Agar (TSA) plates, supplied by Laboratorio Conda, at 37 °C for 24 h. After that, each
166 microorganism was spiked into 2 mL of 7 mM Na₂SO₄, followed by centrifugation at
167 14,000 rpm for 2 min and washing with 1 mL (3 times) of the same electrolyte. The
168 final pellet was resuspended in 1 mL of 7 mM Na₂SO₄, yielding an optical density at
169 600 nm (O.D. 600) of 0.7±0.1, corresponding to about 10⁸ CFU mL⁻¹.

170 2.4. Inactivation experiments

171 The disinfection assays by PEC were carried out in a conventional cylindrical three-
172 electrode cell of Schott Duran[®] glass of 150 mL capacity. Thermostated water was
173 recirculated through a jacket surrounding the cell to keep the suspension temperature at
174 25 °C. The anode was a 3 cm² Ag-TiO₂ thin-film photoelectrode synthesized as
175 described above, the cathode was a Pt spiral and the reference electrode was Ag|AgCl (3
176 M KCl). The back side of the coated ITO substrate was illuminated with an Omnilux
177 125 W UVA light bulb ($\lambda_{\max} = 360$ nm), placed in parallel at a distance of 6 cm. An
178 Amel 2053 potentiostat-galvanostat was used to provide a bias potential (E_{bias}) to the
179 photoanode vs. the reference electrode. [A scheme is shown in Fig. S2](#). For comparison,
180 PC assays were performed under the same experimental conditions, with the prepared
181 Ag-TiO₂ photoelectrode acting as photocatalyst, but without applying a bias potential.

182 The trials were carried out with 100 mL of aqueous solutions containing 7 or 25
183 mM Na₂SO₄ at natural pH 5.9, always under vigorous stirring at 700 rpm with a
184 magnetic PTFE stirring bar. Each sulfate solution was spiked with a 10⁸ CFU mL⁻¹ of *P.*
185 *aeruginosa* or *B. atrophaeus* to obtain a bacterial suspension with 10⁶ CFU mL⁻¹.
186 Before each assay, the cell was cleaned with a H₂O₂:H₂SO₄ mixture (30:70 v/v) for 10

187 min, rinsed with Milli-Q water and dried in an oven at 80 °C. After usage, the electrodes
188 were immersed in ultrapure water at 100 °C for 10 min and finally, air dried.

189 2.5. Analytical procedures

190 The pH and O.D. 600 of the bacterial suspensions were measured with a Crison
191 GLP 22 pH-meter and a Camspec M108 spectrophotometer, respectively.

192 To assess the inactivation of *P. aeruginosa* and *B. atrophaeus* suspensions, 1-mL
193 samples were withdrawn at different times for 45 min of treatment as maximal. They
194 were diluted in 7 mM Na₂SO₄, cultured in duplicate on TSA plates and incubated at 37
195 °C for 24 h. The percentage of inactivation was calculated as follows:

$$196 \quad \% \text{ Inactivation} = \frac{N_0 - N_t}{N_0} (100) \quad (7)$$

197 where N_0 is the initial CFU value and N_t is its content at time t . The theoretical detection
198 limit was 1 bacterium per mL. All the trials were made in triplicate and average values
199 of percentage of inactivation are shown in figures, along with the error bars associated
200 to a 95% confidence interval.

201 The morphological features of each microorganism before and after the inactivation
202 trials were analyzed by SEM using a JEOL JSM-7001F system at 15 kV [12]. To do
203 this, each bacterial suspension was filtered through a 0.2 μm polycarbonate membrane
204 filter purchased from Millipore. The filter was immersed in a 2.5% glutaraldehyde and
205 0.1 M cacodylate solution at pH 7.4 for 30 min and further, it was post-fixed in 1%
206 OsO₄, washed with 0.2 M sodium cacodylate, dehydrated with a graded series of
207 ethanol solutions from 30% to 100% and dried. The final samples were coated with gold
208 to carry out the observation by SEM.

209 The morphological characteristics and surface composition of the raw TiO₂ and the
210 synthesized Ag(4%)-TiO₂ materials were examined by HRTEM and EDX. The latter

211 was powder scratched from the thin-film photoelectrode surface. Both powders were
212 ultrasonically dispersed in isopropyl alcohol and supported on holey carbon-coated
213 copper grids. These analyses were performed with a JEOL JEM-2100 LaB6
214 transmission electron microscope at 200 kV, and the EDX spectra were acquired using
215 an INCA X-sight analyzer (Oxford Instruments) with Microanalysis Suite version 4.09
216 software. The crystal structure of the photoelectrode was analyzed by XRD with a
217 PANalytical X'Pert PRO MPD Alpha1 powder diffractometer in Bragg-Brentano $\theta/2\theta$
218 geometry with 240 mm radius, using Cu $K\alpha_1$ radiation ($\lambda = 1.5406 \text{ \AA}$). The reference
219 XRD patterns were selected from ICDD database.

220 The absorbance of the ITO substrate, the ITO/TiO₂ material and the Ag-TiO₂ thin-
221 film photoelectrodes in the UV/Vis region was determined by UV/Vis spectroscopy
222 using a Shimadzu UV-1800 UV/Vis spectrophotometer.

223 The generation of hydroxyl radicals under PEC conditions was confirmed by spin
224 trapping from the analysis of its adduct with 5,5-dimethyl-1-pyrroline-*N*-oxide (DMPO)
225 by EPR [45]. This was made by treating 10 mL of a 9.2 mM DMPO solution with 25
226 mM Na₂SO₄ at pH 5.9 in a three-electrode cell with a 1 cm² Ag(4%)-TiO₂ photoanode
227 under 125 W UVA illumination and a 1 cm² stainless steel spiral cathode, with vigorous
228 stirring provided by a magnetic PTFE bar at ambient temperature. An $E_{\text{bias}} = 1.70 \text{ V}$ vs.
229 Ag|AgCl (3 M KCl) was applied for 15 min. The EPR analysis of the resulting solution
230 was performed with a Bruker ESP300E spectrometer, using Win-EPR and SimFonia 2.3
231 software.

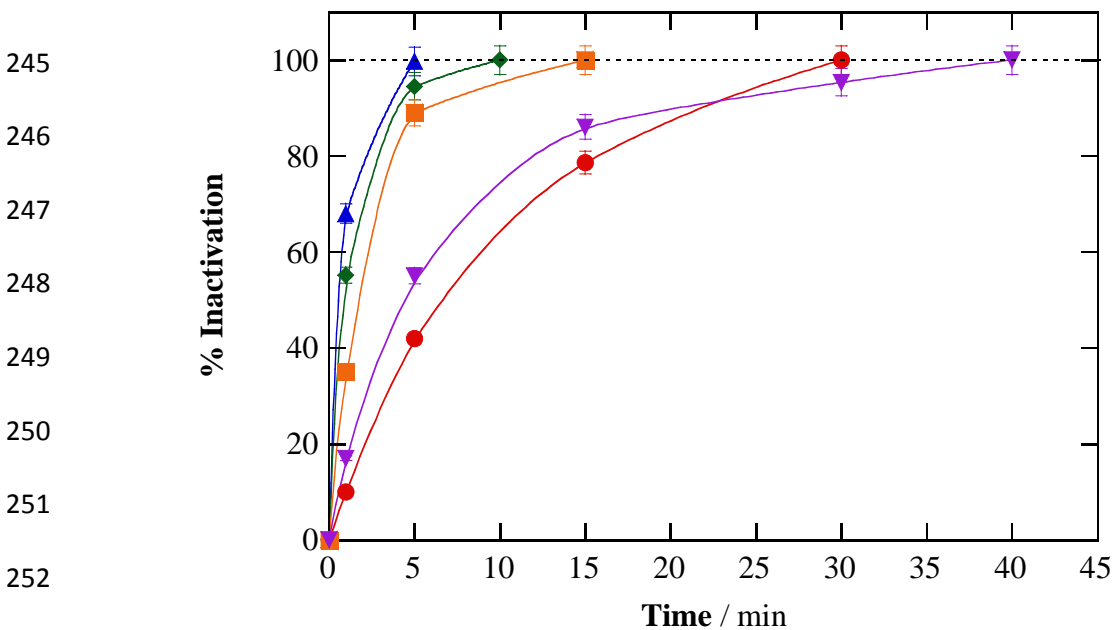
232 **3. Results and discussion**

233 *3.1. Inactivation of Pseudomonas aeruginosa*

234 A set of trials was made to assess the influence of the Ag content in the
235 photoanode, the supporting electrolyte and the E_{bias} applied on the photoelectrocatalytic

236 inactivation of *P. aeruginosa*. The initial pH of 5.9 remained practically unchanged
237 during the trials due to the low current flowing through the electrical circuit and the
238 short electrolysis times utilized.

239 Fig. 1 depicts an evident enhancement of the inactivation rate as the amount of Ag
240 decorating the TiO₂ deposit onto the ITO substrate was increased. As can be seen, for an
241 Ag(1%)-TiO₂ photoanode in 25 mM Na₂SO₄ at $E_{\text{bias}} = 1.70$ V, complete disinfection
242 was reached after 15 min of PEC (filled square), whereas only 5 min (filled triangle)
243 were required when an Ag(4%)-TiO₂ photoanode film was used instead under similar
244 conditions.



253 **Fig. 1.** Percentage of inactivation of 10^6 CFU mL⁻¹ *Pseudomonas aeruginosa* aqueous
254 suspensions with electrolysis time for the photoelectrocatalytic (PEC) treatment of 100
255 mL with (●) 7 mM and (▲, ◆, ■, ▼) 25 mM Na₂SO₄ at pH 5.9 and 25 °C. Trials were
256 run in an undivided cell equipped with a 3 cm² photoanode illuminated with a 125 W
257 UVA lamp and a stainless steel spiral as the cathode. Photoelectrode: (■) Ag(1%)-TiO₂
258 and (▼, ◆, ▲, ●) Ag(4%)-TiO₂. Bias potential (E_{bias}): (◆) 1.00 and (■, ▲, ●) 1.70 V
259 vs. Ag|AgCl (3 M KCl). In (▼), photocatalysis (PC) without the application of a bias
260 potential.

261 This behavior can be related to the larger transfer of electrons from the VB of the
262 TiO₂ nanoparticles, formed from reaction (1), to the more numerous Ag nanoparticles

263 [17,18]. Hence, the deposited Ag assists the external electric field to migrate the
264 photogenerated electrons from the TiO₂ film anode toward the counter electrode [46],
265 thus lowering the electron–hole recombination rates [47]. This phenomenon inhibits the
266 recombination of the generated holes with the e⁻_{CB}, facilitating the production of a
267 larger amount of oxidant •OH from reaction (2) and, finally, promoting the faster
268 inactivation of the microorganisms.

269 Fig. 1 also reveals a quicker inactivation when the Na₂SO₄ concentration rose from
270 7 (filled circle) to 25 mM (filled triangle) operating with the Ag(4%)-TiO₂ photoanode
271 at $E_{\text{bias}} = 1.70$ V. Up to 30 min were needed to completely inactivate the bacterium
272 when the lowest amount of electrolyte was employed, being 6-fold slower as compared
273 to PEC performed with 25 mM. This is due to the much higher j value circulating
274 through the photoelectrolytic cell because of the greater conductivity of the solution as
275 the supporting electrolyte content was increased. The upgrade of j then favors the
276 extraction of higher quantities of electrons accumulated in the Ag, which are transferred
277 from the VB of the TiO₂ by photoinduction. Consequently, the •OH generation is
278 enhanced by the larger amount of holes available, thus accelerating the disinfection
279 process.

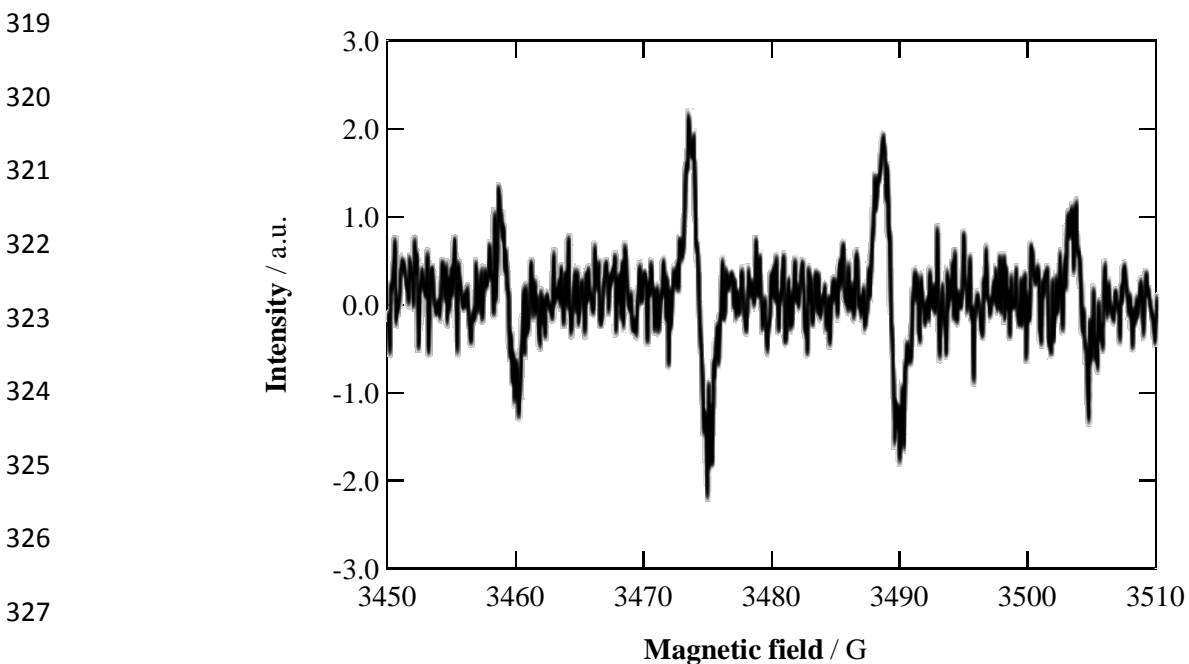
280 Another key parameter in PEC is the applied E_{bias} because it determines the ability
281 of the anode to extract the photoexcited electrons toward the cathode, controlling the j
282 value and the potential difference between the electrodes (E_{cell}). Fig. 1 reveals that using
283 the Ag(4%)-TiO₂ photoanode at $E_{\text{bias}} = 1.00$ V with 25 mM Na₂SO₄ (filled diamond),
284 the time for total inactivation was about 10 min, which is twice the value required at a
285 higher E_{bias} of 1.70 V (filled triangle). No greater inactivation rate was achieved by
286 further applying a higher E_{bias} of up to 2.00 V (data not shown). The higher j attained
287 upon gradual rise of E_{bias} up to 1.70 V favored the extraction of the photoinduced

288 electrons transferred to the Ag from the VB of the TiO₂. This led to a more rapid
289 inactivation of the *P. aeruginosa* cells thanks to the greater generation of •OH from the
290 higher amount of holes formed in the TiO₂, as stated above.

291 The aforementioned findings indicate that, under the experimental conditions
292 tested, the best disinfection rate was achieved using a synthesized Ag(4%)-TiO₂ thin
293 film photoanode at the optimum $E_{\text{bias}} = 1.70$ V with 25 mM Na₂SO₄. These conditions
294 yielded a $j = 0.8$ mA cm⁻² and $E_{\text{cell}} = 5.0$ V. The energy consumption for total
295 inactivation in 5 min was 104.18 kWh m⁻³. This value mainly arises from the UVA
296 lamp consumption (104.17 kWh m⁻³), since the energy requirements of the electrolytic
297 system only accounted for 0.01 kWh m⁻³.

298 Several comparative PC trials were also performed to demonstrate the superior
299 disinfection power of PEC, as shown in Fig. 1. The gradual but slow inactivation of the
300 bacterium by PC under the best PEC conditions (Ag(4%)-TiO₂ photoanode in 25 mM
301 Na₂SO₄) required 40 min for the complete activity loss (upside-down filled triangle).
302 This entails a total energy consumption of 833.34 kWh m⁻³ for the PC treatment, a value
303 much greater than that determined for PEC at $E_{\text{bias}} = 1.70$ V, as pointed out above. This
304 result confirms the very important bactericidal role of the Ag particles added to the TiO₂
305 film in the PEC process, confirming its contribution to the minimization of the
306 recombination of the $e^-_{\text{CB}}/h^+_{\text{VB}}$ pairs photogenerated in the semiconductor surface. This,
307 in concomitance with the application of an optimum E_{bias} , induces a much greater
308 production of •OH from reaction (2). Note that the synthesized Ag(4%)-TiO₂
309 photocatalyst led to faster disinfection in PC as compared to the similar treatment of 10⁸
310 CFU mL⁻¹ of *P. aeruginosa* reported by Li et al. [36], since they needed 120 min for
311 achieving 99% inactivation of suspensions with 200 mg L⁻¹ of Ag-TiO₂ microspheres.

312 The generation of $\bullet\text{OH}$ during the PEC treatment with an Ag(4%)-TiO₂ photoanode
313 at $E_{\text{bias}} = 1.70$ V was confirmed by spin-trapping upon photoelectrolysis of a 9.2 mM
314 DMPO solution with 25 mM Na₂SO₄ at pH 5.9 for 15 min. Fig. 2 presents the EPR
315 spectrum found for the resulting solution, which clearly highlights the four
316 characteristic bands (1:2:2:1) of the $\bullet\text{OH}$ -DMPO adduct. This corroborates the attack of
317 $\bullet\text{OH}$ onto the outer layer of the *P. aeruginosa* in the above PEC trials, eventually
318 inactivating the cells and probably causing their lysis.

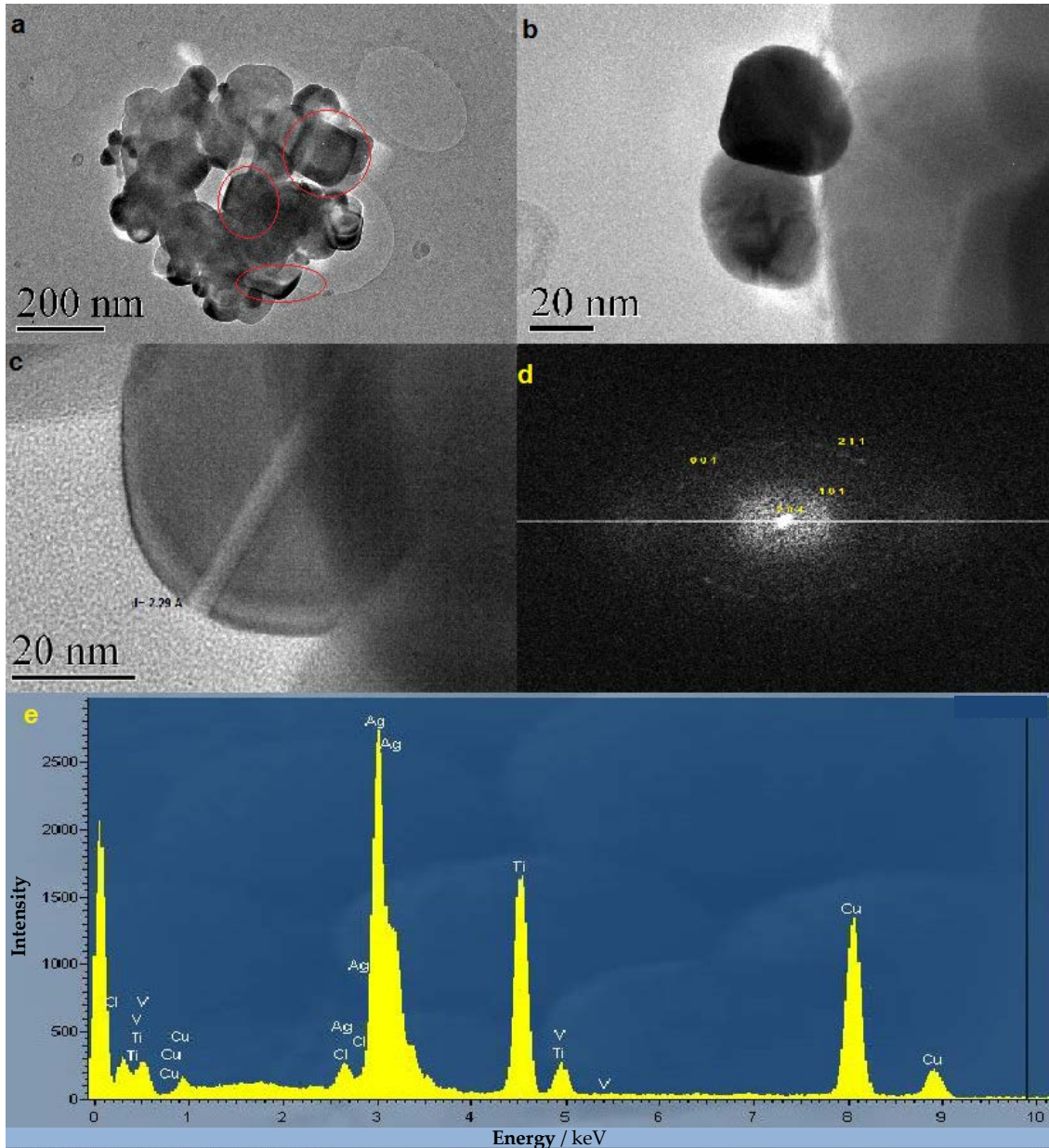


328 **Fig. 2.** EPR spectrum of the $\bullet\text{OH}$ -DMPO adduct detected in 10 mL of a 9.2 mM DMPO
329 solution with 25 mM Na₂SO₄, at pH 5.9 and 25 °C. The electrolysis was made using an
330 Ag(4%)-TiO₂ photoanode illuminated with a 125 W UVA light and a stainless steel
331 spiral cathode, both of 1 cm², at $E_{\text{bias}} = 1.70$ V vs. Ag|AgCl (3 M KCl) for 15 min.

332 3.2. Characterization of the synthesized Ag-TiO₂ photoanodes

333 The morphology of the Ag(4%)-TiO₂ film, the best photoanode synthesized for
334 PEC in the present work, was assessed by HRTEM. Fig. 3a-c highlights the images
335 obtained at increasing magnification, revealing the presence of non-spherical Ag and
336 TiO₂ particles. These micrographs also suggest that the Ag nanoparticles are quite small
337 but with a high degree of crystallinity. Fig. 3a clearly confirms the presence of cuboid-

338 shaped Ag in TiO₂ deposited powder. The P25 nanocrystals showed a size of about 100
339 nm. In Fig. 3b, the color contrast between gray TiO₂ and black Ag particles can be
340 observed, being the average particle size of Ag nanoparticles around 45 nm.



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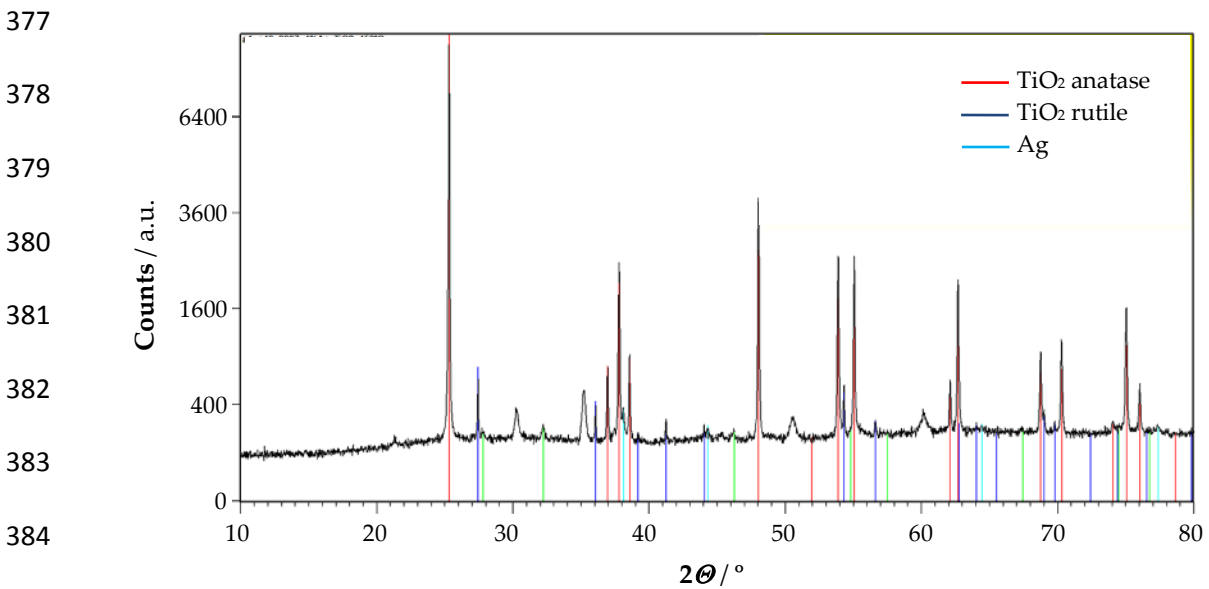
342 **Fig. 3.** HRTEM images of Ag(4%)-decorated TiO₂ powder, synthesized via
343 electrophoresis method, at a magnification of (a) x20,000, (b) x150,000 and (c)
344 x400,000. (d) SAED patterns of nano-TiO₂ powders revealing anatase or rutile phases.
345 (e) EDX analysis of the Ag(4%)-TiO₂ sample shown in (c).

346 Fig. 3c shows the equal-thickness fringes of the anatase phase observed in a square-
347 shape TiO₂ particle, indicating the presence of (101) facets with a lattice spacing of 2.29
348 Å. Selected area electron diffraction (SAED) patterns of nano-TiO₂ powders confirmed
349 the predominance of anatase phase, as shown in Fig. 3d, where the ring pattern indicates
350 the polycrystalline nature of TiO₂ nanoparticles. From the same diffraction pattern, it
351 can be demonstrated that the ring consists of well distinct spots due to crystalline nature
352 of TiO₂, which is an important prerequisite for the improvement of photocatalytic
353 activity. The brightness and intensity of the polymorphic ring are quite weak, which can
354 be related to a rather poor crystallinity with the presence of amorphous material. The
355 observed (204), (004) and (211) planes agree with the standard data (JCPDS 75-1537)
356 of TiO₂ anatase phase, whereas the (101) plane agrees with the standard data (JCPDS
357 88-1175) of TiO₂ rutile phase. The elemental composition of the photoanode was
358 determined by EDX and the corresponding spectrum is shown in Fig. 3e. It evidences
359 the pre-eminent presence of Ag and Ti with their characteristic peaks, along with those
360 related to the Cu mesh and Cl or V impurities.

361 The crystalline planes of Ag and TiO₂ after annealing the electrode at 450 °C were
362 confirmed by XRD analysis. The diffractogram of Fig. 4 shows the peaks obtained for
363 2θ values between 10° and 80°, which can be ascribed to:

- 364 (i) The primary TiO₂ crystalline phase, i.e., anatase, with expected peaks at 25.2°,
365 37.3°, 47.6°, 53.5°, 55.1° and 62.2° for (101), (004), (200), (105), (211) and (204)
366 planes (JCPDS Card No. 21-1272), respectively.
- 367 (ii) Rutile, with peaks at 27.3°, 35.6°, 41.2°, 54.3° and 67.2° corresponding to (110),
368 (101), (111), (211) and (310) planes (JCPDS Card No. 21-1276), respectively.
- 369 (iii) Metallic silver, with peaks at 38.3°, 44.5°, 64.6° and 77.5° related to (111), (200),
370 (220) and (311) planes (JCPDS Card No. 04-0783), respectively.

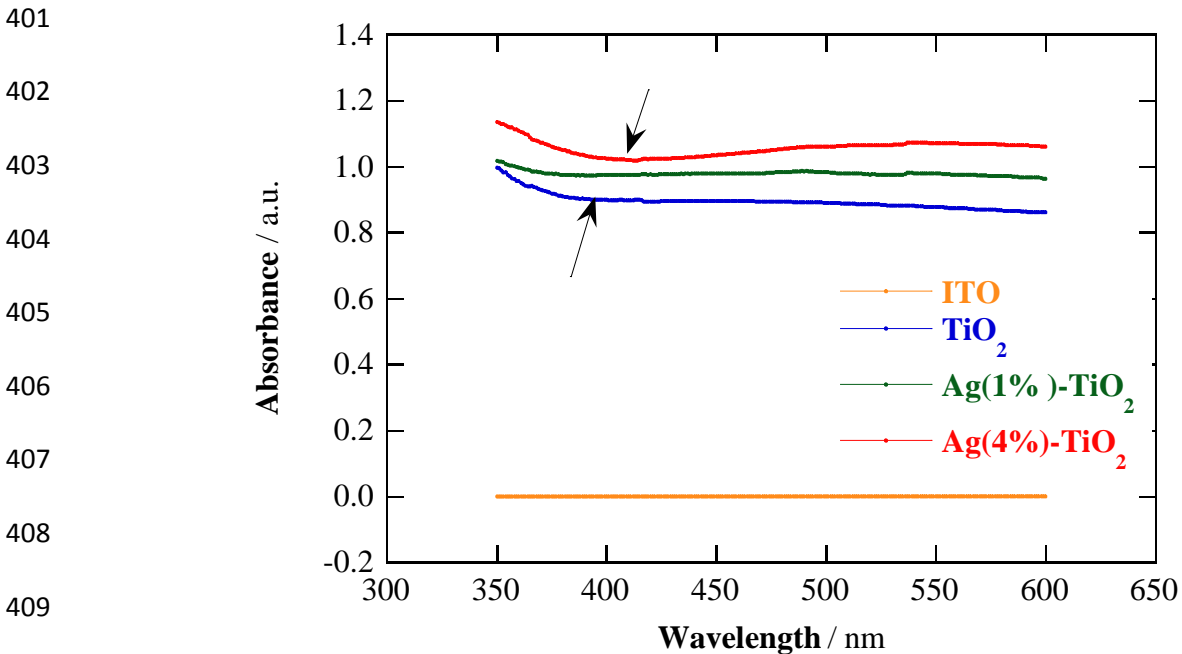
371 Undoped powder presented peaks at the same diffraction angles for both TiO₂
372 phases. Hence, it can be concluded that photodeposition of the silver does not modify
373 the basic crystal structure of TiO₂, which can be related to a uniform decoration of the
374 surface. Furthermore, it is noticeable that no peaks ascribed to AgO were found,
375 meaning that Ag was only detected as pure metal, without oxides formation after
376 annealing at 450 °C.



385 **Fig. 4.** XRD pattern of the ITO/Ag(4%)-TiO₂ photoanode after annealing at 450 °C.

386 In addition, the optical properties of all materials were assessed from their UV/Vis
387 absorption spectra, collected in Fig. 5. As can be seen, the ITO film did not absorb at
388 any wavelength between 350 and 600 nm. In contrast, the TiO₂ deposited onto ITO
389 absorbed the light in all the wavelength range checked, with higher intensity in the UV
390 region up to near $\lambda = 375$ nm, whereupon the absorbance decreased very slowly within
391 the visible region. The high absorption in the near-visible region is ascribed to the
392 electron transition from VB to the CB, with an E_{bg} of 3.0-3.2 eV. The same behavior
393 can be observed for the two synthesized Ag coating TiO₂ deposits, although with larger
394 absorption in the visible region as the Ag content in the photocatalyst was increased,
395 being accompanied by a shift to a higher λ where the visible absorption begins. The

396 higher absorbance in the visible region occurred from near $\lambda = 405$ nm for Ag(4%)-
397 TiO₂, with a maximum shoulder at $\lambda \sim 490$ nm, thereby demonstrating its higher ability
398 to absorb the visible light as compared to the other materials. Therefore, this is due to
399 the enhanced photoexcitation of the electrons from the VB of the TiO₂ to the optimized
400 number of Ag nanoparticles [48,49].



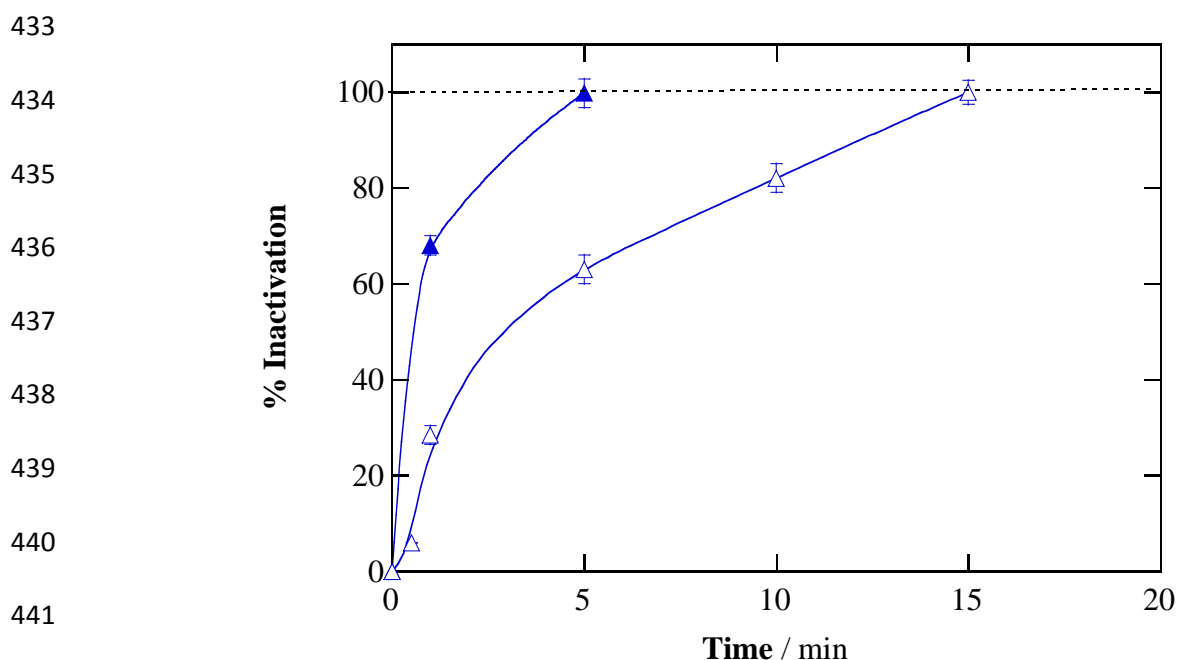
410 **Fig. 5.** UV-Vis absorption spectra of ITO, ITO/TiO₂ and ITO/Ag(4%)-TiO₂ after
411 annealing at 450 °C.

412 3.3. Comparative inactivation of *Bacillus atrophaeus*

413 The results of Fig. 1 demonstrate the effective attack of •OH produced at the
414 Ag(4%)-TiO₂ photoanode onto the outer layer of *P. aeruginosa* bacteria. This layer is
415 composed of peptidoglycans, consisting of a glycan backbone of muramic acid and
416 glucosamine partially cross-linked with peptide chains. To clarify and extend the
417 disinfection power of the PEC treatments tested, the process was also applied to the
418 inactivation of a ubiquitous rod-shaped Gram-positive bacterium like *B. atrophaeus*. In
419 this microorganism, the cell wall is more compact because it contains a densely
420 crosslinked structure of peptidoglycans.

421 Fig. 6 evidences that a suspension with 10^6 CFU mL⁻¹ of *B. atrophaeus* and 25 mM
 422 Na₂SO₄ can be completely disinfected after 15 min of PEC with an Ag(4%)-TiO₂ thin
 423 film photoelectrode at $E_{\text{bias}} = 1.70$ V. This time is much longer than 5 min required for
 424 the total inactivation of *P. aeruginosa* under comparable conditions (see Fig. 1). This
 425 finding agrees with the less compact outer layer of the Gram-negative bacterium, which
 426 can be more easily destroyed by $\bullet\text{OH}$. The trend previously reported by us for the
 427 inactivation of 10^6 CFU mL⁻¹ suspensions of both bacteria in Na₂SO₄ at pH 7.0 by
 428 means of EO with a BDD anode at $j = 33.3$ mA cm⁻² was not so clear, since *B.*
 429 *atrophaeus* was very sensitive to significant pH variations at such high j [12].

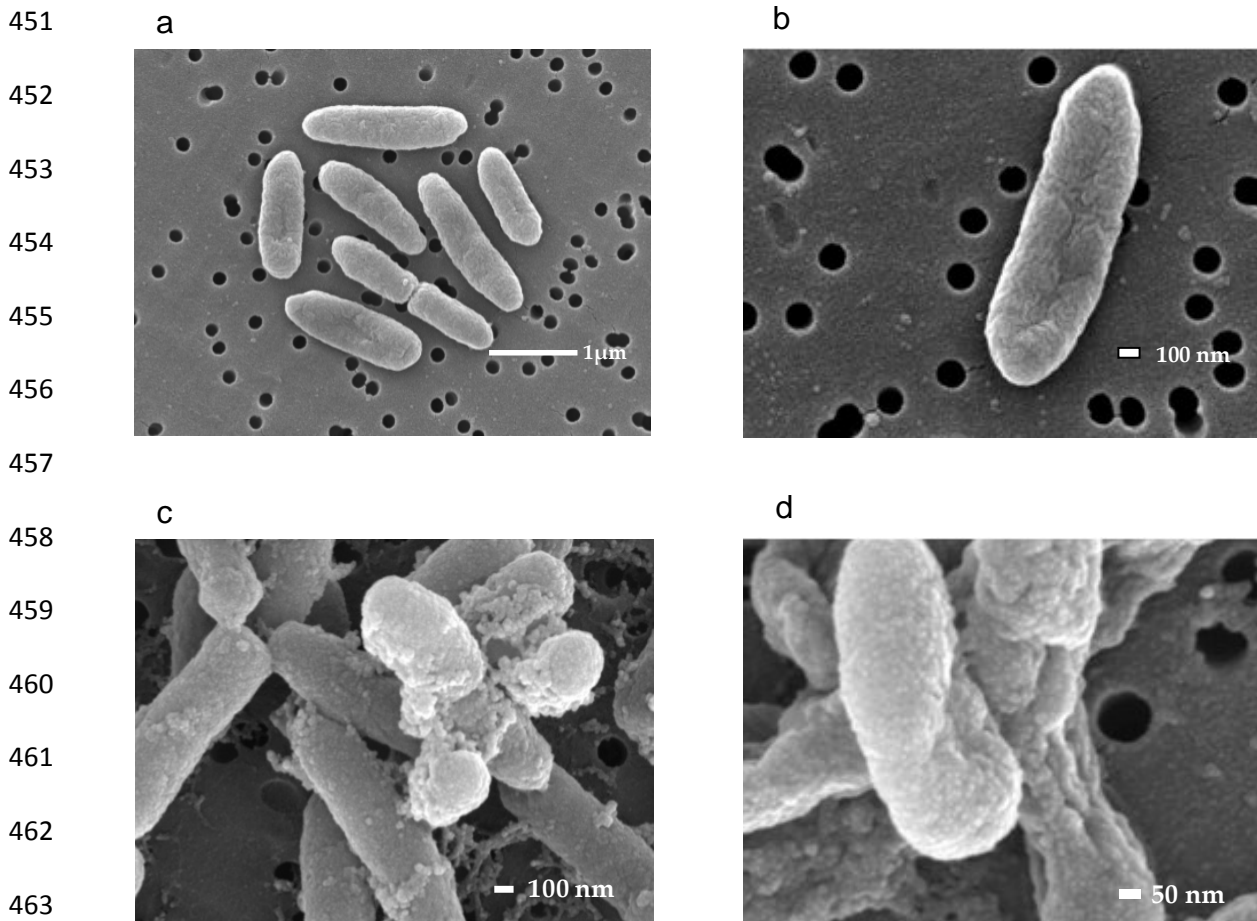
430 The above results allow inferring that the synthesized Ag(4%)-TiO₂ photoanode
 431 can be useful for the fast and total disinfection of water contaminated with both, Gram-
 432 negative and Gram-positive bacteria, by means of PEC.



442 **Fig. 6.** Percentage of inactivation of 10^6 CFU mL⁻¹ of (\blacktriangle) *Pseudomonas aeruginosa*
 443 and (\triangle) *Bacillus atrophaeus* aqueous suspensions vs. electrolysis time for the PEC
 444 treatment of 100 mL with 25 mM Na₂SO₄, at pH 5.9 and 25 °C. Trials were made using
 445 an undivided cell with a 3 cm² Ag(4%)-TiO₂ photoanode illuminated with a 125 W
 446 UVA lamp at $E_{\text{bias}} = 1.70$ V vs. Ag|AgCl (3 M KCl).

447 3.4. Changes in bacterial morphology upon PEC treatment

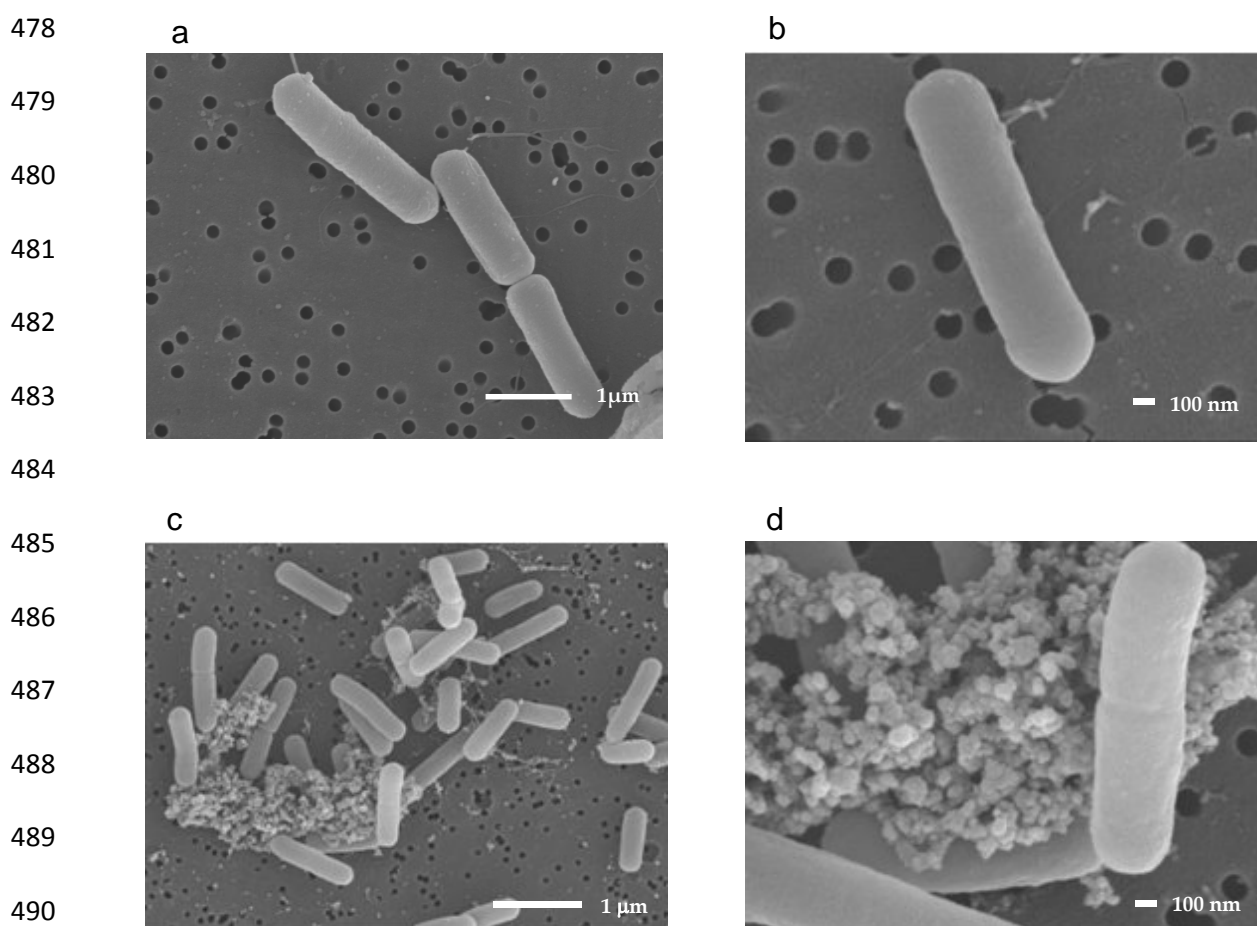
448 Fig. 7a and b and Fig. 8a and b show the morphological features of the two bacteria
449 prior to carrying out the PEC treatments, as obtained by SEM. As expected, a rod-
450 shaped morphology with length of about 1-2 μm can be observed for both cell types.



464 **Fig. 7.** SEM images for *Pseudomonas aeruginosa* supported on a polycarbonate
465 membrane, corresponding to the assays of Fig. 6. (a, b) Initial suspension and (c, d)
466 after 5 min of PEC. Magnification: (a) x20,000, (b, c) x40,000 and (d) x80,000.

467 Worth noting, the micrographs recorded after the total photoelectrocatalytic
468 inactivation, at 5 min for *P. aeruginosa* (Fig. 7c and d) and at 15 min for *B. atrophaeus*
469 (Fig. 8c and d), did not present significant alterations in their overall form and size,
470 although their external surface became much rougher. In addition to such surface
471 alteration, some cells were lysed and, consequently, cellular debris was accumulated in

472 the suspensions. This confirms the oxidation of some molecules of both cell walls by
473 $\bullet\text{OH}$, eventually weakening the structure. In conclusion, the inactivation can be
474 accounted for by two phenomena promoted by the presence of such weaker wall: (i) the
475 molecules within the cytoplasm are more exposed to external conditions, altering the
476 cell metabolism, and (ii) collapse of the wall architecture that ends in the cell lysis with
477 release of cytoplasmic material.



491 **Fig. 8.** SEM images for *Bacillus atrophaeus* supported on a polycarbonate membrane,
492 corresponding to the assays of Fig. 6. (a, b) Initial suspension and (c, d) after 15 min of
493 PEC. Magnification: (a, c) x10,000 and (b, d) x40,000.

494 **4. Conclusions**

495 It has been shown that *P. aeruginosa* suspensions at natural pH 5.9 can be rapidly
496 disinfected by PEC using an Ag-TiO₂ thin film photoanode under UVA illumination.

497 The optimum experimental conditions were found with 4 wt.% Ag, in 25 mM Na₂SO₄
498 as supporting electrolyte and applying an $E_{\text{bias}} = 1.70$ V, which yielded complete
499 inhibition of this Gram-negative bacterium at a very short time of 5 min. The
500 comparative PC treatment required up to 30 min. This suggests the positive bactericidal
501 action of the photoanode under current supply in PEC, minimizing the recombination of
502 the photoinduced electron/hole pairs and upgrading the $\bullet\text{OH}$ production, whose
503 presence was confirmed by EPR. The analysis of the synthesized Ag(4%)-TiO₂ revealed
504 that the photoanode was mainly composed of Ag nanoparticles on crystalline anatase,
505 and it possessed greater absorption in the visible region as compared to the unmodified
506 TiO₂ film. The PEC process was also useful to inactivate the Gram-positive bacterium
507 *B. atrophaeus*, which was more resistant due to its different cell wall formed by a thick
508 outer peptidoglycan layer.

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