

1 **Derisking Development by a Cocrystallization Screen of a Novel Selective Inhaled JAK-STAT**
2 **inhibitor**

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29 **ABSTRACT:**

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31 The discovery and detailed characterization of several new solid forms of a novel selective inhaled JAK-
32 STAT inhibitor are described. Using a holistic cocrystallization screen approach to explore its
33 formulation landscape, we decrease the risk of future potential development failures due to a nonoptimal
34 pharmacokinetic lung profile or undesired lung effects in humans.

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37 1. INTRODUCTION

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39 Compound 1 is a novel and selective Janus kinase-signal transducers and activation of transcription
40 (JAK-STAT) inhibitor. JAKs belong to one intracellular subgroup of the nonreceptor protein tyrosine
41 kinases involved in cell growth, survival, development, and differentiation of a variety of cells, critically
42 important for immune and hematopoietic cells. Type I and II cytokine receptors are constitutively
43 associated with JAKs, and the binding of ligand (cytokine) initiates a transphosphorylation cascade:
44 receptor-JAK-STAT. Phosphorylated STATs dissociate from the receptor, dimerize, translocate to the
45 nucleus, and bind to specific sequences to regulate the expression of target genes.^{1,2}

46 Given the importance of JAKs inhibitors (“jakinibs”) to modulate cytokine signaling, they may be useful
47 for the treatment of various diseases or conditions in which the functions of the innate and/or adaptive
48 immune system are involved.^{3–5} JAK inhibitors are currently being evaluated in a diverse range of
49 disorders (rheumatoid arthritis, psoriasis, inflammatory bowel disease, and myeloproliferative
50 disorders), and many more trials are underway in other autoimmune disorders (juvenile idiopathic
51 arthritis, ankylosing spondylitis, systemic lupus erythematosus, Sjögren’s syndrome), chronic kidney
52 disease and diabetic nephropathy, breast cancer, lymphoma, and the prevention of graft rejection. For a
53 review of the pharmaceutical intervention of the JAK/STAT pathway, see references 6–9.

54 In view of the numerous conditions and disorders susceptible to “jakinibs” treatment, it is expected that
55 the new compounds, new forms of an existing compound, and new routes of administration for these
56 compounds provide significant therapeutic benefits to a variety of patients.

57 When developing poorly soluble drugs for oral inhaled delivery, special caution needs to be paid to the
58 design of the inhalation product. For inhaled compounds, the mean absorption time from the lung is not
59 usually correlated to a single physicochemical property,¹⁰ but there is some evidence that the absorption
60 from the lung of a poorly soluble neutral inhaled compound into the central circulation may be
61 correlated to dissolution.¹¹ Controlling the dissolution rate is therefore of utmost importance to achieve
62 an optimal lung pharmacokinetic profile. When a poorly soluble inhaled compound dissolves too fast,
63 the desired effect can be too short, assuming neither permeability nor transporter cell uptake is limiting
64 the absorption. On the other hand, if it dissolves too slowly, the compound might accumulate in the lung
65 and be a reason for unexpected adverse effects. The rate and extent of the absorption from the lung (and
66 the safety profile) of a poorly soluble inhaled compound will depend (excluding physiological
67 differences or disease-related changes) on many factors, mainly the physicochemical properties of the
68 delivered drug and its material properties (solubility, dissolution rate, size, shape, charge, crystallinity,
69 and chemical composition). It is generally accepted that any undissolved particulate material in the lungs
70 can result in adaptive adverse/ tox effects.¹² It is therefore advisable to explore the formulation
71 landscape as early as possible in the discovery phase so several forms (salts, cocrystals, solvates,

72 polymorphs...), with different physicochemical properties, are available to the team for in vivo
73 assessment and in this way decrease the risk of costly surprises during the following development phase.

74 In this paper, we describe the discovery and detailed characterization of several new forms of 1 using a
75 holistic approach to explore the formulation landscape of 1 to decrease the risk of potential development
76 failures due to a nonoptimal pharmacokinetic lung profile or undesired lung effects in humans. To do so,
77 we selected a list of 20 coformers from a database containing more than 2300 compounds, including 860
78 products regarded as “safe” by the FDA (GRAS list). The selection was performed according to the
79 virtual prediction results in combination with a factor obtained from a multiparameter assessment, which
80 included melting point of the coformers, safety and tox profile of the coformers, and solubility in water
81 stipulation. We gave an important relative weight to the safety/tox profile for each coformer, since the
82 new forms discovered were intended for human use. The safety/tox profile was performed as a
83 combination of an in house AZ in silico assessment, experimental safety end points from multiple
84 databases, and different structural alerts. The resulting safety/tox profile contained information about
85 hERG activity, phospholipidosis, AhR, Nav1.5, CaV1.2, potential to form reactive metabolites, genotox,
86 AMES, carcinogenicity, mouse lymphoma, chromosomal aberration and micro nucleus, among others.
87 The solubility stipulation was assigned a high contribution factor to the multiparameter assessment since
88 it had been shown previously that the solubilities of compounds formulated as cocrystals increase in
89 proportion to the solubility of the coformer.^{13,14} The experimental cocrystal screen was then performed
90 on 20 coformers with a good chance of forming a cocrystal with different, and hopefully better,
91 dissolution rate profiles (compared to the free base 1), which are stable and safe for human oral inhaled
92 dosing.

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94 **2. MATERIALS AND METHODS**

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96 **2.1. Experimental Screen.** A comprehensive cocrystal screening has been conducted by using different
97 combinations of solvents at several concentrations and temperatures, with variable cooling rates, in both
98 thermodynamic and kinetic conditions. Solubility of 1 was initially determined in 36 solvents, and
99 accordingly drop grinding, reaction crystallization, and solvent mediated transformation techniques were
100 applied to each 1/coformer combination. All solids were analyzed by powder X-ray diffraction (PXRD)
101 to assess the formation of a new solid form.

102

103 **2.2. Virtual Cocrystal Screening.** For each compound, the molecule was drawn in an extended
104 conformation and energy minimized using the molecular mechanics methods implemented in
105 TorchLite.¹⁵ Gaussian 09 was used to optimize the geometry and calculate the MEPS on the 0.002 Bohr
106 Å⁻³ electron density isosurface using density functional theory (DFT) and a B3LYP/6-31G* basis
107 set.¹⁶ The MEPS was converted into SSIPs using in-house software.¹⁷

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109 **2.3. Powder X-ray Diffraction (PXRD).** Powder X-ray diffraction patterns were obtained on a
110 PANalytical X'Pert PRO MPD diffractometer in transmission configuration using Cu K α 1+2 radiation
111 ($\lambda = 1.5418 \text{ \AA}$) with a focalizing elliptic mirror, a PIXcel detector working at a maximum detector's
112 active length of 3.347°. Flat geometry has been used for routine samples sandwiched between
113 lowabsorbing films (polyester of 3.6 μm of thickness) measuring $2\theta/\theta$ scans from 2 to 40° in 2θ with a
114 step size of 0.026° and a measuring time of 80–300 s per step. The indexation of the PXRD diagrams
115 was carried out by means of Dicvol04.¹⁸ The unit cell parameters were refined by Le Bail fit¹⁹ using
116 the Fullprof program,²⁰ and the most probable space groups were determined from the systematic
117 absences.

118

119 **2.4. Differential Scanning Calorimetry (DSC).** Differential scanning calorimetry analyses were
120 carried out by means of a Mettler- Toledo DSC-822e calorimeter. Experimental conditions: aluminium
121 crucibles of 40 μL volume, atmosphere of dry nitrogen with 50 mL/min flow rate, heating rate of 10
122 °C/min. The calorimeter was calibrated with indium of 99.99% purity. (m.p.: 156.8 °C ΔH : 28.68 J/g).

123

124 **2.5. Thermogravimetric Analysis (TGA).** Thermogravimetric analyses were performed on a Mettler-
125 Toledo TGA-851e thermobalance. Experimental conditions: alumina crucibles of 70 μL volume,
126 atmosphere of dry nitrogen with 50 mL/min flow rate, heating rate of 10 °C/min.

127 **2.6. X-ray Crystallographic Analysis.** The single crystal structures were collected using a D8 Venture
128 system equipped with a multilayer monochromator and a Mo or Cu microfocus ($\lambda = 0.71073 \text{ \AA}$ or $\lambda =$
129 1.54178 \AA) has been used too. Frames were integrated with the Bruker SAINT software package using a
130 SAINT algorithm. Data were corrected for absorption effects using the multiscan method (SADABS).²¹
131 The structures were solved and refined using the Bruker SHELXTL Software Package, a computer
132 program for automatic solution of crystal structure and refined by fullmatrix least-squares method with
133 ShelXle Version 4.8.0, a Qt graphical user interface for the SHELXL computer program.²²

134

135 3. RESULTS AND DISCUSSION

136

137 **3.1. Virtual Cocrystal Screen.** The selection of the coformers used in the cocrystal screen was based on
138 the computational cocrystal screen method developed by Prof. Hunter, which has been validated using
139 experimental cocrystal data extracted from the literature.^{23,24} This computational method has been
140 recently applied to several active pharmaceutical ingredients (APIs) with a remarkable success by
141 Hunter's group, including nalidixic acid,²⁵ griseofulvin and spironolactone,²⁶ and some of us have
142 recently used it to guide the discovery of new cocrystals of zafirlukast^{27,28} and sildenafil.²⁹ The in
143 silico method is based on the calculation of a cocrystal pairing energy between the API and the
144 coformer. This calculation is performed by using the surface site interaction points (SSIPs), which can
145 be extracted from molecular electrostatic potential surfaces computed at the DFT level of computation
146 as described in reference 17 or estimated with a faster method based on the molecular electrostatic
147 potential surfaces (MEPS) calculated from MMFF94 atomic partial charges.^{30,31} We have followed the
148 methodology at the DFT level of computation with compound 1, and 20 coformers have been chosen
149 among the 100 coformers with the highest probability of cocrystallization from a database containing
150 more than 2400 compounds (including 860 products from the GRAS list) according to the virtual
151 prediction ranking. Table 2 shows the 20 coformers along with their corresponding ΔE values.

152 The so-called "rule of 3" is frequently used to predict the outcome of a salt or a cocrystal. The rule is
153 based on the calculation of the difference of pKa between the protonated base and the acid. When this
154 value is less than 0, a cocrystal is expected, and when it is greater than 3 a molecular salt is the expected
155 form. However, with intermediate values predictions are less reliable.^{32,33} Recently, a linear
156 relationship between the ΔpK_a value and the probability of salt/cocrystal formation has been derived by
157 Cruz-Cabeza from more than 6000 component systems (eq 1).³⁴ This equation allows a statistical
158 prediction of proton transfer (P, %) around the "salt-cocrystal continuum" region of $\Delta pK_a \approx 1$, which
159 lies in a range of values between -1 and 4. We have applied this simple calculation to the coformers
160 chosen for the screening in order to assess the probability of proton transfer, and values can be found in
161 Table 2.

$$162 \quad P(\%) = 17 \Delta pK_a + 28 \text{ for } -1 \leq \Delta pK_a \leq 4 \quad (1)$$

163 Given that 1 contains a pyrimidine and a benzoxazolinone group (measured pKa values of 5.9 and 8.9
164 respectively) and most of the coformers are carboxylic acids and strong organic bases, according to the
165 pKa rule, both salts and cocrystals were expected to be obtained, although with a higher probability of
166 salt formation. However, since both types of multicomponent solid forms could improve physicochemical
167 properties, no coformers were filtered according to proton transfer probability in order to increase
168 diversity of crystal forms.

169 **3.2. Experimental Salt/Cocrystal Screen.** A total number of 130 experiments using selected
170 combinations between 36 solvents and 20 cofomers have been conducted, distributed mainly in two
171 methodologies (drop grinding and reaction crystallization techniques) to test the formation of cocrystals
172 with compound 1. Our high-throughput methodology consists of the initial and qualitative solubility
173 assessment of compound 1 and each of the 20 cofomers in 36 solvents.³⁶ Then, four solvents were
174 selected according to the solubility information, which is a key issue for a rational design of the
175 screening conditions and that allows the optimized exploration of the cocrystallization landscape for
176 each 1/coformer combination with the highest probability of success and the lowest number of
177 experiments. Evidence of cocrystallization is detected by measuring PXRD diffractograms and DSC
178 thermograms for each solid obtained during the screen. When it has not been possible to solve the
179 crystal structures, ¹H NMR has been used to determine API/coformer stoichiometry, and
180 diffractograms have been indexed when possible to confirm the crystal form purity.

181

182 **3.3. New Solid Forms of Compound 1.** During the solubility determination of compound 1 in 36
183 solvents in a range of 30–90 °C, new solvate forms of compound 1 have been discovered and
184 characterized with formic acid (Form A), DMF (Form B), DMSO (Form C), and acetic acid (Forms D-1
185 to D-4). Compound 1 is soluble at 25 °C in formic acid, DMF, DMSO, and dimethylamine (40% in
186 water). At 40 °C, it is soluble in acetic acid. It is insoluble in methanol, ethanol, isopropanol, butanol,
187 ethylene glycol, ACN, MEK, acetone, MiBK, water, pentane, heptane, cyclohexane, toluene, xylene,
188 AcOEt, diethyl ether, THF, dimethyl ethylene glycol, diisopropyl ether, dioxane, 1,2-dichloroethane,
189 chloroform, benzylalcohol, diethylamine, triethylamine, NH₃ (2 M in MeOH), dimethylamine (2 M in
190 MeOH), mixture of MeOH/ DCM (10:90) and trifluoroethanol. The solutions obtained °C until
191 crystallization of a solid. The new forms have been isolated and characterized by means of DSC, ¹H
192 NMR, PXRD, and TGA in some cases. All of them show a 1:1 stoichiometry except the system formed
193 by compound 1 and acetic acid, which is a multicomponent solid forms set composed of four different
194 modifications showing polymorphism and different stoichiometries (1:1, 1:2, and 1:4). A complete
195 characterization of each new form is included in the Supporting Information.

196 All new solvate forms were heated up to a temperature in which the desolvation was ensured under
197 nitrogen atmosphere, then cooled down to room temperature and measured by PXRD, and in all cases
198 the same anhydrous crystal form of compound 1 was obtained. These results together with the fact that
199 no new anhydrous forms of compound 1 were discovered during the solid forms screening suggested
200 initially that compound 1 does not present polymorphism. However, one of the acetic acid solvates
201 (Form D-1) shows a DSC thermogram with a melting point, once the acetic acid is removed on heating,
202 20 °C lower than the previously observed for compound 1 (Figure 5), which suggests that another

203 metastable polymorph of compound 1 can exist although with a rapid conversion to the stable one since
204 it has not been isolated so far.

205

206 **3.4. New Salts/Cocrystals of Compound 1.** New multicomponent forms of compound 1 have been
207 identified after the cocrystal screening with 8 out of the 20 cofomers tested: 1,4,8,11-
208 tetrazacyclotetradecane, 3,5-dinitrobenzoic acid, gallic acid, orotic acid, 5-nitroisophthalic acid, 3,5-
209 dihydroxybenzoic acid, 1-methyl-2-pyrrolidone, and 4-nitrobenzenesulfonic acid. Some of these new
210 phases were isolated as different polymorphs and solvates. In those cases where crystal structure has not
211 been solved, the definition of the form as a salt or a cocrystal has been done based on the probability of
212 proton transfer determined with eq 1.

213 • 1:1,4,8,11-Tetrazacyclotetradecane salt (Form I). It has been obtained by reaction crystallization. A
214 2:1 stoichiometry has been deduced according to ¹H NMR and single X-ray diffraction.

215 • 1:3,5-Dinitrobenzoic acid salt isopropanol solvate (Form II-A): it has been obtained by reaction
216 crystallization in IPA. A 1:1:2 stoichiometry has been deduced according to single X-ray diffraction.

217 • 1:3,5-Dinitrobenzoic acid salt (Form II-B): it has been obtained by reaction crystallization in acetone.
218 A 1:1 stoichiometry has been deduced according to ¹H NMR.

219 • 1:3,5-Dinitrobenzoic acid salt (Form II-C): it has been obtained by reaction crystallization in THF. A
220 1:1 or 1:1.5 stoichiometry can be deduced according to ¹H NMR.

221 • 1:3,5-Dinitrobenzoic acid salt dioxane solvate (Form IID): it has been obtained by reaction
222 crystallization in dioxane. A 1:1:1 stoichiometry has been deduced according to ¹H NMR.

223 • 1:3,5-Dinitrobenzoic acid salt (Form II-E): it has been obtained by slurry in water. A 1:1 stoichiometry
224 has been deduced according to ¹H NMR.

225 • 1:Gallic acid salt THF solvate (Form III): It has been obtained by reaction crystallization in THF. A
226 1:1:1 stoichiometry has been deduced according to ¹H NMR. The XRD pattern shows broad
227 diffraction peaks (Figure 6). Further attempts to obtain higher crystallinity solids were unsuccessful.

228 • 1:Orotic acid salt (Form IV-A): it has been obtained by slurry in IPA (4 days), acetone (4 days), THF
229 (4 days) or dioxane (7 days). A 1:1 stoichiometry has been deduced according to ¹H NMR.

230 • 1:Orotic acid salt (Form IV-B): it has been obtained not pure, as a mixture with Form IV-A, by slurry
231 in dioxane (4 days).

- 232 • 1:Orotic acid salt (Form IV-C): it has been obtained not pure, as a mixture with Form IV-A, by slurry
233 in water (4 days).
- 234 • 1:5-Nitroisophthalic acid salt (Form V-A): it has been obtained by reaction crystallization in IPA (4
235 days). A 1:1 stoichiometry has been deduced according to ¹H-RMN.
- 236 • 1:5-Nitroisophthalic acid salt (Form V-B): it has been obtained by reaction crystallization in acetone
237 (4 days). A 1:1 stoichiometry has been deduced according to ¹H-RMN.
- 238 • 1:5-Nitroisophthalic acid salt (Form V-C): it has been obtained by heating up to 215 °C form V-B in a
239 TGA crucible. A 1:1 stoichiometry has been deduced according to ¹H-RMN.
- 240 • 1:5-Nitroisophthalic acid salt dioxane solvate (Form VD): it has been obtained by reaction
241 crystallization in dioxane (4 days).
- 242 • 1:5-Nitroisophthalic acid salt (Form V-E): it has been obtained by heating up to 230 °C form V-D in a
243 TGA instrument. A 1:1 stoichiometry has been deduced according to ¹H-RMN.
- 244 • 1:3,5-Dihydroxybenzoic acid salt (Form VI): It has been obtained by reaction crystallization in water.
245 A 1:1 stoichiometry has been deduced according to ¹H NMR.
- 246 • 1:1-Methyl-2-pyrrolidone cocrystal (Form VII): It has been obtained by slurry in THF or heptane. A
247 1:1 stoichiometry has been deduced according to ¹H NMR.
- 248 • 1:4-Nitrobenzenesulfonic acid salt (Form VIII): It has been obtained by reaction crystallization in
249 ipOH, acetone, THF, dioxane, or water. A 1:1 stoichiometry has been deduced according to ¹H NMR
- 250 A comparison of the PXRD of the new multicomponent forms of compound 1 obtained as a single form
251 is shown in Figure 6.
- 252 A comparison of the PXRD of the different forms abovementioned for 1:3,5-dinitrobenzoic acid is
253 shown in Figure 7.
- 254 A comparison of the PXRD of the different forms abovementioned for 1:orotic acid is shown in Figure
255 8.
- 256 A comparison of the PXRD of the different forms abovementioned for 1:5-nitroisophthalic acid is
257 shown in Figure 9.
- 258 When possible, the PXRD diagrams of the new forms were indexed, and the results are shown in Table
259 3. The rest of the forms could not be obtained in pure form, which hindered the indexing process.

260 **3.5. Single Crystal Structures. Crystals** of forms I, II-A, DMF solvate, and acetic acid hybrid salt-
261 cocrystal suitable for SCXRD analysis have been obtained, and their crystallographic data are
262 summarized in Table 4.

263 3.5.1. Form I. 1:1,4,8,11-Tetrazacyclotetradecane (cyclam) salt crystallizes with one molecule of 1 and
264 half molecule of 1,4,8,11-tetrazacyclotetradecane in the asymmetric unit. The ΔpK_a value is 2.0 (pK_a of
265 the cofomer is 10.9) with a probability of salt formation of $P = 62\%$, and the SCXRD data confirm
266 (from a difference synthesis and refined with an isotropic temperature factor) the location of two
267 hydrogens bonded to the cofomer's nitrogen. In the structure, every molecule of 1 anion interacts with a
268 molecule of tetrazacyclotetradecane bis cation and another molecule of 1 via the oxazolidinone ring
269 through charged assisted hydrogen bonds and electrostatic interactions. Moreover,
270 tetrazacyclotetradecane molecules are sandwiched between molecules of 1 using two out of the four
271 amine nitrogens to interact with the 1 oxazolidinone nitrogens.

272

273 3.5.2. Acetic Acid Hybrid Salt-Cocrystal. This multicomponent solid form crystallizes with one
274 molecule of 1 and four molecules of acetic acid in the asymmetric unit. The solid form is a hybrid salt-
275 cocrystal^{37,38} since one molecule of acetic acid has transferred the acidic proton to the pyrimidine
276 nitrogen establishing a charge assisted hydrogen bond and at the same time neutral acetic acid molecules
277 are also present in the crystal structure. Since the ΔpK_a value is 1.1 (pK_a of the acetic acid is 4.8), the
278 probability of salt formation is $P = 47\%$, and thus this structure could be considered as an example of the
279 "salt-cocrystal continuum". Cocrystals and salts formed between carboxylic acids and N-heterocycles
280 have been analyzed in the literature, and it has been suggested that the formation of unexpected hybrid
281 salt-cocrystals can be produced because carboxylate moieties are not totally satisfied by a single
282 hydrogen-bond donor, which makes necessary the presence of neutral carboxylic acids in the crystal
283 structure.³⁹ In the crystal structure, both C–O distances in the acetate molecule are practically the same
284 (1.263(3) Å and 1.263(2) Å), and the transferred proton well located (and refined with an isotropic
285 temperature factor) on the pyrimidine nitrogen, discarding a potential disorder. Three other acetic acid
286 molecules satisfy the two amide hydrogen bond donors and the CO acceptor groups of 1, which could be
287 anticipated by the position and magnitude of the SSIPs of the isolated compound 1 molecule (Figure
288 11).

289

290 3.5.3. DMF Solvate. The DMF solvate crystallizes with one molecule of 1 and one of DMF in the
291 asymmetric unit. In the structure, molecules of 1 interact in a zigzag arrangement with intermolecular
292 contacts between the oxazolidinone amide and the aminopyrimidine groups. In principle, two
293 configurations are possible: amide/amide plus aminopyrimidine/aminopyrimidine or mixed
294 amide/aminopyrimidine interactions. Interestingly, the interaction energy of both configurations are

295 very similar when estimated both by pairing H-bond parameters calculated from MMFF94 atomic
296 partial charges ($E = \sum_{ij} \epsilon_i \epsilon_j = 36.7$ and 37.2 kJ/mol respectively) and from DFT MEPs ($E = \sum_{ij} \epsilon_i \epsilon_j =$
297 40.5 and 42.7 kJ/mol respectively), the observed configuration being the one with the predicted highest
298 interaction energy. DMF molecules complete the sphere of coordination of 1 by establishing H-bond
299 interactions with the second best donor of 1, Figure 12.

300

301 3.5.4. Form II-A. Form II-A crystallizes with one molecule of 1, one of 3,5-dinitrobenzoic acid, and two
302 of isopropanol in the asymmetric unit. Again the crystal form corresponds to a salt in which the
303 pyrimidine ring is protonated and interacts strongly with the carboxylate moiety of the cofomer. In the
304 structure only one of the strong SSIPs of 1 (the oxazolidinone NH) is not involved in any relevant
305 intermolecular interaction.

306

307 **3.6. Polymorphism in the Multicomponent Crystals of 1.** Although it has been traditionally suggested
308 that polymorphism in multicomponent crystals is a phenomenon observed with less frequency than in
309 single component crystals,⁴⁰ it has been put in doubt,⁴¹ and recently some of us discovered new
310 cocrystals of agomelatine with polymorphism increasing the list of compounds showing cocrystal
311 polymorphism.⁴² The present case study shows polymorphism in at least two of the new salts of 1 with
312 3,5-dinitrobenzoic acid and 5-nitroisophthalic acid, which we believe can contribute to new data to
313 enrich the debate about whether multicomponent crystals are less prone to exhibit polymorphism than
314 single component crystals. In particular, solvates of the 1:3,5- dinitrobenzoic salt II-A and II-D show a
315 DSC thermogram with recrystallization after desolvation of an anhydrous form with a different melting
316 point than anhydrous forms II-B, II-C, and II-E, demonstrating that this salt exists in at least four
317 different polymorphs and two solvates (Figure 14). A similar behavior is observed in solvated salts with
318 5- nitroisophthalic acid in which forms V-B and V-D are desolvated upon heating in a DSC experiment
319 exhibiting a recrystallization exothermic process which produced two different solid forms according to
320 the melting points.

321

322 **4. CONCLUSIONS**

323 By using a holistic cocrystallization screen approach, we have explored the formulation landscape of the
324 first inhaled JAKSTAT inhibitor 1 and have generated multiple solid forms covering a broad
325 physicochemical space and therefore decreased the risk of future potential development failures due to a
326 nonoptimal pharmacokinetic lung profile or undesired lung effects in humans. This comprehensive
327 cocrystal/salts screening was conducted using different combinations of solvents at several
328 concentrations and temperatures, with variable cooling rates, in both thermodynamic and kinetic
329 conditions. Solubility of 1 was initially determined in 30 solvents, and accordingly drop grinding,
330 reaction crystallization, and slurry techniques were applied to each 1/conformer combination. Despite 1
331 not showing polymorphism, eight new forms of 1 (and multiple solvates) were identified: 1,4,8,11-
332 tetrazacyclotetradecane, 3,5-dinitrobenzoic acid, gallic acid, orotic acid, 5-nitroisophthalic acid, 3,5-
333 dihydroxybenzoic acid, 1-methyl-2-pyrrolidone, and 4-nitrobenzenesulfonic acid. Many of these new
334 phases were isolated as different polymorphs and solvates. All solids were analyzed by PXRD to assess
335 the formation of a new solid form. After a careful comparison and risk assessment of the in vivo
336 pharmacokinetics, lung deposition, clearance, pulmonary response, effect and safety profile, “the best”
337 one will be progressed as the candidate drug into human trials. We are now assessing their in vivo
338 potential where the most promising ones will be scaled up and brought forward to the next phase, and
339 this will be the subject of a future publication.

340

341 **References**

- 342 Leonard, W. J.; Lin, J.-X. Cytokine receptor signaling pathways. *J. Allergy Clin. Immunol.* 2000, 105,
343 877–888.
- 344 Rawlings, J. S.; Rosler, K. M.; Harrison, D. A. The JAK/STAT signaling pathway. *J. Cell Sci.* 2004,
345 117, 1281–1283.
- 346 Kudlacz, E.; Perry, B.; Sawyer, P.; Conklyn, M.; McCurdy, S.; Brissette, W.; Flanagan, M.; Changelian,
347 P. The novel JAK-3 inhibitor CP-690550 is a potent immunosuppressive agent in various murine
348 models. *Am. J. Transplant.* 2004, 4, 51–57.
- 349 Changelian, P. S.; Flanagan, M. E.; Ball, D. J.; Kent, C. R.; Magnuson, K. S.; Martin, W. H.; Rizzuti, B.
350 J.; Sawyer, P. S.; Perry, B. D.; Brissette, W. H.; McCurdy, S. P.; Kudlacz, E. M.; Conklyn, M. J.;
351 Elliott, E. A.; Koslov, E. R.; Fisher, M. B.; Strelevitz, T. J.; Yoon, K.; Whipple, D. A.; Sun, J.;
352 Munchhof, M. J.; Doty, J. L.; Casavant, J. M.; Blumenkopf, T. A.; Hines, M.; Brown, M. F.;
353 Lillie, B. M.; Subramanyam, C.; Chang, S.-P.; Milici, A. J.; Beckius, G. E.; Moyer, J. D.; Su, C.;
354 Woodworth, T. G.; Gaweco, A. S.; Beals, C. R.; Littman, B. H.; Fisher, D. A.; Smith, J. F.;
355 Zagouras, P.; Magna, H. A.; Saltarelli, M. J.; Johnson, K. S.; Nelms, L. F.; Des Etages, S. G.;
356 Hayes, L. S.; Kawabata, T. T.; Finco-Kent, D.; Baker, D. L.; Larson, M.; Si, M.-S.; Paniagua, R.;
357 Higgins, J.; Holm, B.; Reitz, B.; Zhou, Y.-J.; Morris, R. E.; O’Shea, J. J.; Borie, D. C. Prevention
358 of Organ Allograft Rejection by a Specific Janus Kinase 3 Inhibitor. *Science (Washington, DC,*
359 *U. S.)* 2003, 302, 875–878.
- 360 Harrison, D. A. The JAK/STAT pathway. *Cold Spring Harbor Perspect. Biol.* 2012, 4 (3), a011205.
- 361 Frank, D. A. STAT signaling in the pathogenesis and treatment of cancer. *Mol. Med. (N. Y.)* 1999, 5,
362 432–456.
- 363 Seidel, H. M.; Lamb, P.; Rosen, J. Pharmaceutical intervention in the JAK/STAT signaling pathway.
364 *Oncogene* 2000, 19, 2645–2656.
- 365 Clark, J. D.; Flanagan, M. E.; Telliez, J.-B. Discovery and Development of Janus Kinase (JAK)
366 Inhibitors for Inflammatory Diseases. *J. Med. Chem.* 2014, 57, 5023–5038.
- 367 Vijayakrishnan, L.; Venkataramanan, R.; Gulati, P. Treating inflammation with the Janus Kinase
368 inhibitor CP-690,550. *Trends Pharmacol. Sci.* 2011, 32, 25–34.
- 369 Baeckman, P.; Tehler, U.; Olsson, B. Predicting Exposure After Oral Inhalation of the Selective
370 Glucocorticoid Receptor Modulator, AZD5423, Based on Dose, Deposition Pattern, and

371 Mechanistic Modeling of Pulmonary Disposition. *J. Aerosol Med. Pulm. Drug Delivery* 2017, 30,
372 108–117.

373 Forbes, B.; Backman, P.; Christopher, D.; Dolovich, M.; Li, B. V.; Morgan, B. In Vitro Testing for
374 Orally Inhaled Products: Developments in Science-Based Regulatory Approaches. *AAPS J.* 2015,
375 17, 837–52.

376 Jones, R. M.; Neef, N. Interpretation and prediction of inhaled drug particle accumulation in the lung
377 and its associated toxicity. *Xenobiotica* 2012, 42, 86–93.

378 Hunter, C. A.; Prohens, R. Solid form and solubility. *CrystEngComm* 2017, 19, 23–26.

379 Good, D. J.; Rodriguez-Hornedo, N. Solubility Advantage of Pharmaceutical Cocrystals. *Cryst. Growth*
380 *Des.* 2009, 9, 2252–2264.

381 Cresset torchV10lite, <http://www.cresset-group.com/products/torch/torchlite/>.

382 Frisch, M. J.; Trucks, G. W.; Schlegel, H. B.; Scuseria, G. E.; Robb, M. A.; Cheeseman, J. R.; Scalmani,
383 G.; Barone, V.; Mennucci, B.; Petersson, G. A.; Nakatsuji, H.; Caricato, M.; Li, X.; Hratchian, H.
384 P.; Izmaylov, A. F.; Bloino, J.; Zheng, G.; Sonnenberg, J. L.; Hada, M.; Ehara, M.; Toyota, K.;
385 Fukuda, R.; Hasegawa, J.; Ishida, M.; Nakajima, T.; Honda, Y.; Kitao, O.; Nakai, H.; Vreven, T.;
386 Montgomery, J. A., Jr.; Peralta, J. E.; Ogliaro, F.; Bearpark, M.; Heyd, J. J.; Brothers, E.; Kudin,
387 K. N.; Staroverov, V. N.; Kobayashi, R.; Normand, J.; Raghavachari, K.; Rendell, A.; Burant, J.
388 C.; Iyengar, S. S.; Tomasi, J.; Cossi, M.; Rega, N.; Millam, J. M.; Klene, M.; Knox, J. E.; Cross,
389 J. B.; Bakken, V.; Adamo, C.; Jaramillo, J.; Gomperts, R.; Stratmann, R. E.; Yazyev, O.; Austin,
390 A. J.; Cammi, R.; Pomelli, C.; Ochterski, J. W.; Martin, R. L.; Morokuma, K.; Zakrzewski, V. G.;
391 Voth, G. A.; Salvador, P.; Dannenberg, J. J.; Dapprich, S.; Daniels, A. D.; Farkas, O.; Foresman,
392 J. B.; Ortiz, J. V.; Cioslowski, J.; Fox, D. J. *Gaussian 09*; Gaussian, Inc.: Wallingford, CT, 2009.

393 Calero, C. S.; Farwer, J.; Gardiner, E. J.; Hunter, C. A.; Mackey, M.; Scuderi, S.; Thompson, S.; Vinter,
394 J. G. Footprinting molecular electrostatic potential surfaces for calculation of solvation energies.
395 *Phys. Chem. Chem. Phys.* 2013, 15, 18262–18273.

396 Boultif, A.; Louer, D. Powder pattern indexing with the dichotomy method. *J. Appl. Crystallogr.* 2004,
397 37, 724–731.

398 Le Bail, A.; Duroy, H.; Fourquet, J. L. The ab-initio structure determination of lithium antimony
399 tungstate (LiSbWO₆) by x-ray powder diffraction. *Mater. Res. Bull.* 1988, 23, 447–52.

400 Rodriguez-Carvajal, J. Recent advances in magnetic structure determination by neutron powder
401 diffraction. *Phys. B (Amsterdam, Neth.)* 1993, 192, 55–69.

402 SADABS; Bruker AXS: Madison, Wisconsin, USA, 2004; SAINT, Software Users Guide, Version 6.0;
403 Bruker Analytical X-ray Systems: Madison, WI, 1999. Sheldrick, G. M. SADABS v2.03: Area-
404 Detector Absorption Correction: University of Göttingen: Göttingen, Germany, 1999; Saint
405 Version 7.60A (Bruker AXS 2008); SADABS V. 2008-1, 2008.

406 Sheldrick, G. M. A short history of SHELX. *Acta Crystallogr., Sect. A: Found. Crystallogr.* 2008, 64,
407 112–122.

408 Musumeci, D.; Hunter, C. A.; Prohens, R.; Scuderi, S.; McCabe, J. F. Virtual cocrystal screening. *Chem.*
409 *Sci.* 2011, 2, 883–890.

410 Grecu, T.; Hunter, C. A.; Gardiner, E. J.; McCabe, J. F. Validation of a Computational Cocrystal
411 Prediction Tool: Comparison of Virtual and Experimental Cocrystal Screening Results. *Cryst.*
412 *Growth Des.* 2014, 14, 165–171.

413 Grecu, T.; Adams, H.; Hunter, C. A.; McCabe, J. F.; Portell, A.; Prohens, R. Virtual Screening Identifies
414 New Cocrystals of Nalidixic Acid. *Cryst. Growth Des.* 2014, 14, 1749–1755.

415 Grecu, T.; Prohens, R.; McCabe, J. F.; Carrington, E. J.; Wright, J. S.; Brammer, L.; Hunter, C. A.
416 *CrystEngComm* 2017, 19, 3592–3599.

417 Llinas, A.; Barbas, R.; Font-Bardia, M.; Quayle, M. J.; Velaga, S.; Prohens, R. Two New Polymorphic
418 Cocrystals of Zafirlukast: Preparation, Crystal Structure, and Stability Relations. *Cryst. Growth*
419 *Des.* 2015, 15, 4162–4169.

420 Corner, P.; Berry, D. J.; McCabe, J. F.; Barbas, R.; Prohens, R.; Du, H.; Zhou, H.; Llinas, A. Property
421 Prediction and Pharmacokinetic Evaluation of Mixed Stoichiometry Cocrystals of Zafirlukast, a
422 Drug Delivery Case Study. *CrystEngComm* 2018, 20, 1346–1351.

423 Calero, C. S.; Farwer, J.; Gardiner, E. J.; Hunter, C. A.; Mackey, M.; Scuderi, S.; Thompson, S.; Vinter,
424 J. G. Footprinting molecular electrostatic potential surfaces for calculation of solvation energies.
425 *Phys. Chem. Chem. Phys.* 2013, 15, 18262–18273.

426 Oliver, A.; Hunter, C. A.; Prohens, R.; Rossello, J. L. A surface site interaction point methodology for
427 macromolecules and huge molecular databases. *J. Comput. Chem.* 2017, 38, 419–426.0

428 Oliver, A.; Hunter, C. A.; Prohens, R.; Rossello, J. L. An improved methodology to compute Surface
429 Site Interaction Points using high density Molecular Electrostatic Potential Surfaces. *J. Comput.*
430 *Chem.* 2018, 39, 2371–2377.

- 431 Childs, S. L.; Stahly, G. P.; Park, A. The salt-cocystal continuum: the influence of crystal structure on
432 ionization state. *Mol. Pharmaceutics* 2007, 4, 323–338.
- 433 Lemmerer, A.; Govindraju, S.; Johnston, M.; Motloun, X.; Savig, K. L. Co-crystals and molecular salts
434 of carboxylic acid/pyridine complexes: can calculated pKa's predict proton transfer? A case study
435 of nine complexes. *CrystEngComm* 2015, 17, 3591–3595.
- 436 Cruz-Cabeza, A. J. Acid-base crystalline complexes and the pKa rule. *CrystEngComm* 2012, 14,
437 6362–6365.
- 438 Apparent pKa's calculated using ACD Labs software, release 2015 Pack2, 09 Jun 2015.
- 439 Methanol, ethanol, IPA, butanol, ethylene glycol, 2,2,2- trifluoroethanol, benzyl alcohol, ACN, MEK,
440 acetone, MiBK, water, DMF, DMSO, pentane, heptane, cyclohexane, toluene, xylene, AcOEt,
441 diethylether, THF, dimethyl ethylene glycol, diisopropyl ether, dioxane, dichloromethane,
442 chloroform, formic acid, acetic acid, NH₃ (32%) in water, NH₃ (2.0 M) in methanol,
443 dimethylamine (2.0 M) in methanol, diethylamine, diethylamine (32%) in water, trimethylamine.
444 and methanol-dichloromethane (10:90).
- 445 Jacobs, A.; Amombo Noa, F. M. Hybrid Salt-Cocystal Solvate: p-Coumaric Acid and Quinine System.
446 *J. Chem Crystallogr.* 2014, 44, 57–62.
- 447 Mahieux, J.; Gonella, S.; Sanselme, M.; Coquerel, G. Crystal structure of a hybrid salt–cocystal and its
448 resolution by preferential crystallization: ((±)-trans-N,N'-dibenzylidiaminocyclohexane)(2,3-
449 dichlorophenylacetic acid)₄. *CrystEngComm* 2012, 14, 103–111.
- 450 Aakeröy, C. B.; Fasulo, M. E.; Desper, J. Cocystal or Salt: Does It Really Matter? *Mol. Pharmaceutics*
451 2007, 4 (3), 317–322.
- 452 Reddy, L. S.; Babu, N. J.; Nangia, A. *Chem. Commun. (Cambridge, U. K.)* 2006, 0, 1369–1371.
- 453 Lemmerer, A.; Admond, D. A.; Esterhuysen, C.; Bernstein, J. Polymorphic Co-crystals from
454 Polymorphic Co-crystal Formers: Competition between Carboxylic Acid···Pyridine and
455 Phenol···Pyridine Hydrogen Bonds. *Cryst. Growth Des.* 2013, 13, 3935–3952.
- 456 Prohens, R.; Barbas, R.; Portell, A.; Font-Bardia, M.; Alcobe, X.; Puigjaner, C. Polymorphism of
457 Cocystals: The Promiscuous Behavior of Agomelatine. *Cryst. Growth Des.* 2016, 16,
458 1063–1070.
- 459

460 **Legends to figures**

461

462 **Fig. 1** Molecular structure of compound 1.

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464 **Fig. 2** SSIPs calculated for 1. Blue spheres correspond to H-bond donors and red spheres to H-bond
465 acceptors

466

467 **Fig. 3** Powder X-ray diffractograms of compound 1 solvates (blue: Form A, red: Form B, green: Form
468 C).

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470 **Fig. 4** Powder X-ray diffractograms of compound 1 acetic acid solvates (blue: Form D-1, red: Form D-
471 2, green: Form D-3, brown: Form D-4).

472

473 **Fig. 5** DSC (top) and TGA (bottom) of 1/acetic acid forms.

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475 **Fig. 6** Powder X-ray diffractograms of the new multicomponent forms of compound 1 obtained as a
476 single form (blue: Form I, red: Form III, green: Form VI, brown: Form VII, purple: Form VIII).

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478 **Fig. 7** Powder X-ray diffractograms of the new forms of 1:3,5-dinitrobenzoic acid (blue: Form II-A, red:
479 Form II-B, green: Form II-C, brown: Form II-D, purple: Form II-E).

480

481 **Fig. 8** Powder X-ray diffractograms of the new forms of 1:orotic acid (blue: Form IV-A, red: mixture
482 Form IV-A and Form IV-B, green: mixture Form IV-A and Form IV-C).

483

484 **Fig. 9** Powder X-ray diffractograms of the new forms of 1:5-nitroisophthalic acid (blue: Form V-A, red:
485 Form V-B, green: Form V-C, brown: Form V-D, purple: Form V-E).

486

487 **Fig. 10** (a) Electrostatic interactions in the sandwiched 1,4,8,11-tetrazacyclotetradecane cation and (b)
488 ribbons of 1 molecules assembled by charge assisted hydrogen bonds.

489

490 **Fig 11** Contacts observed in the crystal structure of acetic acid hybrid salt-cocrystal.

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493 **Fig. 12** (a) 1 H-bond parameters from DFT calculations and (b) contacts observed in the crystal structure
494 of DMF solvate

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497 **Fig. 13** Interactions of 1 in the crystal structure of Form II-A.

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499 **Fig. 14** DSC (top) and TGA (bottom) of 1:3,5-dinitrobenzoic salts.

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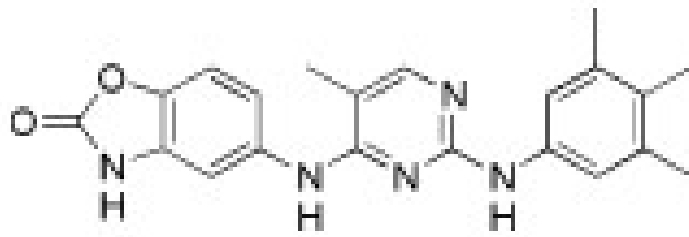
501 **Fig. 15** DSC (top) and TGA (bottom) of 1:5-nitroisophthalic acid salts.

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FIGURE 1

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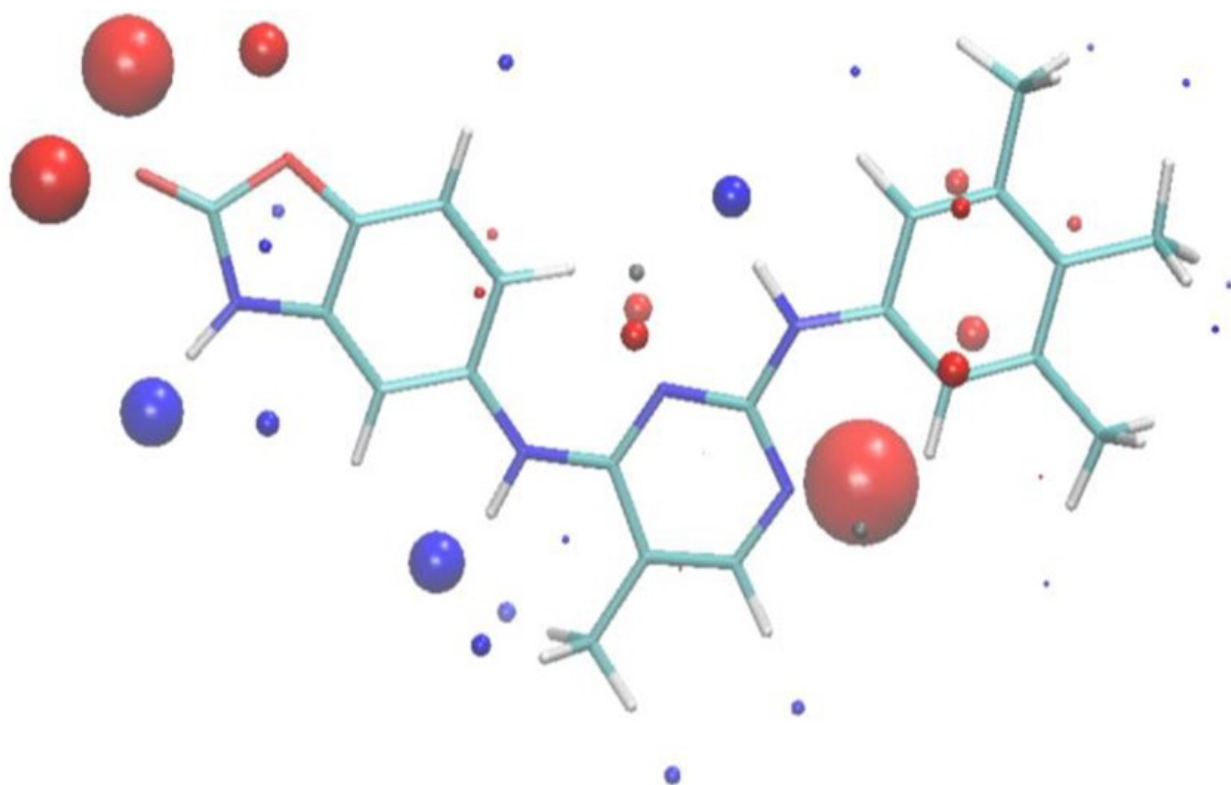


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FIGURE 2



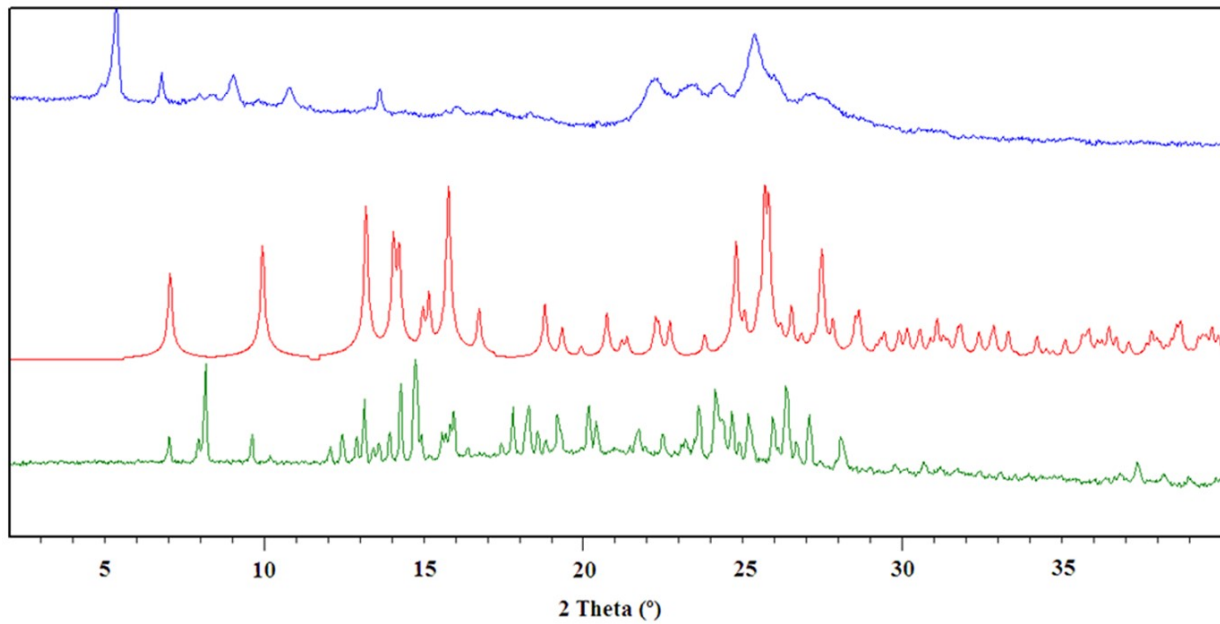
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FIGURE 3

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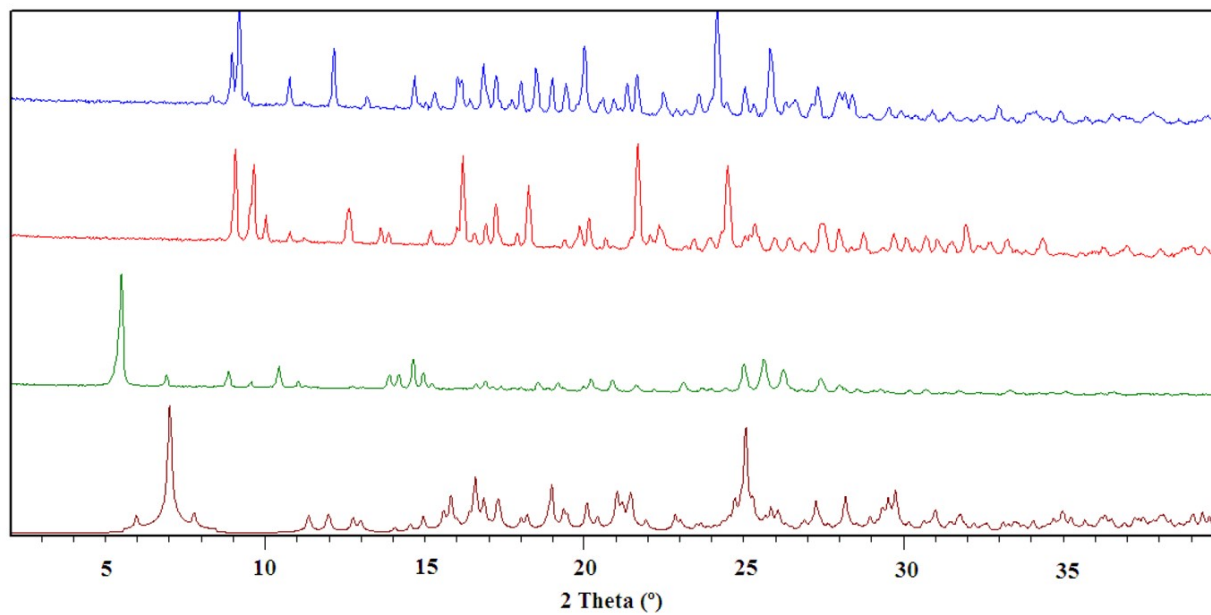
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FIGURE 4

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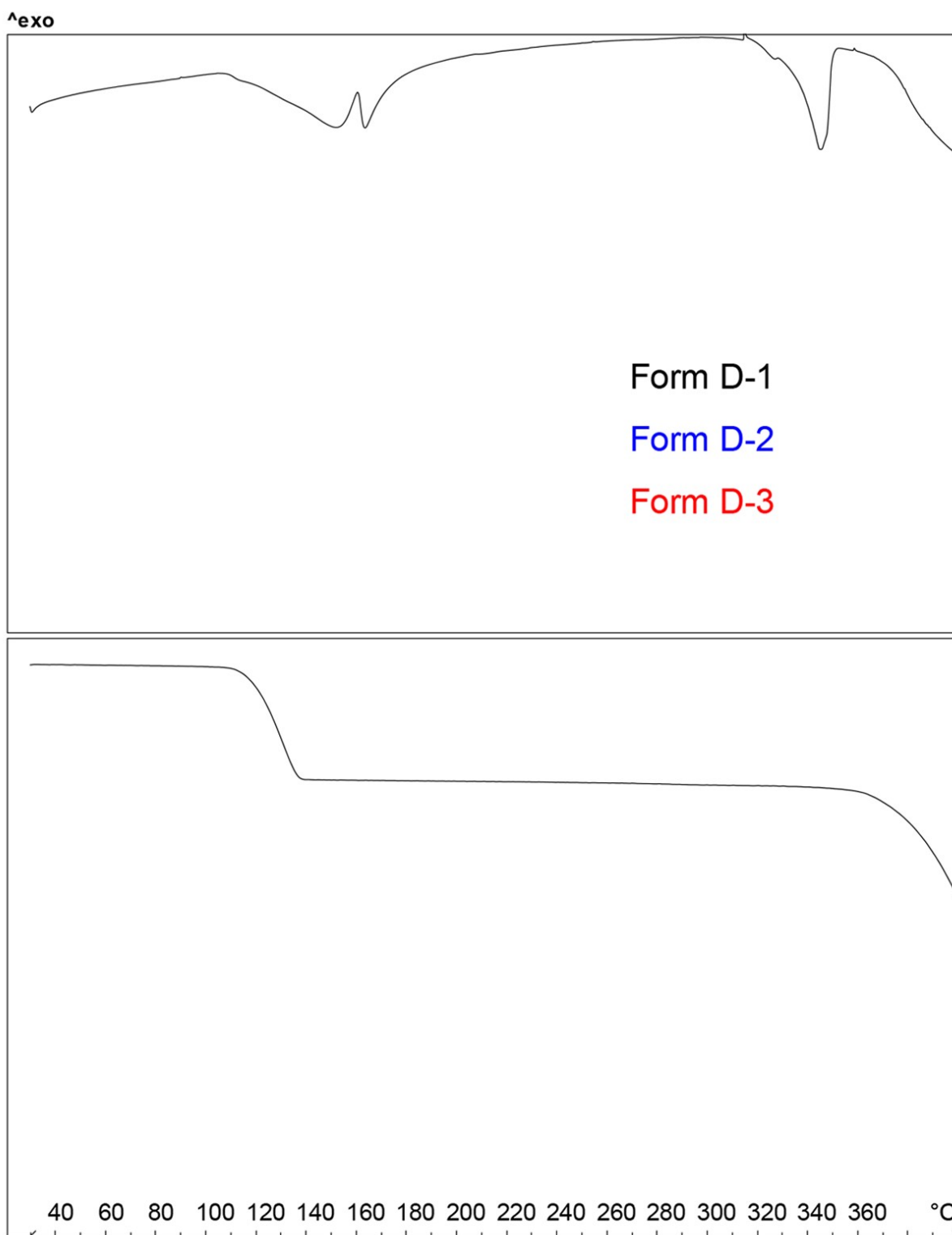


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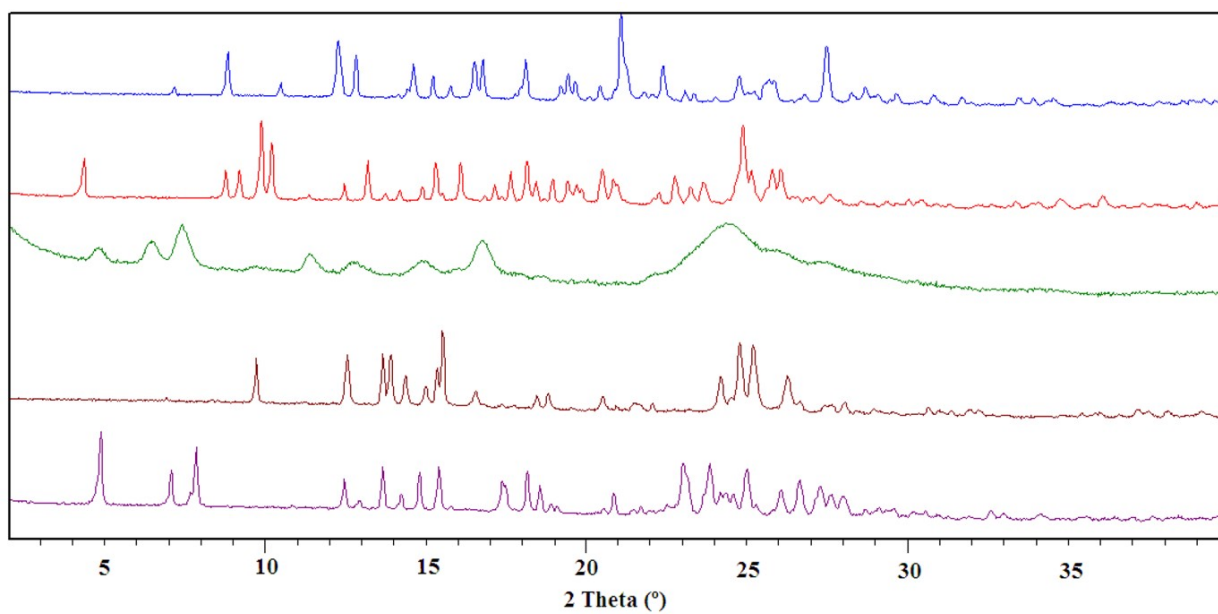
FIGURE 5



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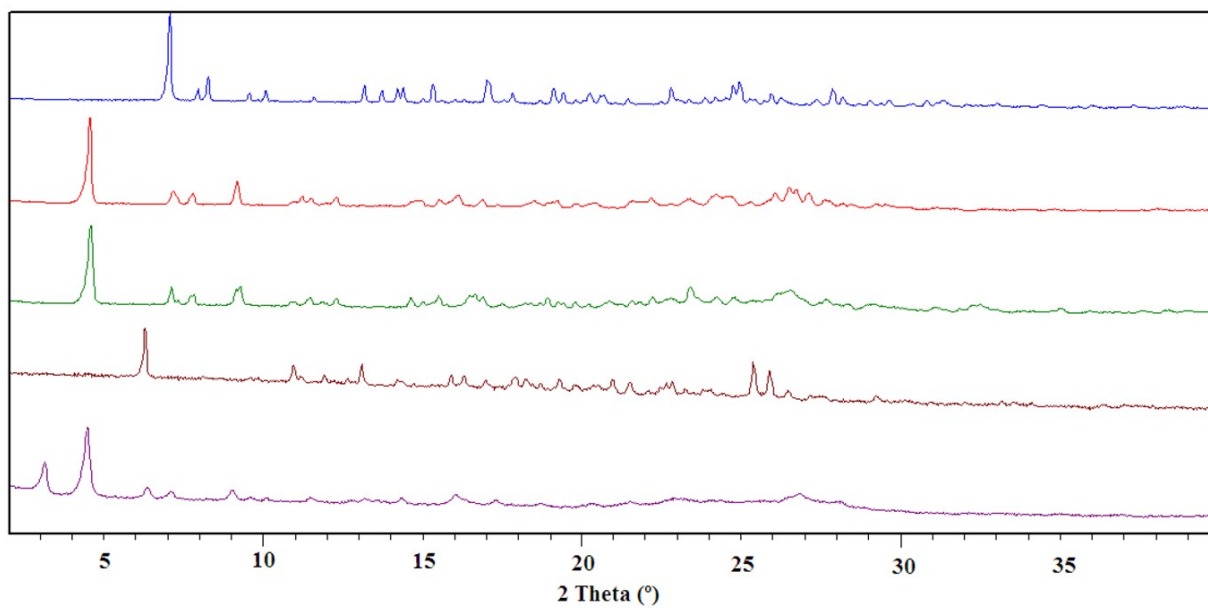
FIGURE 6



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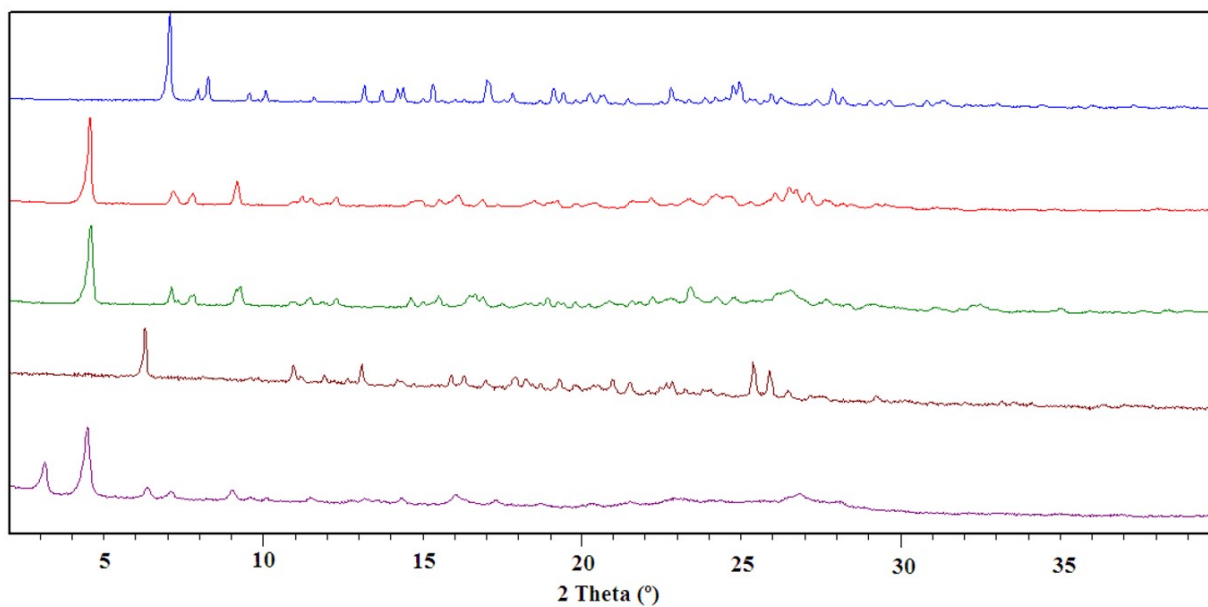
FIGURE 7



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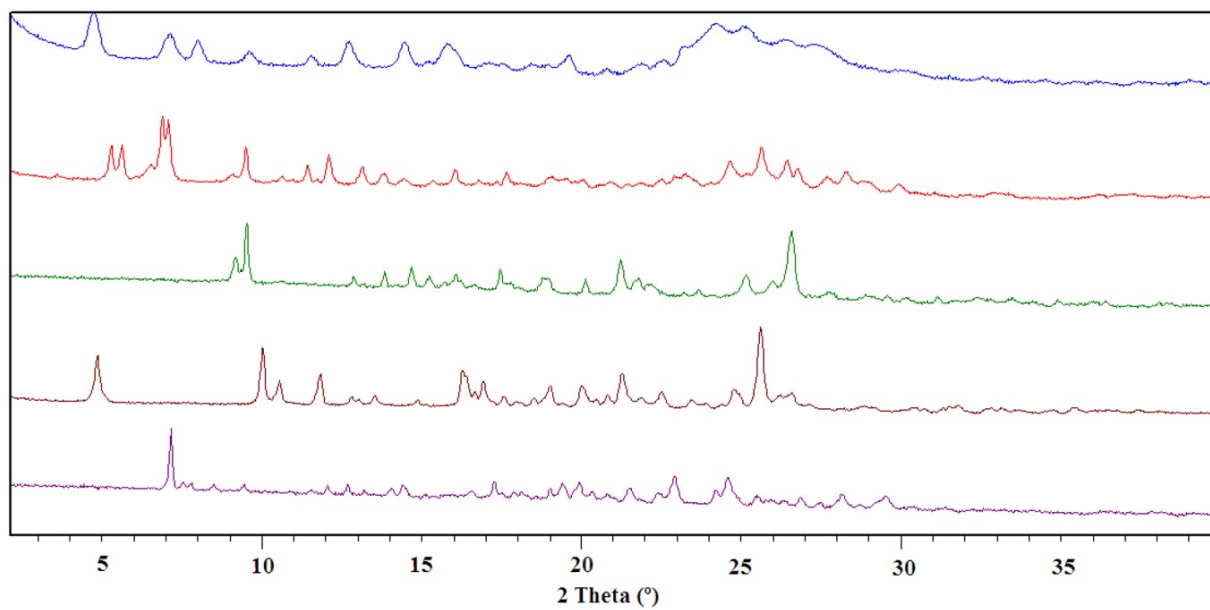
FIGURE 8



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FIGURE 9

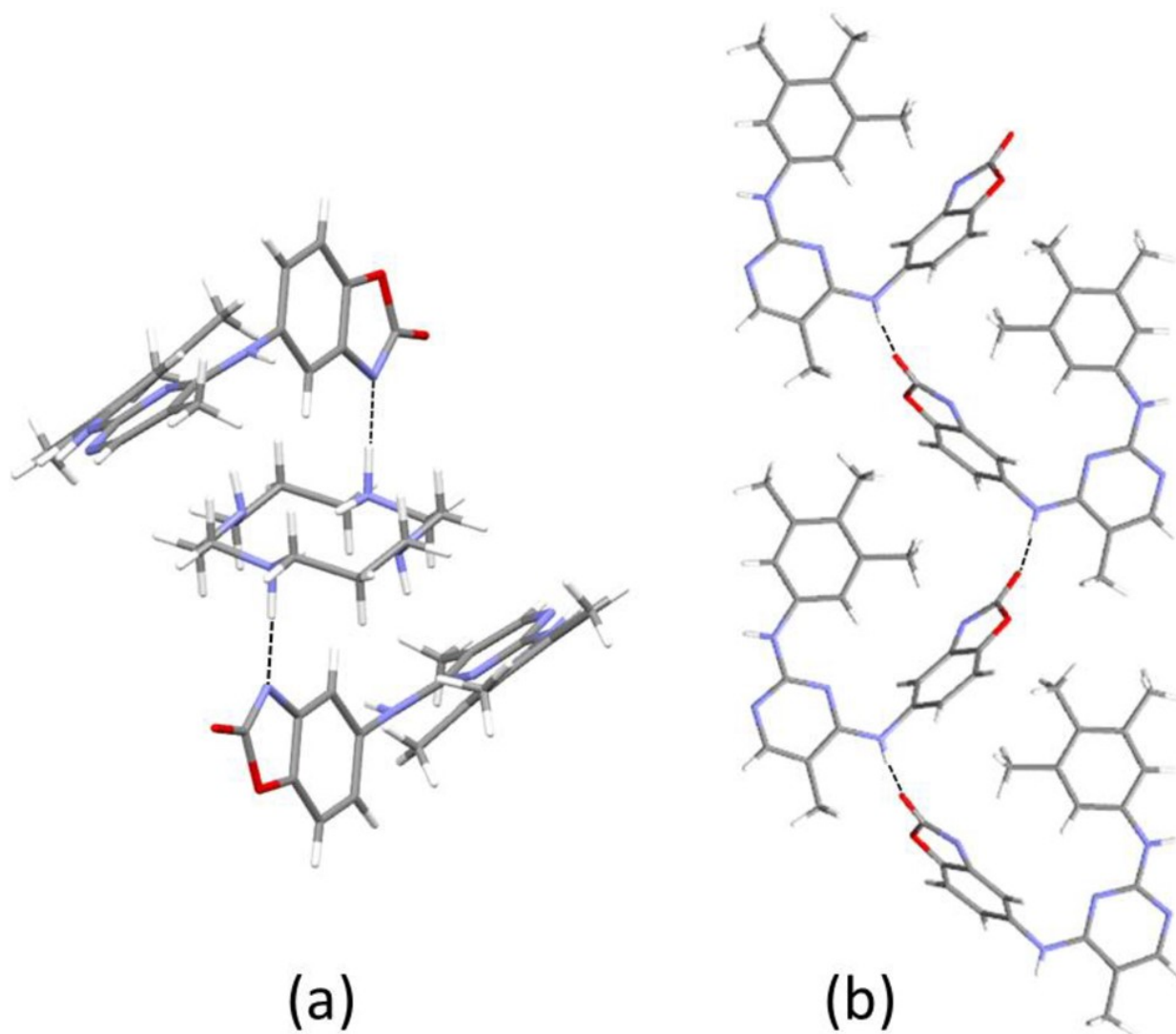
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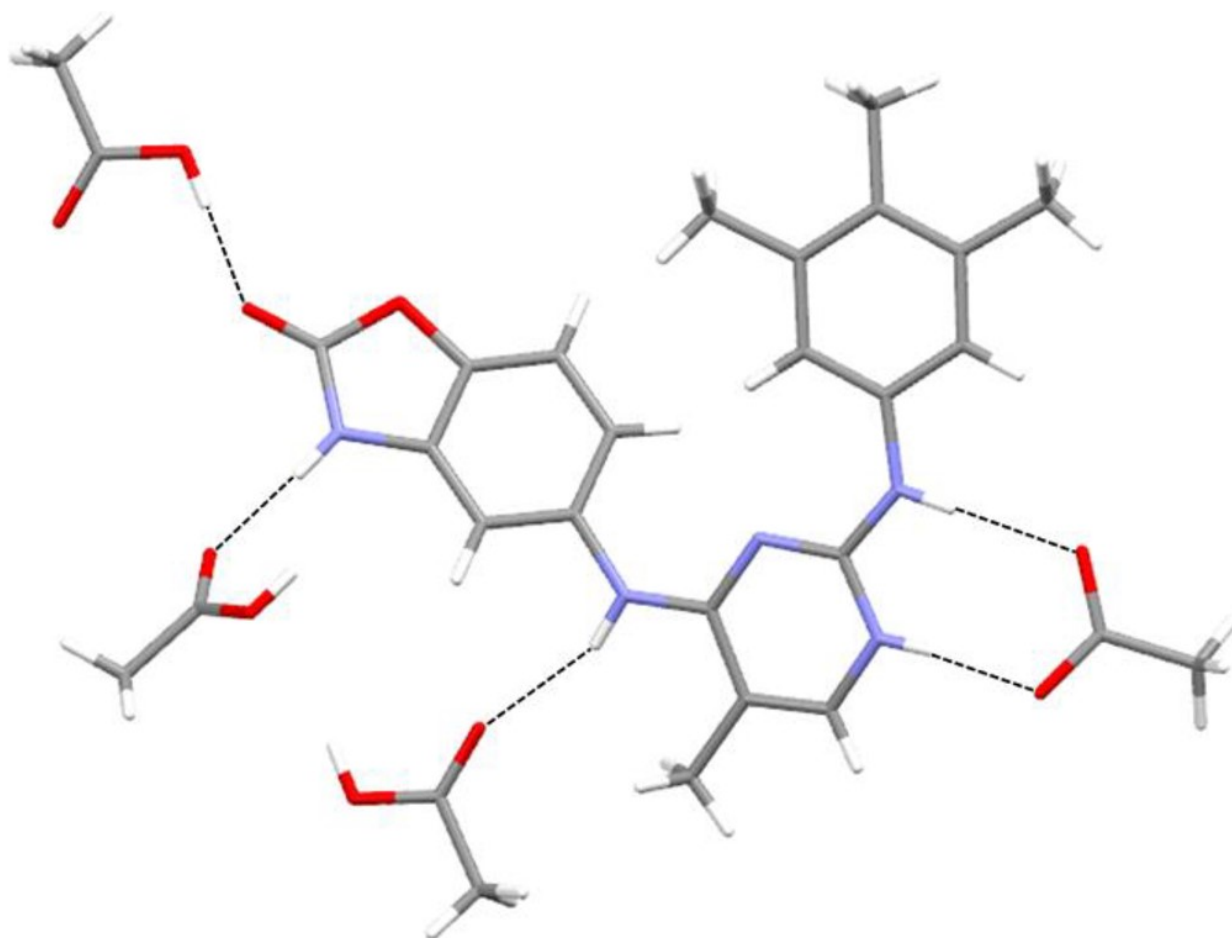
FIGURE 10



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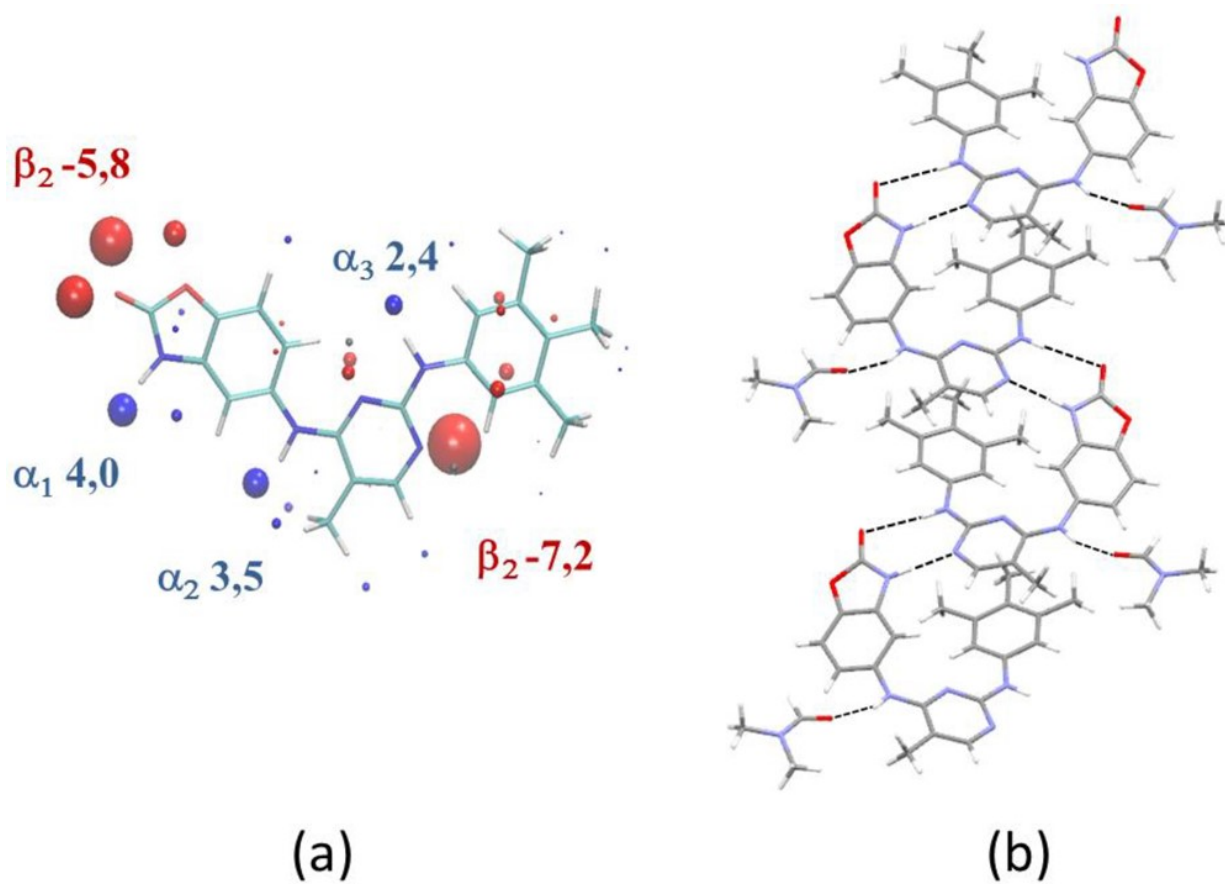
FIGURE 11



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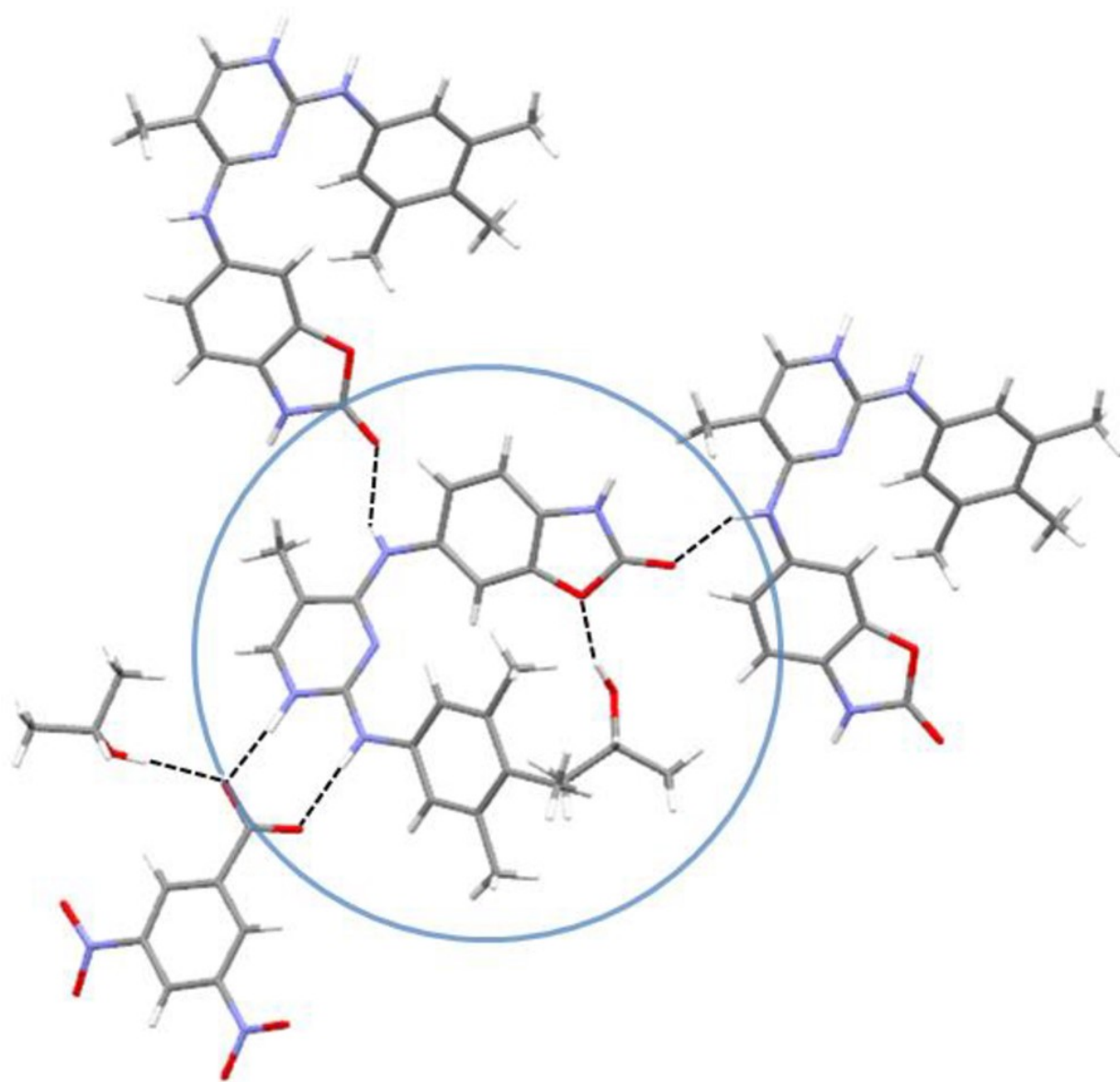
FIGURE 12



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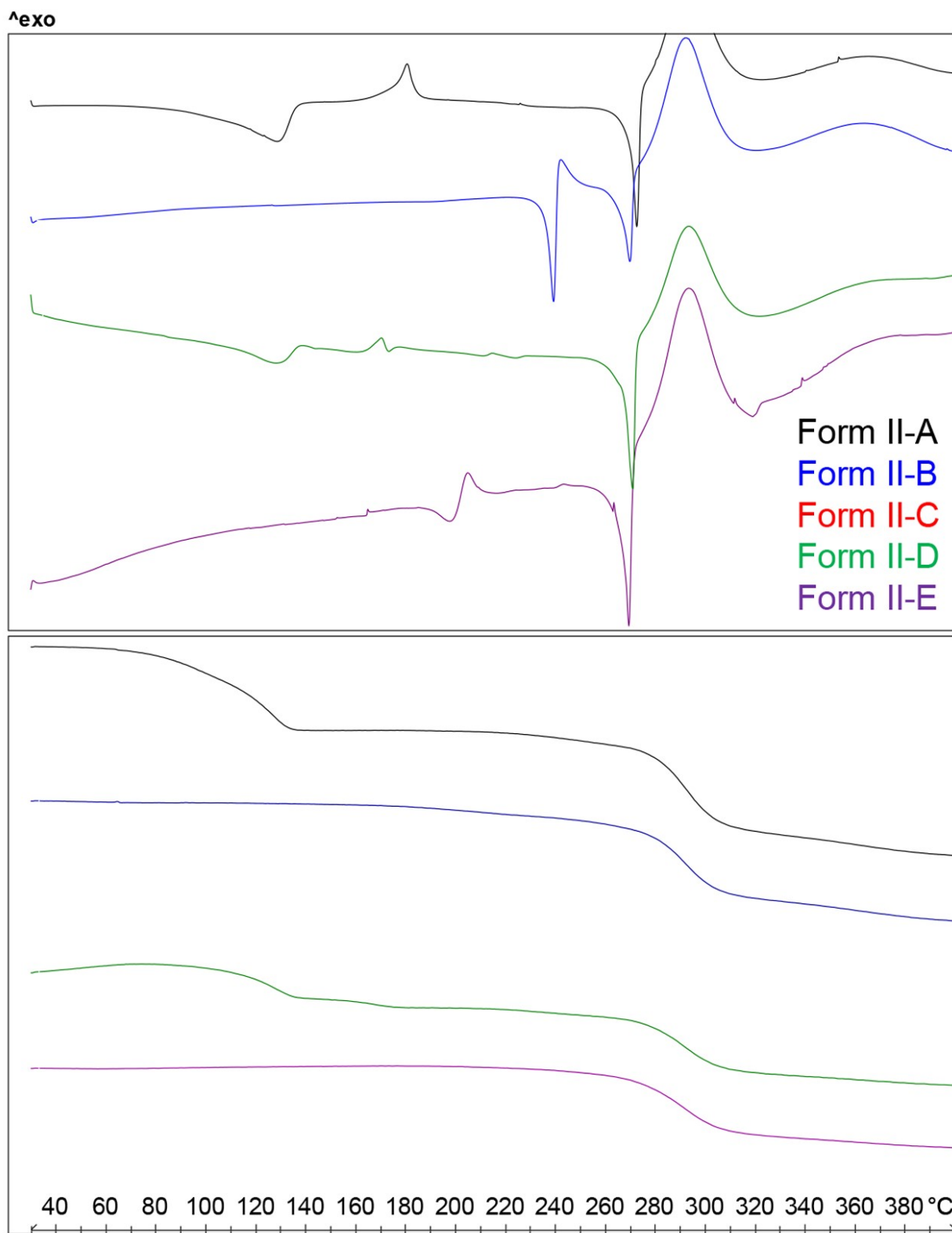
FIGURE 13



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FIGURE 14



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579 **Table 1** Table 1. Summary of Physicochemical Properties of the Free Base of 1

molecular weight (g/mol)	375.4
LogD	4.2
crystalline solubility (μM)	0.03
experimental $\text{p}K_{a1}$, $\text{p}K_{a2}$	5.9, 8.9
Hu PPB (% free)	0.16
Hu Caco2 pH = 6.5 with inhibitors ($\times 1 \times 10^{-6}$ cm/s)	11.2

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582 **Table 2** Cocrystal Screening Coformers Ranked by ΔE , $\text{p}K_a$ 'sa

Coformer	$\Delta E / \text{kJ mol}^{-1}$	Calculated coformer $\text{p}K_a^{35}$	$\Delta \text{p}K_a$	P (%)
Triphenylphosphine Oxide	28.2	-	-	-
1,10-Phenanthroline	27.6	4.9	-4.0	0
1,4-Diazabicyclo[2.2.2]octane	22.3	8.2	-0.7	16
1,4,8,11-Tetrazacyclotetradecane	18.0	10.9	2.0	62
N,N-Dimethylpiperazine	14.5	8.0	-0.9	13
3,5-Dinitrobenzoic acid	14.4	2.8	3.1	81
(+) - Nootkatone	13.7	-	-	-
1,1-Diethoxyethane	13.3	-	-	-
1-Methyl-2-pyrrolidinone	13.0	-0.4	-9.3	0
1-Methylimidazole	12.9	7.0	-1.9	0
4-Nitrobenzenesulfonic acid	12.0	-1.4	7.3	100
Gallie acid	11.1	4.3	1.6	55
Quercetin	10.7	6.3	-0.4	21
Acetylpyrazine	10.7	0.3	-8.6	0
5-Nitroisophthalic acid	10.6	2.8	3.1	81
Orotic acid	10.4	2.8	3.1	81
Tetracyanoethylene	9.0	-	-	-
Pentaflorophenol	8.8	5.5	0.4	35
Resorcinol	8.0	9.4	-3.5	0
3,5-Dihydroxybenzoic acid	7.2	3.9	2.0	62

^aRed = acidic group, blue = basic group.

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585 **Table 3.** Indexed Unit Cell Parameters Data of Compound 1 Multicomponent Forms

form	a (Å)	b (Å)	c (Å)	α (deg)	β (deg)	γ (deg)	V (Å ³)	R_{wp} (%)	space group
D-2	12.123(2)	17.606(3)	11.136(2)	90	105.95(1)	90	2285.3(8)	8.2	$P2_1/n$
III	20.793(4)	10.906(1)	16.091(3)	90	104.80(1)	90	3527(1)	6.8	$P2_1/c$
IV-A	30.07(1)	22.592(9)	4.308(2)	90	116.47(4)	90	2619(2)	8.3	$P2_1/a$
V-D	18.422(8)	10.011(5)	10.972(3)	116.80(2)	87.75(2)	99.29(3)	1781(1)	7.8	$P\bar{1}$
VII	13.201(5)	12.683(4)	7.498(2)	90	101.73(2)	90	1229.2(7)	9.3	$P2/n$

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595 **Table 4.** Crystal Data for the Different Crystal Forms of Compound 1

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structure	Form I	Form II-A	DMF solvate	acetic acid hybrid salt-cocrystal
empirical formula	C ₂₆ H ₃₀ N ₇ O ₂	C ₃₄ H ₄₁ N ₇ O ₁₀	C ₂₄ H ₂₉ N ₆ O ₃	C ₂₉ H ₃₇ N ₅ O ₁₀
formula weight	475.59	707.74	448.52	615.63
temperature (K)	100(2)	100(2)	100(2)	100(2)
wavelength (Å)	0.71073	0.71073	1.54178	0.71073
crystal system	monoclinic	monoclinic	monoclinic	triclinic
space group	<i>P</i> 2 ₁ / <i>c</i>	<i>P</i> 2 ₁ / <i>n</i>	<i>P</i> <i>n</i>	<i>P</i> $\bar{1}$
<i>a</i> , <i>b</i> , <i>c</i> (Å)	12.4888(17) 13.8659(18) 14.4933(19)	11.4419(8) 15.2684(12) 21.4335(17)	7.2802(5) 12.5393(9) 12.7893(10)	7.8386(12) 13.257(2) 15.457(3)
α , β , γ (deg)	90 103.322(5) 90	90 103.182(3) 90	90 99.226(4) 90	72.782(6) 86.818(6) 82.655(7)
volume (Å ³)	2442.2(6)	3645.8(5)	1152.41(15)	1521.4(4)
<i>Z</i> , density (calc.) (Mg/m ³)	4, 1.293	4, 1.289	2, 1.293	2, 1.344
crystal size (mm ³)	0.501 × 0.312 × 0.261	0.522 × 0.091 × 0.040	0.862 × 0.488 × 0.305	0.460 × 0.072 × 0.070
reflections collected/unique data/parameters	77204/9331 [R(int)= 0.0448] 9331/0/335	123319/9088 [R(int) = 0.1079] 9088/397/522	20741/4169 [R(int) = 0.0681] 4169/3/322	53225/7000 [R(int) = 0.1322] 7000/0/409
goodness-of-fit on F ²	1.046	1.036	1.023	1.032
final <i>R</i> indices [<i>I</i> > 2σ(<i>I</i>)]	<i>R</i> ₁ = 0.0479, <i>wR</i> ₂ = 0.1409	<i>R</i> ₁ = 0.0601, <i>wR</i> ₂ = 0.1510	<i>R</i> ₁ = 0.0465, <i>wR</i> ₂ = 0.1026	<i>R</i> ₁ = 0.0571, <i>wR</i> ₂ = 0.1067
CCDC	1868808	1868809	1868807	1868806

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