A comprehensive study on the electrochemical advanced oxidation of antihypertensive captopril in different cells and aqueous matrices

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10 Abstract

11 Anodic oxidation with electrogenerated H_2O_2 (AO- H_2O_2) of 130 mL of antihypertensive drug captopril was studied in sulfate medium, urban wastewater and synthetic urine using a BDD, Pt 12 13 or IrO₂ anode and an air-diffusion cathode. Oxidants were •OH formed during O₂ evolution and active chlorine formed via anodic oxidation of Cl⁻. Drug removal decreased as: BDD > Pt > 14 15 IrO₂. The effect of pH and current density was examined. Further, 2.5 L of drug solutions in the same matrices with Fe²⁺ at pH 3.0 were treated by solar photoelectro-Fenton (SPEF) using a 16 17 solar pre-pilot flow plant with a Pt/air-diffusion cell and a planar photoreactor. In sulfate 18 medium, SPEF outperformed AO-H₂O₂ and electro-Fenton because of the efficient Fe³⁺ 19 photoreduction. Low mineralization was achieved by the small generation of photoactive 20 Fe(III)-carboxylate complexes. In urban wastewater, captopril was more rapidly removed due 21 to active chlorine Two heterocyclic derivatives and four aliphatic acids were detected.

Keywords: Anodic oxidation; Pharmaceutical; Solar photoelectro-Fenton; Urban wastewater;
Urine

24 **1. Introduction**

Recently, the great effectiveness of the solar photoelectro-Fenton (SPEF) process for the removal of recalcitrant and persistent organic pollutants from synthetic and real wastewater has been shown [1-3]. SPEF is an environmental-friendly electrochemical advanced oxidation process (EAOP) that destroys the organic molecules by the simultaneous action of hydroxyl radical (•OH) generated on site and UVA photons directed toward the solution. The •OH possesses a great ability to non-selectively attack the organic structures due to its high standard redox potential (E°) of 2.8 V|SHE [4,5].

In SPEF process, the photolytic action of sunlight enhances the performance of a simpler but more widespread EAOP like electro-Fenton (EF) [6-8]. In EF, the weak oxidant H₂O₂ (E° = 1.76 V|SHE) is generated from O₂ reduction at the cathode of an electrolytic cell via reaction (1). This reaction becomes very efficient using carbonaceous electrodes, including carbonpolytetrafluoroethylene (PTFE) [9-11] and carbon felt [12-14]. In a typical undivided cell, H₂O₂ is partly oxidized to O₂ at the anode M, originating the weaker physisorbed hydroperoxyl radical (M(HO₂•), $E^{\circ} = 1.44$ V|SHE) as intermediate from reaction (2) [5,15].

$$39 \quad O_2 + 2H^+ + 2e^- \to H_2O_2 \tag{1}$$

40
$$M + H_2O_2 \rightarrow M(HO_2^{\bullet}) + H^+ + e^-$$
 (2)

41 EF was developed from anodic oxidation process with cathodic H_2O_2 electrogeneration 42 (AO- H_2O_2), which involves the destruction of organics either by H_2O_2 or, to a much larger 43 extent, by physisorbed M(•OH) formed from water discharge according to reaction (3) [16-18]:

44
$$M + H_2O \rightarrow M(^{\bullet}OH) + H^+ + e^-$$
 (3)

The nature of the anode M is the most influential factor controlling reaction (3). Non-active boron-doped diamond (BDD) electrodes are known to possess a higher oxidation power than active Pt and dimensionally stable anodes like IrO₂ [19,20]. This general trend can be modulated upon production of other oxidizing agents in selected media in which the action of M(•OH) is
affected. For example, in chloride medium, the oxidant active chlorine can be additionally
formed, which can be either beneficial or detrimental [16,21].

In systems equipped with the aforementioned types of cathodes and anodes, the presence of Fe²⁺ as catalyst in the acidic solution promotes the occurrence of Fenton's reaction (4) [7,21,22]. The catalyst can be continuously regenerated from cathodic Fe³⁺ reduction, but the oxidation power is substantially enhanced under SPEF conditions. The exposure of the treated solution to sunlight radiation allows that the incident UVA photons photoreduce the main generated Fe³⁺ species via reaction (5) and photodecompose the Fe(III)-carboxylate complexes according to the general reaction (6) [1,11,23,24].

58
$$\operatorname{Fe}^{2+} + \operatorname{H}_2\operatorname{O}_2 \to \operatorname{Fe}^{3+} + {}^{\bullet}\operatorname{OH} + \operatorname{OH}^{-}$$
 (4)

59
$$\operatorname{Fe}(\operatorname{OH})^{2+} + h\nu \to \operatorname{Fe}^{2+} + {}^{\bullet}\operatorname{OH}$$
 (5)

$$60 \quad \operatorname{Fe}(\operatorname{OOCR})^{2+} + h\nu \to \operatorname{Fe}^{2+} + \operatorname{CO}_2 + \operatorname{R}^{\bullet}$$
(6)

Several works have shown the viability of the SPEF treatment to remove aromatic and 61 62 heteroaromatic drugs from acidic wastewater [7,25-28]. One of the main conclusions is that, 63 unlike AO-H₂O₂, similar results can be obtained using active and non-active anodes because of 64 the concomitant action of UV photons on reaction intermediates [7,21,29]. However, less is 65 known about the destruction of heterocyclic drugs, which are expected to be more recalcitrant. ((2S)-1-[(2S)-2-methyl-3-sulfanylpropanoyl]pyrrolidine-2-carboxylic 66 Captopril acid. 67 $C_9H_{15}NO_3S$) is a widely used heterocyclic thiolated antihypertensive drug. It acts as an inhibitor 68 of the angiotensin-converting enzyme to regulate blood pressure, also being used in the 69 management of diabetic nephropathy and heart failure after myocardial infarction [30]. 70 However, about 40-50% of administered captopril is excreted unmetabolized via urine, whereas 71 the rest is converted to disulfide and other metabolites [31]. This results in a large presence of 72 this drug in influents of municipal wastewater treatment facilities, attaining high contents up to 73 2.3 μ g L⁻¹ [32,33]. In surface water, it can induce toxicity in crustaceans like daphnids and the 74 common carp Cyprinus carpio [34]. Several works have described the chemical oxidation of 75 captopril by bromate and bromine [35], quinolinium dichromate [36] and laccase [37]. As far as we know, only Freitas et al. [30] have assessed its removal by AOPs, addressing the study 76 77 of heterogeneous photocatalysis with TiO₂/UVC. Total degradation of captopril was achieved 78 after 120 min, although the mineralization was only 9.1%, suggesting the generation of highly 79 recalcitrant by-products. In contrast, direct UVC photolysis under analogous conditions yielded 80 93% degradation with 2.9% mineralization. The use of powerful EAOPs could then become an 81 alternative to achieve a faster degradation and mineralization.

82 This work aims to clarify the electrochemical destruction of captopril in different aqueous 83 matrices. First, the experiments were performed under AO-H₂O₂ conditions using a small 84 volume of 130 mL, in order to test the oxidation ability of BDD, Pt and IrO₂ anodes in synthetic 85 sulfate medium as a model matrix. The process was further evaluated in urban wastewater and 86 synthetic urine, examining the effect of pH and current density (j) on the oxidation power. 87 Afterwards, the SPEF process was investigated in the three aqueous matrices using a 2.5 L solar 88 pre-pilot flow plant equipped with a Pt/air-diffusion cell coupled with a planar photoreactor. 89 The effect of *j* and drug content on the SPEF performance was studied, and comparative AO-90 H₂O₂ and EF assays were made to confirm the superiority of SPEF process. Finally, the main 91 reaction by-products were identified by gas chromatography-mass spectrometry (GC-MS).

92 2. Materials and methods

93 2.1. Chemicals

Captopril (CAS number 62571-86-2, M = 217.29 g mol⁻¹, 98% purity) was purchased from Sigma-Aldrich and used as received. Analytical grade sulfuric acid (98% purity) and sodium hydroxide pellets needed for pH adjustment were from Acros and Panreac, respectively.

97	Analytical grade sodium sulfate and iron(II) sulfate heptahydrate were acquired from Prolabo.							
98	The analytical solutions were prepared using a Merck Millipore Milli-Q ultrapure water system							
99	(resistivity >18.2 M Ω cm). The reagents for the preparation of the synthetic urine and the							
100	analytical solutions were of analytical or HPLC grade purchased from Merck and Panreac.							
101	2.2. Aqueous matrices							
102	The aqueous matrices used to dissolve captopril were:							
103	(i) A 0.050 M Na ₂ SO ₄ solution, prepared with Milli-Q or deionized water to perform the							
104	trials at 130-mL and 2.5-L scale, respectively;							
105	(ii) Urban wastewater collected from the secondary clarifier of a municipal facility. The							
106	water was preserved at 4 °C once obtained and its main characteristics were: $pH = 7$.							
107	conductivity = 1.3 mS cm ⁻¹ , total organic carbon (TOC) = 12.0 mg C L ⁻¹ ; cation							
108	contained (mg L ⁻¹): Fe ²⁺ (0.11), NH ₄ ⁺ (37.5), Mg ²⁺ (41.1), K ⁺ (47.4), Ca ²⁺ (130) ar							
109	Na ⁺ (213); anions (mg L ⁻¹): NO ₂ ⁻ (2.03), NO ₃ ⁻ (5.21), SO ₄ ²⁻ (131) and Cl ⁻ (415);							
110	(iii) Synthetic urine prepared with Milli-Q water for the AO-H ₂ O ₂ treatment at 130-m							
111	scale, with the following composition (in mM) [38]: urea (NH_2CONH_2 , 55.6), uric action (in mM) [38]: urea (NH_2CONH_2, 55.6), uric action (in mM) [38]: urea (NH_2CONH_2, 55.6), urea (NH_2CONH_2CONH_2)] [38]: urea (NH_2CONH_2CONH_2CONH_2)] [38]: urea (NH_2CONH_2CONH_2CONH_2CONH_2CONH_2CONH_							
112	(C ₅ H ₄ N ₄ O ₃ , 0.29), creatinine (C ₄ H ₇ N ₃ O, 1.47), KCl (13.4), MgSO ₄ (1.41), (Ca) ₃ (PO ₄							
113	(0.10) , $(NH_4)_2$ HPO ₄ (0.63) and Na_2CO_3 (1.57) ;							
114	(iv) Three synthetic urine solutions prepared with deionized water for SPEF trials:							
115	• Urine 1: 13.9 mM urea + 0.07 mM uric acid + 0.37 mM creatinine;							
116	• Urine 2: 27.8 mM urea + 0.15 mM uric acid + 0.73 mM creatinine; and							
117	• Urine 3: 55.6 mM urea + 0.29 mM uric acid + 1.47 mM creatinine.							
118	2.3. Electrolytic assays							

The AO-H₂O₂ trials were carried out at 130-mL scale using an undivided two-electrode
cell. The solution was stirred with a magnetic follower at 600 rpm and its temperature was kept

121 at 35 °C by recirculating thermostated water through a jacket surrounding the cell. The anode 122 was a BDD thin film deposited onto Si, purchased from NeoCoat, Pt (99.999% purity) 123 purchased from SEMPSA or IrO2-coated Ti plate purchased from NMT Electrodes. The 124 cathode was a carbon-PTFE air-diffusion electrode purchased from Sainergy Fuel Cell, which was fitted in a cylindrical polypropylene support and fed with air at $1 \text{ L} \text{ h}^{-1}$ for H₂O₂ production 125 126 from reaction (1). The interelectrode gap was always 1 cm and each electrode area was 3 cm^2 . 127 Galvanostatic assays were made with an Amel 2049 potentiostat-galvanostat, connected to a 128 601BR digital multimeter for continuous Demestres cell voltage monitoring. 129 Cleaning/activation of electrodes was carried out as described elsewhere [29].

130 The SPEF experiments were performed in a solar pre-pilot flow plant constructed in our 131 laboratory [11,39]. A filter-press one-compartment electrochemical reactor containing two 132 electrodes in parallel, separated 1.2 cm, was employed. The anode was a 20 cm² Pt plate (99.999% purity) from SEMPSA and the cathode was a 20 cm² carbon-PTFE air-diffusion 133 134 electrode from Sainergy Fuel Cell. The back side of this cathode was exposed to a gas chamber in which air was pumped at 4.5 L h⁻¹ for H₂O₂ production. A solar planar photoreactor was 135 136 connected to the outlet of the electrolytic cell. It consisted in a polycarbonate box of 24.0 cm \times 24.0 cm × 2.5 cm (i.e., 600 mL irradiated volume), tilted 41° (latitude of our balcony in 137 138 Barcelona) and with a mirror at the bottom to enhance the photon usage. The solution of 2.5 L 139 placed in the reservoir was recirculated through the plant using a centrifugal pump, 140 consecutively flowing toward the flowmeter, two heat exchangers, the electrolytic cell and the photoreactor before being received back into the reservoir. A liquid flow rate of 180 L h⁻¹ and 141 142 a temperature of 35 °C were kept constant in all the assays. These were carried out under 143 galvanostatic conditions and the constant *j* was provided by a Grelco GVD310 power supply, 144 which displayed the instantaneous cell voltage. Comparative AO-H₂O₂ and EF trials were made 145 by covering the elements with an opaque cloth. The SPEF runs were performed in sunny and 146 clear days of August-September of 2019. The incident average UV irradiance was of 32.3 W
 147 m⁻², as measured with a Kipp & Zonen CUV 5 radiometer.

148 2.4. Apparatus and analytical procedures

149 The solution pH, conductivity and concentrations of cations, anions and chlorine active 150 were measured as reported in earlier work [29]. The H₂O₂ content was determined by the 151 standard Ti(IV) colorimetric method using a Shimadzu 1800 UV/Vis spectrophotometer set at 152 $\lambda = 408$ nm [40]. All the samples withdrawn from treated solutions were immediately 153 conditioned by using PTFE filters (0.45 µm) from Whatman.

154 The decay of captopril concentration was monitored by reverse-phase HPLC using a 155 Waters system that included a photodiode array detector (PDA). The liquid chromatograph was 156 equipped with a BDS Hypersil C18 5 μ m, 250 mm × 4.6 mm, column at 25 °C and the detection 157 was made at $\lambda = 290$ nm. The mobile phase was a 60:40 (v/v) acetonitrile/water (10 mM 158 KH₂PO₄, pH 3) mixture eluted at 1.0 mL min⁻¹. The chromatograms displayed a defined peak 159 for captopril at retention time (*t*_r) of 3.6 min.

160 The mineralization of drug solutions was followed by the decay of their solution TOC, 161 evaluated with a Shimadzu VCSN TOC analyzer using the non-purgeable organic carbon 162 (NPOC) method (reproducibility of $\pm 1\%$). Duplicate assays were made for captopril and TOC 163 analyses, and average values with their error bars (95% confidence interval) are depicted.

164 The specific energy consumption per unit TOC mass (EC_{TOC}) in each run at electrolysis 165 time *t* (in h) was obtained according to Eq. (7) [15]:

166
$$\operatorname{EC}_{\operatorname{TOC}}(\operatorname{kWh}(\operatorname{g}\operatorname{TOC})^{-1}) = \frac{E_{\operatorname{cell}}It}{V\triangle(\operatorname{TOC})_{\exp}}$$
 (7)

167 where E_{cell} is the average cell voltage (in V), *I* is the applied current (in A), *V* is the solution 168 volume (in L) and \triangle (TOC)_{exp} is the TOC abatement (in mg C L⁻¹). 169 The main by-products formed from captopril degradation upon SPEF treatment of a 0.230 170 mM drug solution with 0.050 M Na₂SO₄ at j = 50 mA cm⁻² were identified by GC-MS with the 171 same equipment and procedure described in earlier work [28]. The analysis was made with an 172 Agilent Technologies system containing either a polar HP INNOWax or a non-polar 173 Teknokroma Sapiens-X5ms column, both of 0.25 µm, 30 m × 0.25 mm. The compounds were 174 identified by comparison with the NIST05 MS database.

175 **3. Results and discussion**

176 *3.1. AO*-*H*₂*O*₂ treatment of captopril solutions at small scale

177 3.1.1. Captopril removal using different anodes in acidic aqueous matrices

178 The first AO-H₂O₂ trials were carried out at small scale, treating 130 mL of 0.230 mM 179 captopril solutions prepared either in pure sulfate medium, urban wastewater or synthetic urine at pH 3.0 using a BDD, Pt or IrO₂ anode at i = 33.3 mA cm⁻². The solution pH did not change 180 181 during these electrolyses. Fig. 1a-c show the corresponding normalized decays of drug 182 concentration. Table 1 summarizes the maximum degradation percentage achieved at a given 183 electrolysis time. Based on all these results, it can be concluded that the degradation was always 184 enhanced in the order: $IrO_2 < Pt < BDD$, regardless of the matrix. Moreover, captopril was 185 much more slowly removed in the Na₂SO₄ solution (see Fig. 1a). Considering for example the 186 most powerful anode (i.e., BDD), total drug disappearance was attained after 120 min in this 187 medium, whereas a much shorter time of about 60 min was needed in urban wastewater and 188 synthetic urine (see Table 1). The same trends can be seen using the Pt and IrO₂ anodes in Fig. 189 1a-c. The use of BDD in sulfate medium allows the preeminent drug oxidation upon the attack 190 of BDD(•OH) formed from reaction (3). The slower decay with Pt and IrO₂ can be explained 191 by the lower oxidation power of Pt(•OH) and IrO₂(•OH), respectively [5,16]. The quicker 192 degradation in the other two matrices can then be ascribed to the simultaneous action of chlorine, produced via anodic oxidation of Cl⁻ ion (see content in subsection 2.2) according to reaction (8). Subsequent hydrolysis yields HClO via reaction (9), and this oxidant becomes the predominant active chlorine species at pH 3.0, with $E^{\circ} = 1.49$ V|SHE [3,41].

$$196 \quad 2Cl^- \rightarrow Cl_2 + 2e^- \tag{8}$$

$$197 \quad \text{Cl}_2 + \text{H}_2\text{O} \rightarrow \text{HClO} + \text{Cl}^- + \text{H}^+ \tag{9}$$

198 Fig. 1d-f present the pseudo-first-order kinetic analysis of captopril concentration decays 199 discussed above. These profiles suggest that all the AO-H₂O₂ treatments are limited by the mass 200 transport of Cl⁻ ion and reactants toward the anode, where a small and steady concentration of 201 oxidizing species (HClO and/or BDD(•OH), Pt(•OH), IrO₂(•OH)) is produced. Table 1 202 summarizes the resulting pseudo-first-order rate constants for captopril removal (k_1) , with good 203 R-squared > 0.980. In sulfate medium, the higher oxidation power of BDD(•OH) was confirmed 204 by a k_1 -value that was 3.2-fold and 4.2-fold greater than that determined with Pt and IrO₂, 205 respectively. In wastewater and synthetic urine, the k_1 -value with BDD was increased more 206 than threefold, suggesting the larger destruction of the drug by HClO.

Table 1 also highlights the decreasing TOC removal percentage as BDD was successively replaced by Pt and IrO₂, going down from 17% to 9.4% and 6.2%, respectively, at 120 min in sulfate medium (initial TOC = 25 mg C L^{-1}).

210 3.1.2. Effect of pH and current density on captopril removal

The treatment of 0.230 mM captopril in the three aqueous matrices was extended to neutral pH 7.0 and alkaline conditions (pH 9.0). Fig. S1a-c and Fig. S2a-c depict the corresponding time course of the normalized drug concentration removals by AO-H₂O₂ with BDD, Pt and IrO₂, at j = 33.3 mA cm². The excellent pseudo-first-order kinetic analysis of such trends is shown in Fig. S1d-f and Fig. S2d-f, respectively. These results corroborate the upgrading of the oxidation ability of AO-H₂O₂ in the sequence: IrO₂ < Pt < BDD, as was found at pH 3.0. This 217 means that BDD stands out as the most powerful anode, owing to the action of active chlorine 218 and/or BDD(•OH). The role of the former oxidant was evidenced in the two chloride media, 219 with much greater k_1 -values compared to those in sulfate medium (see Table 1). Table 1 also 220 highlights a decrease of k_1 as pH was increased, regardless of the matrix or the anode. This 221 phenomenon was less significant in sulfate medium, remaining even invariable using IrO₂. The 222 k_1 drop upon pH change from pH 3.0 to 9.0 was more evident in the two chloride media, 223 especially in the case of synthetic urine, undergoing a 0.41-fold, 0.32-fold and 0.27-fold 224 decrease using BDD, Pt and IrO₂, respectively. The larger influence of pH in these matrices can be accounted for by the gradual conversion of HClO into ClO^{-} (p $K_a = 7.45$) and the loss of 225 oxidation ability of the M(•OH) [5,15]. The progressive formation of the weak oxidant ClO-, 226 227 with $E^{\circ} = 0.89$ V/SHE [3], promoted a larger inhibition of captopril removal.

228 The lower performance of AO-H₂O₂ upon pH increase also caused a poorer TOC removal, 229 with the concomitant rise in EC_{TOC} (see Table 1). In this regards, another interesting finding 230 from Table 1 is the gradual mineralization enhancement as the organic load of the matrix was 231 increased. For example, at pH 7.0, the use of BDD at i = 33.3 mA cm⁻² yielded a mineralization 232 percentage of 12% at 120 min in sulfate medium (initial TOC = 25 mg C L^{-1}), 20% at 120 min in urban wastewater (initial TOC = 37 mg C L^{-1}) and 23% at 60 min in synthetic urine (initial 233 234 TOC: 745 mg C L^{-1}). This represents an increasing loss of 3.0, 7.4 and 171.4 mg C L^{-1} of TOC. accounting for decreasing EC_{TOC} values of 5.28, 1.26 and 0.055 kWh (g TOC)⁻¹, respectively. 235 These results reveal the partial destruction of the 12 mg L^{-1} TOC present in the urban 236 237 wastewater associated to the natural organic matter (NOM), which includes humic, fulvic and 238 tannic acids [7,29]. Furthermore, there is clear evidence of partial mineralization of the aliphatic 239 urea and the two heterocyclic compounds, uric acid and creatinine, of the synthetic urine matrix. 240 In the literature, the destruction of urea by AO with Pt in sulfate medium has been reported 241 [41], but the removal of uric acid and creatinine has not been described yet. On the other hand, Cotillas et al. [38] have reported a large mineralization of synthetic urine matrices thanks to theaction of HClO formed at a BDD anode.

244 The current density is a key parameter that modulates the generation of oxidizing agents in 245 the AO-H₂O₂ process. A higher degradation rate is expected with raising *j*, in agreement with 246 the increasing rate of the electrode reactions (1), (3) and (8), thus ending in a greater quantity 247 of H₂O₂, M(•OH) and HClO, respectively [3,29]. This effect was assessed in the three matrices 248 at pH 7.0 using the BDD anode. Fig. 2a-c confirm the enhancement of captopril removal upon rise of *j* from 16.7 to 66.7 mA cm⁻². This trend was less substantial in urine, probably by the 249 250 dominant role of active chlorine due to its higher Cl⁻ content. The drug always obeyed a pseudo-251 first-order reaction, as corroborated from the good fittings shown in Fig. 2d-f. The behavior 252 was confirmed from the higher k_1 -value and percentage of TOC removal determined as *i* was 253 increased, as listed in Table 1. However, note that the EC_{TOC} values also became progressively 254 higher, owing to the greater E_{cell} , which is detrimental if the cost must be minimized. 255 Additionally, the increase in rate of parasitic reactions of the oxidants, as detailed below, caused 256 a relatively smaller \triangle (TOC)_{exp} with the consequent negative impact on EC_{TOC}.

257 In all matrices, the degradation rate also became lower as *j* rose. Thus, the k_1 increase was 1.9-fold in pure sulfate, 3.0-fold in wastewater and 1.8-fold in urine upon a fourfold rise of *j*, 258 from 16.7 to 66.7 mA cm⁻² (see Table 1). This disagreement can be related to the gradually 259 260 larger extent to which parasitic reactions occurred. In the case of BDD(•OH), these reactions 261 include its immediate conversion to O₂ gas and H₂O₂ because of the lack of drug molecules on 262 the anode surface, and its reaction with H_2O_2 to produce the weaker oxidant HO_2^{\bullet} [7,10-12]. 263 Regarding active chlorine, it can be mainly undergo oxidation to yield ClO₂⁻, ClO₃⁻ and ClO₄⁻ 264 ions [3,29]. From these findings, it is evident that a higher *j* causes a larger destruction of the 265 organic matter, although at the expense of current efficiency.

266 3.1.3. Inorganic ions formed during captopril treatment in synthetic urine at neutral pH

267 To corroborate the aforementioned destruction of the organic components of the urine 268 matrix, the inorganic species generated with each anode at the different pH values were 269 analyzed. Fig. 3a-c depict the concentration of nitrogenated species detected after 60 min at j =33.3 mA cm⁻². A large amount of NO₃⁻ and NH₄⁺ ions was accumulated under all conditions, 270 271 whereas NO₂⁻ ion only appeared at pH 7.0 and 9.0 and its concentration was smaller. The NO₂⁻ 272 accumulation was always greater at pH 7.0, increased in the order: $IrO_2 < Pt < BDD$, and attained a maximum value of 7.6 mg L^{-1} (0.16 mM N). This tendency agrees with the higher 273 274 oxidation ability of BDD to destroy the N-compounds. The same sequence can be observed for the NO₃⁻ ion with the three anodes at pH 3.0, dropping from 63.2 mg L^{-1} (1.02 mM N) with 275 BDD to 34.9 mg L⁻¹ (0.56 mM N) with IrO₂. In contrast, similar NO₃⁻ concentrations were 276 277 found at pH 7.0 and 9.0 using BDD and IrO₂, being slightly lower with Pt. Conversely, in the case of NH₄⁺ ion, higher concentrations were always obtained using Pt, reaching the greatest 278 concentration of 42.6 mg L⁻¹ (2.37 mM N). Note that the N content associated to captopril 279 280 spiked into the synthetic urine (i.e., 0.230 mM) was much lower than that corresponding to the 281 sum of accumulated NO_3^- and NH_4^+ , revealing that the two latter ions pre-eminently proceeded 282 from the destruction of urea, uric acid and creatinine, the *N*-components of the synthetic urine. 283 The active chlorine concentration accumulated in treated urine during the above trials is 284 depicted in Fig. 3d. The greatest accumulation was obtained using Pt, regardless of the pH 285 tested, suggesting that this anode fostered reaction (8). Its maximum concentration was 6.4 mg L^{-1} at pH 3.0. The attack of such a high amount of active chlorine on the *N*-compounds could 286 287 justify the prevalence of NH₄⁺ ion, alongside the lower production of NO₃⁻ ion at pH 7.0 and 9.0, as compared to BDD and IrO₂ (see Fig. 3a-c). 288

- 289 3.2. SPEF treatment of captopril solutions
- 290 $3.2.1. H_2O_2$ accumulation in the solar pre-pilot flow plant

291 The ability of the flow plant to accumulate H₂O₂ electrogenerated with a Pt/air-diffusion 292 cell was first assessed by treating 2.5 L of 0.050 M Na₂SO₄ solutions at pH 3.0 and 35 °C under 293 AO-H₂O₂ conditions at different *j* values. Fig. S3a shows a progressive increase of the H_2O_2 294 concentration at each given *j*, attaining a final value of 4.3, 9.5 and 14.1 mM at 10, 30 and 50 mA cm⁻², respectively. For each trial, the current efficiency for H₂O₂ accumulation calculated 295 from Faraday's law decreased over time. At i = 30 mA cm⁻², for instance, the efficiency was 296 297 69.5% at 60 min, 60.7% at 150 min and 42.4% at 300 min. This gradual decay can be related 298 to the acceleration of H_2O_2 destruction from reaction (2), which results from the use of an 299 undivided cell [15]. Fig. S3b also shows a decrease of current efficiency as *j* became higher, varying from 57.8% at 10 mA cm⁻² to 34.9% at i = 50 mA cm⁻². The increase of *i* accelerated 300 301 all electrode reactions, thereby promoting the H₂O₂ production via reaction (1) but 302 comparatively favoring its destruction via reaction (2), finally causing the efficiency drop 303 shown in Fig. S3b.

304 The H₂O₂ accumulation was also determined under EF and SPEF conditions, at i = 50 mA cm^{-2} , in the presence of 0.50 mM Fe²⁺ as catalyst. Fig. S3a shows that, in EF, the H₂O₂ content 305 306 decayed strongly and only reached 3.1 mM at 300 min (7.7% of current efficiency). This means 307 that, apart from the H_2O_2 loss via reaction (2), this compound was more largely removed via 308 Fenton's reaction (4), with the consequent production of 'OH that could upgrade the water 309 treatment. A higher drop of H₂O₂ concentration occurred under SPEF conditions, with a final 310 value of 1.4 mM (3.5% of current efficiency). This is due to the acceleration of Fenton's reaction (4) resulting from Fe^{2+} regeneration via reaction (5). Based on all these results, EF and 311 312 SPEF processes can be employed for the oxidation of organic pollutants and their by-products.

313 3.2.2. Captopril removal by different EAOPs in sulfate medium

Solutions with 0.230 mM captopril (i.e., 25 mg C L⁻¹ TOC) and 0.050 M Na₂SO₄ at pH 3.0
were comparatively treated by AO-H₂O₂, EF and SPEF processes using the pre-pilot flow plant,

at j = 50 mA cm⁻². A Pt anode was utilized as example of active anodes, which are less expensive than the non-active BDD and show similar degradation efficiency in SPEF [7,21,29]. The Fenton-based EAOPs were made in the presence of 0.50 mM Fe²⁺ since this is the optimum content found for similar treatments of aromatic pollutants with an air-diffusion cathode [7]. The value of pH change remained quite constant throughout these trials.

321 The decay of normalized drug concentration with electrolysis time is presented in Fig. 4a. 322 The degradation of captopril by AO-H₂O₂ process was very slow, with a disappearance as low 323 as 36% at 30 min. This confirms the very low oxidation power of electrogenerated H₂O₂ and 324 Pt(OH) formed from reaction (3) to destroy the drug, as found at small scale. In contrast, a 325 much faster captopril removal was provided by EF, with a complete drug removal in about 20 326 min. This is an evidence of the very effective attack of free 'OH produced via Fenton's reaction 327 (4) in all the volume, being a good alternative to hydroxyl radicals confined within the anode 328 vicinity. The degradation rate was slightly increased in the SPEF process, with total removal in 329 only 15 min thanks to the greater generation of 'OH upon photoreduction reaction (5). These 330 results agree with the behavior described for H₂O₂ accumulation in Fig. S3a.

331 The exponential concentration decays shown in Fig. 4a were analyzed assuming a pseudo-332 first-order kinetics for captopril removal. Excellent linear regressions were obtained, as can be 333 seen in Fig. 4b. This means that also in Fenton-based EAOPs a steady quantity of oxidizing 334 agents was generated within the range of conditions tested. Table 2 collects the k_1 -values obtained, always with $R^2 > 0.99$. The lowest k_1 -value was determined in the less powerful 335 336 process (i.e., AO-H₂O₂), which experienced a 22-fold and 30-fold increase in EF and SPEF, 337 respectively. The impressive rise confirms the relevance of homogeneous •OH formed from 338 reaction (4) and/or (5).

The TOC abatement in the above trials was measured for 300 min (data not shown). The final percentages of TOC removal are listed in Table 2. As can be seen, the mineralization

degree increased in accordance with the oxidation power of the EAOPs, i.e., $AO-H_2O_2 < EF < C$ 341 342 SPEF. Note the very low TOC decay achieved by AO-H₂O₂, as expected from the very slow 343 drug disappearance (see Fig. 4a). Surprisingly, TOC was only reduced by 25% (6.2 mg C L⁻¹ 344 TOC) in SPEF, suggesting that the by-products formed were pre-eminently oxidized by 345 homogeneous •OH, but with a very small generation of photoactive Fe(III) complexes that were 346 rapidly removed via reaction (6). This behavior has not been observed for aromatic and 347 heteroaromatic pollutants, whose mineralization by SPEF can be almost complete and fast 348 because of the occurrence of such active complexes that are quickly transformed into CO₂ 349 [1,15,22]. The corresponding EC_{TOC} values calculated at 300 min were as high as 30.1 kWh (g 350 TOC)⁻¹ in AO-H₂O₂, being much smaller in EF (5.9-fold) and SPEF (8.9-fold) (see Table 2).

351 3.2.3. Effect of experimental variables on the SPEF process in sulfate medium

352 The influence of key experimental variables such as applied *j* and drug concentration on 353 the performance of the SPEF process was examined. Fig. 5 depicts the gradual enhancement of captopril degradation when *j* was increased from 10 to 50 mA cm⁻² to treat 0.230 mM drug 354 355 solutions, owing to the larger presence of ${}^{\bullet}OH$ resulting from the acceleration of H_2O_2 electrogeneration. A change from 10 to 30 and 50 mA cm⁻² led to a shorter time span of 60, 40 356 357 and 15 min for total removal. The inset panel of Fig. 5 presents the good linear profiles obtained 358 for these concentration abatements. The resulting k_1 -values, shown in Table 2, evidence that the change from 30 to 50 mA cm⁻² was beneficial, as deduced from the 3.3-fold increase of k_1 . 359 Therefore, at 50 mA cm⁻², the attack of •OH on the drug was more favored than the parasitic 360 361 reactions mentioned above. The percentage of TOC removal at 300 min was also upgraded, 362 from 16% at i = 10 mA cm⁻² to 25% at i = 50 mA cm⁻², as shown in Table 2. In turn, this table 363 reveals a strong increase of EC_{TOC} at higher *j*, resulting from the higher *j* and E_{cell} .

The influence of drug content was studied from 0.100 to 0.460 mM (from 11 to 50 mg C L^{-1} TOC) in 0.050 M Na₂SO₄ solutions, at pH 3.0 and j = 50 mA cm⁻². The rise of the initial

366 concentration led to a longer electrolysis time for total removal. Fig. 6 highlights that captopril 367 disappeared from the medium at increasing time, from 5 min at 0.100 mM to 30 min at 0.460 368 mM. A positive feature was that the degradation process became more efficient. Thus, after 5-369 6 min, a concentration of 0.100 mM was removed from the less concentrated solution, whereas 370 0.212 and 0.314 mM were removed from the solutions initially containing 0.230 and 0.460 mM, 371 respectively. This is correlated with the enhanced oxidation power of the system thanks to the 372 larger number of reactive events between captopril molecules and the oxidants. Obviously, this 373 tendency is opposite to that of k_1 , determined from the excellent linear regressions presented in 374 the inset panel of Fig. 6. The data of Table 2 show the drop of the k_1 -value from 1.128 min⁻¹ at 0.100 mM to 0.186 min⁻¹ at 0.460 mM, in agreement with the longer time required for total 375 376 drug abatement. This fact is also deduced from Table 2 considering the percentage of TOC 377 removed at 300 min in SPEF, since it decayed significantly from 33% to 19% when treating 378 0.100 and 0.460 Mm, respectively. Nevertheless, the amount of TOC abated varied from 3.6 to 9.5 mg C L⁻¹, in agreement with a greater mineralization current efficiency. This trend was 379 380 reflected in the EC_{TOC} values, which were 2.7-fold lower at 0.460 mM as compared to 0.100 381 mM, attaining a minimal of 2.27 kWh (g TOC)⁻¹ (see Table 2). These findings demonstrate that 382 the SPEF treatment was more cost-effective when the solutions were more contaminated.

383 *3.2.4. Effect of the aqueous matrix on captopril removal performance*

Once clarified the behavior of the SPEF process in sulfate medium at pH 3.0, the study was extended to aqueous matrices where captopril has been detected, namely urban wastewater and urine. The latter medium was simulated as described in subsection 2.2 (matrices called urine 1, urine 2 and urine 3).

The urban wastewater contained 12 mg C L⁻¹ TOC. Captopril was spiked at a concentration of 0.230 mM into 2.5 L of this matrix, yielding 37 mg C L⁻¹ TOC, and the resulting solution was adjusted to pH 3.0 with H₂SO₄. After addition of Fe²⁺ (0.50 mM), it was treated by SPEF

in the pre-pilot flow plant, at i = 50 mA cm⁻². Fig. 7 shows a rapid disappearance of the drug in 391 392 only 15 min, a time analogous to that needed in sulfate medium (see Fig. 4). From the pseudo-393 first-order kinetic analysis depicted in the inset panel of Fig. 7, a higher k_I -value of 0.551 (vs. 0.452 min⁻¹) was obtained in wastewater (see Table 2). Since the natural organic components 394 395 of this matrix are supposed to partially scavenge the 'OH [7], a deceleration of the drug decay 396 was presumed, but the opposite behavior was observed. This can be accounted for by the 397 generation of active chlorine (HClO) in SPEF. The simultaneous degradation of captopril by 398 both oxidants (•OH and HClO) in urban wastewater enhanced the SPEF performance. The 399 existence of these oxidants also justified the higher percentage of TOC removal (28%, i.e., 10.4 mg C L⁻¹ of TOC) found in urban wastewater as compared to 6.2 mg C L⁻¹ determined in sulfate 400 401 medium, which can be ascribed to the partial mineralization of NOM. Note also in Table 2 the 402 much higher EC_{TOC} value for the trial in wastewater, because of the much greater E_{cell} resulting 403 from its smaller conductivity.

404 Captopril was also spiked into each synthetic urine matrix at a concentration of 0.230 mM, the solution pH was adjusted to pH 3.0 and Fe^{2+} (0.50 mM) was added as catalyst. Fig. 7 depicts 405 406 the change of normalized drug concentration with electrolysis time for the assays performed at $i = 50 \text{ mA cm}^{-2}$. As expected, the larger competition between captopril and a gradually higher 407 408 quantity of organic components in the urine matrix caused a deceleration of drug degradation. 409 Anyway, overall captopril abatement was always achieved, requiring 15, 20 and 30 min using 410 urine 1, 2 and 3, respectively. The content of oxidants remained constant in each matrix, which 411 explains the good pseudo-first-order kinetic analysis of the concentration decays (inset panel of 412 Fig. 7). The corresponding k_1 -values given in Table 2 dropped significantly, about 50% when 413 moving from the diluted urine 1 to the most concentrated urine 3.

The concomitant oxidation of the organic components of the urine matrices was also observed when monitoring their TOC abatement. Fig. 8a shows a rapid mineralization with 416 TOC reduction by 70% using urine 1 (205 mg C L⁻¹ of initial TOC, including the 25 mg C L⁻¹ 417 of captopril), which decreased down to a 44% using urine 2 (385 mg C L⁻¹) and to a 26% using 418 urine 3 (745 mg C L⁻¹). Despite this, increasing quantities, i.e., 143, 169 and 194 mg C L⁻¹ of 419 TOC, were removed, confirming the suitability of the EAOPs to efficiently treat concentrated 420 solutions. These results clearly evidence that the aliphatic urea and the heterocyclic molecules 421 of the aqueous matrices can be largely mineralized by the SPEF process.

422 The EC_{TOC} values decreased as the organic load in the urine matrix became higher. This 423 trend can be seen inferred from the EC_{TOC}-time plots of Fig. 8b as well as from the final values 424 listed in Table 2, because similar E_{cell} values between 10.1 and 10.7 V were measured in these 425 trials. Fig. 8b also shows an atypical profile of EC_{TOC}. During the treatment of aromatic an 426 heteroaromatic pollutants, this parameter always grows as the electrolysis progresses, due to 427 the gradual formation of more recalcitrant by-products [1,5]. The opposite tendency can be 428 observed in this figure, where EC_{TOC} always decreased as the electrolysis was prolonged, 429 informing about the gradual production of less refractory by-products. Such species could be 430 short-linear aliphatic acids arising from the destruction of uric acid and creatinine, further 431 giving rise to photoactive Fe(III)-carboxylate complexes that are more easily photolyzed by 432 sunlight. In the next subsection, the main reaction by-products are determined.

433 3.2.5. Identification of main by-products of captopril

Table S1 collects the names, chemical structure and characteristics of 6 main by-products detected by GC-MS after 10 min of SPEF treatment of a 0.230 mM drug solution with 0.050 M Na₂SO₄ at pH 3.0 using the solar pre-pilot flow plant at j = 50 mA cm⁻². Considering these compounds, a plausible pathway for captopril (1) degradation is proposed in Fig. 9, where the main oxidant is the homogeneous •OH formed from Fenton's reaction (4), with the important contribution of the photoreduction reaction (5). It should be noted that the four carboxylic acids (compounds **4** to **7**) are either free or in the form of complexes with Fe(III) [5,7,15], but most 441 of them are expected to possess a very low photoactivity, as deduced from the low TOC removal 442 achieved under these conditions (see Table 2). Although the route of Fig. 9 is proposed for the 443 SPEF process, it could be extended to $AO-H_2O_2$ and EF since Pt(•OH) and/or •OH are the main 444 oxidizing agents as well.

The pathway of Fig. 9 is initiated by the attack of $^{\circ}$ OH over 1 causing its decarboxylation, demethylation and loss of a sulfonylmethyl molecule to yield 2, which is further oxidized to the pyrrolidone 3. The subsequent cleavage of the pyrrolidine moiety at the N(1)-C(5) bond originates the acid 4, which is then oxidized and broken to form the acids 5 and 6. Finally, the oxidation of these two acids leads to acid 7. It should be mentioned that acid 7 is very stable to the attack of $^{\circ}$ OH and must be mineralized to CO₂ via oxidation to oxalic and formic acids as intermediates [5,15].

452 **4.** Conclusions

453 The effectiveness of the AO-H₂O₂ process with an air-diffusion cathode to degrade the 454 heterocyclic drug captopril at pH 3.0-9.0 was greater in urban wastewater and urine, as 455 compared to that observed in sulfate, owing to combined action of M(•OH) and active chlorine. 456 BDD anode ensured the highest performance, and in all media the oxidation power decreased 457 at higher pH. The rise of *j* accelerated the degradation but at the expense of current efficiency. 458 A low mineralization was achieved, regardless of the matrix and the anode, being higher in 459 urine. The larger accumulation of NO₃⁻ and NH₄⁺ ions in urine was related to the destruction 460 of its N-compounds. The SPEF process with a Pt/air-diffusion cell clearly outperformed the 461 AO- H_2O_2 and EF treatments, as a result of the effective production of •OH. However, the mineralization tended to be low, suggesting a small production of photoactive Fe(III) 462 complexes with organic molecules. The oxidation power was higher as the initial solution was 463 464 more concentrated. The degradation and mineralization in urban wastewater was slightly

465 accelerated as compared to those in sulfate medium because of the parallel oxidation with active 466 chlorine, but its low conductivity derived in a much higher EC_{TOC} . Total drug decay was also 467 feasible in urine, showing a larger mineralization with lower EC_{TOC} values. Two heterocyclic 468 derivatives and four linear acids were identified as by-products.

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474 **References**

- 475 [1] E. Brillas, A review on the degradation of organic pollutants in waters by UV
 476 photoelectro-Fenton and solar photoelectro-Fenton, J. Braz. Chem. Soc. 25 (2014) 393477 417.
- I. Salmerón, K.V. Plakas, I. Sirés, I. Oller, M.I. Maldonado, A.J. Karabelas, S. Malato,
 Optimization of electrocatalytic H₂O₂ production at pilot plant scale for solar-assisted
 water treatment, Appl. Catal. B: Environ. 242 (2019) 327-336.
- 481 [3] M.F. Murrieta, I. Sirés, E. Brillas, J.L. Nava, Mineralization of Acid Red 1 azo dye by
 482 solar photoelectro-Fenton-like process using electrogenerated HClO and
 483 photoregenerated Fe(II), Chemosphere 246 (2020) 125697.
- 484 [4] M.A. Oturan, J.J. Aaron, Advanced oxidation processes in water/wastewater treatment:
 485 principles and applications. A review. Crit. Rev. Environ. Sci. Technol. 44 (2014) 2577486 2641.

21

- 487 [5] C.A. Martínez-Huitle, M.A. Rodrigo, I. Sirés, O. Scialdone, Single and coupled
 488 electrochemical processes and reactors for the abatement of organic pollutants: A critical
 489 review, Chem. Rev. 115 (2015) 13362-13407.
- 490 [6] J. Vidal, C. Huilñir, R. Salazar, Removal of organic matter contained in slaughterhouse
 491 wastewater using a combination of anaerobic digestion and solar photoelectro-Fenton
 492 processes, Electrochim. Acta 210 (2016) 163-170.
- 493 [7] J.R. Steter, E. Brillas, I. Sirés, Solar photoelectro-Fenton treatment of a mixture of
 494 parabens spiked into secondary treated wastewater e□uent at low input current, Appl.
 495 Catal. B: Environ. 224 (2018) 410-418.
- 496 [8] D.R.V Guelfi, E. Brillas, F. Gozzi, A. Machulek Jr., S.C. de Oliveira, I. Sirés, Influence
 497 of electrolysis conditions on the treatment of herbicide bentazon using artificial UVA
 498 radiation and sunlight. Identification of oxidation products, J. Environ. Manage. 231
 499 (2019) 213-221.
- 500 [9] S. Lanzalaco, I. Sirés, M.A. Sabatino, C. Dispenza, O. Scialdone, A. Galia, Synthesis of
 501 polymer nanogels by electro-Fenton process: investigation of the effect of main operation
 502 parameters, Electrochim. Acta 246 (2017) 812-822.
- 503 [10] A.J. dos Santos, C.A. Martínez-Huitle, I. Sirés, E. Brillas, Use of Pt and BDD anodes in
 504 the electrochemical advanced oxidation of Ponceau SS diazo dye in acidic sulfate
 505 medium, ChemElectroChem (2018) 685-693.
- 506 [11] A.J. dos Santos, I. Sirés, C.A. Martínez-Huitle, E. Brillas, Total mineralization of
 507 mixtures of Tartrazine, Ponceau SS and Direct Blue 71 azo dyes by solar photoelectro 508 Fenton in pre-pilot plant, Chemosphere 210 (2018) 1137-1144.
- 509 [12] M. Panizza, M.A. Oturan, Degradation of Alizarin Red by electro-Fenton process using
 510 a graphite-felt cathode, Electrochim. Acta 56 (2011) 7084-7087.

511	[13] A. El-Ghenymy, R.M. Rodríguez, E. Brillas, N. Oturan, M.A. Oturan, Electro-Fenton
512	degradation of the antibiotic sulfanilamide with Pt/carbon-felt and BDD/carbon-felt cells.
513	Kinetics, reaction intermediates, and toxicity assessment, Environ. Sci. Pollut. Res. 21
514	(2014) 8368-8378.

- 515 [14] O. Ganzenko, N. Oturan, I. Sirés, D. Huguenot, E.D. van Hullebusch, G. Esposito, M.A.
 516 Oturan, Fast and complete removal of the 5-fluorouracil drug from water by electro517 Fenton oxidation, Environ. Chem. Lett. 16 (2018) 281-286.
- 518 [15] E. Brillas, I. Sirés, M.A. Oturan, Electro-Fenton process and related electrochemical
 519 technologies based on Fenton's reaction chemistry, Chem Rev 109 (2009) 6570-6631.
- 520 [16] M. Panizza, G. Cerisola, Direct and mediated anodic oxidation of organic pollutants,
 521 Chem. Rev. 109 (2009) 6541-6569.
- 522 [17] S. Lanzalaco, I. Sirés, A. Galia, M.A. Sabatino, C. Dispenza, O. Scialdone, Facile
 523 crosslinking of poly(vinylpyrrolidone) by electro-oxidation with IrO₂-based anode under
 524 potentiostatic conditions, J. Appl. Electrochem. 48 (2018) 1343-1352.
- 525 [18] E. do Vale-Júnior, A.J. dos Santos, D.R. da Silva, A.S. Fajardo, C.A. Martínez-Huitle,
- 526 Electrochemical technologies for detecting and degrading benzoquinone using diamond
 527 films, ChemElectroChem 6 (2019) 4383-4390.
- 528 [19] F. Sopaj, M.A. Rodrigo, N. Oturan, F.I. Podvorica, J. Pinson, M.A Oturan, Influence of
 529 the anode materials on the electrochemical oxidation efficiency. Application to oxidative
 530 degradation of the pharmaceutical amoxicillin, Chem. Eng. J. 262 (2015) 286-294.
- 531 [20] S. Dbira, N. Bensalah, M.I. Ahmad, A. Bedoui, Electrochemical oxidation/disinfection
 532 of urine wastewaters with different anode materials, Materials 12 (2019) 1254.
- 533 [21] A. Thiam, I. Sirés, E. Brillas, Treatment of a mixture of food color additives (E122, E124
- and E129) in different water matrices by UVA and solar photoelectro-Fenton, Water Res.
- 535 81 (2015) 178-187.

- 536 [22] A. Thiam, R. Salazar, Fenton-based electrochemical degradation of metolachlor in
 537 aqueous solution by means of BDD and Pt electrodes: influencing factors and reaction
 538 pathways, Environ. Sci. Pollut. Res. 26 (2019) 2580-2591.
- 539 [23] C. Espinoza, J. Romero, L. Villegas, L. Cornejo-Ponce, R. Salazar, Mineralization of the
 540 textile dye Acid Yellow 42 by solar photoelectro-Fenton in a lab-pilot plant, J. Hazard.
 541 Mater. 319 (2016) 24-33.
- 542 [24] A.S. Fajardo, A.J. dos Santos, E.C.T. de Araújo Costa, D.R. da Silva, C.A. Martínez543 Huitle, Effect of anodic materials on solar photoelectro-Fenton process using a diazo dye
 544 as a model contaminant, Chemosphere 22 (2019) 880-889.
- 545 [25] L. Feng, E.D. van Hullebusch, M.A. Rodrigo, G. Esposito, M.A. Oturan, Removal of
 546 residual anti-inflammatory and analgesic pharmaceuticals from aqueous systems by
 547 electrochemical advanced oxidation processes. A review, Chem. Eng. J. 228 (2013) 944548 964.
- 549 [26] E. Brillas, I. Sirés, Electrochemical removal of pharmaceuticals from water streams:
 550 reactivity elucidation by mass spectrometry, TRAC Trend. Anal. Chem. 70 (2015) 112551 121.
- 552 [27] Y. Zhang, A. Wang, X. Tian, Z. Wen, H. Lv, D. Li, J. Li, Efficient mineralization of the
 antibiotic trimethoprim by solar assisted photoelectro-Fenton process driven by a
 photovoltaic cell, J. Hazard. Mater. 318 (2016) 319-328.
- G. Coria, T. Pérez, I. Sirés, E. Brillas, J.L. Nava, Abatement of the antibiotic levofloxacin
 in a solar photoelectro-Fenton flow plant: Modeling the dissolved organic carbon
 concentration-time relationship, Chemosphere 198 (2018) 174-181.
- 558 [29] C. Ridruejo, F. Centellas, P.L. Cabot, I. Sirés, E. Brillas, Electrochemical Fenton-based
 559 treatment of tetracaine in synthetic and urban wastewater using active and non-active
 560 anodes, Water Res. 128 (2018) 71-81.

- J.R.L.E. Freitas, F.J.O. Quintão, J.C.C. da Silva, S.Q.D. Silva, S.F. Aquino, R.J.C.F.
 Afonso, Characterisation of captopril photolysis and photocatalysis by-products in water
 by direct infusion, electrospray ionisation, high-resolution mass spectrometry and the
 assessment of their toxicities, Int. J. Environ. Anal. Chem. 97 (2017) 42-55.
- 565 [31] W.M. Mahmoud, K. Kümmerer, Captopril and its dimer captopril disulfide:
 566 Photodegradation, aerobic biodegradation and identification of transformation products

567 by HPLC-UV and LC-ion trap-MS(n), Chemosphere 88 (2012) 1170-1177.

- 568 [32] R. Salgado, R. Marques, J.P. Noronha, J.T. Mexia, G. Carvalho, A. Oehmen, M.A.M.
- Reis, Assessing the diurnal variability of pharmaceutical and personal care products in a
 full-scale activated sludge plant, Environ. Pollut. 159 (2011) 2359-2367.
- 571 [33] Y. Chen, X. Xi, G. Yu, Q. Cao, B. Wang, F. Vince, Y. Hong, Pharmaceutical compounds
 572 in aquatic environment in China: locally screening and environmental risk assessment,
 573 Front. Environ. Sci. Eng. 9 (2015) 394-401.
- 574 [34] M.J.A. Cortes-Diaz, J. Rodríguez-Flores, G. Castañeda-Peñalvo, M. Galar-Martínez, H.
 575 Islas-Flores, O. Dublán-García, L.M. Gómez-Oliván, Sublethal effects induced by
 576 captopril on *Cyprinus carpio* as determined by oxidative stress biomarker, Sci. Total
 577 Environ. 605-606 (2017) 811-823.
- 578 [35] G.P. Kapungu, G. Rukweza, T. Tran, W. Mbiya, R. Adigun, P. Ndungu, B. Martincigh,
 579 R.H. Simoyi, Oxyhalogen-sulfur chemistry: kinetics and mechanism of oxidation of
 580 captopril by acidified bromate and aqueous bromine, J. Phys. Chem. A 117 (2013) 2704581 2717.]
- 582 [36] A.M. Asiri, A.A.P. Khan, A. Khan, Spectroscopic investigation on kinetics and
 583 mechanistic aspects to electron-transfer process into quinolinium dichromate oxidation
 584 of a high blood pressure drug captopril in acidic medium, J. Mol. Liquids 203 (2015) 1585 6.

25

- [37] P. Du, H. Zhao, C. Liu, Q. Huang, H. Cao, Transformation and products of captopril with
 humic constituents during laccase-catalyzed oxidation: role of reactive intermediates,
 Water Res. 106 (2016) 488-495.
- 589 [38] S. Cotillas, E. Lacasa, C. Sáez, P. Cañizares, M.A. Rodrigo, Disinfection of urine by
- conductive-diamond electrochemical oxidation, Appl. Catal. B: Environ. 229 (2018) 6370.
- 592 [39] C. Flox, J.A. Garrido, R.M. Rodríguez, P.L. Cabot, F. Centellas, C. Arias, E. Brillas,
 593 Mineralization of herbicide mecoprop by photoelectro-Fenton with UVA and solar light,
 594 Catal. Today 129 (2007) 29-36.
- 595 [40] F.J. Welcher, Standard Methods of Chemical Analysis, sixth ed, vol. 2, RE Krieger
 596 Publishing Co, Huntington, New York (Part B), 1975.
- 597 [41] C. Carlesi Jara, S. Di Giulio, D. Fino, P. Spinelli, Combined direct and indirect
 598 electroxidation of urea containing water, J. Appl. Electrochem. 38 (2008) 915-922.

599

600 Figure captions

Fig. 1. Normalized concentration decay vs. electrolysis time for the AO-H₂O₂ degradation of 0.230 mM captopril in 130 mL of (a) a 0.050 M Na₂SO₄ solution, (b) urban wastewater and (c) synthetic urine, at pH 3.0 and 35 °C using a stirred undivided cell with different anodes (see legend in plot (a)) and a carbon-PTFE cathode, all of 3 cm² area, at a current density (*j*) of 33.3 mA cm⁻². The corresponding pseudo-first-order kinetic analysis is shown in plots (d), (e) and (f), respectively.

Fig. 2. Influence of current density (see legend in plot (a)) on the time course of the normalized
concentration decay for the AO-H₂O₂ treatment of 0.230 mM captopril in 130 mL of (a) a 0.050
M Na₂SO₄ solution, (b) urban wastewater and (c) synthetic urine, at pH 7.0 and 35 °C using a
BDD/air-diffusion cell. The corresponding pseudo-first-order kinetic analysis is shown in plots
(d), (e) and (f), respectively.

Fig. 3. Concentration of nitrogenated ions (see legend in plot (a)) after 60 min of AO-H₂O₂ treatment of 130 mL of 0.230 mM captopril in synthetic urine at different pH values using a (a) BDD, (b) Pt or (c) IrO₂ anode at j = 33.3 mA cm⁻². (d) Concentration of active chlorine determined under the same conditions.

Fig. 4. (a) Normalized concentration decay and (b) pseudo-first-order kinetic analysis for the treatment of 0.230 mM captopril in 2.5 L of a 0.050 M Na₂SO₄ solution at pH 3.0 and 35 °C using a solar pre-pilot plant with a Pt/air-diffusion cell at j = 50 mA cm⁻² and liquid flow rate of 180 L h⁻¹. Method: (•) AO-H₂O₂ (•) EF with 0.50 mM Fe²⁺ and (▲) SPEF with 0.50 mM Fe²⁺.

Fig. 5. Effect of *j* on the normalized captopril concentration abatement for the SPEF treatment of 0.230 mM drug in 2.5 L of a 0.050 M Na₂SO₄ solution with 0.50 mM Fe²⁺ at pH 3.0 and 35 °C using a solar pre-pilot flow plant with a Pt/air-diffusion cell at liquid flow rate of 180 L h⁻¹. 624 Current density: (\blacklozenge) 10 mA cm⁻², (\bullet) 30 mA cm⁻² and (\blacktriangle) 50 mA cm⁻². The pseudo-first-625 order kinetic analysis of concentration decays is depicted in the inset panel.

Fig. 6. Influence of the initial captopril content on the change of its concentration with electrolysis time for the SPEF treatment of 2.5 L of 0.050 M Na₂SO₄ solutions with 0.50 mM Fe²⁺ at pH 3.0 and 35 °C using a solar pre-pilot flow plant with a Pt/air-diffusion cell at j = 50mA cm⁻² and liquid flow rate of 180 L h⁻¹. Initial captopril content: (**1**) 0.100 mM, (**1**) 0.230 mM and (**1**) 0.460 mM. The inset panel presents the kinetic analysis of the concentration decays assuming a pseudo-first-order reaction.

Fig. 7. Influence of the aqueous matrix on the normalized captopril concentration decay during 632 the SPEF treatment of 2.5 L of 0.230 mM drug solutions with 0.50 mM Fe²⁺ at pH 3.0 and 35 633 °C using a solar pre-pilot flow plant with a Pt/air-diffusion cell at i = 50 mA cm⁻² and liquid 634 flow rate of 180 L h⁻¹. Matrix: (\blacktriangle) Urban wastewater, (\blacksquare) urine 1 (13.9 mM urea + 0.073 mM 635 uric acid + 0.367 mM creatinine), (\bigcirc) urine 2 (27.8 mM urea + 0.146 mM uric acid + 0.734 636 637 mM creatinine) and (\bigtriangledown) urine 3 (55.6 mM urea + 0.292 mM uric acid + 1.47 mM creatinine). 638 The inset panel shows the kinetic analysis of the above concentration decays assuming a 639 pseudo-first-order reaction.

Fig. 8. Time course of (a) TOC and (b) specific energy consumption per unit TOC mass for the
SPEF trials in (■) urine 1, (●) urine 2 and (▼) urine 3 shown in Fig. 7.

642 **Fig. 9**. Proposed route for captopril degradation by SPEF process in sulfate medium.



Fig. 1



Fig. 2



Fig. 3



Fig. 4



Fig. 5



Fig. 6



Fig. 7



Fig. 8





Table 1.

Percentage of degradation, pseudo-first-order rate constant for captopril decay and its *R*-squared, and percentage of TOC removal and specific energy consumption per unit TOC mass for the treatment of 130 mL of a 0.230 mM drug solution in different aqueous matrices and varying pH values at 35 °C using a stirred undivided cell with different anodes and an air-diffusion cathode.

Medium	pН	j	E_{cell}	% Degradation	k_1	R^2	% TOC	EC _{TOC}
		(mA cm ⁻²	²) (V)	(time (min))	$(10^{-2} \text{ min}^{-1})$		removal	(kWh (g TOC) ⁻¹
BDD anode								
$0.050 \text{ M} \text{ Na}_2 \text{SO}_4 ^{a}$	3.0	33.3	10.3	99 (120)	2.073	0.996	17	3.73
	7.0	16.7	6.2	80 (120)	1.181	0.986	6.9	2.76
		33.3	10.3	99 (120)	1.840	0.986	12	5.28
		66.7	13.9	100 (100)	2.196	0.998	23	7.41
	9.0	33.3	10.5	90 (120)	1.060	0.985	8.9	7.26
Urban wastewater ^b	3.0	33.3	12.1	96 (60)	6.684	0.982	_ d	
	7.0	16.7	6.8	82 (60)	2.954	0.988	14	0.504
		33.3	12.1	96 (60)	5.229	0.994	20	1.26
		66.7	13.6	99 (50)	8.891	0.995	27	2.09
	9.0	33.3	10.5	94 (60)	4.765	0.994	_ d	
Synthetic urine ^c	3.0	33.3	12.3	100 (60)	7.050	0.991	_ d	
	7.0	16.7	8.2	97 (60)	5.426	0.994	16	0.026
		33.3	12.3	100 (60)	7.124	0.993	23	0.055
		66.7	15.7	100 (50)	9.783	0.994	33	0.098
	9.0	33.3	12.4	83 (60)	2.923	0.996	_ d	
Pt anode								
0.050 M Na ₂ SO ₄ ^a	3.0	33.3	6.4	56 (120)	0.654	0.989	9.4	2.09
	7.0		7.4	49 (120)	0.568	0.988	5.4	4.22
	9.0		7.6	47 (120)	0.535	0.995	4.3	5.44
Urban wastewater ^b	3.0	33.3	6.9	98 (60)	5.687	0.988	_ d	
	7.0		7.1	93 (60)	4.434	0.994	16	0.922
	9.0		6.7	90 (60)	3.710	0.984	_ d	
Synthetic urine ^c	3.0	33.3	6.7	100 (60)	5.597	0.990	_ d	
	7.0		7.7	93 (60)	4.687	0.996	18	0.044
	9.0		7.8	66 (60)	1.815	0,985	_ d	
IrO ₂ anode								
0.050 M Na ₂ SO ₄ ^a	3.0	33.3	6.1	45 (120)	0.427	0.983	6.2	3.03
	7.0		6.6	41 (120)	0.440	0.993	3.5	5.80
	9.0		6.7	38 (120)	0.417	0.991	2.2	9.37
Urban wastewater ^b	3.0	33.3	6.4	91 (60)	3.290	0.981	_ d	
	7.0		6.5	83 (60)	3.019	0.987	13	1.04
	9.0		6.2	80 (60)	2.830	0.987	_ d	
Synthetic urine ^c	3.0	33.3	6.3	87 (60)	3.429	0.991	_ d	
	7.0		6.1	66 (60)	1.811	0.998	12	0.052

 9.0
 6.1
 43 (60)
 0.941
 0.989
 - d

 Electrolysis time and initial TOC: a 120 min and 25 mg L⁻¹, b 60 min and 37 mg L⁻¹, c 60 min and 745 mg L⁻¹

 d Not determined

Table 2.

Percentage of degradation, pseudo-first-order rate constant for captopril decay and its *R*-squared, and percentage of TOC removal and specific energy consumption per unit TOC mass for the treatment at 300 min of electrolysis for the treatment of 2.5 L of drug solutions in different aqueous matrices at pH 3.0 and 35 °C using a solar pre-pilot flow plant with a Pt/air-diffusion cell connected to a planar photoreactor at liquid flow rate of 180 L h⁻¹.

Medium	[Captopril] (mM)	<i>j</i> (mA cm ⁻²)	E_{cell} (V)	% Degradation (time (min))	k_1 (min ⁻¹)	R^2	% TOC removal	EC _{TOC} (kWh (g TOC) ⁻¹)	
$AO-H_2O_2$									
$0.050~M~Na_2SO_4{}^a$	0.230	50	11.3	36 (30)	0.015	0.993	3.1	30.1	
<i>EF</i> with 0.50 mM Fe^{2+}									
0.050 M Na ₂ SO ₄	0.230	50	10.9	100 (20)	0.326	0.993	17	5.11	
SPEF with 0.50 mM Fe^{2+}									
0.050 M Na ₂ SO ₄	0.100	50	11.3	100 (5)	1.128	0.992	33	6.14	
	0.230	10	4.2	100 (60)	0.083	0.994	16	0.425	
	0.230	30	7.3	100 (40)	0.135	0.995	20	1.75	
	0.230	50	10.5	100 (15)	0.452	0.995	25	3.36	
	0.460	50	10.8	100 (30)	0.186	0.995	19	2.27	
Urban wastewater ^b	0.230	50	24.7	100 (15)	0.551	0.990	28	7.06	
Urine 1 ^c	0.230	50	10.3	100 (15)	0.361	0.991	70	0.144	
Urine 2 ^d	0.230	50	10.1	100 (20)	0.265	0.994	44	0.116	
Urine 3 ^e	0.230	50	10.7	98 (30)	0.185	0.992	26	0.112	

Initial TOC: ^a 25 mg L⁻¹, ^b 37 mg L⁻¹, ^c 205 mg L⁻¹, ^d 385 mg L⁻¹, ^e 745 mg L⁻¹