## Taxonomic Note OpenAccess

International Journal of Systematic and Evolutionary Microbiology (2010), 60, 000-000

### Taxonomic Note

# The reference strain *Aeromonas hydrophila* CIP 57.50 should be reclassified as *Aeromonas salmonicida* CIP 57.50

David Miñana-Galbis, Maribel Farfán, J. Gaspar Lorén and M. Carmen Fusté

Correspondence
David Miñana-Galbis
davidminyana@ub.edu

Departament de Microbiologia i Parasitologia Sanitàries, Facultat de Farmácia, Universitat de Barcelona, Av. Joan XXIII s/n, 08028 Barcelona, Spain

The use of reference strains is a critical element for the quality control of different assays, from the development of molecular methods to the evaluation of antimicrobial activities. Most of the strains used in these assays are not type strains and some of them are cited erroneously because of subsequent reclassifications and descriptions of novel species. In this study, we propose that the reference strain *Aeromonas hydrophila* CIP 57.50 be reclassified as *Aeromonas salmonicida* CIP 57.50 based on phenotypic characterization and sequence analyses of the *cpn60*, *dnaJ*, *gyrB* and *rpoD* genes.

The selection and use of reference strains that are genetically stable is a critical element for achieving quality control goals in antimicrobial susceptibility testing. The reclassification of recognized species and the description of novel species can make it difficult to keep track of the most up to date taxonomic opinion and can lead to confusing citations. As an example, the quality control strain ATCC 43055 can be cited either as Eggerthella lenta or Eubacterium lentum and both names have been validly published. However, the most recent taxonomic opinion places this strain as a member of the genus Eggerthella (Wade et al., 1999). This problem can also occur at an interspecific level if the strain used for quality control is not the type strain, for example, Aeromonas hydrophila ATCC 7965 (=NCTC 7812=CIP 57.50) is a reference strain frequently used to assess antimicrobial susceptibilities, the sensitivity and specificity of PCR protocols and stress responses, among other assays (Isonhood et al., 2002; Huddleston et al., 2006; Skočibušić et al., 2006; Al Amri et al., 2007; Giddens & Bean, 2007; Gordon et al., 2008).

Over the past few years, the taxonomy of the genus *Aeromonas* has became increasingly complex due to the description of several novel species and the rearrangement of species described thus far (Miñana-Galbis *et al.*, 2007; Demarta *et al.*, 2008; Beaz-Hidalgo *et al.*, 2009). An

The GenBank/EMBL/DDBJ accession numbers for the 16S rRNA, cpn60, dnaJ, gyrB and rpoD gene sequences of A. hydrophila CIP 57.50 are FJ936134, FJ936135, FJ936136, FJ936137 and FJ936138, respectively.

The alignments and the distance matrix for the cpn60, dnaJ, gyrB and rpoD gene sequences of A. hydrophila strain CIP 57.50 with the type strains of A. bestiarum, A. hydrophila and A. salmonicida are available with the online version of this paper.

example of this complexity is the difficulty in discriminating between the phenotypically and genetically closely related *A. hydrophila*, *Aeromonas bestiarum* and the mesophilic strains of *Aeromonas salmonicida* (Miñana-Galbis *et al.*, 2002; Martínez-Murcia *et al.*, 2005). Nevertheless, alternative methods for species delineation, e.g. multilocus enzyme electrophoresis (MLEE) and housekeeping gene sequencing, have clarified most of the controversial taxa found in the genus *Aeromonas* (Miñana-Galbis *et al.*, 2004, 2009; Soler *et al.*, 2004; Nhung *et al.*, 2007).

In our laboratory, we have performed an extensive phenotypic characterization, PCR amplification and sequencing of the 16S rRNA, cpn60, dnaJ, gyrB and rpoD genes for A. hydrophila strain CIP 57.50 (=ATCC 7965=NCTC 7812) using previously described methods (Soler et al., 2004; Miñana-Galbis et al., 2007, 2009; Nhung et al., 2007). A. hydrophila strain CIP 57.50, a mesophilic, motile and non-pigmented strain, was phenotypically identified as A. salmonicida after testing positive for sorbitol fermentation and negative for lactate assimilation. These results contradicted those expected for a strain belonging to A. hydrophila (Miñana-Galbis et al., 2002, 2007). Nevertheless, the strain could not be assigned to any of the subspecies of A. salmonicida since most of these are psychrophilic and non-motile (A. salmonicida subsp. achromogenes, A. salmonicida subsp. masoucida, A. salmonicida subsp. salmonicida and A. salmonicida subsp. smithia). A. salmonicida subsp. pectinolytica, the only mesophilic subspecies described to date, is non-motile, pigmented and negative for the lysine decarboxylase test and for aesculin hydrolysis, opposite characteristics to those found for A. hydrophila strain CIP 57.50 (Pavan et al., 2000; Martin-Carnahan & Joseph, 2005).

1

**Table 1.** Sequence distances (%) between A. hydrophila strain CIP 57.50 and the type strains of A. bestiarum, A. hydrophila and A. salmonicida

Sequence alignments and distances (Jukes-Cantor model and pairwise deletion option) were calculated with MEGA4 software.

Type strains	16S rRNA gene		cpn60		dnaJ		gyrB		rpoD	
	%	GenBank nos	%	GenBank nos	%	GenBank nos	%	GenBank nos	%	GenBank nos
A. bestiarum	0.0	X60406	6.0	EU306796	6.3	AB280554	3.0	AY101774	7.5	AY169326
A. hydrophila subsp. hydrophila	1.1	X60404	7.0	EU306804	10.2	AB280561	7.7	AY101778	11.8	AY169325
A. hydrophila subsp. ranae	1.2	AJ508766	8.0	EU306805	11.0	AB280562	8.6	AM262162	10.2	EF465509
A. salmonicida subsp. achromogenes	0.1	X60407	1.1	EU306824	1.8	AB280568	1.3	AY101785	2.3	AY169329
A. salmonicida subsp. masoucida	0.1	X74680	1.1	EU306825	1.7	AB280569	1.2	AY101786	2.4	AY169330
A. salmonicida subsp. pectinolytica	0.1	AF134065	1.3	EU306827	2.4	AB280570	1.9	AY101810	2.3	AY169324
A. salmonicida subsp. salmonicida	0.0	X60405	1.1	EU306828	1.9	AB280571	1.5	AY101773	2.5	AY169327
A. salmonicida subsp. smithia	0.1	AB027544	1.1	EU306829	1.8	AB280572	1.4	AM262159	2.4	AY169331

The 16S rRNA gene sequence of *A. hydrophila* strain CIP 57.50 (GenBank accession no. FJ936134) was identical to those of *A. bestiarum* CIP 74.30<sup>T</sup> and *A. salmonicida* subsp. *salmonicida* NCIMB 1102<sup>T</sup> and showed a single nucleotide difference with respect to *A. salmonicida* subsp. *pectinolytica* 34mel<sup>T</sup>. Differences of two nucleotides were found when strain CIP 57.50 was compared with the type strains of the other subspecies of *A. salmonicida*. Distance values >1% (16–17 nucleotide differences) were obtained between the 16S rRNA gene sequence of *A. hydrophila* CIP 57.50 and those of the type strains of the two subspecies of *A. hydrophila* (Table 1).

In contrast, the sequencing of housekeeping genes allowed us to identify A. hydrophila strain CIP 57.50 to the species level. Previous studies (Soler et al., 2004; Nhung et al., 2007; Miñana-Galbis et al., 2009) have established the following intraspecific threshold values based on divergences of gene sequences: 3.5 % for cpn60, 3.3 % for dnaJ, 2.3% for gyrB and 2.6% for rpoD. The sequences of the cpn60, dnaJ, gyrB and rpoD genes in A. hydrophila strain CIP 57.50 (GenBank accession nos FJ936135, FJ936136, FJ936137 and FJ936138, respectively) showed sequence distances below the interspecific threshold values with respect to those of the type strain of A. salmonicida, while the values were clearly higher than those of the type strains of A. hydrophila and A. bestiarum (Table 1). Moreover, A. hydrophila strain CIP 57.50 could be distinguished from all of the subspecies of A. salmonicida, showing a distance range of 1.1-2.5 % depending on the gene or subspecies analysed (Table 1). The alignments and the distance matrix for the cpn60, dnaJ, gyrB and rpoD gene sequences are available as supplementary material in IJSEM Online.

Although a low discriminatory power between highly related species of the genus *Aeromonas* is obtained from 16S rRNA gene sequence analysis, it has been demonstrated that sequencing of housekeeping genes is an effective method for discriminating between species of the genus *Aeromonas* (Küpfer *et al.*, 2006; Saavedra *et al.*, 2006;

Miñana-Galbis *et al.*, 2009). The results obtained in this study strongly suggest that the reference strain *A. hydrophila* CIP 57.50 (=ATCC 7965=NCTC 7812) belongs to the species *A. salmonicida* rather than *A. hydrophila*.

In order to avoid other possible misclassifications, it would be useful if the reference strains deposited in culture collections were updated and checked using the latest identification tools since 16S rRNA gene sequencing is often insufficient for the identification of species. Additionally, considering the numbers of descriptions of novel species published each year, the number of key phenotypic tests used to identify isolates at the species level should be increased.

### **Acknowledgements**

This work has been supported by a grant from the Ministerio de Ciencia e Innovación, Spain (CGL2008–03281/BOS).

#### References

Al Amri, A., Senok, A. C., Ismaeel, A. Y., Al-Mahmeed, A. E. & Botta, G. A. (2007). Multiplex PCR for direct identification of *Campylobacter* spp. in human and chicken stools. *J Med Microbiol* 56, 1350–1355.

Beaz-Hidalgo, R., Alperi, A., Figueras, M. J. & Romalde, J. L. (2009). *Aeromonas piscicola* sp. nov., isolated from diseased fish. *Syst Appl Microbiol* 32, 471–479.

Demarta, A., Küpfer, M., Riegel, P., Harf-Monteil, C., Tonolla, M., Peduzzi, R., Monera, A., Saavedra, M. J. & Martinez-Murcia, A. (2008). *Aeromonas tecta* sp. nov., isolated from clinical and environmental sources. *Syst Appl Microbiol* 31, 278–286.

**Giddens, S. R. & Bean, D. C. (2007).** Investigations into the in vitro antimicrobial activity and mode of action of the phenazine antibiotic D-alanylgriseoluteic acid. *Int J Antimicrob Agents* **29**, 93–97.

Gordon, K. V., Vickery, M. C., DePaola, A., Staley, C. & Harwood, V. J. (2008). Real-time PCR assays for quantification and differentiation of *Vibrio vulnificus* strains in oysters and water. *Appl Environ Microbiol* 74, 1704–1709.

- **Huddleston, J. R., Zak, J. C. & Jeter, R. M. (2006).** Antimicrobial susceptibilities of *Aeromonas* spp. isolated from environmental sources. *Appl Environ Microbiol* **72**, 7036–7042.
- Isonhood, J. H., Gerard, P., Leenanon, B. & Drake, M. A. (2002). Stress response of *Aeromonas hydrophila* following environmental challenges. *Food Microbiol* 19, 285–293.
- Küpfer, M., Kuhnert, P., Korczak, B. M., Peduzzi, R. & Demarta, A. (2006). Genetic relationships of *Aeromonas* strains inferred from 16S rRNA, *gyrB* and *rpoB* gene sequences. *Int J Syst Evol Microbiol* 56, 2743–2751.
- Martin-Carnahan, A. & Joseph, S. W. (2005). Genus I. Aeromonas Stanier 1943, 213<sup>AL</sup>. In *Bergey's Manual of Systematic Bacteriology*, 2nd edn, vol. 2, part B, pp. 557–578. Edited by G. M. Garrity, D. J. Brenner, N. R. Krieg & J. T. Staley. New York: Springer.
- Martínez-Murcia, A. J., Soler, L., Saavedra, M. J., Chacón, M. R., Guarro, J., Stackebrandt, E. & Figueras, M. J. (2005). Phenotypic, genotypic, and phylogenetic discrepancies to differentiate *Aeromonas salmonicida* from *Aeromonas bestiarum*. *Int Microbiol* 8, 259–269.
- Miñana-Galbis, D., Farfán, M., Lorén, J. G. & Fusté, M. C. (2002). Biochemical identification and numerical taxonomy of *Aeromonas* spp. isolated from environmental and clinical samples in Spain. *J Appl Microbiol* 93, 420–430.
- Miñana-Galbis, D., Farfán, M., Fusté, M. C. & Lorén, J. G. (2004). Genetic diversity and population structure of *Aeromonas hydrophila*, *Aer. bestiarum*, *Aer. salmonicida* and *Aer. popoffii* by multilocus enzyme electrophoresis (MLEE). *Environ Microbiol* 6, 198–208.
- Miñana-Galbis, D., Farfán, M., Fusté, M. C. & Lorén, J. G. (2007). *Aeromonas bivalvium* sp. nov., isolated from bivalve molluscs. *Int J Syst Evol Microbiol* 57, 582–587.

- Miñana-Galbis, D., Urbizu-Serrano, A., Farfán, M., Fusté, M. C. & Lorén, J. G. (2009). Phylogenetic analysis and identification of *Aeromonas* species based on sequencing of the *cpn60* universal target. *Int J Syst Evol Microbiol* 59, 1976–1983.
- Nhung, P. H., Hata, H., Ohkusu, K., Noda, M., Shah, M. M., Goto, K. & Ezaki, T. (2007). Use of the novel phylogenetic marker *dnaJ* and DNA-DNA hybridization to clarify interrelationships within the genus *Aeromonas*. *Int J Syst Evol Microbiol* 57, 1232–1237.
- Pavan, M. E., Abbott, S. L., Zorzópulos, J. & Janda, J. M. (2000). *Aeromonas salmonicida* subsp. *pectinolytica* subsp. nov., a new pectinase-positive subspecies isolated from a heavily polluted river. *Int J Syst Evol Microbiol* **50**, 1119–1124.
- Saavedra, M. J., Figueras, M. J. & Martínez-Murcia, A. J. (2006). Updated phylogeny of the genus *Aeromonas*. *Int J Syst Evol Microbiol* 56, 2481–2487.
- Skočibušić, M., Bezić, N. & Dunkić, V. (2006). Phytochemical composition and antimicrobial activities of the essential oils from *Satureja subspicata* Vis. growing in Croatia. *Food Chem* **96**, 20–28.
- Soler, L., Yáñez, M. A., Chacón, M. R., Aguilera-Arreola, M. G., Catalán, V., Figueras, M. J. & Martínez-Murcia, A. J. (2004). Phylogenetic analysis of the genus *Aeromonas* based on two housekeeping genes. *Int J Syst Evol Microbiol* 54, 1511–1519.
- Wade, W. G., Downes, J., Dymock, D., Hiom, S. J., Weightman, A. J., Dewhirst, F. E., Paster, B. J., Tzellas, N. & Coleman, B. (1999). The family *Coriobacteriaceae:* reclassification of *Eubacterium exiguum* (Poco et al. 1996) and *Peptostreptococcus heliotrinreducens* (Lanigan 1976) as *Slackia exigua* gen. nov., comb. nov. and *Slackia heliotrinireducens* gen. nov., comb. nov., and *Eubacterium lentum* (Prevot 1938) as *Eggerthella lenta* gen. nov., comb. nov. *Int J Syst Bacteriol* 49, 595–600.

http://ijs.sgmjournals.org