# Somatostatin, an *In Vivo* Binder to Aβ Oligomers, Binds to $\beta PFO_{A\beta(1-42)}$ Tetramer

Eduard Puig,[1,2*] James Tolchard,[2] Antoni Riera,[1,3*] Natàlia Carulla[1,2*]

[1]Institute for Research in Biomedicine (IRB Barcelona), The Barcelona Institute of Science and Technology (BIST), Baldiri Reixac 10, 08028 Barcelona, Spain

[2]CBMN (UMR 5248), University of Bordeaux – CNRS – IPB, Institut Européen de Chimie et Biologie, 2 rue Escarpit, 33600, Pessac, France

[3]Departament de Química Inorgànica i Orgànica. Universitat de Barcelona, Martí i Franqués 1, 08028, Barcelona, Spain

[*]Corresponding authors:

Eduard Puig, e-mail: eduard.puig@irbbarcelona.org

Natàlia Carulla, e-mail: natalia.carulla@gmail.com

## Abstract

Somatostatin (SST14) is strongly related to Alzheimer's disease (AD), as its levels decline during aging, it regulates the proteolytic degradation of the amyloid beta peptide (Aβ), and it binds to Aβ oligomers *in vivo*. Recently, the 3D structure of a membrane-associated β-sheet pore forming tetramer ($\beta PFO_{A\beta(1-42)}$ tetramer) has been reported. Here we show that SST14 binds selectively to the $\beta PFO_{A\beta(1-42)}$ tetramer with an a $K_D$ of ~40 μM without binding to monomeric Aβ(1-42). Specific NMR chemical shift perturbations, observed during titration of SST14, define a binding site in the $\beta PFO_{A\beta(1-42)}$ tetramer and are in agreement with a 2:1 stoichiometry determined by both native MS and ITC. These results enabled us to perform driven docking and model the binding mode for the interaction. The present study provides additional evidence on the relation between SST14 and the amyloid cascade, as well as positions the $\beta PFO_{A\beta(1-42)}$ tetramer as a relevant aggregation form of Aβ and as a potential target for AD.

## Introduction

Increased levels of amyloid beta peptide (Aβ) and deposition of amyloid fibrils in neuronal cells constitute a critical part of the etiopathogenesis in Alzheimer's disease (AD).[1,2] Aβ originates from the sequential cleavage of the amyloid precursor protein (APP) by the β-secretase in the extracellular space and the $\gamma$-secretase in the transmembrane domain.[3] In solution, this hydrophobic peptide aggregates in a nucleation-dependent manner into soluble oligomers[4] that gradually increase in molecular-weight until insoluble fibrils are formed [5-8]. The presence of amyloid fibrils in the extracellular space has inevitably drawn research interest in Aβ peptides to this location. However, the fact that Aβ's origin lies within APP, a transmembrane protein,[9] together with numerous reported work of Aβ interacting with the cellular membrane,[10-15] strongly suggests this environment as an alternative location for Aβ accumulation and aggregation.

We have previously studied the aggregation of Aβ within detergent micelles to mimic the membrane environment and reported the preparation[16] and the three-dimensional (3D) structure[17] of a membrane-associated β-sheet pore forming tetramer ($\beta PFO_{A\beta(1-42)}$ tetramer) (Figure S1). Interestingly, the formation of this oligomer was specific for Aβ(1-42), the variant most related to AD but not Aβ(1-40) which is the variant most abundant in the brain. $\beta PFO_{A\beta(1-42)}$ tetramer comprises a β-sheet core formed by six β-strands. Molecular dynamics showed that when $\beta PFO_{A\beta(1-42)}$ tetramer incorporated into lipid bilayers and water molecules were able to permeate the membrane through the hydrophilic edges of the β-sheet core tetramer. This work not only represented the resolution of the first 3D structure of an Aβ oligomer but also the definition of a new mechanism of membrane disruption that could explain the neurotoxic activity of Aβ oligomers in the context of AD.

The screening of potential interactors constitutes an essential step to better understand the function of Aβ and its implication in AD. Schmitt-Ulms *et al*. recently performed an extensive

screening of proteins that bound to Aβ oligomers in human brain extracts.[18] From over 50 proteins detected, somatostatin (SST14) stood out for delaying Aβ aggregation and binding specifically to Aβ oligomers. To the best of our knowledge, the aforementioned work represents the largest Aβ monomeric and oligomeric *in vivo* interactome performed so far. The authors suggested that further investigations should be performed to improve the understanding of the SST14-Aβ interaction. In the present work, we used well established biophysical techniques to assess whether the specific Aβ oligomer binder, somatostatin-14 (SST14) bound to the $\beta PFO_{A\beta(1-42)}$ tetramer.

SST14 is a cyclic tetradecapeptide that is produced in neuroendocrine cells in the hypothalamus as well as in other tissues, including pancreas, intestinal tract and regions of the central nervous system.[19,20] In a clinical context, SST14 is the neuropeptide most highly depleted in both the brain and cerebrospinal fluid (CSF) of AD patients.[21,22] The relation to AD was further described by the work of Saido *et al.* as they found that SST14 regulates the metabolism of Aβ in the brain through the modulation of neprilysin which catalyses its proteolytic degradation.[23] Moreover, a positive correlation between SST14 and Aβ(1-42) levels was established in the CSF of elderly patients with mild cognitive impairment.[24] Undoubtedly, previous work in the literature has established a strong link between SST14, Aβ and AD. Therefore, studying the potential interaction between SST14 and $\beta PFO_{A\beta(1-42)}$ tetramer could deliver evidence to better understand the role of SST14 in the context of AD and point towards the relevance of $\beta PFO_{A\beta(1-42)}$ tetramer in a biological context.

## Results

### SST14 coelutes with βPFO$_{Aβ(1-42)}$ tetramers

We initially relied on size exclusion chromatography (SEC)[25-27] to characterize the potential interaction between SST14 and βPFO$_{Aβ(1-42)}$ tetramer in a membrane mimicking environment. As control samples, we followed the evolution of βPFO$_{Aβ(1-42)}$ tetramer formation and SST14 independently, following its incubation in the dodecyl phosphocholine (DPC) solution used as a membrane mimicking environment. Analysis of both samples showed that βPFO$_{Aβ(1-42)}$ tetramers and SST14 eluted, respectively, at 13.5 mL and 17.5 mL (Supporting information; Figure S2A, B). In the case of the SST14 sample, its evolution over time revealed the appearance of wide peaks near the void volume, which were attributed to aggregated forms as previously reported for this peptide.[28] Coincubation of Aβ(1-42) with SST14 under conditions that lead to βPFO$_{Aβ(1-42)}$ tetramer formation resulted in an increase of 65% of the area under the peak assigned to βPFO$_{Aβ(1-42)}$ tetramers (Figure 1A). Such a change could be explained either due to an increase in the formation of βPFO$_{Aβ(1-42)}$ tetramer or the binding of SST14. Interestingly, we did not observe precipitates when both peptides were coincubated suggesting that the interaction between them increased the stability of SST14 in a membrane-mimicking environment. To assess whether binding occurred specifically during βPFO$_{Aβ(1-42)}$ tetramer formation, we first incubated Aβ(1-42) alone for 24 h under conditions that lead to βPFO$_{Aβ(1-42)}$ tetramer formation and then added SST14. Analysis of this sample by SEC resulted in a 20% increase of the area under the peak assigned to βPFO$_{Aβ(1-42)}$ tetramer (Supporting information, Figure S2D). This result suggested that SST14 binding was not exclusively occurring during βPFO$_{Aβ(1-42)}$ tetramer formation but also when putting in contact the two binding partners after the oligomer was assembled.

## SST14 interacts with the Aβ(1-42) tetramer at a 2:1 ratio

To further study the interaction between $\beta PFO_{A\beta(1-42)}$ tetramer and SST14, we used isothermal titration calorimetry (ITC) to measure the heat exchange and obtain information about the energetic profile of the binding event. Titration of SST14 to the $\beta PFO_{A\beta(1-42)}$ tetramer showed an exothermic interaction with a $K_D$ of ~40 µM and a stoichiometry of approximately 2:1 (SST14 to $\beta PFO_{A\beta(1-42)}$ tetramer) (Figure 1B, C). Such a binding ratio would indeed be in agreement with the symmetric structure of the $\beta PFO_{A\beta(1-42)}$ tetramer. However, we could not exclude the possibility of SST14 interacting with remaining monomeric Aβ(1-42) in the sample.

To better understand the specificity and stoichiometry of the interaction we analyzed the sample using native mass spectrometry (MS). This technique uses non-denaturing conditions to prepare the sample and soft ionization methods (such as electrospray ionization (ESI)) to preserve the non-covalent interactions within ($\beta PFO_{A\beta(1-42)}$ tetramer) and between ($\beta PFO_{A\beta(1-42)}$ tetramer-SST14) molecular complexes.[29,30] To prepare the $\beta PFO_{A\beta(1-42)}$ tetramer sample for MS analysis, we used lauryldimethylamine *N*-oxide (LDAO) instead of DPC to mimic the membrane environment as this detergent also supports $\beta PFO_{A\beta(1-42)}$ tetramer formation and is compatible with MS analysis.[31] Direct infusion of the resulting sample using nanoESI-MS delivered a clean spectrum displaying four consecutive charge states for the tetramer (+3, +4, +5 and +6) confirming that it was the major species in the sample (Figure 2A; Supporting information, Table S2). Infusion of a $\beta PFO_{A\beta(1-42)}$ tetramer sample prepared in the presence of SST14 revealed consecutive charge states that were assigned to one (+3, +4 and +5) and two (+4, +5 and +6) SST14 molecules bound to the $\beta PFO_{A\beta(1-42)}$ tetramer (Figure 2B; Supporting information, Table S2). Both ITC (Figure 1C) and native MS (Figure 2B) data pointed towards a 2:1 ratio for SST14 and $\beta PFO_{A\beta(1-42)}$ tetramer interaction. Moreover, we did not observe any consecutive charge states corresponding to monomeric Aβ(1-42) bound to one or two SST14

molecules in agreement with SST14 binding specifically to oligomeric forms of Aβ (Supporting information; Figure S3, Table S1).

### SST14 binds to the flexible edges of the Aβ(1-42) tetramer

The results obtained by SEC indicated that the binding event was stable over 24 h in the membrane-mimicking environment (Supporting information, Figure S2C), which encouraged us to further study the interaction by solution NMR. We therefore decided to pursue a deeper characterization by titrating SST14 into a $^{15}$N-βPFO$_{Aβ(1-42)}$ tetramer sample and perform 2D [$^{1}$H,$^{15}$N]-SOFAST-HMQC experiments over time. The βPFO$_{Aβ(1-42)}$ tetramer consists of a six-stranded β-sheet comprising two types of Aβ(1-42) subunits referred to orange and green, respectively (Supporting information, Figure S1). The orange subunit contributes with two β-strands (β1 and β2) and the green subunit contributes with one β-strand (β3) and a small α-helix (α1).[17]

The spectrum for βPFO$_{Aβ(1-42)}$ tetramer displayed a well-dispersed set of signals as previously reported for this sample (Figure 3).[16] Upon addition of SST14 to the NMR sample, several changes in chemical shifts were observed in the resulting spectrum indicating that SST14 bound to βPFO$_{Aβ(1-42)}$ tetramer. Indeed, specific shift changes in residues V12, F20, V24 G29, V40 and A42 of the orange subunit and in residues V12, V18, A21, E22, D23, G29, I41 and A42 of the green subunit of the βPFO$_{Aβ(1-42)}$ tetramer were observed (Figure 4A; Supporting information, Figure S4; Source data 1). Chemical Shift Perturbations (CSPs) were considered significant if the values were greater than one standard deviation ($\sigma$) of the Euclidean chemical shift change represented as a grey dashed line (Figure 4A, Source data 1).[32] Both the observation of smooth peak migrations between the free and bound states, and the derivation of $k_{off}$ rates using the maximum change in chemical shift, indicated that somatostatin binding occurred in the fast-exchange regime (Figure 3C).[32,33] $K_D$ rates were determined for each clearly tractable residue by fitting the change in chemical shift as a function of the ligand

concentration. The tightest binding residues (Y10, F20) exhibited $K_D$'s of ~35 and ~52 μM, respectively.

The CSPs induced by SST14 were represented in the $\beta PFO_{A\beta(1-42)}$ tetramer 3D structure (PDB code 6RHY) where the amide protons of the affected residues are represented as red spheres (Figure 4B). The affected residues showed to be close in space and defined a specific binding site within the tetramer structure. These CSPs were used to perform driven docking of SST14 with the $\beta PFO_{A\beta(1-42)}$ tetramer structure using the high ambiguity driven docking approach (HADDOCK).[34] The best-scoring structural clusters that we obtained suggested a binding mode of SST14 in the $\beta PFO_{A\beta(1-42)}$ tetramer where the peptide interacted with the flexible edges of the tetramer and interestingly also tightly with the alpha helix of the green subunit (Figure 4C).

The binding site defined in our study, enabled us to rationalize the specificity of the interaction between both entities since the spatial distribution of the residues in the $\beta PFO_{A\beta(1-42)}$ tetramer is completely different to that of monomeric Aβ(1-42). The localization of the site was also in accordance with the 2:1 stoichiometry of the binding observed by ITC and native MS. Indeed, the symmetric topology of the tetramer contains two possible binding sites in the superior and inferior flexible ends that are solvent-exposed and thus, accessible to SST14.

## Discussion

In summary, our findings show that SST14 binds to $\beta PFO_{A\beta(1-42)}$ tetramers with an affinity in the low micromolar range. Native MS experiments prove the binding to be specific for this oligomeric form with a 2:1 (SST14 to $\beta PFO_{A\beta(1-42)}$ tetramer) stoichiometry, in accordance with ITC data. Our NMR experiments reveal two symmetric binding sites near the flexible ends of the tetramer. Restraint-driven *in silico* docking enables us to propose a binding mode of SST14 to the tetramer structure. Altogether, we conclude that SST14, an *in vivo* binder to Aβ oligomers,[18] specifically binds to $\beta PFO_{A\beta(1-42)}$ tetramer.

We observed an important difference when comparing our results with previously reported work on the binding of SST14 to soluble oligomers of Aβ(1-42).[18] Indeed, work by Schmitt-Ulms and collaborators postulated that SST14 did not bind to Aβ(17-42) oligomers which led them to conclude that the N-terminus was involved in the binding site. Our data, on the contrary, suggests that residues 18-29 are mainly involved in the binding with special emphasis on the ones forming the short alpha helix, residues L17 to F20. We recently showed that while Aβ(1-42) incorporates both as the orange and green subunit in the $\beta PFO_{A\beta(1-42)}$ tetramer arrangement, Aβ(17-42) only incorporates as the green subunit,[17] which prevents Aβ(17-42) to form $\beta PFO_{A\beta(1-42)}$ tetramer by itself. Thus, for $\beta PFO_{A\beta(1-42)}$ tetramer to form it is required that at least 50% of the peptides contain the N-terminus. These results evidence that using shortened versions of proteins can have a huge impact in protein self-assembly and structure. Moreover, work by Schmitt-Ulms *et al.* was performed on soluble oligomers while ours on a membrane-associated oligomer. Therefore, the binding to SST14 may be different for each oligomer type. The authors also emphasized the importance of W8 of SST14 for the binding to occur. Indeed, this residue has been described to play an important role in the activity of the peptide when binding SSTRs.[35]

Whilst our CSP-driven docking did not converge to a singular configuration for the SST14: $\beta PFO_{A\beta(1-42)}$ tetramer interaction, a frequent observation across the best performing poses was that SST14 docks into the concave interior at the solvent-exposed edges of the $\beta PFO_{A\beta(1-42)}$ tetramer (Figure 4C, Figure S5). In many of our calculated models, W8 of SST14 was found to interact with the hydrophobic residues of the $\beta PFO_{A\beta(1-42)}$ tetramer, suggesting that W8 of SST14 could possibly mediate an interaction with the membrane-associated Aβ(1-42) tetramer.

In the AD context, the critical role of SST14 in the metabolism of Aβ(1-42) through the regulation of neprilysin[23] inevitably points towards the potential degradation of oligomeric forms of Aβ(1-42). Two important conclusions of the work by Saido and collaborators are the location of the SST14-neprylisin interaction situated near or in the cellular membrane and the fact that somatostatin-regulated neprilysin activity selectively depleted Aβ(1-42) but not Aβ(1-40). Interestingly, the aforementioned facts also apply to $\beta PFO_{A\beta(1-42)}$ tetramer since it is able to incorporate into membranes[17] and is exclusively formed by Aβ(1-42) but not Aβ(1-40).[16] Proteolytic activities are tightly controlled biological processes that can be regulated at different levels such as through the formation of an activation complex.[36] Therefore, we cannot exclude that binding of SST14 to the $\beta PFO_{A\beta(1-42)}$ tetramer could induce its degradation.

In the present study, we show at a structural level how SST14, which has been reported to bind to Aβ oligomers in human brain extracts, also binds to $\beta PFO_{A\beta(1-42)}$ tetramer. We think these results strengthen the relation of SST14 with the amyloid cascade and due to the clear implication of SST14 in AD, it positions $\beta PFO_{A\beta(1-42)}$ tetramer as relevant oligomer form of Aβ and as a potential target for AD.

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## Competing Interests

The authors declare no competing financial interests.

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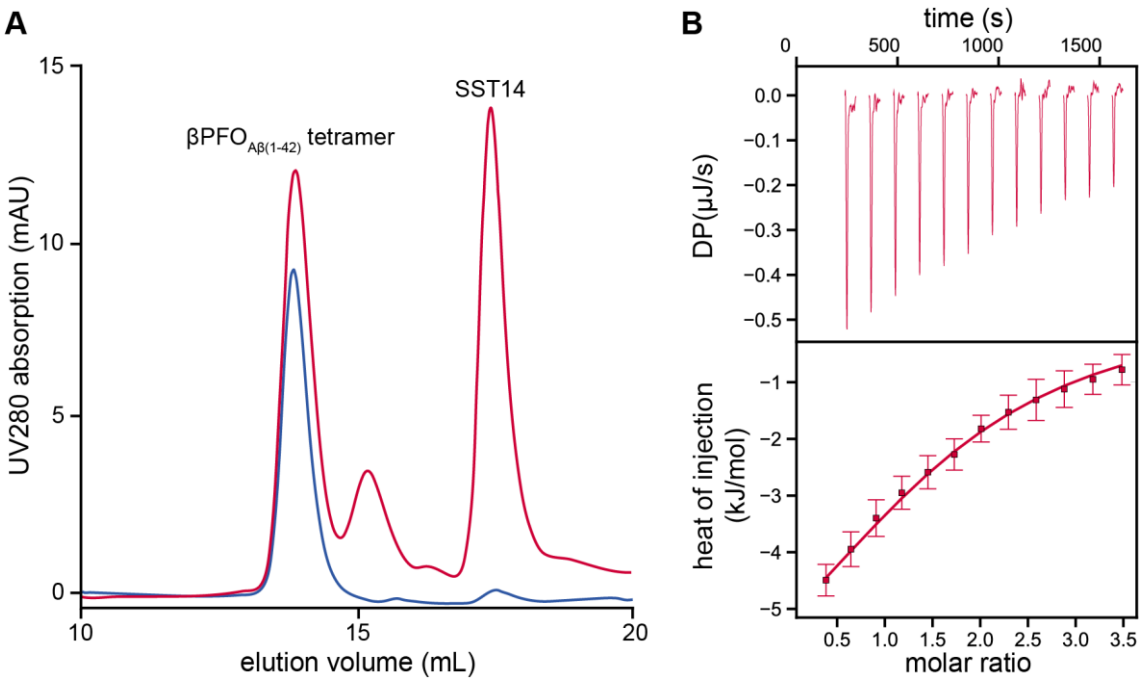
## Figures and Figure Legends

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| Syringe | Cell | $K_d$, μM | ΔH, kcal/mol | -TΔS, kcal/mol | ΔG, kcal/mol | n |
|---|---|---|---|---|---|---|
| SST14 | βPFO | 39.36 ± 23.20 | -2.26 ± 0.14 | -3.82 ± 0.25 | -6.08 ± 0.35 | 0.53 ± 0.11 |

**Figure 1.** SST14 binding to βPFO$_{Aβ(1-42)}$ tetramer assessed by SEC and ITC. (A) SEC elution profile for βPFO$_{Aβ(1-42)}$ tetramer after 24 hours of its formation in the absence (blue) and in the presence of SST14 (red). The peaks have been labeled with the elution volume of βPFO$_{Aβ(1-42)}$ tetramer and SST14. (B) ITC thermogram (top) and analysis of the fitted binding isotherm (bottom) for βPFO$_{Aβ(1-42)}$ tetramer titrated with SST14. Standard deviation values were obtained from three independent replicates. (C) Thermodynamic binding parameters of the interaction determined from ITC experiments at 25°C and pH 9.0.

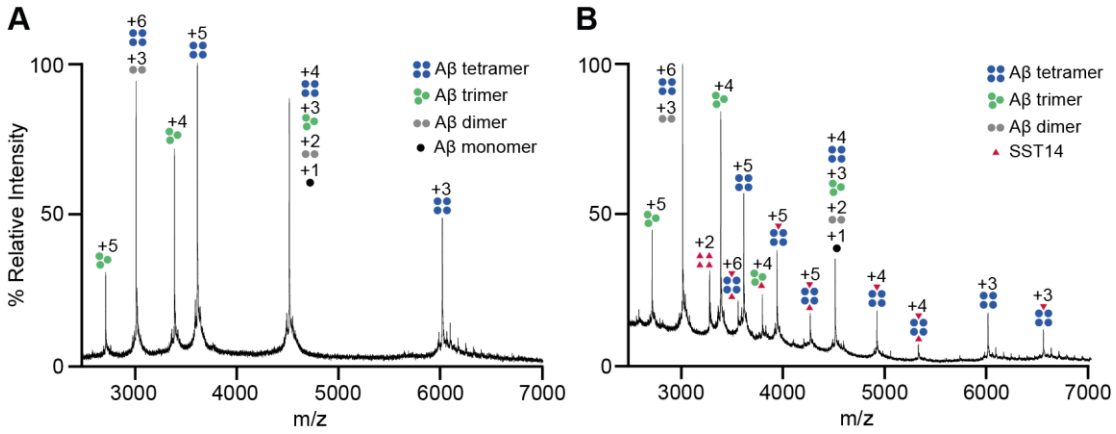


**Figure 2.** SST14 binding to $\beta PFO_{A\beta(1\text{-}42)}$ tetramer assessed by native MS. (A) Electrospray ionization MS (ESI-MS) spectrum of $\beta PFO_{A\beta(1\text{-}42)}$ tetramer (150 μM Aβ(1-42), 7.2 mM LDAO, 200 mM Ammonium Carbonate, pH 9.0 incubated for 24 hours). (B) ESI-MS spectrum of $\beta PFO_{A\beta(1\text{-}42)}$ tetramer coincubated with SST14 (150 μM Aβ42, 150 μM SST14, 7.2 mM LDAO, 200 mM Ammonium Carbonate, pH 9.0 incubated for 24 hours). Charge states corresponding to SST14; Aβ(1-42) monomer, dimer, trimer, and tetramer are indicated with schematic drawings and labelled, respectively, in red, black, grey, green, and blue.

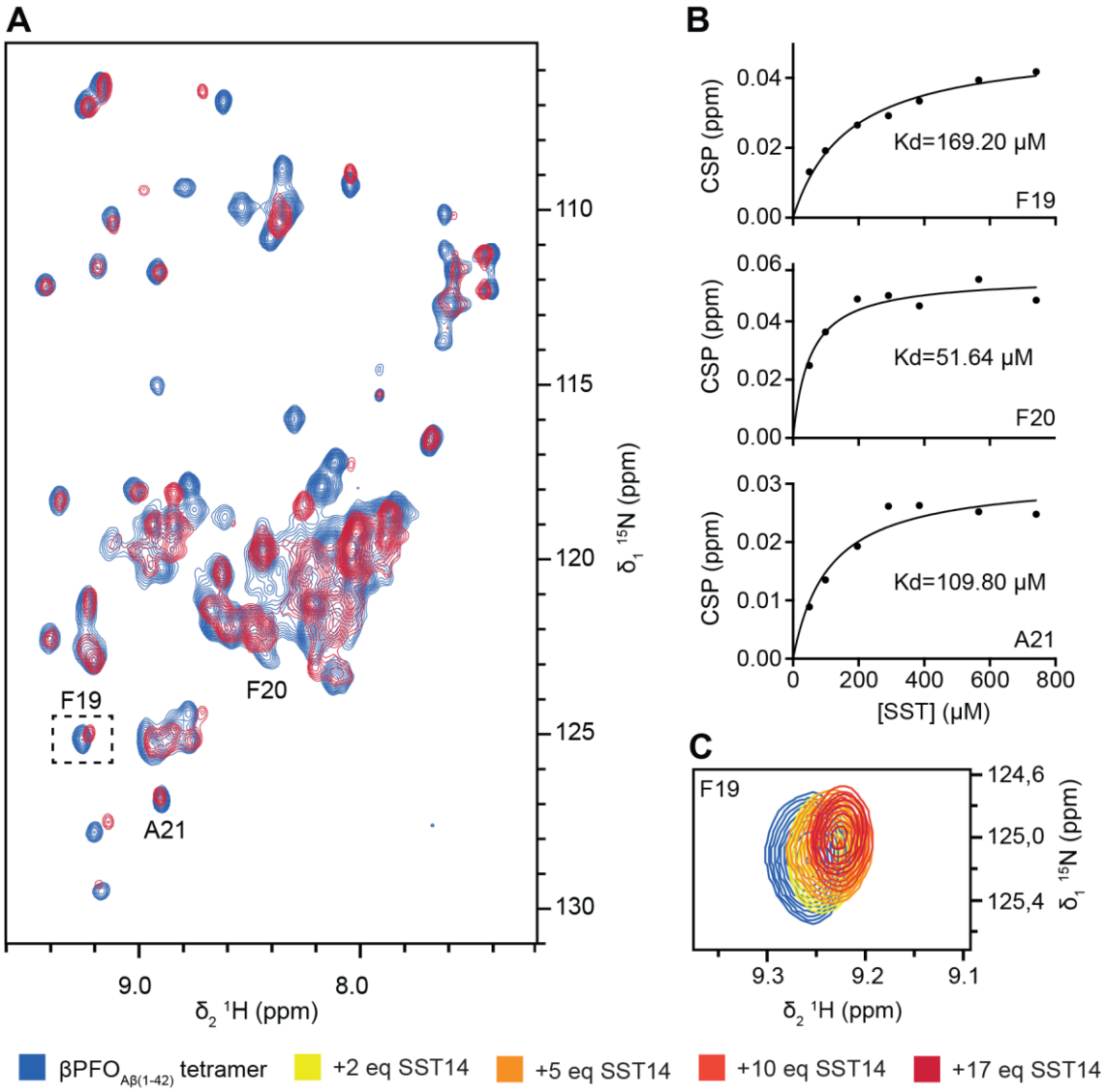


**Figure 3.** NMR titration of SST14 to $\beta PFO_{A\beta(1-42)}$ tetramer. (A) Two-dimensional [$^{15}N$, $^{1}H$]-SOFAST-HMQC spectra of $\beta PFO_{A\beta(1-42)}$ tetramer (230 μM Aβ(1-42), 7.71 mM $d_{38}$-DPC, 10 mM $d_{12}$-Tris·DCl, pH 8.5) alone (blue) and in the presence of 17 equivalents of SST14 (red). (B) Fitted saturation curves of residues F19, F20 and A21 during SST14 titration with their respective calculated $K_D$. (C) Close-up view of residue F19 from the titration in the presence of 0 (blue), 2 (yellow), 5 (orange), 10 (coral) and 17 (red) equivalents (eq.) of SST14.

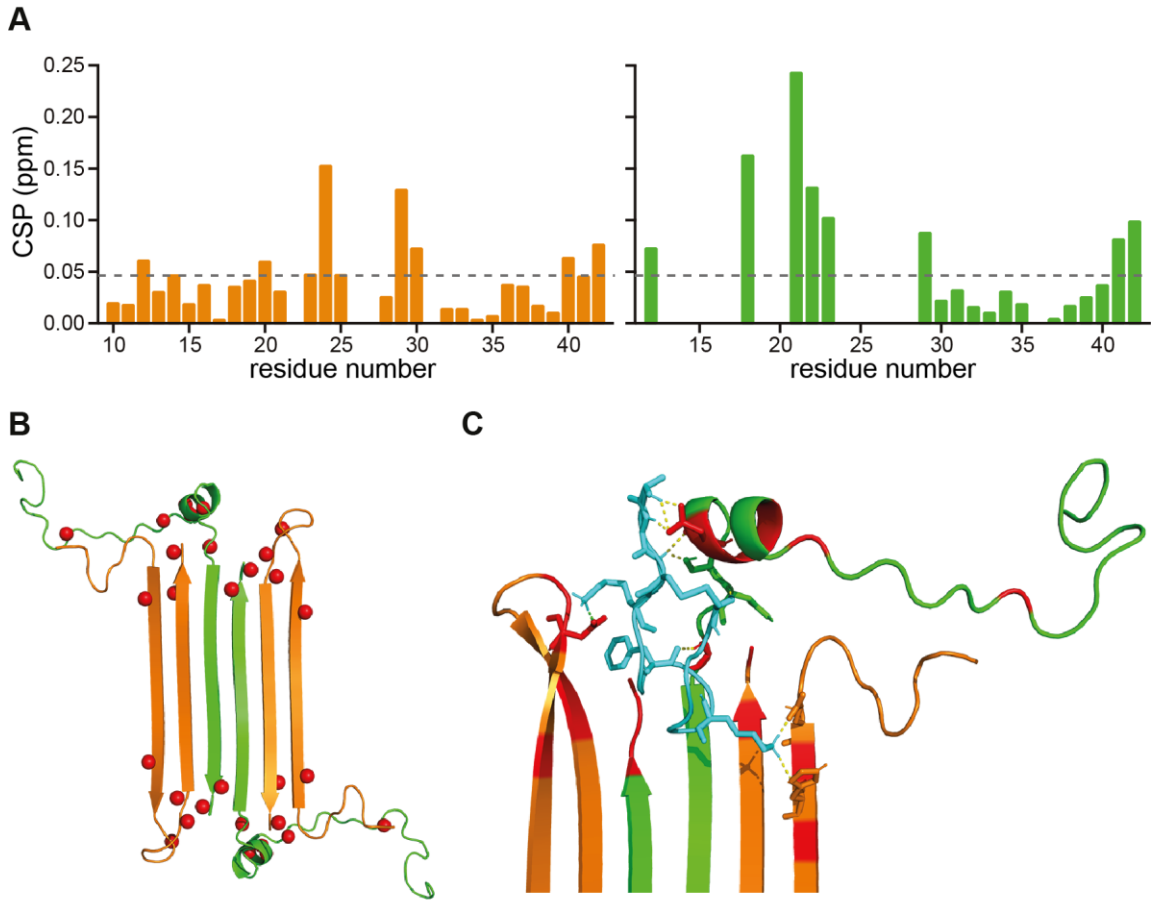


**Figure 4.** Binding site of SST14 to $\beta PFO_{A\beta(1\text{-}42)}$ tetramer. (A) Representation of the CSP of the residues within the orange and green subunits of the $\beta PFO_{A\beta(1\text{-}42)}$ tetramer induced by the presence of SST14. The grey dashed line indicates the threshold dictated by the standard deviation (σ). Source data are provided in Source data 1. (B) Representation of the residues affected by chemical shift changes (red spheres) in the presence of SST14 within the 3D structure of the $\beta PFO_{A\beta(1\text{-}42)}$ tetramer (PDB code 6RHY). (C) Best-ranked structure proposing a binding mode of SST14 (cyan, PDB code 2MI1) with $\beta PFO_{A\beta(1\text{-}42)}$ tetramer (orange and green). Residues introduced as ambiguous interaction restraints (AIRs) are colored in red.