

Tris(triphenylphosphine)Ruthenium(II) dichloride: a versatile catalystin the synthesis of β -, γ - and δ lactams from trichloroacetamides and in the isomerization of protected allylamines toz-enamines under microwave activation

Alexandra Georgiana Şandor

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https://www.journalcra.com/article/microwave-assisted-benzylic-c-h-activation-using-ruthenium-catalysts-synthesis-%EF%81%A2-lactams

INDEX

Abbreviations and Acronyms	
Chapter 1. Introduction and objectives	1
1.1. A short overview on ruthenium complexes	4
1.2. $RuCl_2(PPh_3)_3$ properties and usage in organic synthesis	6
1.2.1. RuCl₂(PPh₃)₃ in ATRC	7
1.2.2. RuCl ₂ (PPh ₃) ₃ in isomerization	13
1.2.3. RuCl ₂ (PPh ₃) ₃ in C-H activation reactions	19
1.2.4. RuCl ₂ (PPh ₃) ₃ in hydrogenation and hydrogen transfer reactions	20
1.2.5. RuCl ₂ (PPh ₃) ₃ in oxidation	_24
1.3. Objectives	_26
Chapter 2. Microwave-assisted benzylic C-H activation using ruthenium	۱
catalysts. Synthesis of β -lactams	_27
2.1. β-Lactams: a short overview	_29
2.2. Synthesis of β -lactams in the presence of RuCl ₂ (PPh ₃) ₃ . Our results	32
2.3. Cytotoxic activity of some of the synthesized lactams	.40
2.3.1. Hemocompatibility studies	40
2.3.2. Cell viability studies	_41
Chapter 3. Microwave assisted isomerization of N-allyl carbamates and	
N-allyl amides to Z-enecarbamates and Z-enamides catalyzed by	
RuCl ₂ (PPh ₃) ₃	.47
3.1. Enamides and enecarbamates en natural compounds and organic	
synthesis	49

3.2. Synthetic precedents in the preparation of enamides and enecarbamates
3.3. Isomerization of <i>N</i> -allyl carbamates and <i>N</i> -allyl amides in the presence of
RuCl ₂ (PPh ₃) _{3.} Our results59
Chapter 4. Microwave assisted synthesis of $\gamma\text{-}$ and $\delta\text{-}\text{lactams}$ from
trichloro- and dichloroacetamides catalyzed by RuCl ₂ (PPh ₃) ₃ 67
4.1. Atom Transfer Radical reactions69
4.2. ATRC catalyzed by RuCl ₂ (PPh ₃) ₃ in the synthesis of γ - and δ -lactams from
trichloro- and dichloroacetamides. Our results70

Chapter 5. Conclusions	
Chapter 6. Experimental part	81
General information	83
6.1. Experimental part of chapter 2	84
6.2. Experimental part of chapter 3	118
6.3. Experimental part of chapter 4	

ABBREVIATIONS AND ACRONYMS

¹³ C NMR	carbon-13 nuclear magnetic resonance
¹ H NMR	proton nuclear magnetic resonance
A431	squamous cell carcinoma
Ac ₂ O	acetic anhydride
ACN	acetonitrile
AIBN	azobisisobutyronitrile
Anisole	methyl phenyl ether
aq	aqueous
Ar	aryl
atm	atmosphere
ATRA	atom transfer radical addition
ATRC	atom transfer radical cyclization
Bn	benzyl
BnNH ₂	benzylamine
br s	broad singlet (spectrum)
Bu	butyl
Bu ₃ P	tributylphosphine
Bz	benzoyl
calcd.	calculated
CDT	cyclododecatriene
COD	cyclooctadiene
COOEt	ethoxycarbonyl
COOMe	methoxycarbonyl
COSY	spectroscopy correlation
d	day (s), doublet (spectrum)

DBU	1,8-diazabicyclo[5.4.0]undec-7-ene
DCE	dichloroethane
DCM	dichloromethane
dd	doublet of doublets (spectrum)
DMEM	Dulbecco's Modified Eagle Medium
DMF	dimethylformamide
DNA	deoxyribonucleic acid
ESI-TOF	Electrospray Ionization Time-of-Flight (Mass Spectrometrometry)
Et	ethyl
Et₃N	triethylamine
EtOH	ethanol
GLC	gas-Liquid Chromatography
h	hour/s
HaCaT	immortal human keratinocyte
HPLC	high performance liquid chromatography
HPPh ₂	diphenylphosphine
HRMS	high resolution mass spectrometry
HSQC	heteronuclear single quantum correlation spectroscopy
IC ₅₀	half maximal inhibitory concentration
ISO	International Organization for Standardization
J	coupling constant
LDA	Lithium diisopropylamide
L-dopa	L-3,4-dihydroxyphenylalanine
LHMDS	Lithium bis(trimethylsilyl)amide solution
М	molar
m	multiplet (spectrum)

m/z	mass to charge ratio
M+	molecular ion
MCF-7	Michigan Cancer Foundation-7 breast cancer cell line
Ме	methyl
MeO	methoxy
MeOH	methanol
mol	mole (s)
MOM	methoxymethyl
m.p.	melting point
MS	mass spectrometry/ molecular sieves
MTT	3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyl tetrazolium bromide
NBS	N-Bromosuccinimide
nM	nanometer (unit of measure)
NMO	N-methylmorpholine N-oxide
NMR	nuclear Magnetic Resonance Spectroscopy
NOESY	Nuclear Overhauser Effect Spectroscopy
ⁿ Pr, <i>n</i> Pr	<i>n</i> -Propyl
NRU	neutral red uptake
Ns	nosyl (4-nitrobenzenesulfonyl)
р	page
P(<i>t</i> Bu) ₃	tri- <i>tert</i> -Butylphospine
Ph	phenyl
Ph₃P	triphenylphosphine
PhH	benzene
PhMe	toluene
PhO	phenolate

ppm	parts per million
<i>n</i> Pr₃P	tripropylphosphine
q	quadruplet (spectrum)
quant.	quantitative
quint	quintuplet (spectrum)
R	generalized alkyl group or substituent
RBC	red blood cells
RCM	ring-closing metathesis
rfx	reflux
rt	room temperature
S	singlet (spectrum)
t	time, triplet (spectrum)
<i>t</i> -Bu (<i>t</i> Bu, ^{<i>t</i>} Bu)	<i>tert</i> -butyl
TEMPO	(2,2,6,6-Tetramethylpiperidin-1-yl)oxyl
Tf ₂ O	trifluoromethanesulfonic anhydride
THAHS	tetrahexylammonium hydrogensulfate
THF	tetrahydrofuran
Ts	tosyl (p-toluenesulfonyl)
UV	ultraviolet radiation
VTMS	vinyloxytrimethylsilane
ZnEt ₂	diethylzinc
β-CF ₃	beta-trifluoromethyl
γ-CF ₃	gamma-trifluoromethyl
μW	microwave activation
δ	chemical shift

CHAPTER 1: Introduction and objectives

In the current PhD thesis are reported the different results related to Tris(triphenylphosphine)ruthenium(II) dichloride (RuCl₂(PPh₃)₃) as a powerful and versatile catalyst for the synthesis of β - γ - and δ -lactams and also for the isomerization of protected allylamines to *Z*-enamines under microwave activation.

The first chapter of this thesis is a short overview that opens with the description of RuCl₂(PPh₃)₃, its preparation and then describes the numerous chemical transformations in which this catalyst is involved.

The second chapter outlines our results concerning the synthesis of β lactams from *N*-tethered benzyl trichloroacetamides through a C(sp³)-C(sp³) bond formation. The latter takes place *via* an unprecedented "Ru" complex promoted activation of a benzylic position in a radical process. In this chapter are also disclosed the results regarding the cytotoxic activity and hemocompatibility studies of six of these lactams.

The third chapter introduces the usage of $RuCl_2(PPh_3)_3$ as a catalyst in the isomerization of *N*-allyl carbamates and *N*-allyl amides to the corresponding *Z*-enecarbamates and *Z*-enamides with very good yields and excellent selectivity.

Finally, the fourth chapter focuses on the synthesis of γ - and δ -lactams from *N*-tethered alkenyl trichloroacetamides in the presence of a catalytic amount of RuCl₂(PPh₃)₃. In this part of the work, a reliable diastereoselective Atom Transfer Radical Cyclization is reported from different substrates in very short reaction time and with good yields. The methodology was successfully applied to the synthesis of functionalized indole and morphan scaffolds present in many natural compounds such as the *Daphniphyllum* alkaloids.

3

1.1. A short overview on ruthenium complexes

The element ruthenium (Ru) was discovered in 1844 in the Ural Mountains by a Russian scientist called Karl Ernst Claus. He named it "Ruthenium" in honor of his native country after the word "Ruthenia" which was the Latin name for Russia until the end of the 17th century.¹

Ruthenium is a rare transition metal found in platinum ores as a minor component. Like other metals of the platinum group it is inert to most of other chemicals.

Ru has the widest scope of oxidation states of all elements of the Periodic table (from -2 valent in Ru(CO)₄²⁻ to octavalent in RuO₄), and various coordination geometries in each electron configuration.

The ruthenium-based catalysts started to be more explored for their great potential in catalytic reactions and synthetic methods at the beginning of 1980s.

Ruthenium forms a wide range of compounds with carbon-ruthenium bonds that have been considered for cancer therapy (figure 1.1). KP1019 and NAMI-A were discovered after an intense synthetic work carried out in the field of anticancer metal complexes.²



NAMI-A (antitumor/antimetastatic)



KP1019 (colorectal cancer treatment)

Figure 1.1. First ruthenium anticancer compounds in clinical trials.

¹ Pitchkov, V. N. The discovery of ruthenium. *Platin. Met. Rev.* **1996**, *40*, 181-188.

² (a) Clarke, M. J. Oncological implications of the chemistry of ruthenium. *Met. Ions. Biol. Syst.* **1980**, *11*, 231-283. (b) Vilar, R. Nucleic acid quadruplexes and metallo-drugs. *Met. Ions Life Sci.* **2018**, *18*, 325-349.

NAMI-A ((ImH)[*trans*-RuCl₄(dmso-S)(Im)], Im = imidazole) and KP1019 ((IndH)[*trans*-RuCl₄(Ind)₂], Ind = indazole) are structurally related Ru(III) compounds, water-soluble with a low systemic toxicity and moderate stability under physiological conditions as a consequence they are classified as prodrugs.

The pharmacological profile of the two is very distinct, KP1019 has a cytotoxic effect for the treatment of platinum-resistant colorectal cancers, whereas NAMI-A has a non-cytotoxic reputation being effective as an antimetastatic drug. The mechanism of action reveals that KP1019 is able to enter the cells in an appreciable amounts, whereas NAMI-A localizes mostly extracellularly or on the cell membrane.³

Among the homogenous catalysts, solutions containing ruthenium trichloride are highly active and used especially for the preparation of other ruthenium complexes that can be divided into five groups according to their supporting ligands:

- Oxo, such as RuO₄, [RuO₂(bpy){IO₃(OH)₃] x 1.5H₂O
- Carbonyl, such as Ru₃(CO)₁₂, [RuCl₂(CO)₃]₂
- Tertiary phosphines, such as RuCl₂(PPh₃)₃, RuH₂(PPh₃)₄
- Cyclopentadienyl, such as RuClCp(PPh₃)₂, RuCl₂Cp(PPh₃)
- Arenes/dienes, such as [RuCl₂(*p*-cymene)]₂, Dichloro(cycloocta-1,5-diene)ruthenium

These species have demonstrated abilities to design specific organic transformations, low redox potential, high electron transfer ability, high coordination ability to heteroatoms, Lewis acid acidity or reactivity of metallic species and intermediates.⁴

A large number of reactions were developed using these ruthenium complexes, one of the most important ones being Grubbs (1st and 2nd generation) that have become extremely important in organic synthesis. Grubbs catalysts,

³ (a) Aitken, J. B.; Antony, S.; Weekley, C.M.; Lai, B.; Spiccia, L.; Harris, H. H. Distinct cellular fates for KP1019 and NAMI-A determined by X-ray fluorescence imaging of single cells. *Metallomics* **2012**, *4*, 1051-1056. (b) Alessio, E.; Messori L. NAMI-A and KP1019/1339, two iconic ruthenium anticancer drug candidates face-to-face: A case story in medicinal inorganic chemistry. *Molecules* **2019**, *24*, 1-20.

⁴ (a) Komiya, S.; Hurano, M. Group 8 (Fe, Ru, Os) metal compounds. Synthesis of organometallic compounds: A practical guide. Komiya, S., Ed. John Wiley & Sons: England, **1997**; pp. 159-203.
(b) Naota, T.; Takaya, H.; Murahashi. S.I. Ruthenium-catalyzed reactions for organic synthesis. *Chem. Rev.* **1998**, *98*, 2599–2660.

some of them presented in Figure 1.2, are usually used in olefin metathesis for the generation of new C-C bonds (Figure 1.3).⁵ Nevertheless many other non-metathetic behaviours have been reported for theses ruthenium complexes.



Figure 1.2. Common Grubbs catalysts used in metathesis.



Figure 1.3. Metathetic behaviour of Ruthenium catalysts.

1.2. RuCl₂(PPh₃)₃ properties and usage in organic synthesis

Tris(triphenylphosphine)ruthenium(II) dichloride or Dichlorotris(triphenylphosphine)ruthenium(II) (Figure 1.4) is a coordination complex and the precursor of Grubbs' 1st generation catalyst. It is a red-brown solid or shiny black crystals soluble in organic solvents such as toluene, benzene and dichloromethane. Its molecular formula is $C_{54}H_{45}Cl_2P_3Ru$ with a molecular weight of 958.83 g/mol and a melting point between 132 and 134 °C.



Figure 1.4. $RuCl_2(PPh_3)_3$ catalyst (Ru3).

⁵ Vougioukalakis, G. C.; Grubbs, R. H. Ruthenium-based heterocyclic carbene-coordinated olefin metathesis catalysts. *Chem. Rev.* **2010**, *110*, 1746–1787.

RuCl₂(PPh₃)₃ is commercially available but can also be prepared in the laboratory following the synthesis described by Wilkinson in 1966.⁶ An excess of triphenylphosphine and ruthenium(III) chloride hydrate are dissolved in boiling ethanol in order to make possible the formation of RuCl₂(PPh₃)₃ complex (scheme 1.1). The solvent could also be replaced by methanol or an acetone-water mixture.

$$RuCl_{3}.xH_{2}O + PPh_{3} \xrightarrow{EtOH} RuCl_{2}(PPh_{3})_{3}$$

quant.

Scheme 1.1. Synthesis of RuCl₂(PPh₃)₃.

RuCl₂(PPh₃)₃ is an effective catalyst used in oxidations, reductions, crosscouplings, Kharasch addition (ATRA) of a range of halogenated derivatives (Scheme 1.2) and isomerization of a variety of organic compounds.⁷

 $R_2C=CH_2 + R'X \longrightarrow R_2CX-CH_2R'$

Scheme 1.2. Basic Kharasch reaction scheme.

1.2.1. RuCl₂(PPh₃)₃ in ATRC

In 1973 Matsumoto *et al.* reported the first intermolecular addition of CCl₄ and CHCl₃ to terminal olefins catalyzed by RuCl₂(PPh₃)₃.^{8a} Later, using the same

⁶ (a) Samouei, H.; Grushin, V. V. New, Highly Efficient, simple, safe, and scalable synthesis of [(Ph₃P)₃Ru(CO)(H)₂]. *Organometallics* **2013**, *32*, 4440-4443. (b) Stephenson, T. A.; Wilkinson, G. New complexes of ruthenium (II) and (III) with triphenylphosphine, triphenylarsine, trichlorostannate, pyridine and other ligands. *J. Inorg. Nucl. Chem.* **1966**, *28*, 945-956. (c) Hallman, P. S; Stephenson, T. A.; Wilkinson, G. Tris(triphenylphosphine)dichlororuthenium(II). Innorganic Syntheses, Volume XII, McGraw-Hill Book Company, **1970**, 237-240.

⁷ (a) Kharasch M.; Jensen E.; Urry, W.; Addition of carbon tetrachloride and chloroform to olefins. *Science* **1945**; 102 (b) Plummer, J. S., Shun-Ichi, M.; Changjia, Z. Dichlorotris (triphenylphosphine) ruthenium(II), e-EROS Encyclopedia of reagents for organic synthesis, John Wiley, **2010**.

⁸ (a) Matsumoto, H.; Nakano, T.; Nagai, Y. Radical reactions in the coordination sphere I. Addition of carbon tetrachloride and chloroform to 1-olefins catalyzed by ruthenium (II) complexes. *Tetrahedron Lett.* **1973**, *14*, 5147-5150. (b) Matsumoto, H.; Nikaido, T.; Nagai, Y. Radical reactions in the coordination sphere II. Stereoselective addition of carbon tetracrloride to cyclohexene catalyzed by dicrlorotris(triphenylphosphine)-rutrenium(II). *Tetrahedron Lett.* **1975**, *16*, 899-902. (c) Matsumoto, H.; Nikaido, T, Nagai, Y. Radical reactions in the coordination sphere. III. Reactions of dichloro- and trichloroacetic acid esters with 1-olefins catalyzed by dichlorotris(triphenylphosphine) ruthenium(II). *J. Org. Chem.* **1976**, *41*, 396–398.

catalyst, the same authors described a stereoselective radical addition of carbon tetrachloride to cyclohexene^{8b} and also the reaction of dichloro- and trichloroacetic acid esters with 1-olefins^{8c} (Scheme 1.3). This research activity encourages the exploration of numerous other Ru complexes for their ability to catalyze ATRA and ATRC reactions.



Scheme 1.3. Intermolecular additions catalyzed by RuCl₂(PPh₃)₃.

RuCl₂(PPh₃)₃ usage for the transformation of trichloroacetamides through ATRC has been outlined in very few examples. Itoh reported the first articles that describe the novel transformation from trichloroacetamides to trichlorinated γ -lactams despite the potential for formation of δ -lactams. The conditions employed involved usage of 5 mol% of **Ru3** catalyst at 110-140 °C obtaining yields between 53-90% of the corresponding *trans* isomers predominantly. Additionally *cis*-fused hexahydro-oxindoles were also prepared using similar conditions (Scheme 1.4).⁹

⁹ (a) Nagashima, H.; Wakamatsu, H.; Itoh, K. A novel preparative method for γ-butyrolactams via carbon-carbon bond formation: copper or ruthenium-catalysed cyclization of *N*-ally trichloroacetamides. *J. Chem. Soc., Chem. Commun.* **1984**, 652-653. (b) Nagashima, H.; Ara, K.; Wakamatsu, H.; Itoh, K. Stereoselective preparation of bicyclic lactams by copper- or ruthenium-catalysed cyclization of *N*-allyltrichloroacetamides: a novel entry to pyrrolidine alkaloid skeletons. *J. Chem. Soc., Chem. Commun.* **1985**, *53*, 518-519. (c) Nagashima, H.; Ozaki, N.; Ishii, M.; Koji,S.; Washiyama, M.; Itoh, K. Transition Metal-Catalyzed Radical Cyclizations: A Low-Temp. process for the cyclization of *n*-protected *n*-allyltrichloroacetamides to trichlorinated γ-lactams and application to the stereoselective preparation of β ,γ-disubstituted γ-lactams. *J. Org. Chem.* **1993**, *58*, 464-470.



Scheme 1.4. RuCl₂(PPh₃)₃ mediated synthesis of γ -lactams (ref. 9).

By the same period, Weinreb *et al.* reported the intramolecular cyclization of unsaturated α,α -dichloro esters, acids and nitriles catalyzed by RuCl₂(PPh₃)₃ for the synthesis of functionalized carbocycles (Scheme 1.5).¹⁰



Scheme 1.5. A sample of RuCl₂(PPh₃)₃ mediated synthesis of carbocycles (ref. 10).

Additionally, Slough reported the synthesis of γ -lactams from α -chloro *N*tosyl amides in the presence of RuCl₂(PPh₃)₃ as a catalyst. The reaction was achieved from a series of substrates varying the substituent on the radical center generated in the first place (R = Cl, H, CH₃, *I*Pr, CH₂Ph, *tert*-amyl). The reaction takes place with predominant *trans* selectivity and a transition from *trans* to *cis*

¹⁰ (a) Hayes, T. K.; Villani. R.; Weinreb, S. M. Exploratory studies of the transition metal catalyzed intramolecular cyclization of unsaturated α, α -dichloro esters, acids, and nitriles. *J. Amer. Chem. Soc.* **1988**, *110*, 5533-5543. (b) Lee, G. M.; Parvez, M.; Weinreb. S.M. Intramolecular metal catalysed Kharasch cyclizations of olefinic α -halo esters and acids. *Tetrahedron* **1988**, *44*, 4671-4678. (c) Phelps, J. C.; Bergbreiter, D. E.; Lee, G. M.; Villani, R.; Weinreb. S. M. A polyethylene-bound ruthenium(II) catalyst for inter- and intramolecular kharasch reactions. *Tetrahedron Lett.* **1989**, *30*, 3915-3918.

was observed with hindered groups such as *i*-Pr, Bn and *tert*-amyl (Scheme 1.6).^{11,12}



Scheme 1.6. N-tosyl amides radical cyclization catalyzed by RuCl₂(PPh₃)₃.

Parson developed a **Ru3**-mediated ATRC toward the synthesis of β - or γ lactams starting from *N*-vinylic trichloro- or bromoacetamides in parallel to his work with CuCl. The use of **Ru3** catalyst in toluene afforded the corresponding highly functionalized β -lactams resulting from a 4-*exo* cyclization as the main products. The authors suggested that the β -lactams are formed under a kinetic control whereas the γ -lactams are the thermodynamic products (Scheme 1.7).¹³

¹¹ Rachita, M. A.; Slough, G. A. Ruthenium (II) catalyzed ring closure of prochiral α-chloro-*N*-tosyl amides: A diastereoselectivity study. *Tetrahedron Lett.* **1993**, *34*, 6821-6824.

¹² Slough, G. A. $(Ph_3P)_3RuCl_2$ catalyzed equilibration and elimination of α -chloro-*N*-tosyl-2pyrrolidinones: A unique route to unsaturated 2-pyrrolidinones. *Tetrahedron Lett.* **1993**, *34*, 6825-6828.

¹³ Bryans, J. S.; Chessum, N. E. A.; Parsons, A.F.; Ghelfi, F. The synthesis of functionalised β and γ -lactams by cyclisation of enamides using copper(I) or ruthenium(II). *Tetrahedron Lett.* **2001**, *42*, 2901-2905.



Scheme 1.7. Formation of β -lactams catalyzed by RuCl₂(PPh₃)₃.

In 1993 Ikeda *et al.* described the synthesis of (-)-trachelanthamidine and a formal total synthesis of (±)-haemanthidine and (±)-pretazettine. One of the key steps of these syntheses is the RuCl₂(PPh₃)₃ catalyzed atom transfer radical cyclizations of *N*-allylic- α -chloro- α -thioacetamides to provide the corresponding α -thio- β -chloromethyl substituted γ -lactams with acceptable yields (Scheme 1.8).¹⁴

¹⁴ Ishibashi, H.; Uemura, N.; Nakatani, H.; Okazaki, M.; Sato, T.; Nakamura, N.; Ikeda, M. Ruthenium-catalyzed chlorine atom transfer cyclizations of *N*-allylic α-chloro-α-thioacetamides. Synthesis of (-)-trachelanthamidine and formal total synthesis of (±)-haemanthidine and (±)-pretazettine. *J. Org. Chem.* **1993**, *58*, 2360-2368.



Scheme 1.8. Synthesis of γ -lactams by chlorine atom transfer.

In 2009 Sutherland *et al.* published an elegant work where a tandem MOM-ether directed Overman rearrangement and RCM reaction process were developed for the efficient and highly selective synthesis of cyclopentenyl and cyclohexenyl trichloroacetamides. The latter was then submitted to a Kharasch cyclization in the presence of RuCl₂(PPh₃)₃ to provide the corresponding bicyclic amide as a single stereoisomer in 75% yield (Scheme 1.9).¹⁵

¹⁵ Swift, M. D.; Donaldson, A.; Sutherland, A. Tandem aza-Claisen rearrangement and ringclosing metathesis reactions: the stereoselective synthesis of functionalised carbocyclic amides. *Tetrahedron Lett.* **2009**, *50*, 3241-3244.



Scheme 1.9. Overman rearrangement, RCM and kharasch cyclization in the synthesis of functionalized carbocyclic amides.

Finally, in an approach to the pentacyclic core of aspidophylline A, Shi *et al.* described a strategy featuring a CAN-mediated intramolecular azidoalkoxylation of an enecarbamate to construct the fused tetrahydrofuran ring and a $RuCl_2(PPh_3)_3$ -catalyzed intramolecular atom transfer radical cyclization to form the azabicycle[3,3,1] fragment as it is indicated in Scheme 1.10.¹⁶



Scheme 1.10. Construction of the morphan ring present in aspidophylline A.

1.2.2. RuCl₂(PPh₃)₃ in isomerization

Besides ATRC, RuCl₂(PPh₃)₃ was widely used in isomerization processes as it is outlined in the examples reported bellow.

¹⁶ Li, Q.; Li, G.; Ma, S.; Feng, P.; Shi, Y. An approach to the skeleton of aspidophylline A. *Org. Lett.* **2013**, *15*, 2601-2603.

Unsymmetrical diallyl ethers give γ , δ -unsaturated ketones or aldehydes upon heating in the presence of tris(triphenylphosphine)ruthenium dichloride.¹⁷ The selective rearrangement takes place in the presence of 0.1 mol% of catalyst and at 200 °C affording pure carbonyl compounds after the distillation process. Although 2 different isomeric γ , δ -unsaturated carbonyl compounds could be obtained from each diallyl ether, the reaction was highly regiospecific providing the isomers showed in Scheme 1.11 alone.



Scheme 1.11. Examples of RuCl₂(PPh₃)₃ catalyzed rearrangement of unsymmetrical diallyl ethers.

These final aldehydes and ketones are presumably produced by an initial ruthenium catalyzed 1,3-hydrogen shift (Scheme 1.12a) followed by a Claisen rearrangement (Scheme 1.12b).



Scheme 1.12. Rearrangement mechanism of unsymmetrical diallyl ethers.

¹⁷ Reuter, J. M.; Salomon, R. G. Ruthenium(II) catalyzed rearrangement of diallyl ethers. a synthesis of γ ,δ-unsaturated aldehydes and ketones. *J. Org. Chem.* **1977**, *4*2, 3360–3364.

The first example of RuCl₂(PPh₃)₃ catalyzed isomerization of 2-ynols to the simple (2*E*)-enals was reported more than three decades ago by Lu *at al.*¹⁸ The stereoselective transformation proceeds in long reaction time (30-48 h) and in the presence of trialkyl phosphine (R₃P) under reflux of toluene (Scheme 1.13). Curiously in the absence of a ligand, or using Ph₃P as the ligand, the reaction did not occur.

 $R = nC_{6}H_{13}, nC_{5}H_{11}, nC_{7}H_{15}, nC_{4}H_{9}, C_{6}H_{5}$ $R = nBu_{3}P/iPr_{3}P$



RuCl₂(PPh₃)₃ catalyzes the isomerization of α , β -ynones to (E,E)- α , β , γ , δ -dienones. Indeed successful isomerization of 3-undecyn-2-one to (3E,5E)-undeca-3,5-dien-2-one with a 20% rate of conversion took place using 1 mol% of RuCl₂(PPh₃)₃ over a 24 h period of time. The presence of an excess of phosphine ligands such as Ph₃P influenced significantly the activity of the catalyst giving a 98% conversion under the same reaction conditions (Scheme 1.14).



Scheme 1.14. Effect of PPh₃ as a ligand in the isomerization of 3-undecyn-2-one.

The procedure was successfully applied to prepare several natural compounds among them I-phenyl-(2E, 4E)-hexa-2,4-dienone (capillone), a component of artemisia with medicinal properties (Scheme 1.15).¹⁹

 $^{^{18}}$ Ma, D.; Lu, X. A simple route to α,β -unsaturated aldehydes from prop-2-ynols. *J. Chem. Soc. Chem. Commun.* **1989**, *14*, 890-891.

¹⁹ Ma, D.; Yu, Y.; Lu, X. Highly stereoselective isomerization of ynones to conjugated dienones catalyzed by transition-metal complexes. *J. Org. Chem.* **1989**, *54*, 1105-1109.



Scheme 1.15. Stereoselective isomerization in the synthesis of capillone.

 $RuCl_2(PPh_3)_3$ has also been used to convert allylic alcohols to the corresponding saturated ketones. A dramatic rate enhancement was observed when a catalytic amount of a base (K₂CO₃) was added to the reaction mixture (Scheme 1.16).²⁰



Scheme 1.16. Isomerization of 1-octen-3-ol.

Another example of Isomerization from alkenyl alcohols using RuCl₂(PPh₃)₃ as a catalyst in water, was reported by Li *et al.* in 1998.²¹ In this investigation the functional groups of homoallylic and allylic alcohols are repositioned to give allylic alcohols with controlled regioselectivity (Scheme 1.17).



Scheme 1.17. Isomerization of homoallylic and allylic alcohols in water.

²⁰ Baeckvall, J.-E.; Andreasson, U. Ruthenium-catalyzed isomerization of allylic alcohols to saturated ketones. *Tetrahedron. Lett.* **1993**, *34*, 5459-5462.

²¹ Wang, D.; Chen, D.; Haberman, J. X.; Li, C. J. Ruthenium-catalyzed isomerization of homoallylic alcohols in water. *Tetrahedron* **1998**, *54*, 5129-5142.

When the substrate used has both an allylic and a homoallylic functional groups, the reaction occurs exlusively by rearrangement of the homoallylic group to give the conjugated dienol product (Scheme 1.18).



Scheme 1.18. Selective isomerization at the homo-allylic group.

Regioselective synthesis of *trans*-anethole from methyl chavicol via double bond isomerization using RuCl₂(PPh₃)₃ catalyst was reported in parallel with the isomerization of eugenol into *trans*-isoeugenol.²² The highest conversion of both molecules (almost 100%) with 95% of *trans* selectivity was achieved in ethanol using 0.8 mol% of catalyst (Scheme 1.19).



Scheme 1.19. Isomerization of methyl chavicol and eugenol.

Renaud and Cahard described the synthesis of β -trifluoromethylated ketones from trifluoromethylated allylic alcohols in the presence of RuCl₂(PPh₃)₃ as a catalyst. The ruthenium(II) complex was able to conduct the intramolecular 1,3-hydrogen shift at temperatures comprised between 30 and 70 °C in toluene and in the presence of Cs₂CO₃ (Scheme 1.20).²³ In this work the CF₃ group was

²² Sharma, S. K.; Srivastava, V. K.; Jasra, R. V. Selective double bond isomerization of allyl phenyl ethers catalyzed by ruthenium metal complexes. *J. Mol. Catal. A Chem.* **2006**, *245*, 200–209. ²³ Bizet, V.; Pannecoucke, X.; Renaud, J. L.; Cahard, D. Synthesis of β-CF₃ ketones from trifluoromethylated allylic alcohols by ruthenium catalyzed isomerization. *J. Fluor. Chem.* **2013**, *152*, 56-61.

found to strongly accelerate the insertion step compared to non-fluorinated substrates.



Scheme 1.20. Isomerization of β -trifluoromethylated secondary allylic alcohols conducted by **Ru3** catalyst.

The same authors reported a tandem isomerization-transfer hydrogenation of γ -CF₃ allylic alcohols and β -CF₃ enones that allows the synthesis of saturated alcohols in high yields. Instead of hydrogen gas, normally necessary in a classical hydrogenation, in this work, 2-propanol was used as a solvent and as the hydrogen source. After a screening of various Ru catalysts and bases, the best combination to obtain the desired saturated alcohol was RuCl₂(PPh₃)₃/Cs₂CO₃ in a 1:2 ratio (Scheme 1.21).²⁴



* with 2 exceptions: $R_1 = Ph$, *t*-Bu and $R_2 = Bn$, Ph only traces of alcohol

Scheme 1.21. Isomerization-transfer hydrogenation of γ -CF₃ allylic alcohols and β -CF₃ enones to satured alcohols.

²⁴ Bizet, V.; Pannecoucke, X.; Renaud, J. L.; Cahard, D. Ruthenium-catalyzed one-pot tandem isomerization-transfer hydrogenation reactions of γ-trifluoromethylated allylic alcohols and β-trifluoromethylated enones. *Adv. Synth. Catal.* **2013**, *355*, 1394–1402.

1.2.3. RuCl₂(PPh₃)₃ in C-H activation reactions

C-C bond formation involving C-H activation and addition on unsaturated substrates has been investigated in the presence of transition metal catalysts.²⁵

Kakiuchi *et al.* demonstated that RuCl₂(Ph₃)₃ catalyzes the regioselective amino- and alkoxycarbonylations at aromatic C-H bonds using carbamoyl chlorides and alkyl chloroformates respectively without the intervention of an oxidant (Scheme 1.22). A broad generality of amide and ester groups was achieved taking advantage of the wide availability of carbonylating agents.²⁶



Scheme 1.22. Amino- and alkoxycarbonylations through aromatic C–H activation.

Martín-Matute *et al.* described a general tandem isomerization of allylic alcohols/C-H activation catalyzed by the stable RuCl₂(PPh₃)₃ complex. The tandem process affords a variety of ortho alkylated ketones with excellent yields and in very short reaction times (Scheme 1.23).²⁷





²⁵ For a review see: Ritleng, V.; Sirlin, C.; Pfeffer, M. Ru-, Rh-, and Pd-catalyzed C-C bond formation involving C-H activation and addition on unsaturated substrates: Reactions and mechanistic aspects. *Chem. Rev.* **2002**, *102*, 1731–1769.

²⁶ Kochi, T.; Urano, S.; Seki, H. Mizushima, E.; Sato, M.; Kakiuchi, F. Ruthenium-catalyzed aminoand alkoxycarbonylations with carbamoylchlorides and alkyl chloroformates via aromatic C-H bond cleavage. *J. Am. Chem. Soc.* **2009**, *131*, 2792–2793.

²⁷ Bartoszewicz, A.; Martín-Matute, B. Building molecular complexity via tandem Ru-catalyzed isomerization/C-H activation. *Org. Lett.* **2009**, *11*, 1749-1752.

1.2.4. RuCl₂(PPh₃)₃ in hydrogenation and hydrogen transfer reactions

Selective hydrogenation of 3-oxo-1,4-diene steroidal compounds in the presence of RuCl₂(PPh₃)₃ occurs at a relatively low temperature and under a high hydrogen pressure (10-162 kg/cm²) providing high yields of the corresponding 3-oxo-4-enes (Scheme 1.24).²⁸ The activity and selectivity of this Ru-complex in the hydrogenation of 1,4-androstadiene-3,17-dione was improved by adding an optimal amount of triethylamine.²⁹



Scheme 1.24. Ru3-Hydrogenation of 1,4-androstadiene-3,17-dione.

Other examples of selective hydrogenations using RuCl₂(PPh₃)₃ as a catalyst are those of cyclododecatriene(CDT) to cyclododecene, cyclooctadiene (COD) to cyclooctene and ethyl 2-acetyl-4,9-decadienoate to 2-acetyl-4-decenoate obtained with a high selectivity at room temperature.³⁰

Symmetrical³¹ as well as unsymmetrical³² cyclic anhydrides could be converted in good yields into the corresponding lactones by hydrogenation catalyzed with RuCl₂(PPh₃)₃ (Scheme 1.25).

²⁸ Nishimura, S.; Tsuneda, K. Selective homogeneous hydrogenation of 3-Oxo-1,4-diene steroids with a ruthenium complex as catalyst. *Bull. Chem. Soc. Jpn.* **1969**, *42*, 852.

²⁹ Nishimura, S.; Ichino, T.; Akimoto, A.; Tsuneda, K. Selective homogeneous hydrogenation of 3-Oxo-1,4-diene steroids. II. Effects of basic additives and of para Substituents on the hydrogenation with dichlorotris(triphenylphosphine)ruthenium. *Bull. Chem. Soc. Jpn.* **1973**, *46*, 279-282.

³⁰ Tsuji, J.; Suzuki, H. Homogeneous selective hydrogenation of cyclododecatriene, cyclooctadiene, and 1,6-dienes to the corresponding monoenes catalyzed by RuCl₂(PPh₃)₃. *Chem. Lett.* **1977**, *6*, 1083-1084.

³¹ Lyons, J. E. Homogeneous catalytic hydrogenolysis of the C-O bond: The selective reduction of cyclic carboxylic acid anhydrides to γ -lactones catalysed by [RuCl₂(Ph₃P)₃]. *J. C. S. Chem. Commun.* **1975**, *11*, 412–413.

³² Morand, P.; Kayser, M. Highly regioselective reduction of unsymmetrical cyclic carboxylic acid anhydrides to γ -lactones. *J. C. S. Chem. Commun.* **1976**, 314-315.



Scheme 1.25. Ru3-Hydrogenation of 2,2-dimethylsuccinic anhydride.

Following these results, γ -lactones formation was also observed in the RuCl₂(PPh₃)₃-catalyzed hydrogenation of some keto acids such as levulinic acid, 3-benzoylpropionic acid and 2-acetylbenzoic acid.³³ The reactions were performed under hydrogen atmosphere in toluene as a solvent and at elevated temperature (180 °C) with yields from 40 to 86% (Scheme 1.26).



Scheme 1.26. An example of Ru3-catalyzed hydrogenation of keto acids.

Hydrogen transfer reactions are an alternative method for hydrogenation of organic substrates where hydrogen is added to a molecule from a source other than gaseous H₂, being considered a more simple and economical procedure. The donor molecule itself undergoes dehydrogenation during the course of the reaction. Low-valent ruthenium complexes are excellent catalysts for the hydrogen transfer reactions because of their low redox potential and higher affinity toward hetero atom compounds. RuCl₂(PPh₃)₃ has been used in hydrogen transfer reactions with alcohols,³⁴ acids,³⁵

³³ Osakada, K.; Ikariya, T.; Yoshikawa, S. Preparation and properties of hydride triphenylphosphine ruthenium complexes with 3-formyl (or acyl) propionate. *J. Organomet. Chem.* **1982**, *231*, 79–90.

³⁴ (a) Sasson, Y.; Blum, J. Dichlorotris(triphenylphosphine)ruthenium-Catalyzed Hydrogen Transfer from Alcohols to Saturated and Unsaturated Ketones. *J. Org. Chem.* **1975**, *40*, 1887-1896. (b) Sasson, Y.; Albin, P.; Blum, J. Effect of a Ru (II) catalyst on the rate of equilibration of carbinols and ketones. *Tetrahedron Lett.* **1974**, *10*, 833-836.

³⁵ Maruyama, Y.; Sezaki, T.; Tekawa, M.; Sakamoto, T.; Shimizu, I.; Yamamoto, A. Rutheniumcatalysed reductive cleavage of allylic esters with formic acid and triethylamine. Application to short-step synthesis of α-hydroxy acids. *J. Organomet. Chem.* **1994**, *473*, 257-264.

olefins,³⁶ ketones,³⁷ aldehydes,³⁸ α , β -unsaturated carbonyl compounds,³⁹ imines,⁴⁰ nitro compounds and quinolines⁴¹ (Table 1.1).

³⁶ Nishiguchi, T.; Imai, H.; Hirose, Y.; Fukuzumi, K. Transfer hydrogenation and transfer hydrogenolysis. VIII. Hydrogen transfer from amines to olefins catalyzed by heterogeneous and homogeneous catalysts. *J. Catal.* **1976**, *41*, 249-257.

³⁷ (a) Watanabe, Y.; Ohta, T.; Tsuji, Y. Ruthenium-catalyzed reduction of carbonyl compounds using formic acid. *Bull. Chem. Soc. Jpn.* **1982**, 2441-2444. (b) Smith, T. A.; Maitlis, P. M. Methanol as hydrogen-donor in homogeneously catalysed reactions. *J. Organomet. Chem.* **1984**, *269*, 7-9. (c) Smith, T.A.; Maitlis, P.M. Methanol as a hydrogen donor in reactions homogeneously catalysed by Ru and Rh complexes. *J. Organomet. Chem.* **1985**, *289*, 385-395.

³⁸ Darensbourg, D.J.; Joo, F.; Kannisto, M.; Katho, A.; Relbensples, J.H. Water-soluble organometallic compounds. 2. catalytic hydrogenation of aldehydes and olefins by new water-soluble 1,3,5-triaza-7-phosphaadamantane complexes of Ruthenium and Rhodium. *Organometallics* **1992**, *11*, 1990–1993.

³⁹ (a) Sasson, Y.; Blum, J. Homogeneous catalityc transfer-hydrogenenation of α , β -unsaturated carbonyl compounds by dichlorotris(triphenylphosphine)ruthenium (II). *Tetrahedron Lett.* **1971**, *24*, 2167–2170. (b) Blum, J.; Sasson, Y.; Iflah, S. Hydrogen transfer from formyl compounds to α , β -unsaturated ketones by Ru, Rh and Ir Complexes. *Tetrahedron Lett.* **1972**, *11*, 1015–1018. (c) Bar, R.; Sasson, Y. Catalytic reductions with formate ion under phase transfer conditions. *Tetrahedron Lett.* **1981**, *22*, 1709–1710.

⁴⁰ Wang, G.Z.; Backvall, J.E. Ruthenium-catalysed transfer hydrogenation of imines by propan-2ol. *J.C.S. Chem. Comm.* **1992**, 980-982.

⁴¹ Watanabe, Y.; Ohta, T.; Tsuji, Y.; Hiyoshi, T.; Tsuji, Y. Ruthenium catalyzed reduction of nitroarenes and azaaromatic compounds using formic acid. *Bull. Chem. Soc. Jpn.* **1984**, 2440–2444.
Hydrogen donor	Hydrogen acceptor	T °C	Product	Conversion (%)
BnOH		180		96
MeOH	^t Bu————————————————————————————————————	145	^t Bu————————————————————————————————————	77
HCO₂H	O C	97	° C	90
HCO₂H	o	125	OH	54
HCO₂H		18	N H	76
HCOONa -H₂O- THAHS		109		99

Table 1.1: Hydrogen transfer reactions catalyzed by $RuCl_2(PPh_3)_3$

1.2.5. RuCl₂(PPh₃)₃ in oxidation

 $RuCl_2(PPh_{3})_3$ catalyzes oxidation of alcohols to ketones by forming various catalytic systems with oxidants such as *t*-BuOOH,⁴² *N*-methylmorpholine *N*-oxide (NMO),⁴³ iodosylbenzene (PHIO)⁴⁴ or bis(trimethylsilyl)peroxyde (Me₃SiOOSiMe₃)⁴⁵ as shown in Scheme 1.27.



Scheme 1.27. Ru3-catalyzed oxidation of alcohols to ketones.

Additionally, secondary alcohols are readily converted to ketones by an excess of acetone in the presence of a catalytic amount of RuCl₂(PPh_{3)3.}⁴⁶

⁴² Murahashi, S-I.; Naota, T. Ruthenium-catalyzed oxidations for selective syntheses of ketones and acyl cyanides. Selective acylation of amino compounds with acyl cyanides. *Synthesis* **1993**, *4*, 433-440.

⁴³ (a) Sharpless, K.B.; Akashi, K.; Oshima, K. Ruthenium catalyzed oxidation of alcohols to aldehydes and ketones by amine-n-oxides. *Tetrahedron Lett.* **1976**, 17 (29), 2503–2506. (b) Vijayasri, K.; Rajaram, J.; Kuriacose, J.C. RuCl₂(PPh₃)₃ catalyzed oxidation of secondary alcohols with NMO. *J. Proc. Indian Acad. Sci. (Chem. Sci)* **1986**, *97*, 125-132.

⁴⁴ Miiller, P.; Godoy, J. Catalyzed oxidation of alcohols and aldehydes with iodosylbenzene. *Tetrahedron Lett.* **1981**, *22*, 2361-2364.

⁴⁵ Kanemoto, S.; Oshima, K.; Matzsubara, S.; Takai, K.; Nozaky, H. Transition-metal catalyzed oxidation of alcohols to aldehydes and ketones by means of Me₃SiOOSiMe₃. *Tetrahedron Lett.* **1983**, *24*, 2185-2188.

⁴⁶ Wang, G. Z.; Bäckvall, J.E. Ruthenium-catalysed oxidation of alcohols by acetone. *J. C. S. Chem. Comm.* **1992**, *4*, 337-339.

It is worth mentioning that aerobic oxidations of alcohols with RuCl₂(PPh₃)₃ catalyst under mild and ambient conditions constitutes an attractive, highly practical and efficient method for the preparation of aldehydes and ketones.⁴⁷

The low-valent ruthenium complex RuCl₂(PPH₃)₃ forms a catalytic system with *t*-Butyl hydroperoxide and catalyzes the oxidation of secondary and tertiary amines⁴⁸ (Scheme 1.28), amides,⁴⁹ phenols,⁵⁰ and various linear and cyclic alkanes.⁵¹



Scheme 1.28. Ru3/*t*BuOOH catalyzed oxidation of secondary amines/preparation of 3,4-dihydroisoquinoline.

Finally, both linear and cyclic alkanes can be converted efficiently into the corresponding ketones along with a small amount of alcohols.⁵² The system is highly useful for the oxidation of alkylated arenes, one of the examples of these

⁴⁷ (a) Matsumoto, M.; Ito, S. Ruthenium-catalysed oxidation of allyl alcohols by molecular oxygen. *J.C.S. Chem. Comm.* **1981**, 907-908. (b) Hanyu, A.; Takezawa, E.; Sakaguchi, S.; Ishii, Y. Selective aerobic oxidation of primary alcohols catalyzed by a Ru(PPh₃)₃Cl₂/hydroquinone system. *Tetrahedron Lett.* **1998**, *39*, 5557-5560. (c) Takezawa, E.; Sakaguchi, S.; Ishii, Y. Oxidative cleavage of vic-diols to aldehydes with dioxygen catalyzed by Ru(PPh₃)₃Cl₂ on active carbon. *Organic Lett.* **1999**, *1*, 713-715. (d) Dijksman, A.; Arends, I. W. C. E.; Sheldon, R. A. Efficient ruthenium-TEMPO-catalysed aerobic oxidation of aliphatic alcohols into aldehydes and ketones. *Chem. Comm.* **1999**, 1591-1592. (e) Dijksman, A.; Marino-González, A.; Payeras, A. M.; Arends, I. W. C. E.; Sheldon, R. A. Efficient and selective aerobic oxidation of alcohols into aldehydes and ketones using ruthenium/TEMPO as the catalytic system. *J. Am. Chem. Soc.* **2001**, *123*, 6826-6833.

⁴⁸ (a) Murahashi, S.; Naota, T.; Taki, H. Ruthenium-catalysed oxidation of secondary amines to Imines using *t*-Butyl hydroperoxide. *J. Chem. Soc., Chem Commun.* **1985**, 613-614. (b) Murahashi, S.; Naota, T.; Yonemura, K. Ruthenium-catalyzed cytochrome P-450 type oxidation of tertiary amines with alkyl hydroperoxides. *J. Am. Chem. Soc.* **1988**, *110*, 8256-8258.

⁴⁹ Murahashi, S. I.; Naota, T.; Kuwabara, T.; Saito, T.; Kumobayashi, H.; Akutagawa, S. Ruthenium-Catalyzed Oxidation of Amides and Lactams with Peroxides. *J. Am. Chem. Soc.* **1990**, *112*, 7820-7822.

⁵⁰ (a) Murahashi, S.; Naota, T.; Miyaguchi, N.; Noda, S. Ruthenium-catalyzed oxidation of phenols with alkyl hydroperoxides. *J. Am. Chem. Soc.* **1996**, *5*, 2509-2510. (b) Murahashi, S.; Miyaguchi, N.; Noda, S.; Naota, T.; Fujii, A.; Inubushi, Y.; Komiya, N. Ruthenium-catalyzed oxidative dearomatization of phenols to 4-(*tert*-butylperoxy) cyclohexadienones: Synthesis of 2-substituted quinones from *p*-substituted phenols. *Eur. J. Org. Chem.* **2011**, *27*, 5355-5365.

⁵¹ Murahashi. S. Biomimetic oxidation in organic synthesis using transition metal catalysts. *Pure App. Chem.* **1992**, *64*, 403-412.

⁵² (a) Murahashi, S.; Oda, Y.; Naota, T. Iron- and ruthenium-catalyzed oxidations of alkanes with molecular oxygen in the presence of aldehydes and acids. *J. Am. Chem. Soc.* **1992**, *114*, 7914-7916. (b) Murahashi, S.; Komiya, N.; Oda, Y.; Kuwabara, T.; Naota, T. Ruthenium-catalyzed oxidation of alkanes with *tert*-butyl hydroperoxide and peracetic acid. *J. Org. Chem.* **2000**, *65*, 9186-9193.

oxidations is displayed in Scheme 1.31 where the oxidation of 9-methyfluorene gave 9-*t*-butyldioxy-9-methylfluorene selectively with a 98% of yield.⁵³



Scheme 1.31. Ru-catalyzed oxidation of alkylated arene 9-methyfluorene.

1.3. Objectives

The main objective of this PhD thesis is to explore even more the potential of tris(triphenylphosphine)ruthenium(II) dichloride (RuCl₂(PPh₃)₃) and demonstrate that it is indeed a potent and versatile catalyst capable of achieving efficient chemical transformations under microwave activation especially in:

1. The synthesis of β -, γ - and δ -lactams using benzylic CH activation or Atom Transfer Radical Cyclizations.



2. The isomerization of tertiary *N*-allyl amides and *N*-allyl carbamates to the corresponding enamides and enecarbamates.



⁵³ Murahashi, S. I.; Oda, Y.; Naota, T.; Kuwabara, T. Ruthenium-catalyzed cytochrome P-450 type oxidation of alkanes with alkyl hydroperoxides. *Tetrahedron Lett.* **1993**, *34*, 1299-1302.

CHAPTER 2:

Microwave-assisted benzylic C-H activation using ruthenium catalysts. Synthesis of β -lactams

2.1. β-Lactams: a short overview

The β -lactam (2-azetidinone) ring is a four membered lactam found in many natural compounds and medicines especially antibiotics such as penams represented by penicillin G and V, cephems represented by cephalosporin C, monobactams represented by Aztreonam and finally carbapenems represented by Imipenem (Figure 2.1).¹ Nearly all of these antibiotics work by inhibiting bacterial cell wall biosynthesis.



Figure 2.1. Chemical structures of some β -lactam antibacterial drugs.

Besides forming part of the core structure of several antibiotic families, the β -lactam ring is found in many other natural and unnatural compounds with diverse activity.² For example in 2011, O'Boyle *et al.*³ described the synthesis of a series of 2-azetidinone derivatives via the Staüdinger reaction. These were then evaluated for antiproliferative, cytotoxic and tubulin-binding activity finding that

¹ Fernandes, R.; Amador, P.; Prudêncio, C. β-Lactams: chemical structure, mode of action and mechanisms of resistance. *Rev. Med. Microbiol.* **2013**, *24*, 7-17.

² Gupta, A.; Halve, A. K. β-lactams: a mini review of their biological activity. *Int. J. Pharmaceut. Sci. Res.* **2015**, *6*, 978-987.

³ O'Boyle, N.M.; Greene, L. M.; Bergin, O.; Fichet, J. B.; McCabe, T.; Lloyd, D. G.; Zisterer, D. M.; Meegan, M. J. Synthesis, evaluation and structural studies of antiproliferative tubulin-targeting azetidin-2-ones. *Bioorg. Med. Chem.* **2011**; *19*, 2306-2325.

the 3-(2-thienyl) and 3-(3-thienyl) analogs (Figure 2.2a) displayed the highest potency in human MCF-7 breast cancer cells with IC50 values of 7 nM and 10 nM respectively, comparable to combretastatin A-4.⁴ Additionally, Dou *et al.* have demonstrated that *N*-thiolated β -lactams have tumor cell-killing ability through induction of DNA-damage and subsequent apoptosis (Figure 2.2b).⁵ On the other hand, Hussain *et al.*⁶ reported the synthesis of β -lactam derivatives with significant antimycobarterial (antitubercular) activity (Figure 2.2c).



Figure 2.2. Structure of β -lactams with cytotoxic and antitubercular activity.

 β -Lactams have been prepared using a wide range of strategies,⁷ in scheme 2.1 are reported the most outstanding methodologies described so far. Among these are of course the Staudinger procedure,⁸ also called the Staudinger ketene-imine non-photochemical 2+2 cycloaddition, the carbonylative ring-

⁴. Cragg, G. M.; Kingston, D. G.; Newman, D. J. Anticancer Agents from Natural Products; CRC press: Florida, **2005**, pp. 123-135.

⁵ Smith, D. M.; Kazi, A.; Smith, L.; Long, T. E.; Heldreth, B.; Turos, E.; Dou, Q. P. A novel β -lactam antibiotic activates tumor cell apoptotic program by inducing DNA damage. *Mol. Pharmacol.* **2002**, *61*, 1348-1358.

⁶ Hussain, S.; Jadhav, S.; Rai, M.; Farooqui, M. Synthesis, Characterization and Biological Evaluation of N [3-Chloro-2 (aryl)-4-oxoazitidin-1-y] pyridine-4-carboxamide.*Int. J. Drug Des. Discov.* **2011**; *2*, 527-532.

⁷ For reviews see: (a) Hosseyni, S.; Jarrahpour, A. recent advances in β–lactams synthesis. *Org. Biomol. Chem.* **2018**, *16*, 6840-6852. (b) Pitts, C. R.; Lectka, T. Chemical Synthesis of β-Lactams: Asymmetric Catalysis and Other Recent Advances. *Chem. Rev.* **2014**, *114*, 7930-7953.

⁸ (a) Staudinger, H. Zur Kenntniss der Ketene. Diphenylketen. *Liebigs, Ann. Chem.* **1907**, *356*, (1–2), 51-123. (b) Palomo, C.; Aizpurua, J. M.; Ganboa, I.; Oiarbide, M. Asymmetric synthesis of β -lactams through the Staudinger reaction and their use as building blocks of natural and

nonnatural products. *Curr. Med. Chem.* **2004**, *11*, 1837-1872. (c) Berry, S.; Bari, S. S.; Banik, B. K.; Bhalla, A. Stereoselective synthesis of novel monocyclic trans-3-halogenated-4-pyrazolyl-β-lactams: potential synthons and promising biologically active agents. *Synth.Commun.* **2017**, *47*, 2239-2246.

opening of aziridines,⁹ the Kinugasa reaction which takes place between terminal alkyne and a nitrone in the presence of copper(I),¹⁰ and metal catalyzed C–H insertion of diazoamide carbenes activated by a variety of transition metals such as copper, gold, rhodium¹¹ and many others.¹²

⁹ Khumtaveeporn, K.; Alper, H. Transition Metal Mediated Carbonylative Ring Expansion of Heterocyclic Compounds. *Acc. Chem. Res.* **1995**, *28*, 414-422.

¹⁰ For selected examples, see: (a) Kinugasa, M. Hashimoto, S. The reactions of copper(I) phenylacetylide with nitrones. *J. Chem. Soc., Chem. Commun.* **1972**, 466-467; (b) Ahn, C.; Kenninghton, J. W.; DeShong, P. A New Approach to the Synthesis of Monocyclic β -lactam derivatives. *J. Org. Chem.*, **1994**, *59*, 6282–6286. (c) Marco-Contelles, J. *Angew. Chem., Int. Ed.* **2004**, *43*, 2198-2200. (d) Kumar, Y.; Singh, P.; Bhargava, G. Cu(I) mediated Kinugasa reactions of α , β -unsaturated nitrones: a facile, diastereoselective route to 3-(hydroxy/bromo)methyl-1-aryl-4-(-styryl)azetidin-2-ones. *New J. Chem.* **2016**, *40*, 8216-8219.

¹¹ Anada, M.; Watanabe, N.; Hashimoto, S. Highly enantioselective construction of the key azetidin-2-ones for the synthesis of carbapenem antibiotics via intramolecular C–H insertion reactions of α -methoxycarbonyl- α -diazoacetamides catalysed by chiral dirhodium(II) carboxylates. *Chem. Commun.* **1998**, 1517-1518.

¹² For reviews see: (a) Davies, H. M. L.; Beckwith, R. E. J. Catalytic enantioselective C-H activation by means of metal-carbenoid-induced C-H insertion. *Chem. Rev.* **2003**, *103*, 2861-2903. (b) Doyle, M. P.; Duffy, R.; Ratnikov, M.; Zhou, L. Catalytic Carbene Insertion into C-H Bonds. *Chem. Rev.* **2010**, *110*, 704-724.

a. The Staudinger ketene imine cycloaddition



b. Carbonylative ring-opening of aziridines



c. Kinugasa reactions



d. Metal catalyzed C-H insertion of diazoamide carbenes



Scheme 2.1. Selected examples for the synthesis of β -lactams.

2.2. Synthesis of β -lactams in the presence of RuCl₂(PPh₃)₃. Our results

Few years ago we reported the first Grubbs' second-generation (**Ru2**) catalyzed intramolecular dearomative ATRC with trichloroacetamides embodying an electron-rich arene for the preparation of 2-azaspirodecadienes **A2**.¹³ The investigation was an extension to a previous work where the same reaction leading to **A1** was achieved on unactivated benzenes using Cu(I) as a catalyst (Scheme 2.2).¹⁴

¹³ Diaba, F.; Martínez-Laporta, A.; Bonjoch, J. Atom transfer radical cyclization of trichloroacetamides to electron-rich acceptors using Grubbs' catalysts: Synthesis of the tricyclic framework of FR901483. *J. Org. Chem.* **2014**, *79*, 9365-9372.

¹⁴ Diaba, F.; Montiel, J. A.; Martínez-Laporta, A.; Bonjoch, J. Dearomative radical spirocyclization from *N*-benzyltrichloroacetamides revisited using a copper(I)-mediated atom transfer reaction leading to 2-azaspiro[4.5]decanes. *Tetrahedron Lett.* **2013**, *54*, 2619-2622.



Scheme 2.2. Group precedents in the synthesis of γ -lactams and actual work.

The reaction which will be developed in this chapter was accidently discovered when dibenzyl trichloroacetamide **1a** (Table 2.1) was submitted to the same reaction conditions in which spiroderivatives type **A2** were isolated. Thus when **1a** was heated with 10% of grubbs catalyst **Ru2** in toluene (0.1 mL) at 160 °C for 2 h, after chromatography, **2a** was isolated in a 26% yield (Entry 1, Table 2.1). Additionally, *N*,*N*-dibenzyl- and *N*-benzyldichloroacetamides were also isolated in minor yields. To our knowledge, this was an unprecedented ruthenium catalyzed synthesis of β -lactams from readily available *N*-benzyltrichloroacetamides and there is only one example reported in the literature where a saturated C(sp³)-C(sp³) bond formation is achieved through an asymmetric C-H functionalization from chloroacetamides catalysed by Palladium(0). The reaction takes place in the presence of Pd(dba)₂, a bulky taddol

phosphoramidite ligant, adamantly carboxylic acid and Cs₂CO₃ in toluene at 110 °C (Scheme 2.3).¹⁵



Scheme 2.3. Palladium(0) catalyzed synthesis of β -lactams.

It is worth noting that the same reaction, under diluted conditions (1 mL of toluene), did not take place when achieved using standard heating. Nevertheless, under microwave activation and at 100 °C a full conversion was observed after only 30 min obtaining **2a** in a slightly better yield (Entry 2). After these unexpected results, we started an investigation to explore the scope and limitations of this reaction. In order to improve the reaction yield, we switched to **1b** since the bulky *tert*-butyl substituent on the nitrogen atom is well known to lock the substrate in a conformation prone to cyclization.¹⁶

As it was expected, when **1b** was heated at 100 °C under microwave activation for only 10 min, lactam **2b** was isolated with 48% yield (Entry 3). Reducing the quantity of catalyst although prolonging the reaction time did not improve the final result (Entry 4). When the reaction was carried out at 80 °C, 20 min were necessary to obtain **2b** with 48% yield (Entry 5). Switching to acetonitrile, a full conversion was observed after only 5 min although the yield for lactam **2b** was lower (Entry 6). We next sought to explore the scope of the reaction on trichloroacetamides **1c-1h** with different substituents on the benzene ring. Treatment of substrates **1c-1e** all of them with a methyl group in the ortho, meta and para position respectively with **Ru2** (10 mol%) under microwave activation for 10 min and at 100 °C provided the corresponding lactams **2c-2e** with moderate yields (entries 7-9). With anisole derivative **1f**, a similar yield of lactam **2f** was obtained (Entry 10). Even if with these substrates usually some of

¹⁵ Pedroni, J.; Boghi, M.; Saget, T.; Cramer, N. Access to β -lactams by enantioselective palladium(0)-catalyzed C(sp³)-H alkylation. *Angew. Chem. Int. Ed.* **2014**, *53*, 9064-9067.

¹⁶ Stork, G., Mah, R. Radical cyclization of allylic haloacetamides a route to *cis*-fused 2-pyrrolidones and piperidones. *Heterocycles* **1989**, *28*, 723-727.

the starting material was recovered, prolonging the reaction time yielded poorer results.

Completely different results were attained with trichloroacetamides **1g** and **1h** with a deactivated benzene ring. Whereas with **1g** a degradation of the starting material was observed (Entry 11), with **1h** lactam **2h** was isolated with a very low yield together with product **3h** which was not observed in the reaction crude but after chromatography using dichloromethane stabilized with ethanol as the eluent (Entry 12). It is worth noting that when dichloromethane stabilized with amylene was used instead in the purification process, we recovered only degradation compounds.

		Ru2 (10%)			$CI \qquad O$ $CI \qquad N$ R_1	
	R	$R_1 R_2$				R ₂ -
	1a E	Bn H	1f	<i>t</i> -Bu	3-OMe	
	1b <i>t</i> -	-Bu H	1g	<i>t</i> -Bu	4-NO ₂	
	1c t-	-Bu 2-Me	1h	<i>t</i> -Bu	3,5-diF	
	1d t	-Bu 3-Me				
	1e t-	-Bu 4-Me				
Entry	1	Metho	d ^a	Tim	e (min)	2 , 3 (yield %)
1	1a	А			120	2a (26)
2	1a	В			30	2a (30)
3	1b	В			10	2b (48)
4	1b	Bb			20	2b (38)
5	1b	Bc			20	2b (48) ^d
6	1b E		^e 5		5	2b (36)
7	1c B		10		10	2c (30) ^f
8	1d	В			10	2d (38) ^g
9	1e	В			10	2e (28)
10	1f	В			5	2f (25) ^h
11	1g	В			10	2g (-)
12	1ĥ	В			5	2h (11) ⁱ , 3h (24) ^j

Table 2.1. Ru2 promoted formation of lactams 2

^{*a*} A: 100 mg scale in 0.1 mL of toluene (≈ 0.3 M) at 160 °C. B: 100 mg scale in 1 mL of toluene (≈ 0.3 M) at 100 °C, μ W. ^{*b*} 5% of **Ru2** was used, 35% of **1b** was recovered. ^{*c*} At 80 °C. ^{*d*} 7% of **1b** was recovered. ^{*e*} Acetonitrile was used as a solvent. ^{*f*} 22% of **1c** was recovered. ^{*g*} 20% of **1d** was recovered. ^{*h*} 23% of **1f** was recovered. ^{*i*} 8% of **1h** was recovered. ^{*j*} For the structure of **3h** see table 2.

After these results with **Ru2** we decided to investigate Grubbs' 1st generation catalyst (**Ru1**) using the best reaction conditions established with **Ru2** (Table 2.2). With dibenzyl derivative **1a** the reaction provided lactam **2a** with a comparable yield (Entry 1). Nevertheless, with trichloroacetamide **1b** only **3b** was isolated after chromatography (Entry 2). As it was mentioned before, **3b** was not observed in the crude but isolated after purification. A similar result was achieved with substrates **1g** and **1h** with an unactivated benzene where only **3g** and **3h** were isolated respectively (entries 7 and 8). With tolyl derivatives **1c-1e**, besides lactams **2c-2e** isolated in modest yields, amides **3c-3d** were detected in some fractions after chromatography but were not isolated in pure forms (entries 3-5). It's worth noting that with trichloroacetamide **1c** a better yield of lactam **2c** was obtained (Entry 3). Additionally, with anisole **1f** only **3f** was isolated after purification (Entry 6).

$\begin{array}{c} \begin{array}{c} & & \\ $								
Entry	1	Time (min)	2 (yield %)	3 (yield %)				
1	1a	20	2a (28)	3a (-)				
2	1b	5	2b (-)	3b (38)				
3	1c	10	2c (40)	3c (9)				
4	1d	5	2d (16)	3d (8) ^b				
5	1e	10	2e (34)	3e (-) ^c				
6	1f	5	2f (-) ^c	3f (37)				
7	1g	10	2g (-)	3g (17)				
8	1h	5	2h (-)	3h (46)				

Table 2.2. Reactions of trichloroacetamides 1 with Ru1^a

^a All the reactions were carried out using almost 100 mg scale of **1** in 1 mL of toluene (≈ 0.3 M) at 100 °C, μ W.^b 12% of **1d** was recovered. ^c Only traces were detected.

With these results in hand, we set out to explore another more accessible ruthenium catalyst for these unusual reactions (Table 2.3). RuCl₂(PPh₃)₃ (**Ru3**) is a commercially available catalyst, more accessible than **Ru2** and is the precursor of **Ru1**. When **1a** was treated with 10% of **Ru3** at 100 °C in toluene for 15 min using microwaves, we were delighted to see that lactam **2a** was isolated

with a modest yield (Table 3, Entry 1). Running the same reaction at 80 °C improved the yield considerably (41%, Entry 2).

	R ₂ CCl ₃	RuCl ₂ (PPh ₃) ₃ (΄ toluene, μW	10%)		R_1 R_2	H N CHCl ₂ OEt O
	N `O I _R 1		ľ.	R_2	3	
Entry	R ₁	R ₂	1	T °C	Time (min)	2, 3 (yield %)
1	Bn	Н	1a	100	15	2a (20) ^b
2	Bn	н	1a	80	20	2a (41)
3	^t Bu	Н	1b	100	10	2b (49)
4	^t Bu	Н	1b	80	10	2b (58)
5	^t Bu	Н	1b	80	10	2b (55) [◦]
6	^t Bu	2-Me	1c	80	10	2c (56) ^d
7	^t Bu	3-Me	1d	80	10	2d (40), 3d (8)
8	^t Bu	4-Me	1e	80	5	2e (40)
9	^t Bu	3-OMe	1f	80	10	2f (18)
10	^t Bu	4-NO ₂	1g	80	10	-
11	^t Bu	3,5-diF	1h	80	10	2h (21), 3h (18)
12	<i>n</i> -Bu	Н	1i	80	10	2i (30) ^e
13	<i>i</i> -Pr	Н	1j	80	10	2j (28 ^f
14	Су	н	1k	80	10	2k (38) ^g
13	(S)-α- MeBn	Н	11	80	10	6I (40) ^h

Table 2.3. RuCl₂(PPh₃)₃ promoted formation of lactams 2^a



^a All the reactions were carried out using 100 mg scale of **1** in 1 mL of toluene (≈ 0.3 M), μ W.^b6% of **1a** was recovered. ^c With 200 mg of **1b**. ^d 12% of **1c** was recovered. ^e Were also isolated **4i** (4%), **5i** (9%), **1i** (25%). ^{*f*} Traces of **4j** and **5j** were detected. ^g Were also isolated **4k** (8%), **5k** (9%). ^{*h*} 48% of **1I** was recovered, when the reaction was run for 20 min the yield dropped to 22% even if the conversion was 100%.

Better results were also achieved from trichloacetamides **1b-1e** where only lactams **2b-2e** were isolated with 40-58% yields (Entries 3-8). Additionally, with electron-poor derivatives **1g** and **1h** a different behaviour was observed. Whereas with **1g** only degradation compounds were detected in the crude, with **1h** a slight improvement in the yield of lactam **2h** was achieved isolated with **3h** (Entry 11). Using catalyst **Ru3** the reaction worked also with amides **1i-1k** where the substituents on the nitrogen are *n*butyl, isopropyl and cyclohexyl groups respectively (Entries 12-14). In all cases lactams **2i-2k** were isolated with acceptable yields. It is worth noting that with **1i** and **1k**, we isolated also small quantities of the partially reduced lactams **4** and **5**.¹⁷ Finally, with trichloroacetamide **1I**, only dichloroacetamide **6I** was isolated (Entry 13).

In this investigation we have described an unprecedented behaviour of some ruthenium catalysts leading to β -lactams **2** from trichloroacetamides **1** through benzylic CH activation. The presence of the catalyst is necessary to achieve the process since after heating 1b alone in toluene at 80 °C under microwave activation for 10 min no reaction took place and 1b was recovered intact. As it was expected, a radical process is involved in this transformation since running the reaction in the presence of TEMPO either from 1a or 1b inhibits the lactam formation. Whereas under microwave activation only degradation compounds were detected in the crude, when 1a was heated at 160 °C with Ru3 (10 mol%) in the presence of 1 equiv. of TEMPO, besides 1a a small quantity of 7a was detected resulting from a coupling between TEMPO and the dichlorocarbamoil radical followed by hydrolysis (Scheme 2.4).¹⁸ 7a was identified by high-resolution mass spectroscopy (ESI-TOF) which confirmed the molecular formula C₂₅H₃₂N₂O₃ (calculated for C₂₅H₃₃N₂O₃ 409.2486 [M+H]+; found 409.2488). This indicates that the first step of the reaction is abstraction of a chloro atom by the "Ru" catalyst (the modified species) from 1 to generate dichlorocarbamoil radical I and "RuCl" complex.¹⁹ Next, we propose that the

¹⁷ Van Driessche, B.; Van Brabandt, W.; D'hooghe, M.; Dejaegher, Y.; De Kimpe, N. Synthesis and reactivity of trans-2-aryl-3-chloroazetidines. *Tetrahedron* **2006**, *6*2, 6882-6892.

¹⁸ Baker, A. D.; Wong, D.; Lo, S.; Bloch, M.; Horozoglu, G.; Golman, N. L.; Engel, R. The reaction of *N*-aryl nitrones with dichloroketene: a new synthesis of isatins. *Tetrahedron Lett.* **1978**, *19*, 219-222.

¹⁹ Matsumoto, H.; Nikaido, T.; Nagai, Y. Radical reactions in the coordination sphere. III. Reactions of dichloro- and trichloroacetic acid esters with 1-olefins catalyzed by dichlorotris-(triphenylphosphine)ruthenium(II). *J. Org. Chem.* **1976**, *41*, 396-398.

"RuCl" complex abstracts then a hydrogen atom from radical I to generate diradical species II with the concomitant release of HCl and regeneration of "Ru" catalyst. Indeed analysis of some reaction crudes in which **Ru3** was successfully used in the synthesis of lactams **2** showed a strong acidic pH of the reaction medium. Next, transient diradical II undergoes an intramolecular cross coupling to generate β -lactams **2**. On the other hand amides **3** result from cleavage of the *tert*-butyl group in acid medium²⁰ and a 1,4-H shift²¹ in radical I generating radical III followed by a chloro atom transfer from "RuCl" complex or amide **1** to provide chloroderivative IV. The latter undergoes nucleophilic substitution during purification providing ethers **3**.²²



Scheme 2.4. Proposed mechanism for the formation of 2 and 3.

As it was mentioned before, ethers **3** were not present in the reaction crudes but were isolated after purification by chromatography using CH₂Cl₂ stabilized with ethanol. Formation of chloroderivative **IV** was reinforced by analysis of the reaction mixture when achieved from **1h** and **Ru1** (Table 2.2, Entry 8) since ¹H NMR spectrum exhibits 2 determinant signals, a singlet at 5.89 ppm and a doublet at 6.32 ppm consistent with a COCHCl₂ and a benzylic CHCl respectively. This argument is supported by the experiments achieved with **1I**

²⁰ Clayden, J.; Stimson, C. C.; Keenan, M. Contra-Friedel–Crafts *tert*-butylation of substituted aromatic rings *via* directed metallation and sulfinylation. *Chem. Comm.* **2006**, 1393-1394.

²¹ (a) Quirante, J.; Diaba, F.; Vila, X.; Bonjoch, J.; Lago, E.; Molins, E. An unexpected course in the 6-exo radical cyclizations of trichloroacetamides on changing the *N*-substituent from benzyl to α -methylbenzyl. *C. R. Acad. Sci.* **2001**, *4*, 513-521. (b) Nechab, M.; Mondal, S.; Bertrand, M. P. 1,n-Hydrogen-atom transfer (HAT) reactions in which n \neq 5: an updated inventory. *Chem. Eur. J.* **2014**, *20*, 16034-16059.

²² Hiemstra, H.; Fortgens, H. P.; Speckamp, W. N. Intramolecular reactions of acyclic *N*-acyliminium ions II allyl silanes as nucleophiles. *Tetrahedron Lett.* **1985**, *26*, 3155-3158.

where only alkene **6I** was isolated. With this substrate, we believe that after formation of the dichlorocarbamoil radical, a 1,4-H shift takes place to generate a radical followed by a chloro atom transfer and elimination to furnish **6I**.

2.3. Cytotoxic activity of some of the synthesized β -lactams

With the background reported in section 2.1., it was of great interest in this study to investigate and characterize some of the β -lactams prepared, naming **2b**, **2c**, **2e**, **2h**, **2j** and **2k** for their cytotoxic activity and hemocompatibility.²³



Figure 2.3. Selected lactams for cytotoxic and hemocompatibility studies.

2.3.1 Hemocompatibility studies

Following the ISO 10993-4 concerning the biological evaluation of medical devices and their interactions with blood, an *in vitro* hemocompatibility assay was carried out.²⁴ Under the assayed conditions, the determination of the degree of hemolysis produced by the different compounds by incubation with red blood cells (RBC) suspension was performed. The chemical modification on the hemolytic response was evaluated at 2 different concentrations (10 and 80 µg/mL). Table 2.4 shows representative results as a function of the chemical structure. The degree of hemolysis fluctuates slightly, with values ranging between 0.01 and 0.05% for compounds **2b-2j** at concentrations equal to 10 µg/mL. The maximum degree of hemolysis was achieved with compound **2k** with hemolysis close to 0.25 %. By increasing concentration up to 80 µg/mL, no appreciable increase in the hemolytic response was observed. Only under discrete conditions, the hemolysis increase until 2.5 % (compound **2b**). Considering the criteria for which compounds are classified as non-hemolytic (<2%), slightly hemolytic (with values

²³ This part of work was achieved in collaboration with Dr Maria Del Carmen Moran Badenas, Department of Biochemistry and Physiology, University of Barcelona.

²⁴ ISO 10993-4:2017. Biological evaluation of medical devices-Part 4: Selection of tests for interactions with blood.

2-5%) and hemolytic (values > 5%), it could be concluded that the proposed compounds showed non-hemolytic properties.

 Table 2.4. Percentage of hemolysis induced by compounds 2b-2k as a function of the concentration.

Compound	Hemolysis (%)				
	10 µg/mL	80 μg/mL)			
2b	0.01 ± 0.01	2.5 ± 0.97			
2c	0.01 ± 0.01	0.01 ± 0.01			
2e	0.01 ± 0.01	0.2 ± 0.23			
2h	0.05 ± 0.05	0.12 ± 0.09			
2j	0.01 ± 0.01	0.02 ± 0.02			
2k	0.25 ± 0.03	0.30 ± 0.21			

2.3.2 Cell viability studies

After ensuring that the selected β -lactams have non-hemolytic character, we were very interested in investigating the cytotoxic activity they induced. For this purpose we used the immortal human keratinocyte (HaCaT) and the squamous cell carcinoma (A431), as close representative skin cell lines with non-tumoral and tumoral characteristics, respectively. Two different end-points, MTT and NRU, were used to assess differences in cell-induced cytotoxicity. In the former method, this offers details about the modification of the metabolic activity of mitochondria inside the cells. In the latter case, however, the derived information is related with the interaction with the plasmatic membrane. Dose–response curves were determined by the MTT²⁵ and NRU²⁶ assays using HaCaT and A431 cell lines. Cytotoxicity assays were performed at concentrations ranged between 10 and 250 µg/mL.

²⁵ For more details about the protocols see the experimental part related to chapter 2.

²⁶ Repetto, G.; Peso, A.; Zurita, J. L. Neutral red uptake assay for the estimation of cell viability/ cytotoxicity. *Nat. Protoc.* **2008**, *3*, 1125-1131.

The results demonstrated that the cytotoxic response is highly dependent on the structural characteristics of β -lactam compounds and their concentration. In general, cell viability decreases with the increasing of the concentration showing a dose-concentration response (Figure 2.4). However, the final response seems to be a function of the β -lactam derivatives. Compound **2b**, **2e**, **2j**, and **2k** were able to decrease the percentage of viability compared to control cells with values ranged down to 20%, as a function of cell type and endpoint method.



Figure 2.4. Concentration-dependent relative viabilities of HaCaT and A431 cells treated with β -lactams derivatives for 24 h determined by MTT (solid lines) and NRU (dotted lines) assays. The data correspond to the average of three independent experiments ± standard deviation.

Compounds **2c** and **2h** however, promote cell viability up to 50% in all cases. Moreover, the structure of the β -lactam determined the selectivity against the cell type and the mode of action. Hence, compound **2e** promotes higher cytotoxicity in HaCaT cells than in A431 cells regardless the concentration range. Meanwhile, oppositely results were observed in the case of compound **2h**. There are not big differences between the cellular responses when the other β -lactams were considered.

As a general trend, these results suggested minor differences between the mode of action of the tested compounds with the plasma membrane and lysosomal accumulation (NRU method) or the modification of the metabolic activity of mitochondria inside the cells (MTT method). In all cases, these compounds can achieve the mitochondrial compartment after alteration of the plasma membrane.

From the fitting of concentration-dependent viabilities curves, the corresponding half-maximal inhibitory concentration (IC₅₀) was determined. The obtained results have been summarized in Figure 2.5 and Table 6. The structural characteristics of β -lactam derivatives seem to be the main factor in the tested compounds' cytotoxic response. Compound **2b**, as a reference compound, demonstrated to be the most cytotoxic compound showing the lowest IC₅₀ values (20-49 µg/mL and 30-47 µg/mL for HaCaT and A431 cell lines, respectively). The cellular response depends on the nature of the substituent on the benzene ring and its location. Introduction of 2 fluorine atoms (compound **2h**) generates a more biocompatible compound, with IC₅₀ values close to 250 µg/mL (A41 cell line) or higher than 250 µg/mL (HaCaT cell line), which corresponds to the highest tested concentration, in an independent manner of the endpoint method.

The location of the methyl group in the benzene ring is also an important parameter on the cell response. Thus, when the CH₃ is in the ortho position (compound **2c**) IC₅₀ values are close to 250 μ g/mL (HaCaT and A41, MTT endpoint) and higher than 250 μ g/mL (HaCaT and A431, NRU endpoint). However, when the methyl substituent is in the para position (compound **2e**), the compound preserves the cytotoxic activity with IC₅₀ values between 78-93 μ g/mL (HaCaT) and 120-180 μ g/mL (A431).

43

Switching from *t*-Bu to *i*-Pr (compound **2***j*) have a poor influence on the cytotoxic response since IC₅₀ values between 40-42 μ g/mL (HaCaT) and 38-76 μ g/mL (A431) were found. However, in the case of the cyclohexyl derivative **2k**, limited interaction with the proposed cells was observed as it is indicated by the IC₅₀ values close to 220 μ g/mL (HaCaT, NRU) or higher than 250 μ g/mL (HaCaT, MTT and A41 in all cases).



Figure 2.5. IC₅₀ values of the corresponding β -lactams on HaCaT (a) and A431 (b) cell line as a function of endpoint method. The data correspond to the average of three independent experiments ± standard deviation.

When the IC_{50} values as a function of the endpoint method were compared, the obtained results suggested that, in general, the interaction with the cell membrane is favored. In almost all cases, IC_{50} values corresponding to the NRU method showed lower values than those obtained in the MTT endpoint. Moreover, from the values displayed in Table 2.5, selectivity Index (SI) seems to depend on the endpoint method. In the case of MTT method, poor selectivity was detected. In all cases, SI values are equal or lower than 1, demonstrating no selectivity in the mode of action. However, some selectivity among the proposed cell lines was observed when the NRU endpoint was considered, demonstrating discrete selectivity in the mode of action of almost all compounds (except for compound **2e**). **Table 2.5**. IC₅₀values of the corresponding β -lactams as a function of cell line and endpoint method. Selectivity index against the tumor cell line (A431) in comparison with the non-tumor cell line (HaCaT).

Compound	IC ₅₀ HaCaT (ug/mL)		IC 50 A431 (ug/mL)		SI	
	MTT	NRU	MTT	NRU	MTT	NRU
2b	19.82	48.27	46.93	29.54	0.42	1.63
2c	250	>250	250	250	1.0	> 1.0
2e	92.61	78.82	180	120.72	0.51	0.65
2h	>250	>250	250	250	> 1.0	> 1.0
2j	42.10	40.14	76.19	37.88	0.55	1.06
2k	>250	>250	250	220	> 1.0	> 1.13



CHAPTER 3:

Microwave assisted isomerization of N-allyl carbamates and N-allyl amides to Z-enecarbamates and Z-enamides catalyzed by RuCl₂(PPh₃)₃

3.1. Enamides and enecarbamates in natural compounds and organic synthesis

The enamide motif is found in several alkaloids and drug candidates as E and the less thermodynamically stable Z isomers (Figure 3.1). Moreover, enamides and enecarbamates are stable enamine surrogates with high synthetic potential used in many chemical transformations as a result of the *N*-withdrawing substituent present on the nitrogen atom.¹ Indeed, in contrast to enamines, they are stable to hydrolysis and maintain their predisposition to electrophilic activation.



Figure 3.1. Natural compounds with the enamide motif with anti-cancer activity.

Thus enamides and enecarbamates have been successfully used in asymmetric hydrogenations catalyzed by transition metal complexes,^{1d,2} halo

¹ (a) Matsubara, R.; Kobayashi, S. Enamides and enecarbamates as nucleophiles in stereoselective C–C and C–N bond-forming reactions. *Acc. Chem. Res.* **2008**, *41*, 292-301. (b) Carbery, D. R. Enamides: valuable organic substrates. *Org. Biomol. Chem.* **2008**, *6*, 3455-3460. (c) Nugent, T. C.; El-Shazly, M. Chiral Amine Synthesis-Recent developments and trends for enamide reduction, reductive amination, and imine reduction. *Adv. Synth. Catal.* **2010**, *352*, 753-819. (d) Gopalaiah, K.; Kagan, H. Use of nonfunctionalized enamides and enecarbamates in asymmetric synthesis. *Chem. Rev.* **2011**, *111*, 4599–4657. (e) Bernadat, G.; Masson, G. Enamide derivatives: versatile building blocks for highly functionalized α,β -substituted amines. *Synlett* **2014**, *25*, 2842-2867.

² Vineyard, B. D.; Knowles, W. S.; Sabacky, M. J.; Bachman, G. L.; Weinkauff, D. J. Asymmetric hydrogenation. Rhodium chiral bisphosphine catalyst. *J. Am. Chem. Soc.* **1977**, *99*, 5946-5952.

functionalizations,³ transition metal mediated C–C bond formations,⁴ and cyclopropanations⁵ among others (Scheme 3.1).

1. Asymmetric hydrogenation



3. Transition metal mediated C-C bond formation (hydroalkynylation)



4. Cyclopropanation



Scheme 3.1. Examples of chemical transformations from enamides and enecarbamates.

The synthetic potential of enamides and enecarbamates has been readily demonstrated in the total synthesis of several natural and unnatural molecules

³ Alix, A.; Lalli, C.; Retailleau, P.; Masson, G. Highly enantioselective electrophilic α-bromination of enecarbamates: chiral phosphoric acid and calcium phosphate salt catalysts. *J. Am. Chem. Soc.* **2012**, *134*, 10389-10392.

⁴ Bai, X.-Y.; Wang, Z.-X.; Li, B.-J. Iridium-catalyzed enantioselective hydroalkynylation of enamides for the synthesis of homopropargyl amides. *Angew. Chem., Int. Ed.* **2016**, *55*, 9007-9011.

⁵ Song, Z.; Lu, T.; Hsung, R. P.; Al-Rashid, Z. F.; Ko, c.; Tang, Y. Stereoselective Simmons-Smith cyclopropanation of chiral enamides. *Angew. Chem., Int. Ed.* **2007**, *46*, 4069-4072

displaying biological activity.⁶ One of the outstanding examples was reported in the total syntheses of (±)-stemonamide and (±)-isostemonamide. Indeed starting from an enamide through a 7-endo/5-endo radical-cascade cyclization via α -amidoyl radical in the presence of tributyltin hydride and 1,1'-azobis(cyclohexanecarbonitrile) (ACN) advanced tricyclic intermediates of these natural compounds were achieved (Scheme 3.2).⁷



(±)-stemonamide (±)-isostemonamide

Scheme 3.2. Enamide derivative in the total synthesis of (±)-stemonamide and (±)-isostemonamide.

3.2. Synthetic precedents in the preparation of enamides and enecarbamates

Enamides and enecarbamates can be prepared by combining the structure parts, usually two, using traditional methods such as acylation of imines,⁸ the Curtius rearrangement of α , β -insaturated acyl azide derivatives,⁹

⁶ For representative reviews, see: (a) Courant, T.; Dagousset, G.; Masson, G.; Enamide derivatives: versatile building blocks for total synthesis. *Synthesis* **2015**, *47*, 1799-1826.

⁽b) Cai, X.; Yang, M.; Guo, H. Tertiary enamides: versatile and available substrates in synthetic chemistry. *Curr. Org. Synth.* **2019**, 16, 70–97. (c) Beltran, F.; Miesch, L. Tertiary enamides as versatile and valuable substrates to reach chemical diversity. *Synthesis* **2020**, *52*, 2497-2511. (d) Ninomiya, I.; Naito, T. in "The Alkaloids, Chemistry and Pharmacology", ed. Brossi, A., Vol. 22, Academic Press, New York, **1983**, pp. 189-279.

⁷ Taniguchi, T.; Tanabe, G.; Muraoka, O.; Ishibashi, H. Total synthesis of (±)-stemonamide and (±)-isostemonamide using a radical cascade. *Org. Lett.* **2008**, *10*, 197-199.

⁸ (a) Breederveld, H. The chemistry of the *N*-alkylaldimines. I. The reaction of *N*-alkylaldimines with acetic anhydride. *Recl. Trav. Chim. Pays-Bas* **1960**, *79*, 401-407. (b) Cassayre, J.; Quiclet-Sire, B.; Saunier, J.-B.; Zard, S. Z. β - and γ -Lactams by nickel powder mediated 4-*exo* or 5- *endo* radical cyclisations. A concise construction of the mesembrine skeleton. *Tetrahedron* **1998**, *54*, 1029-1040.

⁹ Chuang, T.; Wu, P.J. Synthesis and mechanistic study of isoquinolinones from cinnamoyl azides. *Chin. Chem. Soc.* **2006**, *53*, 413-420.

1. Acylation of imines



2. Curtius rearrangement

Ref. 9



3. Horner-Wittig reaction

Ref. 10



4. Reductive acetylation of ketoximes

Ref. 11



5. Dehydrogenation of amides

Ref. 12



Scheme 3.3. Some of the strategies in the synthesis of enamides.

Horner-Wittig reaction,¹⁰ or transformation of a ketone into an oxime followed by subsequent reduction with iron metal in the presence of acetic anhydride¹¹ (Scheme 3.3). Even though these procedures are still used for the preparation of enamides and enecarbamates, they usually provide a mixture of *E-Z* isomers with low to moderate yields. Recently, a direct synthesis of enamides by dehydrogenation on amides using a combination LiHMDS and triflic anhydride was reported providing exclusively *E*-enamides when the reaction was run from acyclic amides (Scheme 3.3).¹²

Another atom-economic synthesis of enamides is the transposition of the terminal carbon–carbon double bond of the corresponding readily available allyl amides.¹³ This transformation has been carried out with many experimental procedures. Among these are the use of strong bases such as LDA or BuLi.¹⁴ Thus operating from para-substituted *N*-allylbenzamides, with LDA at -78°C, the corresponding (*E*)-*N*-(prop-I-enyl)benzamide was the major product whereas with *n*butyllithium and the mixture warmed to 0 °C the (*Z*)-isomer was essentially furnished (Scheme 3.4).



Scheme 3.4. Isomerization of allylamides in the presence of strong bases.

The transition metal catalyzed isomerization of *N*-allyl amides to enamides is another alternative which has been explored in the presence of many transition metal complexes.¹³

¹⁰ Broekhof, N.L.J.M.; Jonkers, F.L.; van der Gen, A. Enamine synthesis by the Horner-Wittig reaction. *Tetrahedron Lett.* **1979**, 20, 2453-2456.

¹¹ Burk, M. J. Casy, G.; Johnson, N. B. A Three-step procedure for asymmetric catalytic reductive amidation of ketones. *J. Org. Chem.* **1998**, *63*, 6084-6085.

¹² Spieß, P.; Berger, M.; Kaiser, D.; Maulide, N. Direct Synthesis of Enamides via Electrophilic Activation of Amides. *J. Am. Chem. Soc.* **2021**, *143*, 10524–10529.

¹³ Krompiec, S.; Krompiec, M.; Penczek, R.; Ignasiak, H. Double bond migration in *N*-allylic systems catalyzed by transition metal complexes. *Coord. Chem. Rev.* **2008**, *252*, 1819-1841.

¹⁴ Ribereau, P.; Delamare, M.; Celanire, S.; Queguiner, G. Selective preparation of (*Z*)- and (*E*)-prop-1-enylamides from *N*-allylbenzamides. *Tetrahedron Lett.* **2001**, *4*2, 3571–3573.

In the presence of ruthenium complexes naming Grubbs' 2nd generation catalyst (Ru1) from allyl sulfonamide, the isomerization takes place providing a mixture of E/Z isomers with a poor yield (Scheme 3.5a).¹⁵ The reaction was also performed from substrates that might experience metathesis as a competitive isomerization. this process to In case the simple addition of vinyloxytrimethylsilane (1 equiv.) is sufficient to provide the corresponding enamide as a mixture of E/Z isomers with good yields (Scheme 3.5b).¹⁶ Isomerization of N-allyl-N-arylethanamides catalyzed by [RuCIH(CO)(PPh₃)₃] (Ru2) was also investigated. Thus operating from N-allyl-N-arylethanamides only the corresponding (E)-N-aryl-N-(1-propenyl)ethanamides were isolated with good yields (Scheme 3.5c).¹⁷ In a similar work, using the same catalyst, N-allyl 2iodoacetanilide provided only the (E)-enamide with a very good yield (Scheme 3.5d).¹⁸ Additionally, isomerization of *N*-allylphtalimide was achieved in the presence of $RuCl_2(PPh_3)_3$ (**Ru3**) to provide exclusively (*E*)-*N*-propenylphthalimide (Scheme 3.5e).¹⁹ To our knowledge this is the only example where **Ru3** is used as a catalyst for allylamide isomerization. Isomerization of secondary allylamides and alkenes was also performed in the presence of Ru4 also called the "alkene zipper" to give exclusively the E-isomers with supposedly very good NMR yields but confusing experimental data about the catalyst loading and the yields obtained (Scheme 3.5f).²⁰

¹⁵ Cadot, C.; Dalko, P. I.; Cossy, J. Olefin isomerization by a ruthenium carbenoid complex. Cleavage of allyl and homoallyl groups. *Tetrahedron Lett.* **2002**, *43*, 1839-1841.

¹⁶ Arisawa, M.; Terada, y.; Nakagawa, M.; Nishida, A. Selective isomerization of a terminal olefin catalyzed by a ruthenium complex: The synthesis of indoles through ring-closing metathesis. *Angew. Chem. Int. Ed.* **2002**, *41*, 4732-4734.

¹⁷ Krompiec, S.; Pigulla, M.; Szczepankiewicz, W.; Bieg, T.; Kuznik, N.; Leszczynska-Sejda, K.; Kubicki, M.; Borowiak, T. Highly selective synthesis of (*E*)-*N*-aryl-*N*-(1-propenyl) ethanamides via isomerization of *N*-allyl ethanamides catalyzed by ruthenium complexes. *Tetrahedron Lett.* **2001**, *42*, 7095–7098.

¹⁸ Dominguez, G.; Casarrubios, L.; Rodriguez-Noriega, J.; Perez-Castells, J. Indole and Quinoline synthesis *via* intramolecular Pauson-Khand reactions of enamines and allylamines. *Helv. Chim. Acta* **2002**, *85*, 2856-2861.

¹⁹ Delogu, G.; Faedda, G.; Gladiali, S. Hydrocarbonylation of unsaturated nitrogen compounds. Synthesis of *N*-protected aminoacid derivatives from *N*-substituted phtalimides. *J. Organomet. Chem.* **1984**, *268*, 167-174.

²⁰ Larsen, C. R.; Grotjahn, D. B. Stereoselective alkene isomerization over one position. *J. Am. Chem. Soc.* **2012**, *134*, 10357-10360.



a.Ref. 15



b.Ref. 16



R =H, o-Me, p-Me, o-MeO, m-MeO,p-MeO,o-Cl, p-Cl, o-Br, p-Br

d. Ref. 18



Scheme 3.5. Isomerization of allylamides in the presence of Ru catalysts.

The transition metal catalyzed isomerization of *N*-allylamides and *N*-allylcarbamates was also achieved in the presence of Fe (Scheme 3.6a),²¹ Ir (Scheme 3.6b),²² Rh²³ or Ni²⁴ complexes where a mixture of *E*/*Z* isomers or a strong *E*-selectivity in the isomerization was observed (Scheme 3.6).



Scheme 3.6. Isomerizations of *N*-allyl amides and *N*-allyl carbamates in the presence of Fe, Ir, Rh and Ni complexes.

 ²¹ (a) Hubert, A. J.; Moniotte, P.; Goebbels, G.; Warin, R.; Teyssie, P. Catalysed prototropic rearrangements. Part II. Metal carbonylcatalysed isomerization of *N*-allylamides to prop-2-enyl derivatives. *J. Chem. Soc., Perkin Trans.* 2 1973, 1954-1957. (b) Sergeyev, S.; Hesse, M. A new convenient method for the preparation of enamides from *N*-allylamides. *Synlett* 2002, 1313-1317.
 ²² Neugnot, B.; Cintrat, J-C; Rousseau, B. A new highly chemoselective isomerization of allylamides. *Tetrahedron* 2004, *60*, 3575-3579.

²³ Zacuto, M. J.; Xu, F. One-Step *RhCl*₃-catalyzed deprotection of acyclic *N*-allyl amides. J. Org. Chem. **2007**, 72, 6298-6300.

²⁴ Wang,L.; Liu, C.; Bai, R.; Pana, Y.; Lei, A. Easy access to enamides: a mild nickel-catalysed alkene isomerization of allylamides. *Chem. Commun.* **2013**, *49*, 7923.

As it was clearly indicated in the examples mentioned before, in the presence of transition metal catalysts, isomerization of *N*-allyl amides and *N*-allyl carbamates usually provides an E/Z-mixture or the thermodynamically favored Econfigured enamides or enecarbamates. The achievement of thermodynamically less stable Z-configured enamide through double-bond migration is rarely reported, still under-represented and limited to secondary amides (Scheme 3.7).²⁵ Moreover the examples described are not very clear about the exact proportion of the Z/E ratio and needed prolonged reaction time to be completed. Additionally, the stereoselective outcome of theses reactions seems to result from a specific coordination of the metal atom with the amide function and the double bond creating a π complex in which the rigidity is responsible for the high stereoselectivities observed. In a more recent paper, using Ni(dppp)Cl₂ as a catalyst in the presence of HPPh₂, Zn and Znl₂, isomerization of secondary amides was carried out at low temperatures (0 °C-25 °C) and prolonged time providing a high Z-selectivity of the enamides (Scheme 3.7c).²⁶ Curiously from acyclic tertiary amides, under different reaction conditions no conversion to the Z and/or E enamides was observed which make the authors suggest that a free NH-group is beneficial, if not crucial, for the nickel-catalysed translocation of the double bond and its stereoselective outcome. Nevertheless cyclic tertiary N-allyl derivatives undergo the reaction to give predominantly the E-isomers. Finally, an additional example of isomerization leading to Z-enamides was reported also in 2017 using CpRu(CH₃CN)₃PF₆ as a catalyst. Although the procedure describes an effective access to Z-di-, tri-, and tetrasubstituted enamides, it is limited to secondary N-allyl amides and prolonged reaction time (1-16 h) was necessary to get full conversions (Scheme 3.7d).²⁷

²⁵ Only few substrates were reported to provide *Z* isomer see: (a) Krompiec, S.; Pigulla, M.; Kuznik, N.; Krompiec, M.; Marciniec, B.; Chadyniak, D.; Kasperczyk, J. Highly selective isomerization of *N*-allylamides catalyzed by ruthenium and rhodium complexes. *J. Mol. Catal. A: Chem.* **2005**, *225*, 91-101. (b) Krompiec, S.; Pigulla, M.; Krompiec, M.; Baj, S.; Mrowiec-Bialo, J.; Kasperczyk, J. Highly selective isomerization of *N*-allylamides and *N*-allylamines. *Tetrahedron Lett.* **2004**, *45*, 5257-5261 (c) Stille, J. K.; Becker, Y. Isomerization of *N*-allylamides and -imides to aliphatic enamides by iron, rhodium, and ruthenium complexes. *J. Org. Chem.* **1980**, *45*, 2139–2145.

²⁶ Weber, F.; Steinlandt, P. S.; Ballmann, M.; Hilt, g. Structure-dependent nickel-catalysed transposition of *N*-allylamides to *E*- or *Z*-enamides. *Synthesis* **2017**, *49*, 440-450.

²⁷ Trost, B. M.; Cregg, J. J.; Quach, N. Isomerization of *N*-allyl amides to form geometrically defined di-, tri-, and tetrasubstituted enamides. *J. Am. Chem. Soc.* **2017**, *139*, 5133–5139.

a. Ref. 25a, 25b the authors do not provide information about the yield or the Z/E ratio



b. Ref. 25c, only 1 example providing the Z-isomer but no yield



c. Ref. 26, with tertiary acyclic amide no isomerization takes place



R = Me, *n*Hp, Bn, Ph, OtBu, OBn



With cyclic tertiary N-allyl derivatives exclusively the E-isomers



d. Ref. 27



R = Ph, vinyl, CH_2CH_2NHBoc


In conclusion isomerizations in the presence of transiton metal catalysts are limited to secondary amides, prolonged reactions are necessary to achieve full conversion and in most cases a mixture of Z/E isomers is obtained.

3.3. Isomerization of *N*-allyl carbamates and *N*-allyl amides in the presence of RuCl₂(PPh₃)_{3.} Our results

As it was reported in the introduction (Chapter 1), the aim of this part of the thesis is to investigate the isomerization process from tertiary allyl carbamates and allyl amides type I in the presence of RuCl₂(PPh₃)₃ and under microwave activation.



Scheme 3.8. Aim of this investigation.

To start our study we decided to prepare carbamates and amides **8** and **9** since their precursors were already available in the laboratory from a previous work (Scheme 3.9).²⁸

²⁸ For the preparation of **8** we followed the same procedure reported in: Diaba, F.; Bonjoch, J. Asymmetric synthesis of 2-azabicyclo[3.3.1]nonanes by a microwave-assisted organocatalysed tandem desymmetrisation and intramolecular aldolisation. *Org. Biomol. Chem.* **2009**, *7*, 2517-2519.



 $\begin{array}{l} \mathsf{R} = \mathsf{MeO}, \ \mathsf{EtO} \ (\mathsf{K}_2\mathsf{CO}_3, \ \mathsf{ACN}, \ \mathsf{rt} \ \mathsf{overnight} \ \mathsf{ref.} \ 28) \\ \mathsf{R} = \mathit{t}\mathsf{BuO} \ (\mathsf{Boc}_2\mathsf{O}, \ \mathsf{CH}_2\mathsf{Cl}_2, \ \mathsf{rt}, \ \mathsf{overinght}) \\ \mathsf{R} = \mathsf{Ph} \ (\mathsf{Et}_3\mathsf{N}, \ \mathsf{CH}_2\mathsf{Cl}_2, \ \mathsf{rt}, \ \mathsf{overinght}) \\ \mathsf{R} = \mathsf{Ts} \ (\mathsf{TsCI}, \ \ \mathsf{CH}_2\mathsf{Cl}_2, \ \mathsf{at} \ 0 \ ^\circ\!\! \mathsf{C} \ \mathsf{for} \ \mathsf{15} \ \mathsf{min} \ \mathsf{then} \ \mathsf{at} \ \mathsf{rt} \ \mathsf{for} \ \mathsf{2} \ \mathsf{h}) \\ \end{array}$

Scheme 3.9. Synthesis of 8 and 9.

Using the classical procedures, described already in the research group, secondary amines **A** and **B** were treated with methyl or ethyl chloroformate in the presence of a base yielding **8a**, **8b**, **9a** and **9b** with good yields. **9c** and **9d** result from reaction of **B** with di-*tert*-butyl dicarbonate and benzoyl chloride respectively (Scheme 3.9).

Having in hand substrates **8** and **9** we started our investigation by screening the reaction conditions with carbamate **8a** and **8b** (Table 3.1).

Following our previous work (Chapter 2) we decided to run the first reaction from **8a** (0.44 mmol) in toluene as a solvent (1 mL) at 100 °C using microwave activation. After 75 minutes of irradiation, only a modest conversion was observed (33%) providing after purification the *Z* derivative **10a** with a low yield (Entry 1). It's worth noting that prolonging the reaction further than 75 min did not bring better results. Nevertheless, to our big delight, an almost full conversion was achieved by simply increasing the temperature of the reaction under microwave activation to 120 °C (Entry 2). Indeed in the ¹H NMR spectrum of the crude the signals of **8a** almost disappeared revealing mainly the ones belonging to *Z*- derivative **10a** (Figure 3.2). After purification by chromatography we were able to isolate as the major product **10a** with an excellent yield and the minor *E* derivative **11a** in pure forms despite their similar polarity.



Table 3.1. Screening of the isomerization conditions from 8a and 8b.ª

Entry	8	Solvent	Temp.(⁰C)	Time (min)	10 , 11 (yield %)	10/11
1	8a	PhMe	100	75	30 ^b	7/1
2	8 a	PhMe	120	75	82	10/1
3	8a	ACN	120	75	-	-
4	8a	THF	120	75	38	2/1
5	8a	DMF	120	75	76	4/1
6	8a	PhMe	120	60	80	11/1
7	8a ^c	DCM	50	90	-	-
8	8a ^d	PhMe	100	45	55	1/3
9	8a	PhMe	140	30	90	1.1/1
10	8a	PhMe	160	30	94	1/2.3
11	8a	PhMe	180	30	93	1/2
12	8a	PhMe	180	10	93	1/1.9
13	8b	PhMe	120	45	90	11/1

^{*a*} Unless otherwise noted, the reactions were carried out with 0.44 mmol of **8a** or **8b** in 1 mL of solvent under microwave activation. ^{*b*} 67% of starting material (**8a**) was recovered. ^{*c*} Using conventional heating in the presence of vinyloxytrimethylsilane (10 equiv.) and Grubbs' second-generation catalyst (5%). ^{*d*} Using the same conditions reported in c but under microwave activation.



5.10 6.05 6.00 5.95 5.90 5.85 5.80 5.75 5.70 5.65 5.60 5.55 5.50 5.45 5.40 5.35 5.30 5.25 5.20 5.15 5.10 5.05 5.00 4.95 4.90 4.85 4.80 f1 (ppm)



The reaction was also carried out under the same conditions using acetonitrile, tetrahydrofuran and dimethylformamide respectively. Whereas the isomerization in ACN did not take place at all and only **8a** was present in the reaction mixture (Entry 3), the reaction in THF and DMF provided a mixture of **10a** and **11a** with lower yields and worse diastereoselectivities (Entries 4 and 5). With these results in hand we decided to maintain toluene as the solvent of the reaction. Reducing the time from 75 to 60 min affects slightly the conversion ratio and the diastereoselectivity of the process (Entry 6). Additionally, we also checked the isomerization process under the conditions reported by Nishida *et al.*,¹⁶ in the presence of vinyloxytrimethylsilane (10 equiv.) and Grubbs' second-generation catalyst (5%), after 90 min of heating at 50 °C only **8a** was present in the reaction mixture (Entry 7). Nevertheless, using the same conditions in toluene as a solvent instead of DCM and under microwave activation, after 45 min of heating at 100 °C, a full conversion was achieved but both the yield and the diastereoselectivity were modest (Entry 8).

Next we decided to investigate the influence of the reaction temperature on the course of the reaction under microwave activation. At 140 °C, a full conversion was achieved after only 30 min providing a mixture of **10a** and **11a** in a 1.1/1 ratio and a very good yield (Entry 9). At 160 °C and after the same reaction time the *E* isomer **11a** was the main compound of the reaction (Entry 10). At 180 °C the reaction was first carried out for 30 min but then a simple ¹H NMR checking showed that it was over after only 10 min providing a mixture of **10a** and **11a** in similar yields and diastereoselectivity as before (Entries 11 and 12). These results showed clearly that rising the temperature of the reaction favors the formation of the thermodynamically more stable *E*-enamides.

Additionally, the reaction from **8b**, with an ethoxy instead of a methoxy group, followed the same scenario affording after only 45 min of reaction at 120 °C, **10b** as the major isomer in a very good yield (Entry 13).

Identification of **10a** and **11a** as well as **10b** and **11b** was possible by simply comparing the vinylic protons coupling constants. As it was reported before,²⁵ the coupling constant in the *Z*-isomer is smaller (\approx 7 Hz) than the *E*-isomer (\approx 13 Hz). Moreover, the vinylic protons in the *Z*-isomer are shielded (the proton on the carbon linked to the nitrogen) and deshielded (the other proton) in comparison with the *E*-isomers. Additionally, the methyl group in the latter appears at lower field in contrast to the former (Figure 3.3).



Figure 3.3. Comparison of ¹H NMR spectra of 10a and 11a.

The optimized conditions were then applied to carbamates and amides **9a**-**9e** (Table 3.2). From **9a-9d** and as in the previous reactions with **8a**, a full conversion was attained after 75 min of heating at 120 °C to provide mainly the *Z*-derivatives **12a-12d** with excellent yields (Entries 1, 5-8). Curiously, from tosylate derivative 9e, no reaction takes place (Entry 8). The influence of the reaction temperature on the *Z*/*E* ratio was also investigated with **9a** providing similar results as before (Figure 3.4).

R _N				R _N	R. _N	_Me
			He +			
	9			12	13	
Entry	R	9	Temp. ⁰C	Time (min) ^b	Yield (%)	12/13
1	CO₂Me	9a	120	75	89	10/1
2		9a	140	30	92	3.9/1
3		9a	160	30	96	1/2.5
4		9a	180	30	84	1/2.2
5	CO ₂ Et	9b	120	75	88	10/1
			120	45	85	13:1
6	CO₂ <i>t</i> Bu	9c	120	75	95	15/1
7	COPh	9d	120	75	90	29/1
8	Ts	9e	120	75	-	-

Table 3.2. Isomerization reactions from 9a-9e.^a

^a Unless otherwise noted, the reactions were carried out with 0.44 mmol of **9** in 1 mL of toluene under microwave activation. ^bTime necessary for a full conversion .



Figure 3.4. ¹H NMR spectra of the isomerization from **9a** achieved at different temperatures (comparison of the Z/E ratio).

Finally the isomerization process was successfully extended to substrates **14** with different pattern providing mainly or exclusively the corresponding *Z*-enamides with good to excellent yields (Scheme 3.10). In the case of **14a** and **14b** the isomerization takes place only on the terminal double bond providing better *Z*-selectivity with the latter. On the other hand from **14d**, despite the presence of another terminal alkenyl chain only **15d** was isolated with an excellent yield and streoselectivity. Finally, It is worth noting that from **14e**, no ATRC takes place and only *Z*-**15e** was isolated as a single stereoisomer.



Scheme 3.10. Z-Isomerization from substrates 14.

CHAPTER 4: Microwave assisted synthesis of γand δ-lactams from trichloro- and dichloroacetamides catalyzed by RuCl₂(PPh₃)₃

4.1. Atom transfer radical reactions

Among the radical processes, atom transfer radical additions (ATRA) and atom transfer radical cyclizations (ATRC) are considered important and useful tools for the formation of C-C bonds. In contrast to the radical reductive procedures, in this type of reactions, known also as the Kharasch additions or cyclizations,¹ the halogen (X) present in the substrate and which abstraction is part of the propagation step, is retained in the final compound (Scheme 4.1). In consequence the generated intermediates possess a potentially useful carbonhalogen bond that allows post-cyclization manipulation if required to access more advanced structures.²



Scheme 4.1. Reductive radical cyclization vs ATRC.

Atom transfer radical additions and cyclizations in the presence of transition metal catalysts specially derived from iron, copper, ruthenium or nickel are well documented.³ In this last part of the thesis, will be reported our results related to the synthesis of γ - and δ -lactams from alkenyl tethered trichloroacetamides in the presence of RuCl₂(PPh₃)₃ using microwave activation. As it was outlined in chapter 1, this catalyst has been successfully used in the synthesis of the above mentioned heterocycles but the processes involved were

¹ (a) Kharasch, M. S.; Engelmann, H.; Mayo, F. R. Atom transfer radical reactions as a tool for olefin functionalization. *J. Org. Chem.* **1937**, 2, 288-302. (b) Kharasch, M. S.; Jensen, E. V.; Urry, W. H. Addition of carbon tetrachloride and chloroform to olefins. *Science* **1945**, *102*, 128-128.

² For reviews see: a) Iqbal, J.; Bhatia, B.; Nayyar, N.K. Transition metal-promoted free-radical reactions in organic synthesis: the formation of carbon-carbon bonds. *Chem. Rev.* **1994**, 94, 519–564; b) Clark, A.J. Atom transfer radical cyclisation reactions mediated by copper complexes. *Chem. Soc. Rev.* **2002**, 31, 1-11; (c) MuñozMolina, J. M.; Belderrain, T. R.; Pérez, P. J. Atom transfer radical reactions as a tool for olefin functionalization – on the way to practical applications. *Eur. J. Inorg. Chem.* **2011**, 3155–3164. (d) Curran, D. P. The design and application of free radical chain reactions in organic synthesis. Part 1. *Synthesis* **1988**, 417–439.

³ See for example: (a) Minisci, F. Free-radical additions to olefins in the presence of redox systems. *Acc. Chem.Res.* **1975**, *8*, 165-171. (b) Martin, P.; Steiner, E.; Streith, J.; Winkler, T.; Belluš, D. Convenient approaches to heterocycles via coppercatalysed additions of organic polyhalides to activated oleffns. *Tetrahedron* **1985**, *41*, 4057-4078. (c) Severin, K. Ruthenium catalysts for the Kharasch reaction. *Curr. Org.Chem.* **2006**, *10*, 217-224. (d) Grove, M. D.; Van Koten, G.; Verschuuren, A. H. M. New Homogeneous catalysts in the addition of polyhalogenoalkanes to olefins; organonickel(II) Complexes [Ni{C₆H₃(CH₂NMe₂)2-*o*,*o*'}X] (X = Cl, Br, I). *J. Mol.Catal.* **1988**, *45*, 169-174.

achieved at high temperatures and using long reaction times. Thus we decided to investigate the compatibility of the ATRC in the presence of RuCl₂(PPh₃)₃ and microwave activation with the aim of improving the reaction conditions, the yield and also the stereoselectivity.

4.2. ATRC catalyzed by RuCl₂(PPh₃)₃ in the synthesis of γ - and δ lactams from trichloro- and dichloroacetamides. Our results

The investigation started with usage of this catalyst in combination with microwave activation for the preparation of γ - and δ -lactams and also for the preparation of these embedded in bicyclic structures such as indoles and morphans from trichoroacetamides and α , α -dichloroamides with a suitable alkenyl chain. For this purpose we choose the already available substrates (**17a-17p**) listed below to carry out our investigation (Scheme 4.2).



Scheme 4.2. Substrates used in the ATRC mediated by Ru3 catalyst.

To optimize the reaction conditions, we carried out our primary study on trichloroacetamide **17a** with a *t*-butyl group on the nitrogen since the presence of the latter can accelerate the reaction (Table 4.1).

Table 4.1. Screening of the ATRC reaction conditions for 17a.ª



Entry	Ru3 (mol%)	Solvent	Temp (⁰C)	Time (min)	18a Yield (%)	17a (%)
1	5	PhMe	140	75	0	100
2	10	PhMe	160	30	79	traces
3	5	PhMe	160	30	82	traces
4	5	PhMe	160	15	83	traces
5	5	PhMe	160	10	79	traces
6	2.5	PhMe	160	20	61	12
7	3	PhMe	160	60	71	5
8	5	ACN	160	15	0	100

^a Unless otherwise noted, the reactions were carried out with 0.386 mmol of **17a** in 1 mL of solvent under microwave activation. All products were separated through column chromatography and analysed by NMR spectroscopy.

Hence the first reaction from **17a** was achieved using the conditions reported in the literature but with microwave irradiation.⁴ When **17a** was heated at 140 °C in toluene for more than 1 h with 5 mol% of catalyst, the reaction did not take place and **17a** was recovered (Entry 1). Only by increasing the temperature to 160 °C, almost a full conversion was observed after 15 min of reaction leading to γ -lactam **18a** with an excellent yield (Entry 4). Shortening the reaction time or decreasing the catalyst loading did not bring any improvement to the reaction yield (Entries 5-7). It's worth noting that when the reaction was run in acetonitrile under the optimized conditions no reaction takes place (Entry 8). The best conditions were then applied to **17b-17d** where one of the chloro atoms was substituted by a hydrogen, a methyl and an ester group respectively. In all cases the reaction proceeds successfully providing the corresponding γ -lactams (**18** and **19**) with good yields for almost all the substrates (Table 4.2). The diastereomers formed in each case were separated and identified by NMR spectroscopy.

⁴ See for example ref. 9 in the introduction.

	CI RuCl ₂ D PhM	(PPh ₃) ₃ (5 mol%) Me, 160 °C, μW	CI-F	R CI CI ▶CI +	
R = H CH ₃ COOEt	17b 17c 17d		18		19
Entry	17	Time (min)	18/19 (yield %)	18/19	17 (%)
1	17b	15	75	1.7/1 ^b	traces
2	17c	15	52	1.5/1	29
3	17c	30	80	2.5/1	traces
4	17d	15	70	1.8/1	10
5	17d	10	77	2.1/1	traces

Table 4.2. RuCl₂(PPh₃)₃ catalyzed ATRC reaction from substrates 17b-17d.^a

^a Unless otherwise noted, the reactions were carried out with 0.386 mmol of **17b**, **17c** or **17d** in 1 mL of solvent under microwave activation. All products were separated through column chromatography and analysed by NMR spectroscopy. ^b **19b** was not isolated in pure form.

We also investigated the reaction from trichloroacetamides **17e-17j** with different substituents on the nitrogen (Table 4.3). In all cases the corresponding γ -lactams were obtained with good yields even if with some substrates (**17e**, **17g** and **17 j**) 30 to 45 min were necessary to achieve a full conversion.

(R =	CCI ₃ NO R 17 H nBu Ph Bn	RuCl ₂ (PP (5 mol%) PhMe, 160 17e 17f 17g 17h	h ₃) ₃ CI-) °C, μW	
	Allyl 2-Bromoallyl	17i 17j		
F actoria	47	T iana (asia)	10 (data	
Entry	17	Time (min)	18 (yieid %)	17 (%)
1	17e	15	62	8
2	17e	30	68	7
3	17f	15	90	0
4	17g	15	48	45
5	17g	45	84	13
6	17h	15	78	traces
7	17i	15	78	traces
8	17j	15	69	20
9	17j	30	67	11

Table 4.3. RuCl₂(PPh₃)₃ catalyzed ATRC reaction for substrates 17e-17j^a

^a Unless otherwise noted, the reactions were carried out with 0.386 mmol of **17e-17j** in 1 mL of toluene under microwave activation. All products were separated through column chromatography and analysed by NMR spectroscopy.

Additionally, we investigated the efficacy of the process varying the nature of the radical acceptor (the alkenyl chain) in trichloroacetamides **17k-17p**.

In the case of **17k** with a methyl group on the terminal carbon of the allyl double bond, the cyclization occurs providing lactams **18k** and **19k** with an excellent yield and a good diastereoselectivity (Scheme 4.3)



Scheme 4.3. ATRC from 17k.

Operating from butenyl derivative **17I**, only δ -lactam **18I** resulting from a *6-exo-trig* radical cyclization was obtained with a very good yield (Scheme 4.4)



Scheme 4.4. ATRC from 17I.

Furthermore, from trichloroacetamide **17m**, the corresponding δ -lactam as a mixture of diastereomers (**18m** and **19m**) in almost an equimolar proportion was produced (Scheme 4.5).



Scheme 4.5. ATRC from 17m.

The procedure was also successfully extended to trichloroacetamides **17n** and **17p**. In both cases the reaction was completely diastereoselective proving the corresponding indole and morphan derivatives with acceptable to good yields (Scheme 4.6). It is worth mentioning that with substrate **17p**, a fair amount of starting material (20%) was recovered and we believe that the yield could be improved by prolonging the reaction time.



Scheme 4.6. ATRC from 17n and 17p.

Finally, we checked also the reaction from dichloroacetamide **170**. After 15 min of irradiation under the same conditions reported previously, a mixture of diastereomers **180** and **190** was isolated in a 3.6:1 ratio. In this case, the reaction was not complete since some of **170** (about 25%) was recovered after chromatography and due to the lack of time we were not able to optimize the reaction from this substrate (Scheme 4.7).



Scheme 4.7. ATRC from 170.

It is worth mentioning that not all the substrates that we investigated in this part of the thesis gave good results. In Scheme 4.8 are listed the α -haloamides derivatives with which the reaction did not take place. In these cases, instead of the expected lactams a mixture of unidentified products or the initial starting materials were recovered.



Scheme 4.8. Substrates which reactions were unsuccessful.

CHAPTER 5: Conclusions

In the research work presented in this PhD thesis, we have demonstrated that tris(triphenylphosphine)ruthenium(II) dichloride is truly a versatile and potent catalyst capable of achieving different chemical transformations under microwave activation.

First, a new behaviour of RuCl₂(PPh₃)₃ leading to β -lactams from *N*tethered benzyl trichloroacetamides through benzylic CH activation was discovered and established. The simplicity of the process and the challenging unprecedented reaction involved for this C(sp³)-C(sp³) bond formation bring new insights into the chemistry of ruthenium catalysts. Future efforts will be steered toward expanding the reaction to more substrates for the synthesis of other nitrogen heterocycles and also to shed more light on the mechanism of this radical transformation.



11 examples (18-58%)

Some of the β -lactams prepared naming **2b**, **2c**, **2e**, **2h**, **2j** and **2k** were evaluated for their cytotoxic activity and hemocompatibility. The results showed that the proposed compounds have non-hemolytic properties and that the cytotoxic response is highly dependent on the structure and the concentrations of these β -lactams. Compound **2b** was found to be the most cytotoxic of the list showing the lowest IC50 values (20-49 µg/mL and 30-47 µg/mL for HaCaT and A431 cell lines, respectively).

In the second place, RuCl₂(PPh₃)₃ was successfully used in protected allylamines isomerization under microwave activation.



The isomerization process proceeds from tertiary *N*-allyl carbamates and *N*-allyl amides to provide the corresponding enecarbamates and enamides with a very good yields and excellent *Z*-selectivity. The reactions are achieved in very short reaction time and the nature of the protective group on the nitrogen seems to have a big influence on the course of the reaction. It is worth noting that RuCl₂(PPh₃)₃ was used herein for the first time in this type of transformations.

Finally, even if ATRC in the presence of RuCl₂(PPh₃)₃ were already reported in the literature from alkenyl tethered trichloroacetamides, in this last part of the work we have showed that these reactions can be achieved under microwave activation. Hence we were able to access several γ - and δ -lactams in very short reaction times and with very good yields. The process was extended to α , α -dichloroamides and the reactions in almost all cases are diastereoselective. Studies are underway to discover more potential of this ruthenium complex.



CHAPTER 6: Experimental part

General information

NMR spectra were recorded in CDCl₃ and the chemical shifts of ¹H spectra are reported in ppm downfield (δ) from Me₄Si, ¹³C NMR spectra are referenced to the deuterated solvent signal (CDCI₃: 77.00 ppm). All NMR data assignments are supported by gCOSY and gHSQC experiments. The high resolution ESI mass spectra were obtained from an Agilent LC/MSD-TOF mass spectrometer. Analytical thin-layer chromatography was performed on SiO₂ (Merck silica gel 60 F_{254}) with a fluorescent indicator ($\lambda = 254$ nm), and the spots were located by UV light and/or with 1% aqueous KMnO₄ solution or anisaldehyde. Chromatography refers to flash chromatography and was carried out on SiO₂ (Carlo Erba silica gel 60A, 35-70 μm). Drying of organic extracts during workup of reactions was performed over anhydrous Na₂SO₄. Solvent evaporation was accomplished with rotatory evaporator. All yields refer to chromatographically and а spectroscopically (NMR) pure material. Microwave irradiation experiments were performed using a single-mode Discover System from CEM Corporation using standard Pyrex vessel (capacity 10 mL)

6.1. Experimental part of chapter 2

1. Synthesis of trichloroacetamides 1a-1I¹

For the preparation of trichloroacetamides **1e-1g** and **1I** we followed the same procedure reported in ref.1 isolating **1e** (40%), **1f** (50%), **1g** (30%) and **1I** (70%).

N-tert-Butyl-2,2,2-trichloro-*N*-(4-methylbenzyl)acetamide (1e). IR (NaCl, neat): 2998, 2973, 2923, 2867, 2857, 1685 cm⁻¹; ¹H NMR (CDCl₃, 400 MHz): δ 1.41 (s, 9H, *t*-Bu), 2.34 (s, 3H, CH₃), 4.99 (brs, 2H, CCl₃ CH₂Ar), 7.15 (brs, 4H, ArH); ¹³C NMR (CDCl₃, 100 MHz): δ 21.0 (CH₃), 28.1 (CH₃), 50.8 (CH₂Ar), 61.8 (C), 95.4 (CCl₃), 126.4, 129.1 (ArCH), 135.5 (C), 136.8 (C), 160.8 (CO). HRMS (ESI-TOF): Calcd

for $C_{14}H_{19}CI_3NO [M+H]^+ 322.0527$; found 322.0528.

N-tert-Butyl-2,2,2-trichloro-N-(3-methoxybenzyl)acetamide (1f). IR (NaCl,



neat): 3001, 2965, 2937, 1685, 1601 cm⁻¹; ¹H NMR (CDCl₃, 400 MHz): δ 1.44 (s, 9H, *t*-Bu), 3.80 (s, 3H, CH₃), 5.01 (brs, 2H, CH₂Ar), 6.76-6.89 (m, 3H, ArH), 7.26 (t, *J* = 7.8 Hz, 1H, ArH); ¹³C NMR (CDCl₃, 100 MHz): δ 28.0 (CH₃), 50.9 (CH₂Ar), 55.2 (CH₃), 61.9 (C), 95.2 (CCl₃), 112.3, 118.7, 129.5 (ArCH), 140.3 (C), 159.7 (C), 160.7

(CO). HRMS (ESI-TOF): Calcd for C₁₄H₁₉Cl₃NO₂ [M+H]⁺ 338.0476; found 338.0464.

N-tert-Butyl-2,2,2-trichloro-N-(4-nitrobenzyl)acetamide (1g). IR (NaCl, neat):



3081, 2981, 2934, 2676, 1688, 1600 cm⁻¹; ¹H NMR (CDCl₃, 400 MHz): δ 1.44 (s, 9H, *t*-Bu), 5.15 (brs, 2H, CH₂Ar), 7.48 (d, *J* = 6.7 Hz, 2H, ArH), 8.24 (d, *J* = 6.7 Hz, 2H, ArH); ¹³C NMR (CDCl₃, 100 MHz): δ 28.1 (CH₃), 50.6 (CH₂Ar), 62.2 (C), 94.9 (CCl₃), 123.9, 127.2 (ArCH), 146.4 (C), 147.2 (C), 160.6 (CO). HRMS (ESI-TOF):

Calcd for $C_{13}H_{16}CI_{3}N_{2}O_{3}$ [M+H]⁺ 353.0221; found 353.0224.

¹ For the preparation of **1a-1d** and **1h-1k** see: Diaba, F.; Montiel, J. A.; Martinez-Laporta, A.; Bonjoch, J. *Tetrahedron Lett.* **2013**, 54, 2619.

(*S*)-*N*-Benzyl-2,2,2-trichloro-*N*-(1-phenylethyl)acetamide (11). IR (NaCl, neat): 3106, 3088, 3062, 3030, 2988, 2939, 1673, 1602 cm⁻¹; ¹H NMR (CDCl₃, 400 MHz): δ 1.58 (s, 3H, CH₃), 3.88 (brd, *J* = 15.8 Hz, 1H, CH₂Ar), 4.76 (brd, *J* = 15.8 Hz, 1H, CH₂Ar), 5.93 (brs, 1H), 6.96-7.43 (m, 10H, ArH); ¹³C NMR (CDCl₃, 100 MHz): δ 17.9 (CH₃), 49.4 (CH₂Ar), 57.3 (CHAr), 93.7 (CCl₃), 126.6, 126.9, 127.8, 128.4, 128.7 (ArCH), 137.3 (C), 139.1 (C), 161.2 (CO). HRMS (ESI-TOF): Calcd for C₁₇H₁₇Cl₃NO [M+H]⁺ 356.0370; found 356.0369; Calcd for

2. General procedure for the reaction of trichloroacetamides 1 with "Ru" catalysts

C₁₇H₂₀Cl₃N₂O [M+NH₄]⁺ 373.0636; found 373.0636.



In a 10 mL vessel was placed trichloroacetamide **1** (100 mg, 0.32 mmol) and ruthenium catalyst (0.032 mmol, 10%) in toluene (1 mL). The mixture was heated with stirring using microwave irradiation for 5 to 30 min at 80 °C or 100 °C. After chromatography (hexane-CH₂Cl₂, 9:1 to CH₂Cl₂) **2** and/or **3** were isolated.²

1-Benzyl-3,3-dichloro-4-phenylazetidin-2-one (2a). IR (NaCl, neat): 3088,



3064, 3032, 2923, 2852, 1790 cm⁻¹; ¹H NMR (CDCl₃, 400 MHz): δ 3.88 (d, *J* = 15.0 Hz, 1H, CH₂N), 4.77 (s, 1H, H-4), 4.90 (d, *J* = 15.0 Hz, 1H, CH₂N), 7.05-7.10 (m, 2H, ArH), 7.15-7.20 (m, 2H, ArH), 7.23-7.28 (m, 3H, ArH), 7.35-7.40 (m, 3H, ArH); ¹³C NMR

(CDCl₃, 100 MHz): δ 45.0 (CH₂N), 73.2 (C-4), 84.8 (C-3), 128.1, 128.4, 128.8, 129.1, 129.9 (ArH), 131.7, 133.5 (C), 161.7 (C-2). HRMS (ESI-TOF) *m/z*: calcd for C₁₆H₁₄Cl₂NO [M+H]⁺ 306.0447; found 306.0450.

² For the yields of pure compounds from **1a-1l** see the tables in chapter 2.

1-(*tert***-Butyl)-3,3-dichloro-4-phenylazetidin-2-one (2b).** mp 81-83 °C; IR (NaCl, neat): 3065, 3036, 2976, 2931, 2872, 2856, 1778 cm⁻¹; ¹H NMR (CDCl₃, 400 MHz): δ 1.33 (s, 9H, *t*-Bu), 5.05 (s, 1H, H-4), 7.35-7.46 (m, 5H, ArH); ¹³C NMR (CDCl₃, 100 MHz): δ 27.9

(CH₃), 55.7 (C), 73.6 (C-4), 83.4 (C-3), 128.5, 129.7 (ArH), 134.6 (C), 161.6 (C-2). HRMS (ESI-TOF) *m/z*: calcd for C₁₃H₁₆Cl₂NO [M+H]⁺ 272.0603;

found 272.0606.

1-(tert-butyl)-3,3-dichloro-4-(o-tolyl)azetidin-2-one (2c). IR (NaCl, neat):



3074, 3029, 2980, 2937, 2876, 1769 cm⁻¹; ¹H NMR (CDCl₃, 400 MHz): δ 1.37 (s, 9H, *t*-Bu), 2.44 (s, 3H, CH₃), 5.31 (s, 1H, H-4), 7.22-7.37 (m, 4H, ArH); ¹³C NMR (CDCl₃, 100 MHz): δ 19.7 (CH₃), 27.8 (CH₃), 55.5 (C), 69.9 (C-4), 83.2 (C-3), 125.8, 126.9,

129.0, 130.6 (ArCH), 133.2 (C), 136.5 (C), 161.9 (C-2). HRMS (ESI-TOF) *m/z*: calcd for C₁₄H₁₈Cl₂NO [M+H]⁺ 286.0760; found 286.0761.

1-(tert-butyl)-3,3-dichloro-3-(p-tolyl)azetidin-2-one (2d). IR (NaCl, neat):



3029, 2975, 2926, 2871, 1781 cm⁻¹; ¹H NMR (CDCl₃, 400 MHz): δ 1.33 (s, 9H, *t*-Bu), 2.40 (s, 3H, CH₃), 5.00 (s, 1H, H-4), 7.15-7.34 (m, 4H, ArH); ¹³C NMR (CDCl₃, 100 MHz): δ 21.4 (CH₃), 27.9 (CH₃), 55.7 (C), 73.6 (C-4), 83.8 (C-3), 128.4, 130.4 (ArCH), 134.5 (C), 138.3 (C), 161.7 (C-2). HRMS (ESI-TOF) *m/z*: calcd

for C₁₄H₁₈Cl₂NO [M+H]⁺ 286.0760; found 286.0764.

1-(tert-butyl)-3,3-dichloro-4-(p-tolyl)azetidin-2-one (2e). IR (NaCl, neat):



3053, 3028, 2976, 2934, 2873, 1778 cm⁻¹; ¹H NMR (CDCl₃, 400 MHz): δ 1.32 (s, 9H, *t*-Bu), 2.39 (s, 3H, CH₃), 5.01 (s, 1H, H-4), 7.23 (d, *J* = 8.5 Hz, 2H, ArH), 7.26 (d, *J* = 8.5 Hz, 2H, ArH); ¹³C NMR (CDCl₃, 100 MHz): δ 21.3 (CH₃), 27.9 (CH₃),

55.6 (C), 73.5 (C-4), 83.5 (C-3), 128.2, 129.2 (ArCH), 131.5 (C), 139.8 (C), 161.7 (C-2). HRMS (ESI-TOF) m/z: calcd for C₁₄H₁₈Cl₂NO [M+H]⁺ 286.0760; found 286.0766.

1-(tert-Butyl)-3,3-dichloro-4-(3-methoxyphenyl)azetidin-2-one (2f). IR (NaCl,



neat): 3071, 3060, 3064, 2975, 2935, 2876, 2853, 2837, 1778 cm⁻¹; ¹H NMR (CDCl₃, 400 MHz): δ 1.34 (s, 9H, *t*-Bu), 3.84 (s, 3H, CH₃O), 5.01 (s, 1H, H-4), 6.89 (m, 1H, ArH), 6.93-6.98 (m, 2H, ArH), 7.34 (t, *J* = 7.9 Hz, 1H, ArH); ¹³C

NMR (CDCl₃, 100 MHz): δ 27.9 (CH₃), 55.3 (CH₃O), 55.7 (C), 73.5 (C-4), 83.4 (C-3), 113.8, 115.0, 120.5, 129.6 (ArCH), 136.1 (C), 159.7 (C), 161.6 (C-2). HRMS (ESI-TOF) *m/z*: calcd for C₁₄H₁₈Cl₂NO₂ [M+H]⁺ 302.0709; found 302.0709.

1-(tert-butyl)-3,3-dichloro-4-(3,5-difluorophenyl)azetidin-2-one (2h). IR



(NaCl, neat): 3094, 3063, 2977, 2931, 2875, 2855, 1784 cm⁻¹; ¹H NMR (CDCl₃, 400 MHz): δ 1.35 (s, 9H, *t*-Bu), 4.99 (s, 1H, H-4), 6.84-6.96 (m, 3H, ArH); ¹³C NMR (CDCl₃, 100 MHz): δ 27.9 (CH₃), 56.0 (C), 72.4 (C-4), 83.0 (C-3), 105.3 (t, *J* = 25.1 Hz, 1C, ArCH), 111.0 (brs, 2C, ArCH), 138.8 (t, *J* = 8.6 Hz,

ArC), 161.3 (C-2), 163.0 (dd, J = 249.3, 12.4 Hz, 2C, C-F). HRMS (ESI-TOF) m/z: calcd for C₁₃H₁₄Cl₂F₂NO [M+H]⁺ 308.0415; found 308.0412.

1-Butyl-3,3-dichloro-4-phenylazetidin-2-one (2i). IR (NaCl, neat): 3065, 3035,



2960, 2932, 2873, 1789 cm⁻¹; ¹H NMR (CDCl₃, 400 MHz): δ 0.92 (t, J = 7.3 Hz, 3H, CH₃), 1.29-1.41 (m, 2H, CH₂), 1.51-1.60 (m, 2H, CH₂), 2.96 (dt, J = 14.0, 6.6 Hz, 1H), 3.62 (dt, J = 14.0, 7.8 Hz, 1H), 5.05 (s, 1H, H-4), 7.05-7.10 (m, 2H, ArH), 7.28-7.32 (m, 2H, 2H), 3.65 (m, 2H), 7.05-7.10 (m, 2H), 3.65 (m, 2H), 7.28-7.32 (m, 2H), 7.05-7.10 (m, 2H), 7.28-7.32 (m, 2H), 7.28-7.32 (m, 2H), 7.05-7.10 (m, 2H), 7.05-7.10 (m, 2H), 7.28-7.32 (m, 2H), 7.05-7.10 (m, 2H), 7.28-7.32 (m, 2H), 7.05-7.10 (m, 2H), 7.28-7.32 (m, 2H), 7.28-7.38 (m, 2H), 7.28-7.

ArH), 7.43-7.49 (m, 3H, ArH); ¹³C NMR (CDCl₃, 100 MHz): δ 13.5 (CH₃), 20.0 (CH₂), 29.1 (CH₂), 40.9 (CH₂), 74.0 (C-4), 84.7 (C-3), 128.1, 128.8, 129.9 (ArCH), 132.0 (C), 161.8 (C-2). HRMS (ESI-TOF) *m/z*: calcd for C₁₃H₁₆Cl₂NO [M+H]⁺ 272.0603; found 272.0604.

(3RS,4SR)-1-Butyl-3-chloro-4-phenylazetidin-2-one (4i). IR (NaCl, neat):



3063, 3035, 2958, 2925, 2871, 1770 cm⁻¹; ¹H NMR (CDCl₃, 400 MHz): δ 0.89 (t, *J* = 7.4 Hz, 3H, CH₃), 1.26-1.38 (m, 2H), 1.45-1.56 (m, 2H), 2.90-2.99 (m, 2H), 3.55 (dt, *J* = 14.0, 7.7 Hz, 1H), 4.46 (d, *J* = 4.9 Hz, H-4), 5.10 (d, *J* = 4.9 Hz, 1H, H-3), 7.25-7.30 (m, 2H,

ArH), 7.37-7.47 (m, 3H, ArH); ¹³C NMR (CDCl₃, 100 MHz): δ 13.5 (CH₃), 20.2 (CH₂), 29.3 (CH₂), 40.9 (CH₂), 60.9 (C-3), 61.0 (C-3), 128.2, 128.5, 129.1 (ArCH),

133.2 (C), 164.3 (C-2). HRMS (ESI-TOF) *m/z*: calcd for C₁₃H₁₇CINO [M+H]⁺ 238.0993; found 238.0997.

(3*RS*,4*RS*)-1-Butyl-3-chloro-4-phenylazetidin-2-one (5i). IR (NaCl, neat): 3064, 3032, 2958, 2931, 2872, 1772 cm⁻¹; ¹H NMR (CDCl₃, 400 MHz): δ 0.89 (t, *J* = 7.3 Hz, 3H, CH₃), 1.24-1.36 (m, 2H, CH₂), 1.44-1.53 (m, 2H, CH₂), 2.86 (dt, *J* = 14.1, 7.0 Hz, 1H), 3.52 (dt, *J* = 14.1, 7.6 Hz, 1H), 4.49 (brd, *J* = 1.7 Hz, H-4), 4.54 (d, *J* = 1.7

Hz, 1H, H-3), 7.30-7.33 (m, 2H, ArH), 7.37-7.47 (m, 3H, ArH); ¹³C NMR (CDCl₃, 100 MHz): δ 13.5 (CH₃), 20.0 (CH₂), 29.4 (CH₂), 40.8 (CH₂), 63.1 (C-4), 66.0 (C-3), 126.6, 129.3, 129.4 (ArCH), 135.2 (C), 163.8 (C-2). HRMS (ESI-TOF) *m/z*: calcd for C₁₃H₁₇CINO [M+H]⁺ 238.0993; found 238.0993.

1-Isopropyl-3,3-dichloro-4-phenylazetidin-2-one (2j). IR (NaCl, neat): 2954,



2923, 2852, 1789 cm⁻¹; ¹H NMR (CDCl₃, 400 MHz): δ 1.18 (d, J = 6.8 Hz, 3H, CH₃), 1.40 (d, J = 6.8 Hz, 3H, CH₃), 1.75 (hept, J = 6.8 Hz, 1H), 5.03 (s, 1H, H-4), 7.05-7.10 (m, 2H, ArH), 7.33-7.37 (m, 2H, ArH), 7.41-7.47 (m, 3H, ArH); ¹³C NMR (CDCl₃, 100

MHz): δ 20.1 (CH₃), 20.8 (CH₃), 46.5 (CH), 73.3 (C-4), 84.0 (C-3), 128.3, 128.7, 129.9 (ArCH), 133.1 (C), 161.5 (C-2). HRMS (ESI-TOF) *m/z*: calcd for C₁₂H₁₄Cl₂NO [M+H]⁺ 258.0447; found 358.0447.

3,3-Dichloro-1-cyclohexyl-4-phenylazetidin-2-one (2k). IR (NaCl, neat): 3035,



2933, 2857, 1784 cm⁻¹; ¹H NMR (CDCl₃, 400 MHz) δ 1.103-1.36 (m, 4H), 1.58 (m, 1H), 1.65-1.90 (m, 4H), 2.08 (dm, *J* = 12.6 Hz, 1H), 3.40 (tt, *J* = 11.4, 3.8 Hz, 1H), 5.05 (s, 1H, H-4), 7.05-7.10 (m, 2H, ArH), 7.32-7.37 (m, 2H, ArH), 7.42-7.47 (m,

3H, ArH); ¹³C NMR (CDCl₃, 100 MHz): δ 24.7 (CH₂), 25.0 (2 CH₂), 30.2 (CH₂), 30.9 (CH₂), 53.8 (CH), 73.2 (C-4), 84.1 (C-3), 128.3, 128.6, 129.8 (ArCH), 133.2 (C), 161.5 (C-2). HRMS (ESI-TOF) *m/z*: calcd for C₁₅H₁₈Cl₂NO [M+H]⁺ 298.0760; found 398.0762.

(3RS,4SR)-3-Chloro-1-cyclohexyl-4-phenylazetidin-2-one (4k). IR (NaCl,



neat): 3031, 2932, 2855, 1761 cm⁻¹; ¹H NMR (CDCl₃, 400 MHz): δ 0.99-1.30 (m, 4H), 1.51-1.70 (m, 3H), 1.76 (m, 1H), 1.84 (m, 1H), 2.03 (dm, *J* = 12.5 Hz, 1H), 3.43 (tt, *J* = 11.5, 3.9 Hz, 1H), 4.97 (brd, *J* = 5.0 Hz, H-4), 5.03 (brd, *J* = 5.0 Hz, 1H,

H-3), 7.30-7.35 (m, 2H, ArH), 7.38-7.45 (m, 3H, ArH); ¹³C NMR (CDCl₃, 100 MHz): δ 24.9 (CH₂), 25.0 (CH₂), 25.1 (CH₂), 30.4 (CH₂), 31.3 (CH₂), 53.6 (CH), 60.0 (C-4), 60.2 (C-3), 128.3, 128.4, 129.1 (ArCH), 134.4 (C), 164.0 (C-2). HRMS (ESI-TOF) *m/z*: calcd for C₁₅H₁₉CINO [M+H]⁺ 264.1150; found 264.1154.

(3RS,4RS)-3-Chloro-1-cyclohexyl-4-phenylazetidin-2-one (5k). IR (NaCl,



neat): 3031, 2931, 2855, 1768 cm⁻¹; ¹H NMR (CDCl₃, 400 MHz): δ 0.97-1.30 (m, 4H), 1.50-1.70 (m, 3H), 1.71-1.82 (m, 2H), 2.00 (dm, J = 12.5 Hz, 1H), 3.38 (tt, J = 11.4, 3.8 Hz, 1H), 4.44 (brd, J = 1.7 Hz, H-4), 4.53 (brd, J = 1.7 Hz, 1H, H-3),

7.33-7.40 (m, 5H, ArH); ¹³C NMR (CDCl₃, 100 MHz): δ 24.9 (CH₂), 25.0 (CH₂), 25.1 (CH₂), 30.4 (CH₂), 31.2 (CH₂), 53.5 (CH), 62.6 (C-4), 65.1 (C-3), 126.7, 129.1, 129.3 (ArCH), 136.7 (C), 163.6 (C-2). HRMS (ESI-TOF) *m/z*: calcd for C₁₅H₁₉CINO [M+H]⁺ 264.1150; found 264.1153.

2,2-Dichloro-N-(ethoxyphenylmethyl)acetamide (3b). IR (NaCl, neat): 3260,



3060, 3005, 2974, 2922, 2853, 1673 cm⁻¹; ¹H NMR (CDCl₃, 400 MHz): δ 1.30 (t, *J* = 7.0 Hz, 3H, CH₃), 3.68 (dq, *J* = 9.4,

OEt O 7.0 Hz, 1H), 3.81 (dq, J = 9.4, 7.0 Hz, 1H), 5.97 (s, 1H, CHCl₂), 6.22 (d, J = 9.3 Hz, 1H), 6.87 (brd, J = 9.1 Hz, 1H, NH), 7.33-7.48 (m, 5H, ArH); ¹³C NMR (CDCl₃, 100 MHz): δ 15.1 (CH₃), 64.5 (CH₂), 66.4 (CHCl₂), 80.7 (CH), 125.8, 128.7, 128.8 (ArCH), 138.4 (C), 164.3 (CO). HRMS (ESI-TOF) *m/z*: calcd for C₁₁H₁₃Cl₂NNaO₂ [M+Na]⁺ 284.0216; found 284.0227.

2,2-Dichloro-N-[ethoxy(3-methoxyphenyl)methyl]acetamide (3f). IR (NaCl,



neat): 3261, 3061, 3005, 2977, 2925, 2855, 2835, 1674 cm⁻¹; ¹H NMR (CDCl₃, 400 MHz): δ 1.30 (t, *J* = 7.0 Hz, 3H, CH₃), 3.67 (dq, *J* = 9.4, 7.0 Hz, 1H), 3.80 (dq, *J* = 9.4, 7.0 Hz, 1H), 3.82 (s, 3H, CH₃), 5.97 (s, 1H, CHCl₂), 6.18 (d, *J* = 9.4 Hz,

1H), 6.85 (brs, 1H, NH), 6.90 (ddd, J = 8.2, 2.6, 0.7 Hz, 1H, ArH), 6.99-7.05 (m,

2H, ArH), 6.90 (t, J = 8.0 Hz, 1H, ArH); ¹³C NMR (CDCl₃, 100 MHz): δ 15.1 (CH₃), 55.3 (CH₃), 64.5 (CH₂), 66.3 (CHCl₂), 80.6 (CH), 111.4, 114.3, 118.0, 129.9 (ArCH), 140.0, 159.9 (C), 164.2 (CO). HRMS (ESI-TOF) *m/z*: calcd for C₁₂H₁₄Cl₂NO₃ [M-H]⁻ 290.0356; found 290.0360.

2,2-Dichloro-N-[ethoxy(3,5-difluorophenyl)methyl]acetamide (3h). IR (NaCl,

F OEt O

neat): 3276, 3057, 3005, 2981, 2930, 2915, 2901, 2889, 1685 cm⁻¹; ¹H NMR (CDCl₃, 400 MHz): δ 1.31 (t, *J* = 7.0 Hz, 3H, CH₃), 3.68 (dq, *J* = 9.4, 7.0 Hz, 1H), 3.82 (dq, *J* = 9.4, 7.0 Hz, 1H), 5.98 (s, 1H, CHCl₂), 6.19 (d, *J* = 9.4

Hz, 1H), 6.80 (tt, J = 8.8, 2.4 Hz, 1H, ArH), 6.80 (brs, 1H, NH), 6.96-7.03 (m, 2H, ArH); ¹³C NMR (CDCl₃, 100 MHz): δ 15.0 (CH₃), 64.8 (CH₂), 66.2 (CHCl₂), 79.6 (CH), 104.2 (t, J = 25.1 Hz, 1C, ArCH), 109.1 (d, J = 26.4 Hz, 2C, ArCH), 109.1 (d, J = 11.6 Hz, 1C, ArC), 1.63.1 (dd, J = 248.1, 12.5 Hz, 2C, CF), 164.4 (CO). HRMS (ESI-TOF) *m/z*: calcd for C₁₁H₁₀Cl₂F₂NO₂ [M-H]⁻ 296.0062; found 296.0052.

N-Benzyl-2,2-dichloro-*N*-(1-phenylvinyl)acetamide (6I). From 1I (100 mg, 0.28 mmol) and RuCl₂(PPh₃)₃ (27 mg, 0.028 mmol, 10%) in toluene (1 mL). The mixture was heated with stirring to 80 °C using microwave irradiation for 10 min. After chromatography (hexane-CH₂Cl₂, 1:0 to 1:1) 6I was isolated (36 mg, 40%). In this reaction 48 mg of 1I was also recovered. IR (NaCl, neat): 3061, 3029, 2952, 2924, 2854, 1690, 1625, 1576 cm⁻¹; ¹H NMR

(CDCl₃, 400 MHz): δ 4.66 (brs, 2H, CH₂Ar), 4.98 and 5.70 (2 s, 1H each, =CH₂), 6.38 (s, 1H, CHCl₂), 7.21-7.30 (m, 5H, ArH), 7.37-7.45 (m, 5H, ArH); ¹³C NMR (CDCl₃, 100 MHz): δ 50.7 (CH₂Ar), 64.0 (CHCl₂), 116.0 (=CH₂), 126.0, 127.9, 128.5, 129.2, 129.3, 129.9 (ArCH), 134.0 (C), 135.8 (C), 144.5 (C), 164.8 (CO). HRMS (ESI-TOF): Calcd for C₁₇H₁₆Cl₂NO [M+H]⁺ 320.0603; found 320.0603.

3. Cytotoxic activity and hemocompatibility studies of some of the synthesized lactams



Selected lactams for cytotoxic and hemocompatibility studies

3.1. Materials and methods

3.1.1. Materials

Dulbecco's modified Eagle's medium (DMEM), fetal bovine serum (FBS), L-glutamine solution (200 mM), trypsin-EDTA solution (170,000U/L trypsin and 0.2 g/L EDTA), penicillin–streptomycin solution (10,000 U/mL penicillin and 10 mg/mL streptomycin) and phosphate buffered saline (PBS) were obtained from Lonza (Verviers, Belgium). 2,5-Diphenyl-3,-(4,5-dimethyl-2-thiazolyl) tetrazolium bromide (MTT) and neutral red dye (NR) were from Sigma–Aldrich (St.Louis, MO, USA). The 75 cm² flasks and 96-well plates were obtained from TPP (Trasadingen, Switzerland). All other reagents were of analytical grade.

3.1.2. Methods

3.1.2.1. In vitro assay with Human erythrocytes

3.1.2.1.1 Obtention and extraction of the erythrocytes

Human blood samples were obtained from the Banc de Sang i Teixits de Barcelona from the Catalan Department of Health. Blood was deposited in tubes with anticoagulant EDTA-K₃. Blood samples were centrifugated at 3000 rpm at 4 °C for 10 min (Megafuge 2.0 R. Heraeus Instruments) to induce sedimentation. Plasma was extracted with a Pasteur pipette. Next, the residual pellet was washed with PBS pH 7.4. This procedure was repeated three times to remove residual leukocytes and platelets and to concentrate the erythrocytes. Following the last wash, the erythrocytes suspension was diluted (1:1) in PBS pH 7.4, to obtain suitable erythrocytes suspension (cell density of 8 × 10⁹ cell/mL).

3.1.1.2. Hemolysis assay

The hemolysis assay determined the capability of the different compounds to induce lysis of the erythrocyte membrane. A stock solution of 1 mg/mL in PBS pH 7.4.of each compound was prepared. Different volumes (10-80 μ L) were placed in polystyrene tubes, and an aliquot of 25 μ L of the erythrocyte suspensions was added to each tube. The final volume was 1 mL. The tubes were incubated at room temperature by rotatory conditions. After that, the tubes were centrifugated at 10.000 rpm for 5 min. The supernatants absorbance at 540 nm (Shimadzu UV-160A) was compared with those of the control samples hemolysed with distilled water (positive control). Negative control was obtained by incubating an aliquot of 25 μ L of the erythrocyte suspension with PBS pH 7.4.

The hemolysis degree was determined by the following equation:

Hemolysis (%) = $100 \times (Abs - Ab_0)/(Abs_{100} - Abs_0)$ where Abs, Abs_0, and Abs_{100} are the absorbance of test samples, the suspension treated with isotonic physiological buffer saline (PBS), and the suspension of complete hemolysis treated with distilled water, respectively.

3.1.2.2. Cell cultures

The immortal human keratinocyte (HaCaT) and the squamous cell carcinoma (A431) were obtained from Celltec UB. Cells were grown in DMEM medium (4.5 g/L glucose) supplemented with 10 % (v/v) FBS, 2 mM L-glutamine, 100 U/ mL penicillin and 100 μ g/ mL streptomycin at 37 °C, 5% CO₂. Cells were routinely cultured in 75 cm² culture flasks and were trypsinized using trypsin-EDTA when the cells reached approximately 80 % confluence.

3.1.2.3. Cell viability assays

HaCaT cells (1 × 10⁵ cells/ mL) and A431 (5 x 10⁴ cells/mL) were grown at the defined densities into the central 60 wells of a 96-well plate. Cells were incubated for 24 h under 5% CO₂ at 37 °C. Then, the spent medium was removed and cells were incubated for 24 h with the corresponding compound solutions (1 mg/mL) previously diluted in the minimum amount of DMF and then in DMEM medium supplemented with 5% FBS (100 μ L) at the required concentration range (10–250 μ g/mL).

3.1.2.3.1. NRU assay

Neutral red uptake (NRU) assay is based in the accumulation of the dye in the lysosomes of viable cells. After the cells were incubated for 24 h with the corresponding systems, the medium was removed, and the compounds solutions were incubated for 3 h with the NR dye (Sigma-Aldrich) solution (50 μ g/mL) dissolved in the medium without FBS and phenol red (Lonza). Cells were then washed with sterile PBS, followed by adding 100 μ L of a solution containing 50% ethanol absolute and 1% acetic acid in distilled water to extract the dye. To help the total dissolution of the dye, plates were placed in a microtitre-plate shaker for 5 min at room temperature. The absorbance of the resulting solutions was measured at 550 nm (Bio-Rad 550 microplate reader). Finally, the effect of each treatment was calculated as the percentage of tetrazolium salt reduction by viable cells against the control cells (cells without any treatment).

3.1.2.3.2. MTT assay

Only living cells can reduce the yellow tetrazolium salt, 2,5-diphenyl-3, -(4,5-dimethyl-2-thiazolyl) tetrazolium bromide (MTT) to insoluble purple formazan crystals. After 24 h of incubation of the cells with the corresponding NPs, the medium was removed, and 100 μ L of MTT (Sigma–Aldrich) in PBS (5 mg/mL) diluted 1:10 in culture medium without phenol red and absence of FBS (Lonza) was added to the cells. After 3 h of incubation the medium was removed. Thereafter, 100 μ L of DMSO (Sigma–Aldrich) was added to each well to dissolve the purple formazan crystals. Agitation, determination of the absorbance of the extracted solution, and the effect of each treatment measured at the same conditions that on 3.1.2.3.1.

3.1.2.3.3. Selectivity towards cancer cells

The corresponding half-maximal inhibitory concentration (IC₅₀) values for the different formulations as a function of cell line and endpoint were determined from the fitting of concentration-dependent viabilities curves.

The corresponding selectivity indexes toward cancer cells were calculated as the following ratio:

 $SI = IC_{50}$ (non-tumoral cell line)/IC₅₀ (tumoral cell line)

where HaCaT keratinocytes were used as closely representative of skin model cell lines under normal conditions.














110 100 f1 (ppm) ò 210 200



























110 100 f1 (ppm)

Experimental part



110 100 f1 (ppm)











6.2. Experimental part of chapter 3

1. Synthesis of protected allylamines

• Synthesis of 8a and 8b

For the preparation of **8a** and **8b** we followed the classical procedures reported in the literature³ isolating **8a** (93%) and **8b** (98%).

Methyl allyl(1,4-dioxaspiro[4.5]decan-8-yl)carbamate (8a). ¹H NMR (CDCl₃,



400 MHz): δ 1.57-1.84 (m, 4H, H-10, H-9), 3.70 (s, 4H, H-6, H-7), 3.79 (brs, 2H, CH₂N), 3.92-3.96 (m, 4H, H-8, CH₃), 3.98-4.19 (zm, 2H, H-2, H-3), 5.07 (d, *J* = 10.3, 1H, CH₂CH), 5.13 (d, *J* = 17.5 Hz, 1H, CH₂CH), 5.70-5.90 (m, 1H, CHCH₂); ¹³C NMR (CDCl₃, 100 MHz): δ 27.8 (CH₂Ar), 34.0 (CH₂Ar), 44.8 (CH₂N),

52.5 (CH₃), 64.2 (CHAr), 64.3 (C-2, C-3), 107.7 (CAr), 115.6 (CH₂CH), 135.8 (CHCH₂), 156.7 (CO).

Ethyl allyl(1,4-dioxaspiro[4.5]decan-8-yl)carbamate (8b). IR (NaCl, neat):



3080, 2879, 1694, 1467, 1448, 1413 cm⁻¹; ¹H NMR (CDCl₃, 400 MHz): δ 1.25 (t, *J* = 7.08 Hz, 3H, CH₃), 1.53-1.74 (m, 4H, H-10, H-9), 1.74-1.92 (m, 4H, H-6, H-7), 3.75 (brs, 2H, CH₂N), 3.93 (m, 5H, H-8, H-2, H-3), 4.14 (q, *J* = 6.3 Hz, 2H, CH₂O), 5.07 (dd, *J* = 10.3, 1.8 Hz, 1H, CH₂CH), 5.13 (d, *J* = 17.2 Hz,

1H, CH₂CH), 5.79 (tt, J = 10.3, 5.1 Hz, 1H, CHCH₂); ¹³C NMR (CDCl₃, 100 MHz): δ 14.6 (CH₃), 27.8 (CH₂Ar), 34.0 (CH₂Ar), 44.8 (CH₂N), 61.1 (CH₂O), 64.2 (CHAr), 64.3 (C-2, C-3), 107.8 (CAr), 115.5 (CH₂CH), 135.9 (CHCH₂), 156.2 (CO). HRMS (ESI-TOF) *m/z*: calcd for C₁₄H₂₄NO₄ [M+H]⁺ 270.1700; found 270.1705.

• Synthesis of 9a-9e

For the preparation of **9a-9e** we followed the same procedures reported in ref.3 isolating **9a** (73%), **9b** (84%), **9c** (99%) and **9d** (91%), **9e** (93%).

³ Diaba, F.; Montiel, J. A.; Serban, G.; Bonjoch, J. Org. Lett. 2015, 17, 3860-3863.

Methyl allyl (4-oxocyclohexyl) carbamate (9a). IR (NaCl, neat): 3081, 2955,



2873, 2360, 2341, 1698, 1546, 1469 cm⁻¹; ¹H NMR (CDCl₃, 400 MHz): δ 1.81-2.00 (m, 2H, H-6), 2.00-2.15 (m, 2H, H-2), 2.35-2.54 (m, 4H, H-3, H-5), 3.73 (s, 3H, CH₃O), 3.82 (s, 2H, CH₂N), 4.39 (bs, 1H, H-1), 5.03-5.23 (m, 2H, CH₂CH), 5.8 (ddt, J = 16.17, 10.62 Hz, CHCH₂); ¹³C NMR (CDCl₃, 100

MHz): δ 29.8 (CH₂Ar), 39.9 (CH₂Ar), 45.8 (CH₂N), 52.6 (CH₃O), 54.0 (CHAr), 116.1 (CH₂CH), 135.4 (CHCH₂), 156.4 (CO), 209.5 (CAr). HRMS (ESI-TOF) *m/z*: calcd for C₁₁H₁₈NO₃ [M+H]⁺ 212.1281; found 212.1280.

Ethyl allyl (4-oxocyclohexyl) carbamate (9b). IR (NaCl, neat): 3081, 2956,



1693, 1469, 1450, 1414 cm⁻¹; ¹H NMR (CDCl₃, 400 MHz): δ 1.28 (td, *J* = 7.12, 2.78 Hz, 3H, CH₃), 1.94 (d, *J* = 14.45 Hz, 2H, H-6), 2.00-2.10 (m, 2H, H-2), 2.36-2.54 (m, 4H, H-3, H-5), 3.81 (s, 2H, CH₂N), 4.18 (qd, *J* = 7.10, 2.72 Hz, 2H, CH₂O), 4.42 (bs, 1H, H-1), 5.06-5.20 (m, 2H, CH₂CH), 5.80 (dtt, *J* =

15.68, 5.48, 2.64 Hz, CHCH₂); ¹³C NMR (CDCl₃, 100 MHz): δ 14.6 (CH₃), 29.9 (CH₂Ar), 40.0 (CH₂Ar), 45.7 (CH₂N), 53.8 (CHAr), 61.4 (CH₂O), 116.1 (CH₂CH), 135.5 (CHCH₂), 156.0 (CO), 209.6 (CAr). HRMS (ESI-TOF) *m/z*: calcd for C₁₂H₂₀NO₃ [M+H]⁺ 226.1438; found 226.1437.

tert-Butyl allyl(4-oxocyclohexyl)carbamate (9c). IR (NaCl, neat): 3081, 3005,



2974, 2872, 1720, 1693, 1654, 1454 cm⁻¹; ¹H NMR (CDCl₃, 400 MHz): δ 1.47 (s, 9H, *t*-Bu), 2.00-2.09 (m, 4H, H-2, H-6), 2.35-2.53 (m, 4H, H-3, H-5), 3.75 (s, 2H, CH₂N), 4.44 (bs, 1H, H-1), 5.01-5.18 (m, 2H, CH₂CH), 5.70-5.86 (m, 1H, CHCH₂). ¹³C NMR (CDCl₃, 100 MHz): δ 28.4 (CH₃), 30.01 (CH₂Ar),

40.1 (CH₂Ar), 45.7 (CH₂N), 53.4 (C-1), 80.0 (CCH3), 115.7 (CH₂CH), 135.8 (CHCH₂), 155.2 (CO), 209.8 (C-4). HRMS (ESI-TOF) *m/z*: calcd for C₁₄H₂₄NO₃ [M+H]⁺ 254.1751; found 254.1754.

N-Allyl-*N*-(4-oxocyclohexyl)benzamide (9d). IR (NaCl, neat): 3061, 2955, 2360, 2341, 1715, 1632, 1494, 1412 cm⁻¹; ¹H NMR (CDCl₃, 400 MHz): δ 2.11 (s, 4H, H-2, H-6), 2.45 (s, 4H, H-3, H-5), 3.92 (s, 2H, CH₂N), 4.77 (bs, 1H, H-1), 5.15 (d, *J* = 10.42 Hz, 2H, CH₂CH), 5.79 (bs, 1H, CHCH₂), 7.34-7.47 (m, 5H, CHPh); ¹³C NMR (CDCl₃, 100 MHz): δ 29.6 (CH₂Ar), 39.9 (CH₂Ar), 49.0 (CH₂N),

53.1 (C-1), 117.0 (CH₂CH), 126.2(CHPh), 128.5(CHPh), 129.5(CPh), 135.2 (CHCH₂), 136.9 (CO), 209.5 (C-4). HRMS (ESI-TOF) *m/z*: calcd for C₁₆H₂₀NO₂ [M+H]⁺ 258.1485; found 258.1488.

N-Allyl-4-methyl-N-(4-oxocyclohexyl)benzenesulfonamide (9e). IR (NaCl,



neat): 3069, 2955, 2872, 2359, 1716, 1641, 1597, 1494 cm⁻¹; ¹H NMR (CDCl₃, 400 MHz): δ 1.82-1.96 (m, 4H, H-2, H-6), 2.32-2.42 (m, 4H, H-3, H-5), 2.43 (s, 3H, CH₃), 3.84 (d, *J* = 6.06, 1.49 Hz, 2H, CH₂N), 4.22 (m, 1H, H-1), 5.11 (dq, *J* = 10.19, 1.35 Hz, 1H, CH₂CH), 5.19 (dq, *J* = 17.16, 1.24 Hz, 1H, CH₂CH), 5.74-5.89 (m, 1H, CHCH₂), 7.70-

7.75 (m, 2H, CHAr), 7.28-7.34 (m, 2H, CHAr); ¹³C NMR (CDCl₃, 100 MHz): δ 21.5 (CH3), 30.03 (CH₂Ar), 40.0 (CH₂Ar), 46.6 (CH₂N), 56.7 (C-1), 80.0 (CCH3), 117.4 (CH₂CH), 126.9(CHAr), 129.8(CHAr), 136.0 (CHCH₂), 138.0 (CS), 143.4 (CCH₃), 208.7 (C-4). HRMS (ESI-TOF) *m/z*: calcd for C₁₆H₂₂NO₃S [M+H]⁺ 308.1315; found 308.1312.

• Synthesis of 14a and 14b

Allylamines **14a** and **14b** were prepared from **9a** and **9d** reapectively following the experimental reported previously ⁴ isolating **14a** (75%) and **14b** (82%)

Methyl allyl(4-oxocyclohex-2-en-1-yl)carbamate (14a). IR (NaCl, neat): 3364,



2981, 2361, 1689, 1455, 1410, 1382 cm⁻¹; ¹H NMR (CDCl₃, 400 MHz): δ 1.27 (tdf, *J* = 7.09, 1.84 Hz, 1H, H-6) 2.19 (bs, 1H, H-6), 2.19 (s, 1H, H-5), 2.4-2.6 (m, 1H, H-5), 3.82 (s, 3H, CH₃), 4.19 (qd, *J* = 7.08, 1.67 Hz, 2H, CH₂N), 4.7 (s, 1H, H-1), 4.99 (bs, 1H,

⁴ Nicolaou, K. C.; Montagnon, T.; Baran, P. S. ; Zhong, Y.-L. *J. Am. Chem. Soc.* **2002**, *124*, 2245-2258.

CH₂CH), 5.16 (d, J = 11.05 Hz, 1H, CH₂CH), 5.76-5.91 (m, 1H, CHCH₂), 6.04 (d, J = 10.31 Hz, 1H, H-3), 6.85 (dft, J = 10.34, 1.43 Hz, 1H, H-2); ¹³C NMR (CDCl₃, 100 MHz): δ 14.6 (C-6), 36.9 (C-5), 47.2 (CH₂N), 54.1 (CH₃), 61.7 (CHAr), 116.6 (CH₂CH), 130.6 (C-3), 134.9 (CHCH₂), 152.2 (H-2), 156.0 (CO), 197.9 (CAr). HRMS (ESI-TOF) *m/z*: calcd for C₁₁H₁₈NO₃ [M+H]⁺ 224.1281; found 224.1284.

N-Allyl-*N*-(4-oxocyclohex-2-en-1-yl) benzamide (14b). IR (NaCl, neat): 3059, 2954, 1683, 1633, 1601, 1577, 1493, 1405 cm⁻¹; ¹H NMR (CDCl₃, 400 MHz): δ 2.24 (s, 1H, H-6), 2.46 (s, 1H, H-6), 2.59 (s, 2H, H-5), 3.89 (d, *J* = 15.25 Hz, 2H, CH₂N), 4.77 (bs, 1H, H-1), 5.20 (dt, *J* = 10.36, 1.34 Hz, 2H, CH₂CH), 5.78 (bs, 1H, H-2), 6.07 (ddd, *J* = 10.32, 2.84, 1.08 Hz, 1H, H-3), 6.94 (s, 1H, CHCH₂), 7.39-7.47

(m, 5H, CHPh); ¹³C NMR (CDCl₃, 100 MHz): δ 27.6 (C-6), 36.9 (C-5), 50.3 (CH₂N), 53.4 (C-1), 117.9 (CH₂CH), 126.5 (CHPh), 128.6 (CHPh), 129.9 (C-3), 134.3 (C-2), 135.9 (CPh), 152.3 (CHCH₂), 172.0 (CO), 197.5 (C-4). HRMS (ESI-TOF) *m/z*: calcd for C₁₆H₁₈NO₂ [M+H]⁺ 256.1332; found 256.1326.

• Synthesis of *N*-allylacrylamide (14c)

14c was prepared from acryloyl chloride and allylamine following the experimental reported previously.⁵

Methyl allyl(1-phenylbut-3-en-1-yl)carbamate (14d). IR (NaCl, neat): 3077,



3029, 2979, 2953, 1698, 1642, 1602, 1454 cm⁻¹; ¹H NMR (CDCl₃, 400 MHz): δ 2.73 (m, 2H, CH₂CHN), 3.55 (dd, *J* =15.95, 6.58 Hz, 1H, CH₂N), 3.66 (s, 1H, CH₂N), 3.72 (s, 3H, CH₃), 4.89-5.00 (m, 2H, CH₂CH), 5.03-5.16 (m, 2H, CH₂CH), 5.40 (s, 1H,

CHN), 5.51-5.68 (m, 1H, CHCH₂), 5.79 (ddt, *J* =17.01, 10.22, 6.74 Hz, 1H, CHCH₂), 7.26 (t, *J* =4.49 Hz, 3H, CHAr), 7.32 (d, *J* =4.93 Hz, 2H, CHAr); ¹³C NMR

⁵ Qu, Q.; Liu, G.; Lv, X.; Zhang, B.; An, Z. ACS Macro Lett. **2016**, *5*, 316–320.

(CDCl₃, 100 MHz): δ 35.5 (CH₂CHN), 46.2 (CH₂N), 52.6 (CH₃), 58.5 (CHN), 116.0 (CH₂CH), 117.4 (CH₂CH), 127.5 (CHAr), 128.0 (CHAr), 128.3 (CHAr), 134.9 (CHCH₂), 135.3 (CHCH₂), 139.7 (CO), 157.1 (CAr). HRMS (ESI-TOF) *m/z*: calcd for C₁₅H₂₀NO₂ [M+H]⁺ 246.1489; found 246.1481.

Synthesis of N-Allyl-N-(*tert*-butyl)-2-chloroacetamide (14e).
CH₂Cl
N
O

14e was prepared following the procedure reported in the literature.⁶

 General procedure for the isomerization of protected allylamines 8a and 8b in the presence of Ru catalysts



In a 10 mL vessel allylamine **8** was placed (0.443 mmol) and ruthenium catalyst (0.022 mmol, 5%) in toluene (1 mL). The mixture was heated with stirring using microwave irradiation for 45 min at 120 °C. After chromatography (hexane-CH₂Cl₂, 9:1 to CH₂Cl₂) **10** and/or **11** were isolated.

Methyl (Z)-prop-1-en-1-yl(1,4-dioxaspiro[4.5]decan-8-yl)carbamate (10a). IR



(NaCl, neat): 3032, 2949, 2878, 1698, 1659, 1446, 1398 cm⁻¹; ¹H NMR (CDCl₃, 400 MHz): δ 1.54 (dd, *J* = 6.86, 1.79 Hz, 3H, CH₃CH), 1.62-1.81 (m, 8H, CH₂Ar), 3.70 (s, 3H, CH₃O), 3.93 (s, 4H, H-2, H-3), 4.10 (bs, 1H, H-8), 5.53-5.64 (m, 1H, CHCH₃), 6.15 (dq, *J* = 7.94, 1.81 Hz, 1H, CHN); ¹³C NMR

⁶ Zhang, Z.; Zhu, L.; Li, C. *Chin. J. Chem.* **2019**, *37*, 452-456.

(CDCl₃, 100 MHz): δ 12.5 (CH₃CH), 27.2(CH₂Ar), 33.8(CH₂Ar), 52.7 (CH₃O), 54.9 (CHAr), 64.3 (C-2), 64.3 (C-3), 107.8 (CAr), 123.4 (CHN), 127.2 (CHCH₃), 155.3 (CO); HRMS (ESI-TOF) *m/z*: calcd for C₁₃H₂₂NO₄ [M+H]⁺ 256.1543; found 256.1543.

Methyl (E)-prop-1-en-1-yl(1,4-dioxaspiro[4.5]decan-8-yl)carbamate (11a). IR

(NaCl, neat): 3383, 2935, 2360, 2341, 1704, 1662, 1443 cm⁻¹; ¹H NMR (CDCl₃, 400 MHz): δ 1.62-1.68 (m, 4H, H-10, H-9), 1.71 (dd, *J* = 6.67, 1.67 Hz, 3H, CH₃CH), 1.74-1.83 (m, 2H, H-6), 1.93-2.08 (m, 2H, H-7), 3.72 (s, 3H, CH₃O), 3.92-3.97 (m, 4H, H-2, H-3), 3.94 (s, 1H, H-8), 5.40 (dq, *J*

= 13.48, 6.67 Hz, 1H, CHCH₃), 6.15 (d, J = 14.08 Hz, 1H, CHN); ¹³C NMR (CDCl₃, 100 MHz): δ 15.5 (CH₃CH), 27.3 (CH₂Ar), 34.1 (CH₂Ar), 52.7 (CH₃O), 55.2 (CHAr), 64.3 (C-2, C-3), 107.8 (CAr), 117.1 (CHCH₃), 125.8 (CHN), 155.4 (CO). HRMS (ESI-TOF) *m/z*: calcd for C₁₃H₂₂NO₄ [M+H]⁺ 256.1543; found 256.1543.

Ethyl (Z)-prop-1-en-1-yl(1,4-dioxaspiro[4.5]decan-8-yl)carbamate (10b). IR



(NaCl, neat): 3029, 2954, 2871, 1716, 1694, 1660, 1531 cm⁻¹; ¹H NMR (CDCl₃, 400 MHz): δ 1.27 (td, *J* = 7.08, 6.64 Hz, 3H, CH₃CH₂), 1.56 (dd, *J* = 6.86, 1.78 Hz, 3H, CH₃CH), 1.80-1.97 (m, 2H, H-10), 2.00-2.14 (m, 2H, H-9), 2.36-2.50 (m, 4H, H-6, H-7), 2.51 (dd, *J* = 14.25, 5.9 Hz, 4H, H-2, H-3), 4.17 (qt, *J* =

7.13, 1.12 Hz, 2H, CH₂O), 4.52 (t, J = 12.25 Hz, 1H, H-8), 5.6 (dq, J = 7.85, 6.85 Hz, 1H, CHCH₃), 5.75 (dq, J = 7.84, 1.76 Hz, 1H, CHN); ¹³C NMR (CDCI₃, 100 MHz): δ 12.6 (CH₃CH), 14.7 (CH₃CH₂), 29.3 (CH₂Ar), 39.8 (CH₂Ar), 40.0 (C-2, C-3), 54.0 (CHAr), 61.6 (CH₂O), 108.4 (CAr), 123.1 (CHN), 127.5 (CHCH₃), 154.8 (CO). HRMS (ESI-TOF) *m/z*: calcd for C₁₄H₂₄NO₄ [M+H]⁺ 270.1700; found 270.1703.

Ethyl (E)-prop-1-en-1-yl(1,4-dioxaspiro[4.5]decan-8-yl)carbamate.(11b) IR



(NaCl, neat): 2940, 2880, 1703, 1662, 1465, 1432, 1411, 1371 cm⁻¹; ¹H NMR (CDCl₃, 400 MHz): δ 1.28 (tt, *J* = 7.08, 1.15 Hz, 3H, CH₃CH₂), 1.57-1.68 (m, 4H, H-10, H-9), 1.64-1.74 (m, 3H, CH₃CH), 1.74-1.84 (m, 2H, H-6), 1.96-2.11 (m, 2H, H-7), 3.88-4.01 (m, 5H, H-8, H-2, H-3), 4.16 (qt, *J* = 7.13, 2H, H-7), 3.88-4.01 (m, 5H, H-8, H-2, H-3), 4.16 (qt, *J* = 7.13)

1.12 Hz, 2H, CH₂O), 5.38 (dqd, J = 13.42, 6.68, 1.45 Hz, 1H, CHCH₃), 6.19 (d, J = 14.20 Hz, 1H, CHN); ¹³C NMR (CDCl₃, 100 MHz): δ 14.6 (CH₃CH₂), 15.6 (CH₃CH), 27.2 (CH₂Ar), 34.2 (CH₂Ar), 55.1 (CHAr), 61.5 (CH₂O), 64.3 (C-2), 64.3 (C-3), 107.8 (CAr), 115.7 (CHCH₃), 126.0 (CHN), 154.9 (CO). HRMS (ESI-TOF) *m/z*: calcd for C₁₄H₂₄NO₄ [M+H]⁺ 270.1700; found 270.1707.

3. General procedure for the isomerization of protected allylamines 9a-9d in the presence of RuCl₂(PPh₃)₃



R = MeO(9a), EtO(9b)*t*-BuO(9c), Ph (9d), Ts (9e)

In a 10 mL vessel allylamine **9** was placed (0.443 mmol) and ruthenium catalyst (0.022 mmol, 5%) in toluene (1 mL). The mixture was heated with stirring using microwave irradiation for 45-75 min at 120 °C. After chromatography (hexane- CH_2Cl_2 , 9:1 to CH_2Cl_2) 1**2** and/or **13** were isolated.

Methyl (*Z*)-(4-oxocyclohexyl)(prop-1-en-1-yl)carbamate. (12a) IR (NaCl, neat): 3346, 3029, 2953, 2869, 1715, 1658, 1531, 1447 cm⁻¹; ^{Me} O H H H NMR (CDCl₃, 400 MHz): δ 1.56 (dd, *J* = 6.84, 1.75 Hz, 3H, ^{CH₃} CH₃ CH), 1.80-1.96 (m, 2H, H-6), 2.04-2.13 (m, 2H, H-2), 2.36-2.55 (m, 4H, H-3, H-5), 3.73 (s, 3H, CH₃O), 4.51 (t, *J* = 12.17 Hz, 1H, H-1), 5.62 (dq, *J* = 7.82, 6.83 Hz, 1H, CHCH₃), 5.75

(dq, J =7.82, 1.77 Hz, 1H, CHN); ¹³C NMR (CDCl₃, 100 MHz): δ 12.5 (CH₃CH), 29.3 (CH₂Ar), 39.8 (CH₂Ar), 52.9 (CH₃O), 54.2 (CHAr), 123.1 (CH₂CH), 127.8 (CHAr), 155.3 (CO), 209.5 (CAr); HRMS (ESI-TOF) *m/z*: calcd for C₁₁H₁₈NO₃ [M+H]⁺ 212.1281; found 212.1276.





neat): 3355, 2955, 2885, 1713, 1663, 1530, 1445 cm⁻¹; ¹H NMR (CDCI₃, 400 MHz): δ 1.72 (dd, *J* = 6.68, 1.68 Hz, 3H, CH₃CH), 1.98-2.08 (m, 2H, H-6), 2.13-2.28 (m, 2H, H-2), 2.38-2.52 (m, 4H, H-3, H-5), 3.74 (s, 3H, CH₃O), 4.33 (tt, *J* = 11.92, 3.97 Hz, 1H, H-1), 5.36 (dq, *J*=14.13, 6.68 Hz, 1H,

CHCH₃), 6.22 (d, J = 14.09 Hz, 1H, CHN); ¹³C NMR (CDCl₃, 100 MHz): δ 15.4 (CH₃CH), 29.1 (CH₂Ar), 40.0 (CH₂Ar), 52.8 (CH₃O), 54.5 (CHAr), 116.7 (CH₂CH), 126.2 (CHAr), 155.1 (CO), 209.7 (CAr). HRMS (ESI-TOF) *m/z*: calcd for C₁₁H₁₈NO₃ [M+H]⁺ 212.1281; found 212.1276.

Ethyl (Z)-(4-oxocyclohexyl)(prop-1-en-1-yl)carbamate (12b). IR (NaCl, neat):



3350, 3030, 2976, 2915, 2871, 1716, 1660, 1532 cm⁻¹; ¹H NMR (CDCl₃, 400 MHz): δ 1.26 (td, J = 7.11, 0.71 Hz, 3H, CH₃CH₂), 1.56 (ddd, J = 6.84, 1.77, 0.70 Hz, 3H, CH₃CH), 1.80-1.97 (m, 2H, H-6), 2.02-2.14 (m, 2H, H-2), 2.36-2.55 (m, 4H, H-3, H-5), 4.17 (qd, J = 7.12, 0.70 Hz, CH₂O), 4.52 (t, J =

12.03 Hz, 1H, H-1), 5.54-5.66 (m, 1H, CHCH₃), 5.75 (dqd, J=7.83, 1.76, 0.66 Hz, 1H, CHN); ¹³C NMR (CDCl₃, 100 MHz): δ 12.6 (CH₃CH), 14.6 (CH₃CH₂), 29.3 (CH₂Ar), 39.8 (CH₂Ar), 54.0 (CHAr), 61.6 (CH₂O), 123.2 (CHN), 127.5 (CHCH₃), 154.8 (CO), 209.6 (CAr). HRMS (ESI-TOF) *m/z*: calcd for C₁₂H₂₀NO₃ [M+H]⁺ 226.1438; found 226.1436.

Ethyl (E)-(4-oxocyclohexyl)(prop-1-en-1-yl)carbamate (13b). IR (NaCl, neat):



3094, 2959, 2938, 2887, 2050, 1983, 1704, 1662 cm⁻¹; ¹H NMR (CDCl₃, 400 MHz): δ 1.29 (t, *J* = 7.12 Hz, 3H, CH₃CH₂), 1.72 (dd, *J* = 6.65, 1.67 Hz, 3H, CH₃CH), 1.97-2.07 (m, 2H, H-6), 2.15-2.35 (m, 2H, H-2), 2.45 (dd, *J* = 9.38, 4.64 Hz, 4H, H-3, H-5), 4.18 (q, *J* = 7.12 Hz, CH₂O), 4.31 (s, 1H, H-

1), 5.33 (dq, J=13.54, 6.58 Hz, 1H, CHCH₃), 6.27 (d, J=14.12 Hz, 1H, CHN); ¹³C NMR (CDCl₃, 100 MHz): δ 14.6 (CH₃CH₂), 15.4 (CH₃CH), 29.0 (CH₂Ar), 40.1 (CH₂Ar), 54.4 (CHAr), 61.7 (CH₂O), 120.4 (CHCH₃), 126.4 (CHN), 154.6 (CO), 209.8 (CAr). HRMS (ESI-TOF) *m/z*: calcd for C₁₂H₂₀NO₃ [M+H]⁺ 226.1438; found 226.1435.

tert-Butyl (Z)-(4-oxocyclohexyl)(prop-1-en-1-yl)carbamate (12c). IR (NaCl,



neat): 3029, 2973, 2930, 2870, 1719, 1692, 1660 cm⁻¹; ¹H NMR (CDCl₃, 400 MHz): δ 1.46 (s, 9H, *t*-Bu), 1.56 (dd, *J* = 6.85, 1.79 Hz, CH₃CH), 1.85 (qd, *J* = 12.81, 5.04 Hz, 2H, H-6), 2.00-2.13 (m, 2H, H-2), 2.35-2.53 (m, 4H, H-3, H-5), 4.48 (bs, 1H, H-1), 5.54 (dq, *J* = 7.85, 6.85 Hz, 2H, CHCH₃), 5.71

(dq, *J* = 7.83, 1.79 Hz, 1H, CHN); ¹³C NMR (CDCl₃, 100 MHz): δ 12.7 (CH₃), 28.3 (CH₃-*t*Bu), 29.4 (CH₂Ar), 39.9 (CH₂Ar), 53.5 (C-1), 80.0 (CCH3), 123.5 (CHN), 126.7 (CHCH₃), 158.4 (CO), 209.8 (C-4).

(Z)-N-(4-Oxocyclohexyl)-N-(prop-1-en-1-yl)benzamide (12d). IR (NaCl, neat):



3029, 2914, 2870, 1716, 1638, 1578, 1537, 1446 cm⁻¹; ¹H NMR (CDCl₃, 400 MHz): δ 1.24 (dd, *J* = 6.98, 1.79 Hz, 3H, CH₃), 1.93 (qd, *J* = 12.77, 4.54 Hz, 2H, H-2), 2.16-2.27 (m, 2H, H-6), 2.44 (dt, *J* = 4.92, 2.24 Hz, 1H, H-3), 2.48 (dt, *J* = 4.70, 2.32 Hz, 1H, H-3), 2.50-2.64 (m, 2H, H-5), 4.99 (bs, 1H, H-1), 5.35 (q, *J* =

7.18, CHCH₃), 6.02 (dq, J = 7.86, 1.82 Hz, 1H, CHCH) 7.28-7.35 (m, 3H, CHPh), 7.46-7.53 (m, 2H, CHPh); ¹³C NMR (CDCl₃, 100 MHz): δ 12.5 (CH₃), 29.0 (CH₂Ar), 39.8 (CH₂Ar), 52.6 (H-1), 125.7 (CHCH), 127.5 (CHPh), 128.0 (CHPh), 128.0 (CHCH₃), 129.9 (CPh), 170.6 (CO), 209.5 (C-4). HRMS (ESI-TOF) *m/z*: calcd for C₁₆H₂₀NO₂ [M+H]⁺ 258.1485; found 258.1485.

4. Reaction of protected allylamines 14a and 14b with RuCl₂(PPh₃)₃



In a 10 mL vessel allylamine **14** was placed (0.443 mmol) and ruthenium catalyst (0.022 mmol, 5%) in toluene (1 mL). The mixture was heated with stirring using microwave irradiation for 90-120 min at 120-140 °C. After chromatography (hexane-CH₂Cl₂, 9:1 to CH₂Cl₂) **15** and/or **16** were isolated.





(NaCl, neat): 3031, 2954, 2918, 1693, 1660, 1536, 1447 cm⁻¹; ¹H NMR (CDCl₃, 400 MHz): δ 1.61 (dd, *J* = 6.93, 1.77 Hz, 3H, CH₃), 2.13-2.28 (m, 2H, H-6), 2.39-2.53 (m, 1H, H-5), 2.54-2.64 (m, 1H, H-5), 3.75 (s, 3H, CH₃O), 5.03 (s, 1H, H-1), 5.60 (dq, *J* = 7.89, 6.90 Hz, 1H, CHCH₃), 5.80 (dq, *J* =

7.86, 1.78 Hz, 1H, H-3), 6.04 (ddd, J = 10.31, 2.77, 1.13 Hz, 1H, CHCH), 6.84 (ddd, J = 10.40, 2.22, 1.30 Hz, 1H, H-2); ¹³C NMR (CDCl₃, 100 MHz): δ 12.7 (CH₃), 27.4 (C-6), 36.7 (C-5), 53.1 (CH₃O), 54.3 (CHAr), 124.1 (C-3), 126.9 (CHCH₃), 130.5 (CHCH), 151.2 (C-2), 152.3 (CO), 197.8 (CAr). HRMS (ESI-TOF) *m/z*: calcd for C₁₁H₁₆NO₃ [M+H]⁺ 210.1125; found 210.1124.

J = 10.36, 2.97, 1.17 Hz, 1H, H-3), 6.54 (d, J = 14.56 Hz, 1H, CHCH), 6.93 (dt, J = 10.40, 2.11 Hz, 1H, H-2); ¹³C NMR (CDCl₃, 100 MHz): δ 15.5 (CH₃), 26.9 (C-6), 37.1 (C-5), 53.2 (CH₃O), 53.4 (CHAr), 109.8 (CHCH₃), 126.1 (CHCH), 128.8 (C-3), 153.1 (C-2), 154.5 (CO), 197.7 (CAr). HRMS (ESI-TOF) *m/z*: calcd for C₁₁H₁₆NO₃ [M+H]⁺ 210.1125; found 210.1127.

(Z)-N-(4-Oxocyclohex-2-en-1-yl)-N-(prop-1-en-1-yl)benzamide.(15b)



(NaCl, neat): 3059, 3034, 2955, 1716, 1681, 1635, 1601, 1578, 1530 cm⁻¹; ¹H NMR (CDCl₃, 400 MHz): δ 1.26 (dd, J = 7.05, 1.78 Hz, 3H, CH₃), 2.15-2.37 (m, 2H, H-6), 2.50-2.67 (m, 2H, H-5), 5.36 (p, J = 7.18, CHCH₃), 5.57 (bs, 1H, H-1), 6.05 (dq, J = 7.88, 1.83 Hz, 1H, H-2), 6.12 (ddd, J = 10.32, 2.84, 1.08 Hz, 1H, H-3),

6.91 (dt, J = 10.34, 2.02 Hz, 1H, CHCH), 7.51-7.39 (m, 3H, CHPh), 7.51-7.57 (m, 2H, CHPh); ¹³C NMR (CDCl₃, 100 MHz): δ 12.7 (CH₃), 26.9 (CH₂Ar), 36.6 (CH₂Ar), 52.6 (H-1), 126.3 (H-2), 127.6 (CHCH₃), 127.6 (CHPh), 128.3 (CHPh),

130.3 (CHPh), 131.1 (H-3), 135.7 (CPh), 150.05 (CHCH), 170.6 (CO), 197.9 (C-4). HRMS (ESI-TOF) *m/z*: calcd for C₁₆H₁₈NO₂ [M+H]⁺ 256.1332; found 256.1326.

7. Reaction of 14c with RuCl₂(PPh₃)₃

In a 10 mL vessel allylamine **14c** was placed (49.2 mg, 0.443 mmol) and ruthenium catalyst (0.022 mmol, 5%) in toluene (1 mL). The mixture was heated with stirring using microwave irradiation for 30 min at 120 °C. After chromatography (hexane-CH₂Cl₂, 9:1 to CH₂Cl₂) **15** and/or **16** were isolated.



(Z)-N-(Prop-1-en-1-yl)acrylamide (15c). IR (NaCl, neat): 3253, 3194, 3055, 2922, 2855, 2779, 1658, 1625, 1537 cm⁻¹; ¹H NMR (CDCl₃, 400 MHz): δ 1.31 (dd, J =6.8, 1.6 Hz, 3H, CH₃), 5.17-5.31 (m, 2H, CH₂CH₃), 5.69 (dd, J =10.3, 1.3 Hz, 1H, CH₂CH), 6.09 (q, J =12.9 Hz, 1H, CH₂CH), 6.35 (dd, J =16.9, 3.1 Hz, 1H, CHCO), 6.77-6.89 (m, 1H, CHNH), 7.13 (bs, 1H, NH); ¹³C NMR (CDCl₃, 100 MHz): δ 14.8 (CH₃), 108.9 (CH₂CH₃), 123.1 (CH₂CH), 127.5 (CHNH), 130.2 (CHCO), 162.1 (CO). HRMS (ESI-TOF) *m/z*: calcd for C₉H₁₀NO [M+H]⁺ 112.0757; found 112.0757.

8. Reaction of 14d with RuCl₂(PPh₃)₃



In a 10 mL vessel allylamine **14d** was placed (109 mg, 0.443 mmol) and ruthenium catalyst (0.022 mmol, 5%) in toluene (1 mL). The mixture was heated with stirring using microwave irradiation for 75 min at 120 °C. After chromatography (hexane-CH₂Cl₂, 9:1 to CH₂Cl₂) **15d** was isolated.

Methyl (Z)-(1-phenylbut-3-en-1-yl)(prop-1-en-1-yl)carbamate (15d). IR (NaCl,



neat): 3064, 3030, 2978, 2951, 2916, 1698, 1160, 1445 cm⁻¹; ¹H NMR (CDCl₃, 400 MHz): δ 1.31 (dd, *J* =6.8, 1.64 Hz, 3H, CH₃), 2.61-2.78 (m, 2H, CH₂CHN), 3.73 (d, *J* =2.17 Hz, 3H, CH₃O), 5.03-5.17 (m, 2H, CH₂CH), 5.48 (dq, *J* =7.85, 6.73 Hz,

1H, CHCH₃), 5.51 (s, 1H, CHN), 5.57 (dq, J =7.96, 1.66 Hz, 1H, CHCH₂), 5.79 (dddd, J =17.34, 10.22, 7.24, 6.19 Hz, 1H, CHNCO), 7.22-7.36 (m, 5H, CHAr). ¹³C NMR (CDCl₃, 100 MHz): δ 12.8 (CH₃), 35.2 (CH₂CHN), 52.8 (CH₃O), 58.9 (CHN), 117.4 (CH₂CH), 123.0 (CHCH₂), 126.9 (CHCH₃), 127.5 (CHAr), 127.9 (CHAr), 128.2(CHAr), 134.8 (CHNCO), 139.3 (CO), 157.5 (CAr). HRMS (ESI-TOF) *m/z*: calcd for C₁₅H₂₀NO₂ [M+H]⁺ 246.1489; found 246.1484.

9. Reaction of 14e with RuCl₂(PPh₃)₃



In a 10 mL vessel allylamine **14e** was placed (73 mg, 0.386 mmol) and ruthenium catalyst (0.019 mmol, 5%) in toluene (1 mL). The mixture was heated with stirring using microwave irradiation for 30 min at 160 °C. After chromatography (hexane-CH₂Cl₂, 9:1 to CH₂Cl₂) **15e** and/or **16e** were isolated.
























































6.3. Experimental part of chapter 4

1. Synthesis of trichloro- and dichloroacetamides 17

Starting materials of this part of the work (**17a-17p**) were prepared in the context of another project

2. General procedure for the reaction of trichloro- and dichloroacetamides 17 with RuCl₂(PPh₃)₃

In a 10 mL vessel were placed chloroacetamide **17** (0.386 mmol) and $RuCl_2(PPh_3)_3$ (0.019 mmol, 5 mol%) in toluene (1 mL). The mixture was heated with stirring using microwave irradiation for 15 min (**17a**, **17b**, **17f**, **17h-17p**), 30 min (**17c-17e**) or 45 min (**17g**) at 160 °C. After chromatography (hexane-CH₂Cl₂, 9:1 to CH₂Cl₂) **18** and/or **19** were isolated.

1-(tert-Butyl)-3,3-dichloro-4-(chloromethyl)pyrrolidin-2-one (18a). IR (NaCl,



neat): cm⁻¹; 2978, 2937, 2874, 1715, 1667 cm⁻¹; ¹H NMR (CDCl₃, 400 MHz): δ 1.44 (s, 9H, *t*-Bu), 2.95-3.03 (m, 1H, CH), 3.22 (dd, *J* = 10.10, 8.22 Hz, 1H, CH2Cl), 3.68-3.75 (m, 2H, CH₂Cl, CH₂N), 3.99 (dd, *J* = 11.2, 4.2 Hz, 1H, CH₂N); ¹³C NMR (CDCl₃, 100 MHz):

 δ 27.2 (CH₃), 41.2 (C-5), 46.0 (CH2), 51.0 (CH), 55.6 (C), 85.0 (C-3), 165.6 (CO). HRMS (ESI-TOF): Calcd for C₉H₁₅Cl₃NO [M+H]⁺ 258.0214; found 258.0211.

(CDCl₃, 100 MHz): δ 27.4 (CH₃), 43.6 (C-5), 44.3 (C-4), 45.9 (CH₂Cl), 58.1 (C-3), 55.1 (C), 168.4 (CO). HRMS (ESI-TOF) *m/z*: calcd for C₉H₁₆Cl₂NO [M+H]⁺ 224.0603; found 224.0602.

(3RS,4RS)-1-(tert-Butyl)-3-chloro-4-(chloromethyl)-3-methylpyrrolidin-2-



one (18c). IR (NaCl, neat): 3003, 2974, 2930, 2872, 1686, 1650 cm⁻¹; ¹H NMR (CDCl₃, 400 MHz): δ 1.42 (s, 9H, *t*-Bu), 1.73 (s, 3H, CH₃), 2.39-2.50 (m, 1H, H-4), 3.15 (ddd, J = 9.9, 9.1, 0.5 Hz, 1H, CH₂Cl), 3.63 (dd, J = 10.0, 6.9 Hz, 1H, CH₂Cl), 3.66-3.72

(m, 1H, H-5), 3.82 (dd, *J* =11.2, 5.1 Hz, 1H, H-5); ¹³C NMR (CDCl₃, 100 MHz): δ 24.7(CH₃), 27.4(CH₃), 42.3 (C-5), 46.5 (C-4), 47.1 (CH₂Cl), 54.8 (C), 70.6 (C-3), 171.0 (CO). HRMS (ESI-TOF) m/z: calcd for C10H18Cl2NO [M+H]+ 238.076; found 238.0762.

(3RS,4SR)-1-(tert-Butyl)-3-chloro-4-(chloromethyl)-3-methylpyrrolidin-2-

one (19c). IR (NaCl, neat): 3345, 2976, 2929, 1698, 1479, 1406, 1365 cm⁻¹; ¹H NMR (CDCl₃, 400 MHz): δ 1.42 (s, 9H, *t*-Bu), 1.59 (s, 3H, CH₃), 2.79-2.89 (m, 1H,

H-4), 3.31 (ddd, J = 10.2, 4.4, 0.6 Hz, 1H, CH₂Cl), 3.37-3.45 (m, MHz): δ 21.2(CH₃), 27.3(CH₃), 42.5 (C-5), 45.8 (CH₂Cl), 47.5 (C-MHz): δ 21.2(CH₃), 27.3(CH₃), 42.5 (C-5), 45.8 (CH₂Cl), 47.5 (C-4), 54.8 (C), 69.3 (C-3), 170.9 (CO). HRMS (ESI-TOF) m/z: calcd for C₁₀H₁₈Cl₂NO [M+H]⁺ 238.076; found 238.0767.

Ethyl (3RS,4SR)-1-(tert-Butyl)-3-chloro-4-(chloromethyl)-2-oxopyrrolidine-



3-carboxylate (18d). IR (NaCl, neat): 2976, 2935, 1704, 1698, 1478, 1457 cm⁻¹; ¹H NMR (CDCl₃, 400 MHz): δ 1.29-1.37 (m, 3H, CH₃), 1.42 (s, 9H, *t*-Bu), 3.18 (t, *J* = 9.7 Hz, 1H, CH₂Cl), 3.35-3.45 (m, 1H, H-4), 3.56-3.79 (m, 3H, CH₂Cl, H-

5), 4.20-4.42 (m, 2H, CH₂O); ¹³C NMR (CDCl₃, 100 MHz): δ 14.0 (CH₃), 27.3 (CH₃), 41.5 (H-5), 44.1 (C-4), 46.3 (CH₂Cl), 55.4 (C), 63.5 (CH₂O), 72.3 (C-3), 166.3 (C-2), 167.0 (CO). HRMS (ESI-TOF) m/z: calcd for C12H20Cl2NO3 [M+H]+ 296.0815; found 296.0814.

Ethyl (3RS,4RS)-1-(tert-Butyl)-3-chloro-4-(chloromethyl)-2-oxopyrrolidine-



3-carboxylate (19d). IR (NaCl, neat): 2973, 2935, 2908, 2872, 1758, 1705 cm⁻¹; ¹H NMR (CDCl₃, 400 MHz) δ 1.32 (t, J = 7.2 Hz, 3H, CH₃), 1.44 (s, 9H, *t*-Bu), 2.96-3.06 (m, 1H, H-4), 3.32 (t, J = 9.4 Hz, 1H, CH₂Cl), 3.41 (t, J = 10.9 Hz, 1H, H-

5), 3.71-3.79 (m, 1H, CH₂Cl), 3.84 (dd, *J* = 11.0, 4.5 Hz, 1H, H-5), 4.19-4.39 (m,

2H, CH₂O); ¹³C NMR (CDCl₃, 100 MHz): δ 14.0 (CH₃), 27.3 (CH₃), 41.2 (C-5), 47.1 (CH₂Cl), 49.0 (C-4), 56.0 (C), 63.4 (CH₂), 94.9 (C-3), 165.3 (C-2), 166.8 (CO). HRMS (ESI-TOF) *m/z*: calcd for C₁₂H₂₀Cl₂NO₃ [M+H]⁺ 296.0815; found 296.0816.

3,3-Dichloro-4-(chloromethyl)pyrrolidin-2-one (18e). IR (NaCl, neat): 3261,

 $\begin{array}{c} \text{CI} \\ \text{CI} \\$

1-Butyl-3,3-dichloro-4-(chloromethyl)pyrrolidin-2-one (18f). IR (NaCl, neat):



2960, 2932, 2872, 1727, 1483, 1447, 1431 cm⁻¹; ¹H NMR (CDCl₃, 400 MHz): δ 0.95 (t, *J* = 7.3 Hz, 3H, CH₃), 1.27-1.42 (m, 2H, CH₂), 1.49-1.63 (m, 2H, CH₂), 3.05-3.15 (m, 1H, H-4), 3.21-3.36 (m, 2H, CH₂Cl, CH₂N), 3.47 (dt, *J* = 13.7, 7.5 Hz, 1H, CH₂N), 3.60 (dd, *J*

= 10.1, 6.9 Hz, 1H, CH₂Cl), 3.68-3.79 (m, 1H, H-5), 4.01 (dd, J = 11.2, 4.2 Hz, 1H, zH-5); ¹³C NMR (CDCl₃, 100 MHz): δ 13.6 (CH₃), 19.8 (CH₂), 28.9 (CH₂), 41.2 (C-5), 43.7 (CH₂N), 47.8 (CH₂Cl), 51.7 (C-4), 83.8 (C-3), 165.9 (CO). HRMS (ESI-TOF) *m/z*: calcd for C₉H₁₅Cl₃NO [M+H]⁺ 258.0214; found 258.0214

3,3-Dichloro-4-(chloromethyl)-1-phenylpyrrolidin-2-one (18g). IR (NaCl,



neat): 1708, 1594, 1495, 1473, 1459, 1410 cm⁻¹; ¹H NMR (CDCl₃, 400 MHz): δ 3.20-3.31 (m, 1H, H-4), 3.74 (ddd, *J* = 10.0, 8.4, 0.5 Hz, 1H, H-5), 3.80-3.88 (m, 1H, CH₂Cl), 4.01-4.15 (m, 2H, CH₂Cl, H-5), 7.19-7.31 (m, 1H, ArH), 7.33-7.52 (m, 2H, ArH), 7.58-7.68 (m, 2H, ArH); ¹³C NMR (CDCl₃, 100 MHz): δ 41.0 (CH₂Cl), 49.1

(C-5), 51.1 (C-4), 84.0 (C-3), 120.2 (CHAr), 126.2 (CHAr), 129.2 (CHAr), 137.9 (CAr), 16.4 (CO). HRMS (ESI-TOF) *m/z*: calcd for C₁₁H₁₁Cl₃NO [M+H]⁺277.9901; found 277.9899.

1-Benzyl-3,3-dichloro-4-(chloromethyl)pyrrolidin-2-one (18h). IR (NaCl, neat): 3063, 3030, 2926, 2878, 1731, 1604, 1585, 1495 cm⁻¹; ¹H NMR (CDCl₃, 400 MHz): δ 3.01-3.16 (m, 2H, H-4, CH₂Cl), 3.40-3.53 (m, 1H, CH₂Cl), 3.63-3.73 (m, 1H, H-5), 3.95 (dd, *J* = 11.2, 3.72 Hz, 1H, H-5), 4.45 (d, *J* = 14.7 Hz, 1H, CH₂N), 4.63 (d, *J* = 14.7 Hz, 1H, CH₂N), 7.21-7.29 (m, 2H, ArH), 7.30-7.42 (m, 3H, ArH); ¹³C NMR (CDCl₃, 100 MHz): δ 41.0 (H-5), 47.2 (CH₂Cl), 47.9 (CH₂N), 51.6 (H-4), 83.7 (C-

3), 128.2 (CHAr), 128.3 (CHAr), 129.1 (CHAr), 134.5 (CAr), 166.0 (CO). HRMS

(ESI-TOF) *m/z*: calcd for C₁₂H₁₃Cl₃NO [M+H]⁺ 292.0057; found 292.0062.

1-AllyI-3,3-dichloro-4-(chloromethyl)pyrrolidin-2-one (18i). IR (NaCl, neat): $CI \rightarrow CI = 3085, 3015, 2961, 2923, 1731, 1644 \text{ cm}^{-1}; {}^{1}\text{H} \text{ NMR} (CDCl_3, 400)$ MHz): δ 3.06-3.16 (m, 1H, H-4), 3.23 (ddd, J = 10.2, 8.2, 0.5 Hz, 1H, CH₂Cl), 3.59 (ddd, J = 10.2, 7.0, 0.6 Hz, 1H, CH₂Cl), 3.74 (ddd, J = 11.3, 10.1, 0.6 Hz, 1H, H-5), 3.91-4.05 (m, 3H, H-5, CH₂N), 5.24-5.32 (m, 2H, CH₂), 5.66-5.81 (m, 1H, CH); {}^{1}\text{3}\text{C} \text{ NMR} (CDCl_3, 100 \text{ MHz}): δ 41.1 (C-5), 46.4 (CH₂N), 47.4 (CH₂Cl), 51.7 (C-4), 83.6 (C-3), 119.7 (CH₂), 130.5 (CH), 165.7 (CO). HRMS (ESI-TOF) *m/z*: calcd for C₈H₁₁Cl₃NO [M+H]⁺ 241.9901; found 241.9898.

1-(2-Bromoallyl)-3,3-dichloro-4-(chloromethyl)pyrrolidin-2-one (18j). IR (NaCl, neat): 2961, 2920, 2849, 1730, 1629 cm⁻¹; ¹H NMR (CDCl₃, 400 MHz) δ 3.09-3.22 (m, 1H, H-4), 3.31 (ddd, J = 10.1, 8.1, 0.7 Hz, 1H, CH₂Cl), 3.57-3.65 (m, 1H, CH₂Cl), 3.71-3.81 (m, 1H, H-5), 4.02 (ddd, J = 11.3, 4.3, 0.74 Hz, 1H, H-5), 4.17 (d, J = 15.4 Hz, 1H, NCH₂), 4.32 (d, J = 15.4 Hz, 1H, NCH₂), 5.67-5.73 (d, J = 2.24 Hz, 1H, CH₂), 5.86-5.89 (d, J = 2.00 Hz, 1H, CH₂); ¹³C NMR (CDCl₃, 100 MHz): δ 40.9 (C-5), 47.3 (CH₂Cl), 51.6 (NCH₂), 51.8 (C-4), 83.2 (C-3), 121.0 (CH₂), 125.9 (CBr), 165.9 (CO). HRMS (ESI-TOF) *m/z*: calcd for C₈H₁₀BrNO [M+H]⁺ 319.9006; found 319.9003.

(RS)-1-(tert-Butyl)-3,3-dichloro-4-((SR)-1-chloroethyl)pyrrolidin-2-one (18k).

IR (NaCl, neat): 3131, 2979, 2935, 2913, 2873, 1724 cm⁻¹; ¹H NMR
(CDCl₃, 400 MHz): δ 1.44 (s, 9H, *t*-Bu), 1.84 (d, J = 6.5 Hz, 3H, CH₃), 2.81 (ddd, J = 9.9, 9, 7.2 Hz, 1H, H-4), 3.17 (dd, J = 10.3, 9.0, 1H, CH₂), 3.74 (dd, J = 10.3, 7.2 Hz, 1H, CH₂), 4.33 (dq, J = 9.9, 6.5 Hz, 1H, CHCl); ¹³C NMR (CDCl₃, 100 MHz): δ 23.8 (CH₃), 27.2

(CH₃), 46.9 (C-5), 55.5 (C-4), 55.6 (C), 56.9 (CCl), 84.8 (C-3), 165.7 (CO). HRMS (ESI-TOF) *m/z*: calcd for C₁₀H₁₇Cl₃NO [M+H]⁺ 272.0370; found 272.0363

(RS)-1-(tert-Butyl)-3,3-dichloro-4-((RS)-1-chloroethyl)pyrrolidin-2-one (19k).

CI IR (NaCl, neat): 3001, 2981, 2934, 2914, 1715, 1670 cm⁻¹; ¹H NMR (CDCl₃, 400 MHz): δ 1.44 (s, 9H, *t*-Bu), 1.53 (d, *J* = 6.7 Hz, 3H, CH₃), 2.94-3.01 (m, 1H, H-4), 3.18-3.24 (m, 1H, CH₂), 3.52-3.57 (m, 1H, CH₂), 4.39-4.48 (m, 1H, CHCl); ¹³C NMR (CDCl₃, 100 MHz): δ 21.9 (CH₃), 27.2 (CH₃), 44.4 (C-5), 54.3 (CCl), 54.7 (C-4), 55.6 (C), 85.4 (C-3), 165.9 (CO). HRMS (ESI-TOF) *m/z*: calcd for C₁₀H₁₇Cl₃NO [M+H]⁺ 272.0370; found 272.0366.

1-(*tert***-Butyl)-3,3-dichloro-4-(chloromethyl)piperidin-2-one (18l).** IR (NaCl, neat): 3008, 2967, 2931, 2870, 1672, 1631 cm⁻¹; ¹H NMR (CDCl₃, 400 MHz) δ 1.46 (s, 9H, *t*-Bu), 1.79-1.96 (m, 1H, H-5), 2.33-2.43 (m, 1H, H-5), 2.66 (dddd, J = 12.6, 10.1, 3.5, 2.4 Hz, 1H, H-4), 3.31 (dddd, J = 13.1, 11.9, 4.8, 1.1 Hz, 1H, H-6), 3.50-3.61 (m, 2H, H-6, CH₂Cl), 4.13 (dd, J = 11.1, 3.2 Hz, 1H, CH₂Cl); ¹³C NMR (CDCl₃, 100 MHz): δ 23.4 (C-5), 27.6 (CH₃), 43.0 (C-6), 44.0 (CH₂Cl), 51.8 (C-4), 59.0 (C), 87.2 (C-3), 163.2 (CO). HRMS (ESI-TOF) *m/z*: calcd for C₁₀H₁₇Cl₃NO [M+H]⁺ 272.0370; found 272.0363.

(4RS,6SR)-1-Benzyl-3,3-dichloro-4-(chloromethyl)-6-phenylpiperidin-2-one

(18m). IR (NaCl, neat): 3086, 3063, 3029, 2966, 2931, 1681, Bn = Cl Cl Cl Cl Cl Cl Cl Cl Cl 2H, H-5), 2.74-2.83 (m, 1H, H-4), 3.44-3.59 (m, 2H, CH₂Cl, CH_2Bn), 4.04 (dd, J = 11.1, 2.97 Hz, 1H, CH₂Cl), 4.63 (dd, J = 5.5, 2.6 Hz, 1H, H-6), 5.59 (d, J = 14.8 Hz, 1H, CH₂Bn), 7.06-7.23 (m, 5H, ArH), 7.25-7.49 (m, 5h, ArH); ¹³C NMR (CDCl₃, 100 MHz): δ 31.0 (C-5), 43.5 (CH₂Cl), 47.2 (C-4), 49.7 (CH₂Bn), 58.1 (C-6), 85.4 (C-3), 126.0 (ArCH), 127.9, 128.0 (ArCH), 128.3, 128.9 (ArCH), 129.4 (ArCH), 135.8 (ArC), 138.6 (ArC), 164.2 (CO). HRMS (ESI-TOF) *m/z*: calcd for C₁₉H₁₉Cl₃NO [M+H]⁺ 382.0527; found 382.0515.

(4RS,6RS)-1-Benzyl-3,3-dichloro-4-(chloromethyl)-6-phenylpiperidin-2-one

(19m). IR (NaCl, neat): 3086, 3063, 3029, 2926, 2849, 1730, Bn_{V} (19m). IR (NaCl, neat): 3086, 3063, 3029, 2926, 2849, 1730, $1676, 1603 \text{ cm}^{-1}$; ¹H NMR (CDCl₃, 400 MHz) δ 2.0-2.13 (m, 1H, H-4), 3.47-3.6 (m, 2H, CH₂Cl, CH₂Bn), 4.2 (dd, *J* = 11.1, 2.9 Hz, 1H, CH₂Cl), 4.33 (dd, *J* = 11.6, 5.8 Hz, 1H, H-6), 5.35 (d, *J* = 14.6 Hz, 1H, CH₂Bn), 6.94-7.22 (m, 5H, ArH), 7.22-7.48 (m, 5H, ArH); ¹³C NMR (CDCl₃, 100 MHz): δ 32.8 (C-5), 43.8 (CH₂Cl), 48.3 (CH₂Bn), 50.5 (C-4), 60.1 (C-6), 85.3 (C-3), 127.8 (ArCH), 128.4 (ArCH), 128.8 (ArCH), 129.2 (ArCH), 135.8 (ArC), 139.8 (ArC), 164.1 (CO). HRMS (ESI-TOF) *m/z*: calcd for C₁₉H₁₉Cl₃NO [M+H]⁺ 382.0527; found 382.0514.

(3aRS,4RS,7aRS)-1-Benzyl-3,3,4-trichlorohexahydro-2H-indole-2,5(3H)-

dione (18n). IR (NaCl, neat): 3064, 3029, 2949, 2923, 1737, 1731, 1715 cm⁻¹; ¹H NMR (CDCl₃, 400 MHz): δ 2.00-2.15 (m, 1H, H-7), 2.18-2.28 (m, 1H, H-7), 2.46 (dt, J = 17.2, 4.3 Hz, 1H, H-6), 2.70 (ddd, J = 17.0, 13.0, 6.0 Hz, 1H, H-6), 3.44 (dd, J = 8.2, 5.0 Hz, 1H,

CH), 3.82 (m, J = 11.4, 8.2, 5.1 Hz, 1H, NCH), 4.26 (d, J = 15.0 Hz, 1H, CH₂Ar), 4.9 (d, J = 4.6, 1H, CICH), 4.94 (s, 1H, CH₂Ar), 7.24-7.30 (m, 2H, ArCH), 7.33-7.43 (m, 3H, ArCH); ¹³C NMR (CDCl₃, 100 MHz): δ 24.6 (C-7), 32.8 (C-6), 46.2 (CH₂Ar), 52.3 (NCH), 57.2 (CICH), 57.3 (CH), 82.6 (CCl₂), 128.1(ArCH), 128.6(ArCH), 129.2 (ArCH), 134.4 (C), 165.4 (CO), 199.2 (CO). HRMS (ESI-TOF) *m/z*: calcd for C₁₅H₁₅Cl₃NO₂ [M+H]⁺ 346.0163; found 346.0158.

(3RS,3aRS,4SR,7aRS)-1-(tert-Butyl)-3,4-dichlorooctahydro-2H-indol-2-one



Bn 、

(180). IR (NaCl, neat): 2949, 2869, 1704, 1480, 1459, 1393 cm⁻¹; ¹H NMR (CDCl₃, 400 MHz): δ 1.07-1.26 (m, 2H, CH₂), 1.44 (s, 9H, *t*-Bu), 1.56-1.68 (m, 1H, CH₂), 1.76-1.89 (m, 1H, CH₂), 1.94-2.07 (m, 1H, CH₂), 2.15-2.26 (m, 1H, CH₂), 2.71 (dd, *J* =12.3, 6.6 Hz, 1H, H-3a), 3.88-3.99 (m, 1H, H-4), 4.27 (d, *J* =12.2 Hz, 1H, H-7a),

4.65 (bs, 1H, H-3); ¹³C NMR (CDCl₃, 100 MHz): δ 17.1 (CH₂), 28.0 (CH₃), 28.5 (CH₂), 31.7 (CH₂), 51.7 (C-3a), 52.6 (C-4), 55.2 (C), 56.7 (C-3), 58.2 (C-7a), 167.8

(CO). HRMS (ESI-TOF) m/z: calcd for C₁₂H₂₀Cl₂NO [M+H]⁺ 264.0916; found 264.0916.

(1RS,5RS,6RS)-2-benzyl-4,4,6-trichloro-2-azabicyclo[3.3.1]nonan-3-one



The spectroscopic data of this product matches the ones reported previously.⁷

⁷ Diaba, F.; Martínez-Laporta, A.; Bonjoch, J.; Pereira, A.; Muñoz-Molina, J. M.; Pérez, P. J.; Belderrain, T. R.. *Chem. Commun.* **2012**, *48*, 8799–8801.












130 120 f1 (ppm) . 30

VNMRS400A_04022021_as416-21y23-H1 VNMRS400F / Num.Inv. 205984 cdcl3 / Temp: 25C / N.Reg: XXXXXXXXX Usuari: san / Mostra: as416-21y23 Nom: ALEXANDRA SANDOR Data: 04/02/21 / Ope.: A.SANDOR





180 170 160 150 140 130 120 110 100 90 80 70 60 50 40 f1 (ppm)









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VNMRS400A_18122020_as404-2-20-H1 VNMRS400F / Num.Inv. 205984 cdcl3 / Temp: 25C / N.Reg: XXXXXXXXX Usuari: san / Mostra: as404-2-20 Nom: ALEXANDRA SANDOR Data: 18/12/20 / Ope.: A.SANDOR









VNMRS400A_10122020_as397-2-40-H1 VNMRS400F / Num.Inv. 205984 cdcl3 / Temp: 25C / N.Reg: XXXXXXXXX Usuari: san / Mostra: as397-2-40 Nom: ALEXANDRA SANDOR Data: 10/12/20 / Ope.: A.SANDOR



С









120 110 f1 (ppm)





120 110 f1 (ppm)



100 90 f1 (ppm) . 160 . 150 . 120 . 50