ELSEVIER

Contents lists available at ScienceDirect

Journal of South American Earth Sciences

journal homepage: www.elsevier.com/locate/jsames





A new fossil inchworm moth discovered in Miocene Dominican amber (Lepidoptera: Geometridae)

Victor Sarto i Monteys ^{a,b,*}, Axel Hausmann ^c, Mónica M. Solórzano-Kraemer ^d, Jörg U. Hammel ^e, Joaquín Baixeras ^f, Xavier Delclòs ^{g,h}, Enrique Peñalver ⁱ

- a Institut de Ciència i Tecnologia Ambientals (ICTA), Entomology, Plants and Health, Edifici Z, C/ de les columnes s/n. Campus de Bellaterra, Universitat Autònoma de Barcelona, Bellaterra, Spain
- ^b Servei de Sanitat Vegetal, DARP, Generalitat de Catalunya, Barcelona, Spain
- ^c SNSB Zoologische Staatssammlung München, Münchhausenstraße 21, D-81247 Munich, Germany
- d Senckenberg Forschungsinstitut und Naturmuseum, Paleaontologie und Historische Geologie, Senckenberganlage 25, 60325 Frankfurt am Main, Germany
- e Institute of Materials Physics, Helmholtz-Zentrum Hereon, Outstation at DESY, Max-Planck-Str. 1, D-21502 Geesthacht, Germany
- f Institut Cavanilles de Biodiversitat i Biologia Evolutiva, Universitat de València, C/ Catedràtic José Beltran 2, 46980 Paterna (València), Spain
- g Department Dinàmica de la Terra i de l'Oceà, Facultat de Ciències de La Terra, Universitat de Barcelona (UB), Martí i Franquès s/n, 08028 Barcelona, Spain
- ^h Institut de Recerca de la Biodiversitat (IRBio), Universitat de Barcelona (UB), Barcelona, Spain
- i Instituto Geológico y Minero de España (IGME), CSIC, C/ Cirilo Amorós 42, 46004 València, Spain

ARTICLE INFO

Keywords: Lepidoptera Desmobathrinae New species Amber Miocene

ABSTRACT

We report a fossil geometrid moth, a male, virtually complete, preserved in a clear piece of Miocene Dominican amber dating from 19 to 16 Mya. Fore- and hindwings appear partially overlapped, and all body characters are visible externally in dorsal and ventral views, including the outer surface of the valvae of the genitalia. The scale pattern on the wing membrane is preserved, whereas the wing color pattern is not. It belongs to the genus *Dolichoneura* (Geometridae: Desmobathrinae) and is named *Dolichoneura jorelisae* Sarto i Monteys, Hausmann, Baixeras and Peñalver sp. n., based on wing features. Because of the poor fossil record of lepidopterans, both in amber and compression rocks, the description of the available well-preserved specimens is of considerable interest for phylogenetic studies. Furthermore, it could also serve for calibrating molecular clocks and for paleobiogeographic inferences.

1. Introduction

So far, the oldest known fossil lepidopteran, *Archaeolepis mane* Whalley (1985), is from the Lower Jurassic (Sinemurian, *ca.* 190 Mya) of England (Whalley, 1985). Yet, phylogenomic studies by Kawahara et al. (2019) suggest that the most recent common ancestor of crown Lepidoptera appeared perhaps significantly earlier, in the Late Carboniferous, *ca.* 300 Mya, although this estimation might be biased (this age is very close to the initial appearance of the Holometabola) and should be confirmed by further studies.

Although Lepidoptera is one of the megadiverse orders of insects and one of the most abundant around different terrestrial ecosystems, they are, due to taphonomical processes (Martínez-Delclòs and Martinell, 1993), among the rarest findings amid insect orders in deep time, mostly occurring in ambers. Yet, imagoes of macrolepidopterans are rare as

inclusions in ambers, principally because they live and eat far from the trunk or big branches of resiniferous trees, from where the resin would mostly ooze (Solórzano Kraemer et al., 2018). Actually, macrolepidopteran wings can get caught very easily in resin, so their size is not that important. If they get trapped in resin emissions on a tree trunk or branch, the ants will have easy access and will eat them before the next flow arrives covering the carcasses (MMS-K, XD, EP, personal observation, Madagascar). All this contributes to the scarcity of their inclusions in ambers.

Geometridae, whose age was estimated at about 54 Mya, in the Eocene (credibility interval of 62 to 48 Mya corresponding to Middle Paleocene–Early Eocene) (Yamamoto and Sota, 2007; Wahlberg et al., 2013), stand as the second most species-rich family of Lepidoptera, comprising *ca.* 24,000 described species (van Nieukerken et al., 2011; Murillo-Ramos et al., 2019). They belong to the superfamily

E-mail address: victor.sarto@uab.cat (V. Sarto i Monteys).

https://doi.org/10.1016/j.jsames.2022.104055

Received 8 August 2022; Accepted 27 September 2022 Available online 30 September 2022

^{*} Corresponding author. Institut de Ciència i Tecnologia Ambientals (ICTA), Entomology, Plants and Health, Edifici Z, C/ de les columnes s/n. Campus de Bellaterra, Universitat Autònoma de Barcelona, Bellaterra, Spain.





Fig. 1. Habitus of †*Dolichoneura jorelisae* sp. n. (Geometridae: Desmobathrinae), holotype. (A) Dominican amber piece with the geometrid specimen, ventral view. (B) Virtual representation (SR μ CT) of the geometrid specimen in ventral view, with the hind wing venation enhanced.

Geometroidea, which traces back to 83 Mya (Wahlberg et al., 2013). Geometrids are characterized by the presence of paired tympanal organs placed at the base of the abdomen; they occur throughout the family, in both sexes, although they are secondarily reduced or lost in a very few flightless females (Cook and Scoble, 1992). The prolegs of their larvae are usually reduced to two pairs causing the larvae to move by 'looping' (Minet and Scoble, 1999), that is where the names "inchworm", "loopers" and "Geometridae", as for 'earth-measuring', come from. A comprehensive molecular study by Murillo-Ramos et al. (2019, 2021) proposed recognizing nine mostly monophyletic geometrid subfamilies: Sterrhinae, Larentiinae, Archiearinae, Desmobathrinae, Orthostixinae (validity controversial), Epidesmiinae, Oenochrominae, Geometrinae, and Ennominae.

Despite of being one of the most specious family groups of Lepidoptera, the fossil record of Geometridae is comparatively very scarce, with only 19 findings formally reported worldwide, most of them of difficult systematic assignment. The affiliation of two of them is disputed by Sohn et al. (2012). They were all listed and reviewed by Zhang et al. (2020), who noted that *Eogeometer vadens* Fischer, Michalski and Hausmann, 2020 (Bartonian–Priabonian, Baltic amber), *Geometridites larentiiformis* Jarzembowski, 1980 (Upper Priabonian, Bembridge Marls, of the Isle of Wight), and *Hydriomena protrita* Cockerell, 1922 (Priabonian, Florissant Fossil Beds), all from the Late Eocene, were believed to be the earliest representatives of Geometridae.

To date, 31 fossil specimens within seven superfamilies of ditrysian Lepidoptera had been reported from amber collected in the Dominican Republic (Grimaldi and Engel, 2005; Sohn et al., 2012; Zhang et al., 2020), with an estimated age of *ca.* 19–16 Mya (Iturralde-Vinent and MacPhee, 2019). Among the known fossil geometrids worldwide, four

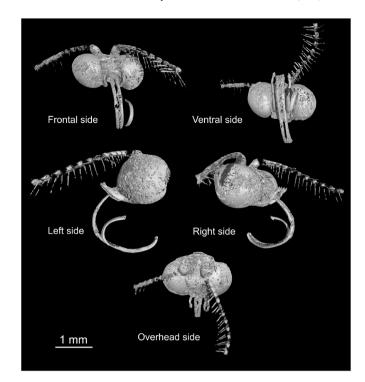


Fig. 2. Virtual representation (SR μ CT) in different views of the head of †*Dolichoneura jorelisae* sp. n. (Geometridae: Desmobathrinae), holotype. Both antennae are distally incomplete due to the limits of the scan; in the amber piece one distal end was kept within it (see Fig. 3B).

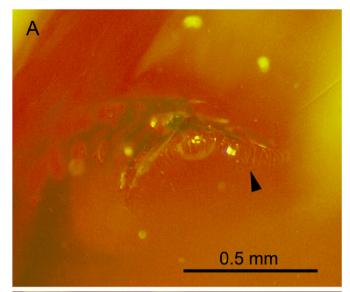
are from amber collected in this area: (1) an adult assigned to Ennominae, plus (2) a caterpillar and (3) another adult, both unassigned, figured but undescribed (Grimaldi and Engel, 2005), and (4) an adult assigned to Ennominae, described as *Miogeometrida chunjenshihi* Zhang et al., 2020 (Zhang et al., 2020). Here we report a fifth geometrid moth included in Dominican amber.

2. Materials and methods

The type specimen in amber described herein is housed at the Museo Nacional de Historia Natural "Prof. Eugenio de Jesús Marcano", Santo Domingo, Dominican Republic, with the accession number MNHNSD FOS 16.01. The piece that includes the moth originates from the eastern amber mines of Hato Mayor-El Valle Region, Dominican Republic (Eastern district in Iturralde-Vinent and MacPhee, 2019); it is oval, weighs 3.58 g, with dimensions 31 mm by 20 mm, and a height of 10 mm–2 mm since it is flattening from one side to the other, no internal cracks are visible and no syninclusions are observed. The moth is situated centrally within it, with its head contacting one outer border (Fig. 1). The piece was delivered to us by Mr. Jorge Caridad, from his private collection in the Amber World Museum. Mr. Jorge Caridad is known to most authors, and is an established and trusted source.

2.1. Spectroscopy

The amber piece was kept at constant temperature in plastic clip bags within metal boxes during the study to exclude oxygen and light. To determine its origin and establish a molecular fingerprint, we carried out a Fourier Transform Infrared Spectroscopy (FTIR) (see results below). To ensure the piece containing the moth was amber, and not copal or Defaunation resin (*sensu* Solórzano-Kraemer et al., 2020), its FTIR spectrum was compared to that of two pieces of amber found by us in the field, one in the Northern district (Palo Quemado) and another in the Eastern district (El Valle - Mina Siete Cañadas), the latter belonging to



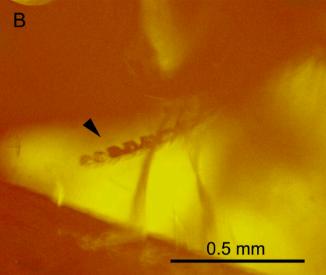


Fig. 3. Cilia on antennal flagellum of †*Dolichoneura jorelisae* sp. n. (Geometridae: Desmobathrinae), holotype. (A) Flagellum densely setose ventrally (arrowhead), seen on the left antenna, sectioned in the amber surface. (B) Cilia shorten progressively as they approach the distal end, where the antennae look serrate (arrowhead). Both images are Z-stacked.

the same location from where the piece including the moth was collected.

A tiny bit was removed from the amber sample for FTIR analysis at one edge. The latter was conducted using an IR PerkinElmer Frontier spectrometer that utilizes a diamond ATR system with a temperature stabilized DTGS detector and a CsI beam splitter at the Molecular Spectrometry Unit of the CCiTUB (Centres Científics i Tecnològics of the Universitat de Barcelona). To avoid breaking the piece of amber containing the moth, the FTIR was performed using a Thermo SCIENTIFIC NICOLET iN10 MX, reflexion, detector MCT, overture 100μ , accumulation 64 scans, spectral resolution 4 cm $^{-1}$, Kramers-Kronic.

2.2. Stereomicroscopy and photography

The specimen was examined and drawn by using an optical microscope Olympus BH-2 equipped with an Olympus camera lucida, and photomicrographs were taken using a digital camera (Leica M80 Camera) attached to a Leica IC90 E stereomicroscope, at the laboratory of the Museu Valencià d'Història Natural (Alginet, València). These devices

used cool white LED illuminators. Images were prepared for illustration using Adobe Photoshop CS2, and some of them were z-stacked. The body length was measured from the frons to the end of the abdomen.

2.3. Synchrotron radiation micro-computed tomography (SRµ-CT)

In order to observe morphological details which could not be properly seen under a stereomicroscope, the inclusion was additionally studied by synchrotron radiation based x-ray micro tomography (SRµCT). Imaging was performed at the Imaging Beamline - IBL P05 -PETRA III at Deutsches Elektronen Synchrotron (DESY) in Hamburg operated by the Helmholtz-Zentrum Hereon (Greving et al., 2014; Wilde et al., 2016). Due to the size of the specimen imaging was done in an extended field of view (eVoF) mode (\sim 12 mm \times 5 mm). 12,602 projections have been recorded at 6001 angular steps equally spaced between 0 and π . Projections were recorded at a photon energy of 18 keV and a sample to detector distance of 30 mm using a custum build CMOS detector (Lytaev et al., 2014). Tomographic reconstruction was done with a beamline specific stitching and processing workflow (Moosmann et al., 2014; Beckmann pers. comm., June 2019) implemented in Matlab (Math Works) and IDL (Harris Inc) using the filtered back projection algorithm implemented in the Astra Toolbox (van Aarle et al., 2015, 2016; Palenstijn et al., 2011). Raw projections have been binned two times for processing, resulting in an effective pixel size of 1.28 μm in the reconstructed tomographic volume.

2.4. Systematics

Family-level classification follows van Nieukerken et al. (2011). Terminology of veins follows Hausmann (2001).

3. Results

3.1. Systematic paleontology

Order Lepidoptera Linnaeus, 1758
Suborder Glossata Fabricius, 1775
Clade Ditrysia Börner, 1925.
Superfamily Geometroidea Leach, 1815
Family Geometridae Leach, 1815
Subfamily Desmobathrinae Meyrick, 1886
Genus Dolichoneura Warren, 1894
†Dolichoneura jorelisae Sarto i Monteys, Hausmann, Baixeras and Peñalver, sp. n.
Figs. 1–7.

LSID:urn:lsid:zoobank.org:act:DC6F5F7F-2CAA-4955-B78E-26CBEEEB59BB

3.2. Diagnosis (male)

The species can be distinguished from other species of the genus by the following combination of wing characters: pointed forewing apexes, relatively straight forewing termen, a well-developed double areole and absence of anastomosis between Sc $+\ R_1$ and Rs in the forewing, the size of the forewing discal cell relatively longer than in other species of the genus, the point where M_2 arises from the forewing discal cell relatively closer to M_1 than to M_3 in comparison to other species of the genus, an evenly rounded hindwing termen, and a rather weak but tubular M_2 vein in the hindwing.

Holotype: Specimen code MNHNSD FOS 16.01, male (Figs. 1-7).

Repository: Museo Nacional de Historia Natural "Prof. Eugenio de Jesús Marcano", Santo Domingo, Dominican Republic.

Etymology: The specific name is dedicated to Ms. Jorelis Caridad, daugther of Mr. Jorge Caridad, who entrusted us with this piece. She is well-known for her dedication to preserving and promoting the study of Dominican amber.

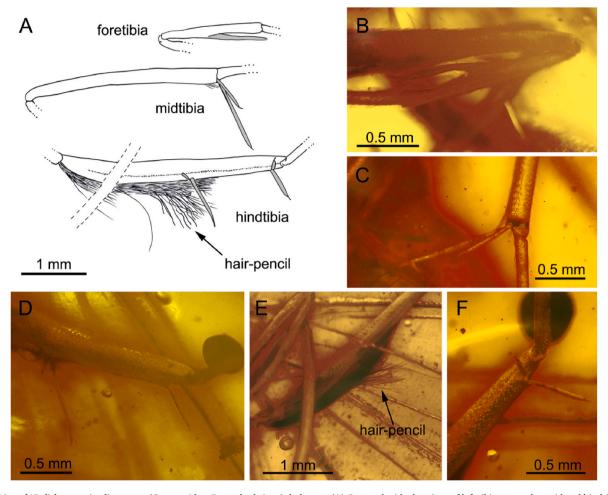


Fig. 4. Tibiae of †Dolichoneura jorelisae sp. n. (Geometridae: Desmobathrinae), holotype. (A) Camera lucida drawings of left tibiae; note that mid and hindtibiae show spurs not present in foretibia. (B) Left foretibia. (C) Left midtibial spurs. (D) Left hindtibia. (E) Left hindtibia: detail of hair-pencil; also two unequal spurs projecting from its central part. (F) Left hindtibia: two unequal spurs projecting from its distal part. Images D and F are Z-stacked.

Locus typicus: Amber mine of Hato Mayor-El Valle Region, Dominican Republic.

Stratum typicum: The piece was found in stratigraphic levels of the Yanigua Formation, in the boundary between Early–Middle Miocene, 17–16 Mya (Iturralde-Vinent and MacPhee, 2019).

3.3. Description. Male

Head: Vertex and frons scaled. Eyes well developed. Antennae dentate, scaled dorsally, densely ciliate-setose ventrally, length of cilia at basal part of antennae *ca.* three times the width of flagellum (Fig. 2); cilia shorten progressively as they approach the distal end, where the antennae look serrate (Fig. 3B). Ocelli not visible. Chaetosemata present, paired. Labial palpi upturned, not extending dorsally beyond frons, first and second segments about the same length, the second being slender, third segment small, about one fourth the length of the first two segments. Maxillary palpi not visible. Haustellum well developed, *ca.* 4.5 mm in length, coiled, unscaled, with distal part broken and kept next to the main part (Fig. 2).

Thorax: Scaled. Legs elongate; foretibia without tibial spurs, but showing a conspicuous single process arising from its centre; spur formula 0-2-4, pairs of spurs unequal; hindtibia with well developed hairpencil, partially retractable in a groove (Fig. 4).

Wings. Forewing length: 15.7 mm, with pointed apexes; no trace of wing color pattern (Fig. 5). Costa slightly convex at base, central portion rather straight, distal part convexly curved from the point where R₂ meets the costa to pointed apex, termen straight, tornus broadly curved;

forewing venation fairly complete with characteristic double areole involving distal part of Sc, basal half of R_1 and base of Rs, R_1 from the areole, R_2 and $R_3 + R_4$ connate in the areole, R_4 to costa, R_5 from areole to termen; discal cell 0.4 the length of the wing, discocellular vein complete, $M_1,\,M_2$ and M_3 approximately equidistant at base; CuA_1 closer at base to M_3 than to CuA_2 ; hindwing venation unmodified, $\text{Sc} + R_1$ and Rs unanastomosed, $\text{Sc} + R_1$ rather straight to apex, Rs sinuous on discal cell, $M_1,\,M_2,\,M_3$ rather equidistant on the discal cell, M_2 tubular although less developed than M_1 and M_3 (Fig. 6), base of CuA_1 closer to M_3 than to CuA_2 .

Abdomen: Long, slender, covered by appressed scales. Male genitalia with valvae closed, well delimited externally but structural details poorly visible, uncus apparently robust, prominent, valva elongate with rounded edges (Fig. 7).

3.4. Infrared absorption spectrum of amber piece studied

The infrared absorption spectrum of the amber piece that includes the moth is shown in Fig. 8 (in red), where the basic skeleton of amber is an aliphatic structure. It is dominated by an asymmetric telescopic vibration of ν (CH2) located at 2928 cm⁻¹ (small C–H stretching band), symmetric telescopic vibration of ν (CH2) located at 2865 cm⁻¹, transformative vibration of ν (CH2, CH3) located at 1465 cm⁻¹ (intense C–H band), the symmetric transformative vibration of ν (CH2, CH3) located at 1380 cm⁻¹, and telescopic vibration of ν (C=O) located at 1725 cm⁻¹ (intense carbonyl band), and telescopic vibration of ν (C–O) located at 1245 cm⁻¹, 1135 cm⁻¹, and 1030 cm⁻¹. Hydroxyl bands near 3500

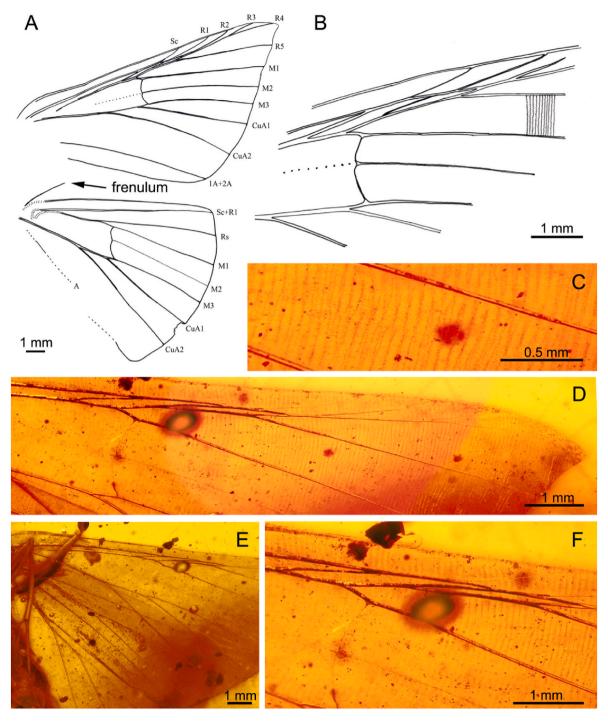


Fig. 5. Wing venation and preserved scale pattern of † Dolichoneura jorelisae sp. n. (Geometridae: Desmobathrinae), holotype. (A) Camera lucida drawing of fore- and hindwing. Note the absence of anastomosis between $Sc + R_1$ and Rs on the hindwing. (B) Same, showing detail of anterior venation and scale pattern of forewing. (C) Detail of scale pattern preserved on forewing. (D) Costal part of left forewing (underside). (E) Detail of left fore- and hindwing (underside) partially overlapped. (F) Detail of the well-developed double areole on forewing. Images D and F are Z-stacked.

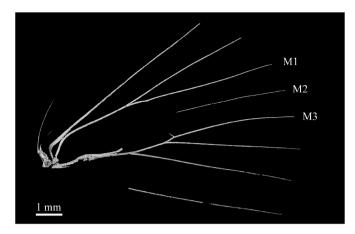


Fig. 6. Virtual representation (SR μ CT) showing hindwing venation of *Doli-choneura jorelisae* sp. n. (Geometridae: Desmobathrinae), holotype. Note that M_2 is tubular although less developed than M_1 and M_3 .

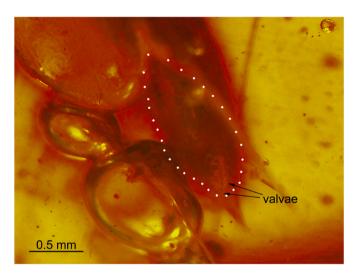


Fig. 7. Micrograph showing the male genitalia, delimited by white dots (ventral view), of $\dagger Dolichoneura\ jorelisae\$ sp. n. (Geometridae: Desmobathrinae), holotype. Z-stacked image.

cm⁻¹ are also present. The spectrum of the piece that contains the geometrid is very similar to those from two geological Dominican amber localities. These are El Valle-Siete Cañadas (2018 own extraction in the Eastern district of the island, in <u>Iturralde-Vinent and MacPhee</u>, 2019, in blue) and Palo Quemado (2019 own extraction in the Northern district, in green), their main difference being the presence of a small band near 1200 cm⁻¹ in the latter.

4. Discussion

As usual in the study of fossil Lepidoptera, characters of the genitalia, which are of most interest for a suitable identification, are not accessible despite 3D preservation in amber and having explored the structure through SR μ -CT.

The wings show pointed forewing apex, relatively straight forewing termen and an evenly rounded hindwing termen, rather unusual wing shape characters that deserve some attention. Except for minor damage

in the posterodistal margin of the left hindwing, the wings are quite well preserved. We may conjecture that the moth became trapped in resin shortly after emerging and was rapidly and entirely covered by subsequent resin flows. Yet, their venation, usually of supraspecific value, is reasonably well marked and allows, in some detail, a taxonomical assignment.

The most striking venation characters of this fossil moth are the presence of a well-developed double areole in the forewing, and the absence of an astomosis between $\mathrm{Sc} + \mathrm{R}_1$ and Rs , as well as a rather weak but tubular M2 vein in the hindwing. A forewing areole is common in some subfamilies of Geometridae, mostly Orthostixinae, Desmobathrinae, Larentiinae and Sterrhinae (Hausmann, 2001). The areole is formed by a variable degree of anastomosis of R1 and Rs veins, occasionally involving Sc. In this fossil, the areole is clearly affecting distally Sc as well as basally R₁ and Rs veins. Although Sc and R veins meet the wing edge separately, all of them arise from the areole. In addition, the areole is subdivided by a cross vein between basal Rs and R₁ giving rise to two areoles. The presence of a tubular M2 in the hindwing is a generalized feature in the above-mentioned subfamilies, but the absence of anastomosis between Sc + R₁ and Rs rules out our fossil moth from being a Larentiinae member. Holloway (1996) included representatives of slender-bodied moths with long and narrow appendages in the subfamily Desmobathrinae. Pointed forewing apex, among other wing shape traits, as present in our fossil, are common among representatives of this subfamily. Examination of this subfamily representatives, and especially of the genus Dolichoneura Warren, 1894 (e.g., D. oxypteraria (Guenée, 1858)), revealed striking similarities in venation and wing shape with our fossil. Indeed, we compared the wing venation of the type species of the genus Dolichoneura, D. oxypteraria, with that of our fossil. Minor differences are found in fore- and hindwing venations. These are (1) the relative size of the forewing discal cell (being longer and narrower in D. oxypteraria vs. shorter and broader in D. jorelisae), (2) the relative size of the hindwing discal cell (being shorter and broader in D. oxypteraria vs. longer and somewhat narrower in D. jorelisae), and (3) the point (in both, fore- and hindwings) where M2 arises from the discal cell (relatively closer to M_1 than to M_3 in D. oxypteraria vs. equidistant in D. jorelisae) (Fig. 9). There is also a coincidence between D. jorelisae and other Dolichoneura spp. concerning the spur number in legs and antennal features (dentate flagellum, densely ciliate-setose in males, Figs. 2 and

Although the monophyly of Desmobathrinae has been challenged by recent multigene analysis (Murillo-Ramos et al., 2019, 2021), the phylogenetic position of *Dolichoneura* remains unquestionable. The similarity between our fossil and the extant representatives of the genus *Dolichoneura* is so high that the description of a new genus does not seem appropriate. Actually, taking into account the relatively little antiquity of Dominican amber (19–16 Mya), there is no surprise that arthropods preserved in it have been assigned to extant genera several times and sometimes even to living species (e.g., Arillo and Ortuño, 2005; Hoernschemeyer et al., 2010). Based on this evidence, the former paleoenvironment is considered to be not so different from a typical extant lowland Neotropical rainforest (Grimaldi, 1996). Therefore, we conclude that, based on wing shape and venation, our amber geometrid is congeneric with *Dolichoneura*.

According to Iturralde-vinent and MacPhee (2019) the main amberiferous deposits in the Dominican Republic were formed in a single sedimentary basin in the boundary between Early and Middle Miocene, 19–16 Mya, due to accumulation of abundant resin produced by an extinct legume tree, *Hymenaea protera* Poinar, 1991 (Leguminosae). The genus *Hymenaea* consists today of fourteen extant species, all but one native to the tropics of the Americas, with one additional species

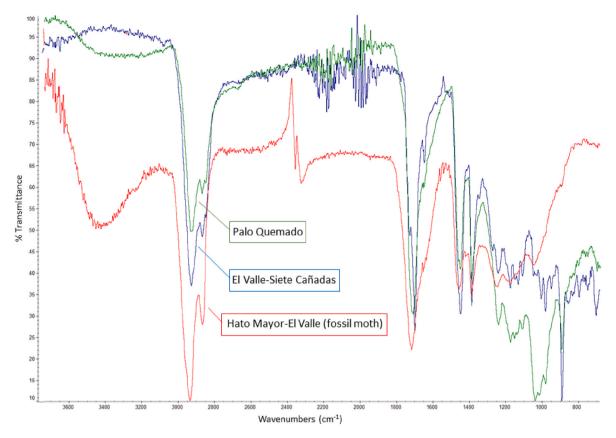


Fig. 8. Infrared absorption spectrum of the amber piece containing the moth (red line) from Hato Mayor-El Valle and spectra of two geological samples from El Valle-Siete Cañadas (blue line) and Palo Quemado (green line), for comparison.

(Hymenaea verrucosa Gaertner, 1791) on the east coast of Africa and Madagascar, considered sister to all other Neotropical species (Pinto et al., 2018). Although, so far, fossils of Dolichoneura or Desmobathrinae have not been reported anywhere from the Miocene, our finding of †Dolichoneura jorelisae sp. n. evidences that they were already present at this time in Central America.

The genus Dolichoneura, in which the fossil herein has been classified, includes eleven extant species distributed from Central to South America (between Costa Rica and Argentina) (Scoble and Hausmann, 2007; GBIF Secretariat, 2021). The sister genus Pycnoneura Warren, 1894 includes ten species exclusively distributed in South America (Scoble and Hausmann, 2007). Prout (1932) stated that both genera might have to be merged into one. The biology of Dolichoneura is poorly known; all species are nocturnal and have been collected in well sunny slope forest and ridge forest in the lowland rainforest of Costa Rica, where the resiniferous legume trees Hymenaea courbaril Linnaeus, 1753 and H. osanigraseminae Aguilar Fernández, Poveda Álvarez and Santamaría-Aguilar, 2018 live today, but not in creek forest (Rabl, 2016). Two larvae of an unidentified Dolichoneura species were collected by canopy fogging a Pouteria caimito (Ruiz & Pav.) Radlk, 1882 (Sapotaceae) tree in Peru; molecular gut content analysis suggested this tree to be the feeding plant.

5. Conclusions

We report a fossil geometrid moth, a male, virtually complete, preserved in a clear piece of Miocene Dominican amber dating from 19 to 16 Mya. Despite of being one of the most specious family groups of

Lepidoptera, the fossil record of Geometridae is comparatively very scarce, with only 19 findings formally reported worldwide, most of them of difficult systematic assignment, so that this fossil is of remarkable interest for phylogenetic studies. It belongs to the genus *Dolichoneura* (Geometridae: Desmobathrinae) and is named *Dolichoneura jorelisae* Sarto i Monteys, Hausmann, Baixeras and Peñalver sp. n., based on wing features. Furthermore, it could also serve for calibrating molecular clocks and for paleobiogeographic inferences. The head and wings of this specimen were analyzed through SR μ CT which provided superb virtual representations.

CRediT authorship contribution statement

Victor Sarto i Monteys: Conceptualization, Investigation, Writing – original draft. Axel Hausmann: Conceptualization, Investigation, Writing – review & editing. Mónica M. Solórzano-Kraemer: Data curation, Funding acquisition, Investigation, Methodology, Writing – review & editing. Jörg U. Hammel: Methodology. Joaquín Baixeras: Investigation, Writing – review & editing. Xavier Delclòs: Funding acquisition, Investigation, Writing – review & editing. Enrique Peñalver: Conceptualization, Investigation, Writing – review & editing.

Declaration of competing interest

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

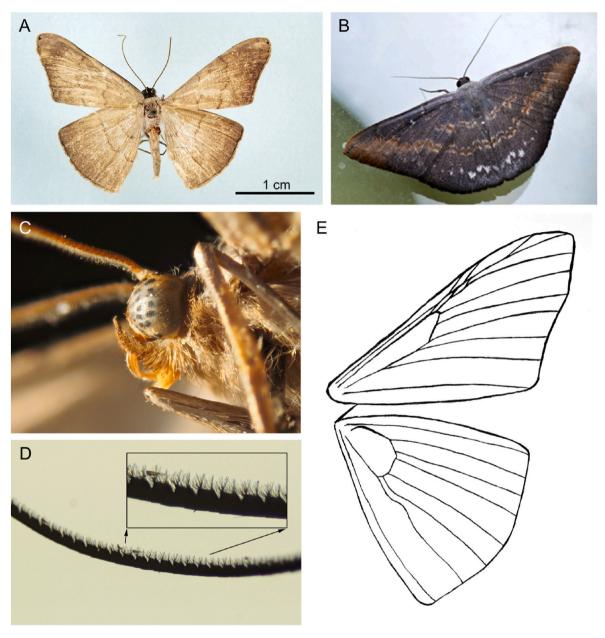


Fig. 9. Dolichoneura oxypteraria (Guenée, 1858). (A) male, from French Guyana, Cayenne, Rte. Regina, PK 32.5, Piste de Nancibo, 8.II.1986, ex coll. Herbulot; wingspan 29 mm, forewing length 15 mm. (B) Live specimen of *Dolichoneura* spp. from Canelos, Pastaza, Ecuador, Eerika Schulz, 15.02.2020, Source iNaturalist, http://www.inaturalist.org/pages/privacy. (C) Close-up of head, proboscis comparatively short, labial palpi strongly curved, length equal to diameter of eye. (D) Close-up of male antenna, slightly dentate, ciliate-fasciculate. (E) Forewing venation: typical characters are the double areole with R5 arising from second areole; Hindwing venation: typical characters are the lack of anastomosis between Sc + R₁ and Rs and the full number of M-veins (both characters pointing to Desmobathrinae).

Data availability

Data will be made available on request.

Acknowledgements

We thank Mr. Jorge Caridad (World Amber Museum, Santo Domingo, Dominican Republic) and his family for the loan of the piece MNHNSD FOS 16.01. for studying purposes, and for its donation to the Museo Nacional de Historia Natural "Prof. Eugenio de Jesús Marcano", Santo Domingo. We thank Carlos Suriel (Museo Nacional de Historia

Natural "Prof. Eugenio de Jesús Marcano") for including it in the collection. Robin Kunz (Senckenberg Research Institute, Frankfurt) supported us with the segmentation of the Synchrotron data. Dr. Alberto Martínez Ortí, director of the Museu Valencià d'Història Natural (Alginet, València, Spain), facilitated the use of the optical microscope and stereomicroscope of that scientific institution. We thank the Spanish Ministry of Economy and Competitiveness (project "CRE", Spanish AEI/FEDER, UE CGL 2017-84419) and VolkswagenStiftung (project 90946) for financial support. Funding sources were not involved in study design, analysis and interpretation of data, the writing of the report and in the decision to submit the article for publication.

References

- Arillo, A., Ortuño, V., 2005. Catalogue of fossil insect species described from Dominican amber (Miocene). Stuttgarter Beiträge zur Naturkunde 352, 1–68.
- Cook, M.A., Scoble, M.J., 1992. Tympanal organs of geometrid moths: a review of their morphology, function, and systematic importance. Syst. Entomol. 17 (3), 219–232. https://doi.org/10.1111/j.1365-3113.1992.tb00334.x.
- GBIF Secretariat, 2021. Dolichoneura Warren, 1894. GBIF Backbone Taxonomy. Checklist dataset:. accessed via GBIF.org on 2022-03-16. https://www.gbif.org/species/ 1982158.
- Greving, I., Wilde, F., Ogurreck, M., Herzen, J., Hammel, J.U., Hipp, A., Friedrich, F., Lottermoser, L., Dose, T., Burmester, H., Müller, M., Beckmann, F., 2014. P05 imaging beamline at PETRA III: first results. Proc. SPIE 9212, 921200.
- Grimaldi, D.A., 1996. Amber: Window to the Past. Harry N. Abrams, New York, p. 216.
 Grimaldi, D.A., Engel, M.S., 2005. Evolution of the Insects. Cambridge University Press,
 New York, p. 755.
- Hausmann, A., 2001. Introduction. Archiearinae, Orthostixinae, Desmobathrinae, alsophilinae, Geometrinae. In: Hausmann, A. (Ed.), The Geometrid Moths of Europe, vol. 1. Apollo Books, Stenstrup, p. 282. https://doi.org/10.1007/978-1-4757-3423-2-1
- Holloway, J.D., 1996. The moths of borneo: Geometridae (incl. Orthostixini), Oenochrominae, Desmobathrinae, Geometrinae, Ennominae, addenda, Part 9. Malay. Nat. J. 49, 1–180.
- Hoernschemeyer, T., Wedmann, S., Poinar, G., 2010. How long can insect species exist? Evidence from extant and fossil *Micromalthus* beetles (Insecta: Coleoptera). Zool. J. Linn. Soc. 158 (2), 300–311.
- Iturralde-Vinent, M.A., MacPhee, R.D., 2019. Remarks on the age of Dominican amber. Palaeoentomology 2 (3), 236–240. https://doi.org/10.11646/ palaeoentomology.2.3.7.
- Kawahara, A.Y., Plotkin, D., Espeland, M., Meusemann, K., Toussaint, E.F.A., Donath, A., Gimnich, F., Frandsen, P.B., Zwick, A., dos Reis, M., Barber, J.R., Peters, R.S., Liu, S., Zhou, X., Mayer, C., Podsiadlowski, L., Storer, C., Yack, J.E., Misof, B., Breinholt, J. W., 2019. Phylogenomics reveals the evolutionary timing and pattern of butterflies and moths. Proc. Natl. Acad. Sci. USA 116 (45), 22657–22663. https://doi.org/10.1073/pnas.1907847116.
- Lytaev, P., Hipp, A., Lottermoser, L., Herzen, J., Greving, I., Khokhriakov, I., Meyer-Loges, S., Plewka, J., Burmester, J., Caselle, M., Vogelgesang, M., Chilingaryan, S., Kopmann, A., Balzer, M., Schreyer, A., Beckmann, F., 2014. Characterization of the CCD and CMOS cameras for grating-based phase-contrast tomography. SPIE Optical Engineering + Applications. Proc. SPIE 9212, 10.
- Martínez-Delclòs, X., Martinell, J., 1993. Insect taphonomy experiments. Their application to the Cretaceous outcrops of lithographic limestones from Spain. Kaupia, Darmstadter Beitrage zur Naturgeschichte 2, 133–144.
- Minet, J., Scoble, M.J., 1999. The Drepanoid/Geometroid assemblage. In: Kristensen, N.
 P. (Ed.), Handbook of Zoology, Part 35, Lepidoptera, Moths and Butterflflies,
 Evolution, Systematics, and Biogeography, vol. 1. De Gruyter, Berlin, pp. 301–320.
- Moosmann, J., Ershov, A., Weinhardt, V., Baumbach, T., Prasad, M.S., LaBonne, C., Xianghui, X., Kashef, J., Hofmann, R., 2014. Time-lapse X-ray phase-contrast microtomography for in vivo imaging and analysis of morphogenesis. Nat. Protoc. 9 (2), 294–304.
- Murillo-Ramos, L., Brehm, G., Sihvonen, P., Hausmann, A., Holm, S., Reza Ghanavi, H., Õunap, E., Truuverk, A., Staude, H., Friedrich, E., Tammaru, T., Wahlberg, N., 2019. A comprehensive molecular phylogeny of Geometridae (Lepidoptera) with a focus on enigmatic small subfamilies. PeerJ 7, e7386. https://doi.org/10.7717/peerj.7386.
- Murillo-Ramos, L., Friedrich, E., Williams, S., Wahlberg, N., Brehm, G., Sihvonen, P., 2021. A morphological appraisal of the new subfamily Epidesmiinae (Lepidoptera: Geometridae) with an overview of all geometrid subfamilies. Zool. J. Linn. Soc. 193, 1205–1233. https://doi.org/10.1093/zoolinnean/zlaa189.

- Palenstijn, W.J., Batenburg, K.J., Sijbers, J., 2011. Performance improvements for iterative electron tomography reconstruction using graphics processing units (GPUs). J. Struct. Biol. 176 (2), 250–253.
- Pinto, R.B., Lusa, M.G., Mansano, V.D., Tozzi, A.M., Mayer, J.L., 2018. Morphoanatomy of the leaflets of the *Hymenaea* clade (Fabaceae: detarioideae) reveals their potential for taxonomic and phylogenetic studies. Bot. J. Linn. Soc. 187, 87–98.
- Prout, L.B., 1932. The American Geometridae. In: Seitz, A. (Ed.), The Macrolepidoptera of the World, vol. 8, pp. 1–149 pls. 1–13, 15, 17.
- Rabl, D., 2016. Small-Scale Topographic Differences Affect Moth Communities Across Three Forest Types in the Golfo Dulce region, SW Costa Rica. Masterarbeit. Universität Wien. https://doi.org/10.25365/thesis.44985. Fakultät für Lebenswissenschaften.
- Scoble, M.J., Hausmann, A., 2007. Online list of valid and nomenclaturally available names of the Geometridae of the World. Available from. http://www.herbulot.de/gl obalspecieslist.htm. (Accessed 12 June 2022).
- Sohn, J.C., Labandeira, C., Davis, D., Mitter, C., 2012. An annotated catalog of fossil and subfossil Lepidoptera (Insecta: Holometabola) of the world. Zootaxa 3286, 1–132. https://doi.org/10.11646/zootaxa.3286.1.1.
- Solórzano Kraemer, M.M., Delclòs, X., Clapham, M., Arillo, A., Peris, D., Jäger, P., Stebner, F., Peñalver, E., 2018. Arthropods in modern resins reveal if amber accurately recorded forest arthropod communities. Proc. Natl. Acad. Sci. USA 115 (26), 6739–6744. https://doi.org/10.1073/pnas.1802138115.
- Solórzano-Kraemer, M.M., Delclòs, X., Engel, M.S., Peñalver, E., 2020. A revised definition for copal and its significance for palaeontological and Anthropocene biodiversity-loss studies. Sci. Rep. 10, 19904 https://doi.org/10.1038/s41598-020-76808-6
- van Aarle, W., Palenstijn, W.J., De Beenhouwer, J., Altantzis, T., Bals, S., Batenburg, K.J., Sijbers, J., 2015. The ASTRA Toolbox: a platform for advanced algorithm development in electron tomography. Ultramicroscopy 157, 35–47.
- van Aarle, W., Palenstijn, W.J., Cant, J., Janssens, E., Bleichrodt, F., Dabravolski, A., De Beenhouwer, J., Batenburg, J., Sijbers, J., 2016. Fast and flexible X-ray tomography using the ASTRA toolbox. Opt Express 24 (22), 25129–25147.
- van Nieukerken, E.J., Kaila, L., Kitching, I.J., Kristensen, N.P., Lees, D.C., Minet, J., Mitter, C., Mutanen, M., Regier, J.C., Simonsen, T.J., Wahlberg, N., Yen, S.-H., Zahiri, R., Adamski, D., Baixeras, J., Bartsch, D., Bengtsson, B.Å., Brown, J.W., Bucheli, S.R., Davis, D.R., De Prins, J., De Prins, W., Epstein, M.E., Gentili-Poole, P., Gielis, C., Hättenschwiler, P., Hausmann, A., Holloway, J.D., Kallies, A., Karsholt, O., Kawahara, A.Y., Koster, S.J.C., Kozlov, M.V., Lafontaine, J.D., Lamas, G., Landry, J.F., Lee, S., Nuss, M., Park, K.-T., Penz, C., Rota, J., Schintlmeister, A., Schmidt, B.C., Sohn, J.-C., Solis, M.A., Tarmann, G.M., Warren, A.D., Weller, S., Yakovlev, R.V., Zolotuhin, V.V., Zwick, A., 2011. Order Lepidoptera Linnaeus, 1758. In: Zhang, Z.Q. (Ed.), Animal Biodiversity: an Outline of Higher-Level Classification and Survey of Taxonomic Richness, vol. 3148. Zootaxa, pp. 212–221. https://doi.org/10.11646/zootaxa.3148.1.41.
- Wahlberg, N., Wheat, C.W., Peña, C., 2013. Timing and patterns in the taxonomic diversification of Lepidoptera (butterflies and moths). PLoS One 8 (11), e80875. https://doi.org/10.1371/journal.pone.0080875.
- Whalley, P.E.S., 1985. The systematics and palaeogeography of the lower jurassic insects of dorset, England. Bull. Br. Mus. (Nat. Hist.) Geol. 39, 107–189.
- Wilde, F., Ogurreck, M., Greving, I., Schreyer, A., 2016. Micro-CT at the imaging beamline P05 at PETRA III. AIP Conf. Proc. 1741 (1), 030035.
- Yamamoto, S., Sota, T., 2007. Phylogeny of the Geometridae and the evolution of winter moths inferred from simultaneous analysis of mitochondrial and nuclear genes. Mol. Phylogenet. Evol. 44 (2), 711–723 https://doi.org/10.1016/j.ympev.2006.12.027.
- Zhang, W., Shih, C., Shih, Y.H., Ren, D., 2020. A new macrolepidopteran moth (Insecta, Lepidoptera, Geometridae) in Miocene Dominican amber. ZooKeys 965, 73–84. https://doi.org/10.3897/zookeys.965.54461.