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Evaluation of two rapid ultrafiltration-based methods for SARS-CoV-2 concentration from wastewater --Manuscript Draft--

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Abstract:	<p>Quantitative measurements of the severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) in raw wastewater have been implemented worldwide since the beginning of the pandemic. Recent efforts are being made to evaluate different viral concentration methodologies to overcome supplier shortages during lockdowns. A set of 22-wastewater samples seeded with murine hepatitis virus (MHV), a member of the Coronaviridae family, and the bacteriophage MS2, were used to characterize and compare two ultrafiltration-based methods: a centrifugal ultrafiltration device (Centricon®Plus-70) and the automated concentrating pipette CP-Select™. Based on the recovery efficiencies, significant differences were observed for MHV, with Centricon® Plus-70 (24%) being the most efficient method. Nevertheless, concentrations of naturally occurring SARS-CoV-2, Human adenoviruses and JC polyomaviruses in these samples did not result in significant differences between methods suggesting that testing naturally occurring viruses may complement the evaluation of viral concentration methodologies. Based on the virus adsorption to solids and the necessity of a pre-centrifugation step to remove larger particles and avoid clogging when using ultrafiltration methods, we assessed the percentage of viruses not quantified after ultrafiltration. Around 23% of the detected SARS-CoV-2 would be discarded during the debris removal step. The CP-Select™ provided the highest concentration factor (up to 333x) and the lowest LoD (6.19 x103 GC/L) for MHV and proved to be fast, automatic, highly reproducible and suitable to work under BSL-2 measures.</p>
Response to Reviewers:	<p>Reviewer 1:</p> <p>We understand the concern of the first reviewer regarding the rationale of the method decision and we must say that feedback on a manuscript helps the most when it comes from people who don't agree with you. When coping with having to test a high amount of wastewater samples for SARS-CoV-2 quantification, we decided to use an ultrafiltration method because it does not require a pre-acidification step and are usually faster methodologies than the flocculation-precipitation methods. We think this rapid processing of the samples and the minimum process steps might favour the stability of enveloped viruses and thus their integrity which may also favour their further</p>

detection.

To date, the published SARS-CoV-2 surveillance studies have been conducted by applying mainly centrifugal ultrafiltration devices (Centricon and Amicon) [Medema et al., 2020, Ahmed et al., 2020], and other methods including polyethylene glycol (PEG) or aluminium hydroxide (Al(OH)₃) flocculation-precipitation are also being used [Randazzo et al., 2020, Zhang et al 2020, LaRosa et al., 2020]. On the first revision of this manuscript, we already included a paragraph stating that: "Ultrafiltration is an interesting method since samples do not need preacidification....Centricon® devices 10, 30 and 100kDa as well as Amicon Ultra-15 have been successfully applied for virus concentration". Thus, we selected the 30kDa filters because they were the most frequently used even in pre-published studies.

In March, centricon shortage made us move to an alternative ultrafiltration method, but before doing that, we decided to test if the obtained results would be equivalent.

Moreover, and in the context of wastewater-based epidemiology applied to routine SARS-CoV-2 monitoring in wastewater samples, there is a claim for easy, safe, fast and reproducible methods as the one characterized in this study.

The tips selected for the CP-Select™ were the smallest tips available. Those tips and the CP-Select™ were previously proposed for virus concentration and preliminary results from our group (Rusiñol et al., 2020 "Concentration methods for the quantification of coronavirus and other potentially pandemic enveloped virus from wastewater" demonstrated their utility for SARS-CoV-2 surveillance in WW. Later, other authors have also used this device in WBE (Gonzalez et al., 2020 "COVID-19 surveillance in South-eastern Virginia using wastewater-based epidemiology" but to date an evaluation of this new system performance has not yet been published.

But, despite the pore size, the most remarkable novelty this method provides is the elution mechanism not seen in other ultrafiltration procedures: according to what is stated in Innovaprep's webpage, "The elution fluid, is conveniently packaged in a single-use canister pressurized by carbon dioxide gas dissolved into the fluid. During the extraction process, the fluid passes from a high-pressure environment, to a low-pressure environment causing the dissolved CO₂ to expand and come out of solution to form into a high-quality microbubble foam. These microbubbles expand the volume of the fluid sevenfold or more, behaving as a solid body as it moves down the inner bore of the hollow fibres in the filter cell creating uniform flow without channelling. The process gently exfoliates and lifts the particles that adhere to the filter cell wall into the concentrate. The elution process is instant and the foam collapses into a liquid in seconds – ready for analysis. An additional benefit of Wet Foam Elution is the simultaneous clean buffer matrix exchange which in many cases, removes unwanted inhibitory substances".

For all these reasons we believe that providing another ultrafiltration-based option useful for SARS-CoV-2 detection with equivalent efficiency than the one applied (centricon) is relevant for the scientific community. CP-Select™ is also a handy equipment that can be used to concentrate at the point-of-use by simply connecting the equipment to a power supply.

We do not agree with the reviewer that the approach is poor. We have compared a substantial amount of wastewater samples which characteristics were added in the revised manuscript as a new Table 1. That proves both methods are useful for SARS-CoV-2 detection independently of the of organic matter content present in the sample or other sample characteristics. And yes, the recovery varies among samples. This fact has also been seen when evaluating viral recoveries in many other studies focusing on concentration of viruses from environmental samples and as it has been also observed when using Centricon devices.

Also, in this manuscript, other aspects, in our opinion very valuable, have been included such as the way the concentration of the viral stock used for spiking is estimated since as shown may have great influence in this kind of studies and should be considered.

In the previous revision, we included additional information on enzymatic inhibition and what has been made regarding this. Conclusion section was already added in the previous revised version of the manuscript.

Reviewer 3:

As answered above, the manuscript main aim is not to compare methods but to provide an alternative ultrafiltration-based method to ultrafiltration devices more commonly used for SARS-CoV-2 concentration. The CP-Select™ uses a new elution concept based on the use of pressurized eluent. Thus, our aim hasn't been to compare

two different ultrafiltration systems because of their different pore sizes but two different ultrafiltration systems reported to be useful for SARS-CoV-2 detection that differ in the pore size, the concentration factor and the elution system among other features. Moreover, we choose this alternative since it is a rapid method that can be performed into a BL2 cabinet and could be performed in the point of use if needed. We have proved that the method provides equivalent results to the ones obtained by using Centricon devices. Moreover, we believe the method could be easily used in routine testing.

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2 **concentration from wastewater**

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4 **M.3, Borrego C.M.4⁵, Corominas LL.4, ⁶, Girones R.1², Rusiñol M. 7**

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1 Abstract

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3 (SARS-CoV-2) in raw wastewater have been implemented worldwide since the beginning
4 of the pandemic. Recent efforts are being made to evaluate different viral concentration
5 methodologies to overcome supplier shortages during lockdowns. A set of 22-wastewater
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20 GC/L) for MHV and proved to be fast, automatic, highly reproducible and suitable to
21 work under BSL-2 measures.

22

Reviewer #1: Dear Editor,

I believe that the content of the manuscript does not comply with the aim of this highly impact journal "...for publication of novel, hypothesis-driven and high-impact research on..." In addition, the authors did not reply to all my comments, while the given answers are still not well addressed, i.e. it seems that the rationale behind this work is the shortage of a specific product in the market (1st paragraph of response), which may be true, however there are many other companies that provide filters with the same specs. In addition, the authors did not comment on the following: "There are some studies revealing the role of not only the concentration of suspended solids, but also the content of organic matter, nutrients, etc.; factors that should also be considered when dealing with wastewater samples. In addition, a significant constrain when implementing such analyses that may influence the recovery rates is the presence of inhibitors and this is not discussed in the manuscript, although the authors tested wastewater samples from different plants and in some cases recovery rates presented high variability. The presented results are in line with already published data and new contributions are limited to the CP-Select commercial concentration method. "Conclusions" section is absent. All in all, the approach is rather poor, the methodology not well justified, the results inadequately presented, and the discussion is very weak..." I regret to say that to my opinion the scientific quality of the manuscript does not fit with the scientific value of the papers published in this journal.

We understand the concern of the reviewer regarding the rationale of the method decision and we must say that feedback on a manuscript helps the most when it comes from people who don't agree with you. When coping with having to test a high amount of wastewater samples for SARS-CoV-2 quantification, we decided to use an ultrafiltration method because it does not require a pre-acidification step and are usually faster methodologies than the flocculation-precipitation methods. We think this rapid processing of the samples and the minimum process steps might favour the stability of enveloped viruses and thus their integrity which may also favour their further detection.

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Reviewer #3: On the whole I'm satisfied by how Authors managed the first round of review and upgraded their paper.

Nevertheless, I still share the same concern of Rev.1 about the (unsupported) choice of different molecular weight cut offs of tested devices. However, I'd say that the manuscript could be accepted in its current form, after a short paragraph is added in the Discussion, plainly stating and debating this methodological important limitation and maybe expliciting the possibility/need of further comparison among devices with the same MWCO (independently from their stock availability during the ongoing pandemic times).

As answered above, the manuscript main aim is not to compare methods but to provide an alternative ultrafiltration-based method to ultrafiltration devices more commonly used for SARS-CoV-2 concentration. The CP-Select™ uses a new elution concept based on the use of pressurized eluent. Thus, our aim hasn't been to compare two different ultrafiltration systems because of their different pore sizes but two different ultrafiltration systems reported to be useful for SARS-CoV-2 detection that differ in the pore size, the concentration factor and the elution system among other features. Moreover, we choose this alternative since it is a rapid method that can be performed into a BL2 cabinet and could be performed in the point of use if needed. We have proved that the method provides equivalent results to the ones obtained by using Centricon devices. Moreover, we believe the method could be easily used in routine testing.

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19 **Abstract**

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40

41 **Keywords:** SARS-CoV-2, wastewater, viral concentration method, ultrafiltration, viral
42 recovery

43

44 **1. Introduction**

45 There is increasing evidence that untreated wastewater is a promising unbiased indicator
46 of the presence of SARS-CoV-2 virus in the population as it has been reported by different
47 research groups as a possible way to monitor trends and the approximate overall
48 prevalence of COVID-19 in the population (Kitajima et al., 2020; Medema et al., 2020a)

49 Given the coronavirus pandemic impacts, the method to detect SARS-CoV-2 RNA in
50 wastewater had, by necessity, to be developed and implemented at warp-speed. One of
51 the major challenges in SARS-CoV-2 research in wastewater has been the lack of
52 standardized protocols for its detection. The way the virus is concentrated seems to be
53 crucial in order to avoid false negative results or inaccurate reported concentrations.

54 On the lack of much data regarding coronavirus recovery efficiency when using common
55 methods for viral concentration, we should rely on what it is known for other enveloped
56 viruses considering that every single virus will have a different behaviour during viral
57 concentration. Alone or combined, electropositive and electronegative filtration,
58 centrifugal ultrafiltration, organic flocculation and PEG/Al(OH)₃ precipitation methods
59 have been used in different studies targeting enveloped viruses' in environmental waters
60 as recently reviewed (Rusiñol et al., 2020).

61 Preliminary data obtained by our research group in a study evaluating different
62 concentration methods for the detection of SARS-CoV-2 in wastewater showed no
63 significant differences between skimmed milk organic flocculation and Centricon® Plus-
64 70- and CP-Select™ ultrafiltration devices (Rusiñol et al., 2020). Centricon® Plus-70
65 ultrafilters have been described as a useful method for SARS-CoV-2 concentration from
66 wastewater. Ultrafiltration is an interesting method since: i) samples do not need
67 preacidification, ii) nor a long time of precipitation, which could not favour the stability

68 of enveloped viruses, and iii) their concentration relies mainly on their size. However,
69 and due to COVID-19 pandemic, there has been a shortage of these ultrafiltration devices.
70 For this reason, this study was focused on the evaluation of two ultrafiltration methods
71 described as useful for SARS-CoV-2 concentration from wastewater. Centricon® Plus-
72 70 30kDa devices and the Concentrator Pipette CP-Select™ from Innovaprep were tested
73 to concentrate raw wastewater samples in artificially spiked with MS2 bacteriophage and
74 Murine Hepatitis Coronavirus (MHV) ~~raw wastewater samples~~ and presenting also
75 naturally occurring SARS-CoV-2, Human adenoviruses (HAdV) and JC polyomaviruses
76 (JCPyV). Centricon® of different cut-off size (10, 30 and 100kDa) have been applied to
77 concentrate SARS-CoV-2 (Medema et al., 2020a; Rusiñol et al., 2020). In this issue
78 30kDa were the filters of election, trying to favour viral retention while avoiding the
79 retention of smaller molecules that could act as enzymatic inhibitors. Regarding filter tips
80 to be coupled to CP-Select™, the smaller pore size tips available (150kDa) were used.
81 The novelty of this method resides in the use of a pressurized eluent in the form of wet
82 foam.

83

84 **2. Material and methods**

85 *2.1. Viruses and cell lines*

86 Bacteriophage MS2 (ATCC 23631), a model for non-enveloped RNA viruses and
87 Murine Hepatitis Virus-A59 (MHV-A59), a model for enveloped betacoronaviruses (like
88 SARS-CoV-2), were propagated using the following protocols. Bacteriophage MS2 was
89 cultured in *Salmonella typhimurium* strain WG49 (NCTC 12484) following ISO 10705-
90 1 indications. MHV-A59 and DBT murine cell line were kindly provided by Wigginton
91 Group Research, Michigan University, Michigan. MHV were propagated by infecting
92 confluent monolayers of DBT cells following previously described instructions

93 (Leibowitz et al., 2011). Viruses were clarified from the supernatant by centrifugation at
94 3,000xg for 15 min and the supernatants were kept at - 80°C.

95 *2.2. Sample collection*

96 A total of 22 24-hours-composite raw wastewater samples (500 ml) were collected
97 between March and September 2020 from 6 WWTPs, located in Catalonia (Spain) (Table
98 1). The selected WWTPs treat urban and industrial wastewater from approximately 20%
99 of the Catalan population. Samples were either shipped to the laboratory under cool
100 conditions or alternatively stored after collection at - 20°C.

101 Additionally, to determine the relation between the viral recovery and wastewater
102 physicochemical characteristics, the turbidity was measured using a turbidimeter
103 HI98703 (Hanna Instruments Inc.), the pH was measured using a pHmeter 902/4 (Nahita
104 Inc.) and the BOD_5 values were provided by WWTP managers.

105 *2.3. Viral concentration methods*

106 An aliquot of 200ml of each wastewater sample was seeded with 10^7 GC/ml of MS2 and
107 MHV (1:100, v/v). Samples were centrifuged at 4,750xg for 30 minutes in order to
108 remove suspended solids that may interfere with the ultrafiltration. The resulting
109 supernatant was divided into two aliquots of 100 ml and subjected to two different viral
110 concentration methods:

111 1) Concentration Pipette CP-Select™ using Hollow Fiber Polysulfone PVP high-flow
112 pipette ultrafilter tips (CPT) with a cut-off of 150 KDa (InnovaPrep) and 2) Centricon®
113 Plus-70 centrifugal ultrafiltration (CeUF) devices, with a cut-off of 30 KDa (Millipore).
114 CP-Select™ method began with filtration of 80 ml of supernatant through single-use
115 CPT. Viral particles were eluted with 0.075% Tween 20/Tris using *Wet Foam Elution™*
116 canisters (Innovaprep) into a final volume of between 240 μl and 600 μl .

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117 The CeUF devices were pre-rinsed before use, following manufacturer instructions, and
118 then 70 ml of supernatant ~~were~~ centrifuged at 3,000xg for 30 minutes. Viruses were
119 eluted inverting the CeUF device and centrifuged at 1,000xg for 3 minutes to obtain the
120 final concentrate of approximately 280-900 µL.

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121 *2.4. Nucleic acid extraction and q(RT)PCR quantification*

122 Viral nucleic acids (NA) were extracted using the QIAmp Viral RNA Mini kit (Qiagen,
123 Inc., Valencia, CA) according to the manufacturer's protocol in an automated QIAcube
124 platform (Qiagen, Inc., Valencia, CA). The volume of the concentrates used for the
125 extraction were 140 µL and the elution volumes were 60 - 80 µL. A negative control of
126 the viral nucleic acid extraction was added per batch of samples.

127 Specific real-time qPCR and RT-qPCR assays previously described were used to quantify
128 SARS-CoV-2 N1 and N2 (probes, primers and cycling conditions described in the CDC-
129 006-00019 CDC/DDID/NCIRD/ Division of Viral Diseases protocol), MS2
130 bacteriophage (Pecson et al., 2009), MHV (Ahmed et al., 2020), HAdV (Hernroth et al.,
131 2002) and JCPyV (Pal et al., 2006) by using TaqMan™ Environmental Master Mix 2.0
132 and RNA UltraSense™ One-Step RT-qPCR System (Invitrogen) for DNA and RNA
133 viruses respectively. Quantification was performed in a StepOne plus Real-Time PCR
134 System (Applied Biosystems, USA). Undiluted and 10-fold dilutions of the nucleic acid
135 extracts were analysed in duplicate. All the qPCR and RT-qPCR assays included non-
136 template controls to demonstrate that the mix did not produce fluorescence and bovine
137 serum albumina (BSA) (1mg/ml), was added to RT-qPCR assays to reduce PCR
138 inhibitors. The standards for viruses were prepared using synthetic gBlocks® Gene
139 Fragments (IDT) and quantified with a Qubit® fluorometer (Thermo Fisher Scientific)
140 except for SARS-CoV-2 standard which was constructed using the EURM-019 single
141 stranded RNA fragments of SARS-CoV-2, provided by the European Commission Joint

142 Research Centre. For all the standards, ten-fold dilutions were prepared from 10^0 to 10^7
143 copies per reaction.

144 As for enzymatic inhibition we performed previous tests, when setting up qPCR for N1
145 and N2 assays for SARS-CoV-2 detection, by adding known amounts of target RNA into
146 wastewater. Inhibition was reduced when including BSA to the qPCR master mix. Every
147 tested sample was previously spiked with MS2 bacteriophages that were used as a process
148 control as well as for controlling inhibition by analysing tenfold dilutions of every nucleic
149 acid extraction.

150 *2.5.LOD/LOQ determination*

151 The limit of detection (LoD) of the whole method (including ultrafiltration, extraction
152 and RT-qPCR detection) was calculated by running six replicate tenfold dilutions of
153 target DNA/RNA suspensions around the detection end point (2.5, 5, 25 and 50
154 GC/reaction), for each analysed virus. The concentration that produced at least 95%
155 positive replicates was assumed to be the LoD of the qPCR assay, which was transformed
156 to LoD of the entire method using the sample volume tested in each of the methodologies.
157 The limit of quantification (LoQ) was estimated using the procedure described by
158 Foorotan and colleagues (Foorotan et al., 2017).

159 *2.6.Evaluation of viral recovery*

160 Viral recovery percentage was calculated according to experimental values obtained by
161 spiking samples with MS2 and MHV viral stocks, shaking for 10 min and using as input
162 viral concentration the direct quantification of the viral stock added:

$$163 \quad \text{Virus recovery (\%)} = \frac{\text{Concentrate Titer (GC/ml)} \times \text{Sample Volume (ml)}}{\text{Inoculum Titer (GC/ml)} \times (\text{Sample Volume (ml)}/100)} \times 100$$

164 To shed some light into the role that the matrix into which viral stock is embedded may
165 play when calculating viral recoveries, four different quantification strategies were
166 conducted: 1) direct quantification of the viral stocks; 2) quantification of raw wastewater
167 spiked with known concentrations of the viral stocks; 3) same as 2 but after debris
168 removal, and 4) quantification of the viral stocks in a concentrated wastewater sample.
169 All these quantifications were assayed in triplicate.

170 *2.7. Virus attachment to suspended solids*

171 To investigate the percentage of coronaviruses which could remain attached to suspended
172 material and not be properly quantified using ultrafiltration methods, viruses present in
173 pellets obtained after centrifugation of 9 raw wastewater samples were further eluted in
174 3.5 ml of glycine buffer pH9.5 for 30 minutes and after the addition of 3.5 ml of 2xPBS
175 centrifuged at 3000xg for 20 minutes. The resulting supernatant (6.5 - 7.5 ml) was filtered
176 using Amicon® Ultra-15 Centrifuge Filters Ultracell® 50KDa (Merck Millipore) and
177 eluted for further viral quantification. Simultaneously supernatants obtained after first
178 centrifugation were further concentrated as described in section 2.3 using Centricon®
179 Plus-70 devices.

180 *2.8. Tween-20 addition in the pre-concentration step before ultrafiltration with CP- 181 Select™*

182 CP-Select™ manufacturer recommends the addition of Tween-20 before ultrafiltration in
183 order to increase viral recovery. The appropriateness of including this step to the CP-
184 Select™ concentration protocol step was evaluated in 3 selected wastewater samples (100
185 ml). Prior to ultrafiltration, 5% Tween 20 (1:100, v/v) was added to raw wastewater and
186 processed as described above.

187 *2.9. Data visualization and statistical analysis*

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188 Data visualization, plotting and statistical test was done using R version 4.0.2 (R Core
189 Team, 2020). For each virus, Wilcoxon signed rank tests for paired data were used to
190 evaluate whether there were statistically significant differences between both
191 ultrafiltration methods. To evaluate potential associations between viral recovery and raw
192 wastewater turbidity we run Pearson's correlation coefficient tests.

193

194 **3. Results**

195

196 *3.1. Comparison between CP-Select™ and Centricon® Plus-70 devices*

197 The MS2 phage, a non-enveloped RNA virus frequently used as a process control in
198 environmental studies (Coulliette et al., 2014; Ikner et al., 2011; Ye et al., 2016) and the
199 MHV, an enveloped RNA surrogate for human coronavirus (Ahmed et al., 2020;
200 Casanova et al., 2009; Ye et al., 2016), were seeded to calculate viral concentration
201 methods recovery efficiencies.

202 Mean recovery values for MS2 and MHV in wastewater are represented in Figure 1. No
203 statistically significant differences were observed between concentration methods
204 regarding MS2 recovery (p -value = 0.75) but CeUF provided significant highest mean
205 recoveries for MHV (p -value = 0.004). However, no statistical differences were observed
206 between the two methods when naturally occurring viruses were quantified (Figure 2):
207 SARS-CoV-2 (p -value of 0.27 and 0.73 for N1 and N2, respectively), HAdV, JCPyV (p -
208 values > 0.05). CeUF provided higher mean recovery percentages for MHV whereas CP-
209 Select™ provided higher recovery rates for MS2.

210 Table 2 summarizes equivalent sample volumes analyzed and the resulting concentration
211 factors by using CP-Select™ or CeUF methods as well as the limits of detection and

212 quantification (LoD_{95%} and LoQ), calculated mean recoveries, standard deviations and
213 coefficients of variation of the compared concentration methods based on MS2 and MHV
214 quantifications. By using the concentrating pipette, a higher concentration factor was
215 obtained, and a larger sample volume was analyzed in each RT-qPCR reaction.

216 After addition of Tween-20 into wastewater previously to concentration with CP-
217 Select™, no statistical differences were observed when adding Tween-20 (p -value =
218 0.105), obtaining mean values of 50.7 and 20.9 GC/ml SARS-CoV-2 with and without
219 Tween-20 addition respectively. However, the ultrafiltration time when adding Tween-
220 20 was reduced.

221 *3.2. Viral stock quantification*

222 When evaluating if calculation of viral recoveries could be biased by the effect of the
223 matrix in which viral stocks were embedded, no significant differences were observed
224 when quantifying MS2 stocks directly or within different wastewater matrices (p -values
225 >0.05) (Figure 3). On the other hand, MHV stock quantification showed a matrix effect
226 suggesting that the way the viral stock, used for spiking recovery assays, is quantified
227 may influence recovery values obtained. In this study, the recoveries represented in
228 Figure 1 were calculated according to the direct quantification of the MHV used for
229 spiking whereas MHV stock quantification in wastewater matrices would have showed
230 higher viral recoveries (data not shown).

231 *3.3. Virus attachment to suspended solids*

232 Seeded MS2 and naturally occurring SARS-CoV-2 (N1 gene) were quantified from
233 sample concentrates and in the generated pellets at the debris removal step (Figure 4).
234 For MS2, similar fractions were measured from the pellet (49%) and the supernatant
235 (51%). For the naturally occurring SARS-CoV-2 (N1 assay), those samples that could be

236 quantified showed more variability. In samples 1-9, most of the detectable SARS-CoV-2
237 fraction (mean values of 77%) was measured in the supernatant whereas the remaining
238 23% was detected in the pellets.

239 The turbidity of the wastewater samples was highly variable, ranging from 106 to 830
240 NTU (Nephelometric Turbidity Units). Weak correlations were observed between sample
241 turbidity and viral quantifications obtained by using the CP-Select™ method (Pearson's
242 correlation coefficients of 0.2 and 0.4 for MS2 and MHV, respectively) and inverse
243 relation with sample turbidity was observed when using CeUF (Pearson's correlation
244 coefficients of 0.2 and 0.1 for MS2 and MHV, respectively). No correlations between
245 viral concentrations and pH and BOD₅ were observed (<0.3).

246

247 4. Discussion

248 In the actual pandemic scenario, viral concentration methods showing acceptable
249 performance for both enveloped and non-enveloped viruses have received increased
250 attention. As recently reviewed, a wide variety of strategies are being used to study viral
251 presence in wastewater (Corpuz et al., 2020) but few of those concentration
252 methodologies has been implemented for SARS-CoV-2 surveillance (Rusiñol et al.,
253 2020). When comparing methodologies, ultrafiltration achieves higher MHV recoveries
254 (25%) than PEG precipitation (5%), but the ultrafiltration devices are less used than
255 flocculation/precipitation methods (Ye et al., 2016). This has been mainly caused by the
256 shortage of supplies and the lack of readily material in many countries during lockdowns.
257 Nevertheless, the one-step centrifugal ultrafiltration techniques enable the detection of
258 viruses from relatively small sample volumes (70–80 mL).

259 Three ultrafiltration devices: the Centricon® Plus-70 (Medema et al., 2020b), the
260 Amicon® Ultra-15 (Ahmed et al., 2020) and the new automatic Concentrating Pipette

261 (CP-Select™) from Innovaprep (Gonzalez et al., 2020; Rusiñol et al., 2020) have been
262 successfully used to detect SARS-CoV-2 from wastewater. The first two devices have
263 also been used to concentrate other human enteric viruses from water (Qiu et al., 2016;
264 Sidhu et al., 2018). Viruses are retained based on size exclusion and backwashed from
265 the ultrafilters. Both CeUF devices (Centricon® and Amicon®) contain an Ultracell®
266 regenerated cellulose membrane that results in 19 cm² and 7.6 cm² respectively, whereas
267 the CP-Select™ with Hollow Fiber Polysulfone ultrafiltration tips has a surface area of 98
268 cm², which is 5 to 13 times larger than those of the other CeUF devices. To our knowledge
269 this is the first study that compares the performance of the CP-Select™ with Centricon®
270 Plus-70 to concentrate SARS-CoV-2 and other viruses from wastewater samples. It
271 should be noticed that this system includes a wet foam elution step which according to
272 the manufacturer's improves viral elution from filter cells.

273 When applying ultrafiltration to wastewater, samples need to be pre-centrifuged to
274 remove larger particles and avoid clogging. The resulting supernatant (70 - 80ml) is then
275 passed in a single-step through the ultrafilter. Viruses have been reported to adsorb to the
276 solid fraction of wastewater (Ye et al., 2016). According to our results, 23% of total
277 detected SARS-CoV-2 would be discarded during the debris removal step while higher
278 percentage of the detectable MS2 (49%) would be retained in the pellet. Ye et al. (2016)
279 reported MHV to adsorb to the solid fraction of wastewater samples in higher percentages
280 (26%) than MS2 (6%) while Ahmed et al., reported similar loss for seeded MHV (30%)
281 at the pre-filtration step (Ahmed et al., 2020). According to our results and considering
282 the need of easy and fast method for SARS-CoV-2 detection in wastewater as an early
283 warning tool, a straightforward and routinely adopted method shouldn't consider
284 including viruses attached to the debris. This would suppose simply e an extra elution step,
285 from the debris, and addition to the wastewater sample, a procedure which has not a 100%

286 efficiency, which would suppose an addition of only a percentage of viruses attached to
287 solid material. Thus, in our opinion, this step is not worth doing for routine testing and
288 only when very high sensitivities and accurate quantifications are needed. Regarding the
289 two ultrafiltration methods evaluated in this study, significant differences were only
290 observed for MHV for which CeUF devices performed better than CP-Select™. In
291 contrast, for naturally occurring SARS-CoV-2 both methods provided similar results
292 showing that, as expected, each single virus behave differently under the same
293 concentration procedure. Despite MHV is also a member of the Genus *Betacoronavirus*
294 (as SARS-CoV-2), it did not show equivalent recovery rates to CeUF. Interestingly,
295 however, the concentration of naturally occurring SARS-CoV-2 from wastewater using
296 both concentration methods resulted in equivalent outcomes. This suggest that the best
297 way to compare concentration methods for SARS-CoV-2 could be testing real
298 environmental samples since, as observed for other viruses and other concentration
299 methods, each virus has a particular behaviour for each of the methodologies applied. The
300 way the MHV stock was quantified seemed to affect the recovery value obtained thus
301 pointing at a clear effect of the matrix into which the viral stock is suspended. This could
302 be probably due to different RNA protection/degradation phenomena or to the
303 presence/absence of enzymatic inhibition in the different matrices assayed. This is
304 another reason to consider when evaluating viral concentration methods and another
305 argument in favour of using naturally occurring virus to complement concentration
306 methods comparison studies, although this strategy does not allow the estimation of
307 recovery rates.

308 Overall, CeUF devices were confirmed as an efficient ultrafiltration procedure for SARS-
309 CoV-2 as it has been previously reported by others (Ahmed et al., 2020; Medema et al.,
310 2020b). Moreover, CP-Select™ with Hollow Fiber Polysulfone tips showed to be useful

311 for SARS-CoV-2 concentration from wastewater as well as for the concentration of other
312 wastewater occurring viruses independently of the turbidity of the samples. It is worth
313 mentioning that equipment fits into a BSL-2 cabinet which makes this procedure strongly
314 recommended for viruses requiring biosafety containment. In turn, CeUF devices should
315 be used in a superspeed centrifuge that is difficult to fit into BSL-2 facility especially in
316 routine laboratories that require extreme security measures to avoid spill overs.

317 Also, CP-Select™ provides with good concentration factor and equivalent LoD, LoQ and
318 variance than CeUF devices. The use of Tween-20, as it has been recommended by
319 manufacturers, has not proven to increase SARS-CoV-2 recovery although it has been
320 observed it may help to filtrate samples reducing the time required for ultrafiltration.

321 CP-Select™ is a handy equipment that can be applied without previous debris elimination
322 or by only using syringe filters or vacuum filtration devices. This device allows
323 concentration at the point-of-use by simply connecting the CP-Select™ equipment to a
324 power supply. The number of methods available for SARS-CoV-2 concentration from
325 wastewater is increasing, as well as data on their performance, which will be relevant for
326 researchers and routine laboratories in order to make a good election on their SARS-CoV-
327 2 testing strategies. Detection of other potential pandemic enveloped viruses, that could
328 emerge soon, would require optimized and well characterized viral concentration
329 methods.

330 **Conclusions:**

- 331 • Ultrafiltration devices (Centricon® and CP-Select™) performed equally for
332 different naturally occurring viruses, including SARS-CoV-2, whereas for the
333 spiked MHV, used as a model of enveloped viruses of the genus betacoronavirus,
334 the CeUF achieved higher recoveries.
- 335 • The way the viral stock is quantified may influence recovery values calculations.

- 336 • Up to 23% of detected SARS-CoV-2 adsorb to the solid fraction and is not
337 considered in the further detection by quantitative PCR.
- 338 • The CP-Select™ fits into a BSL-2 cabinet enabling to work under biosafety
339 containment

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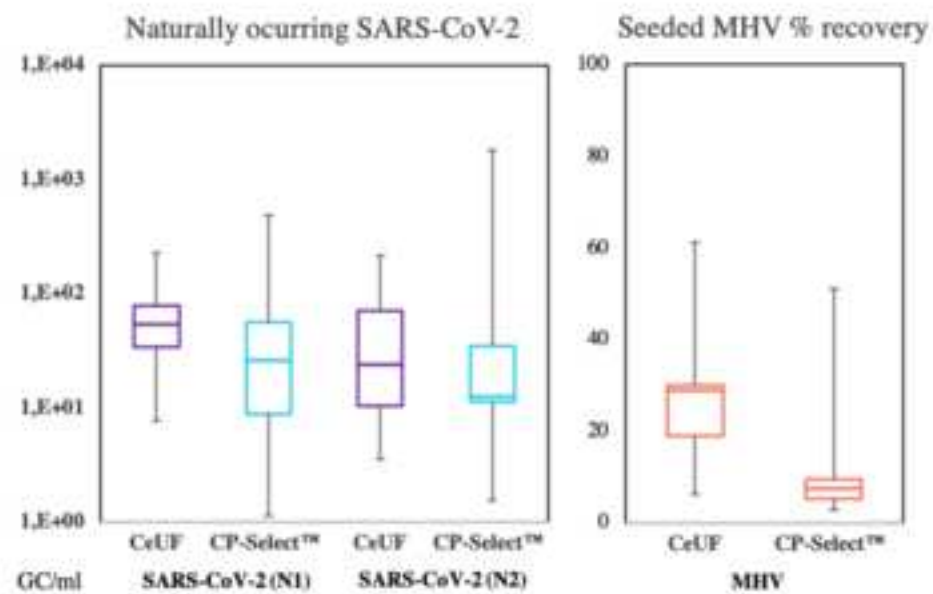
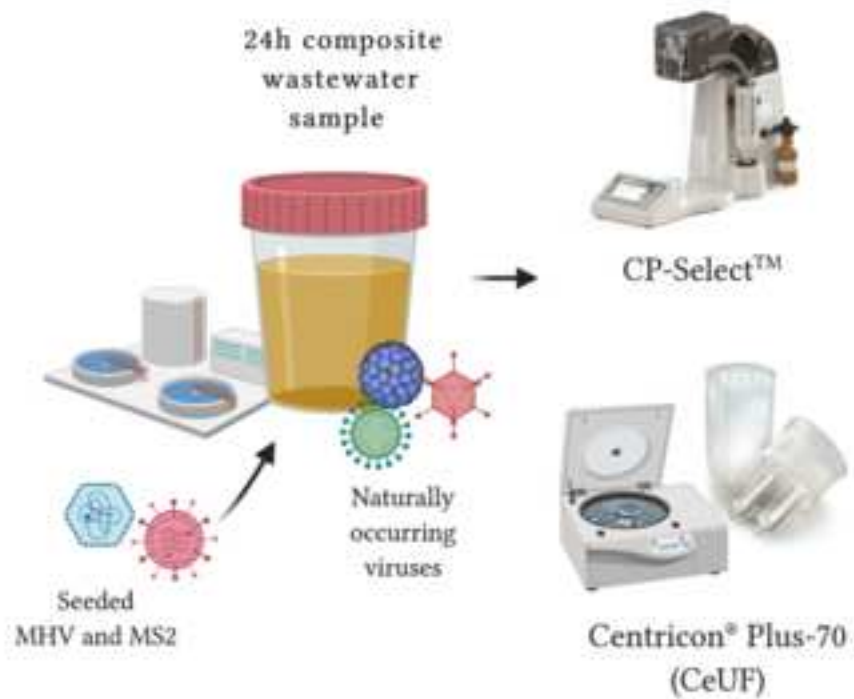
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424



1 **Highlights**

- 2 Centricon[®] and CP-Select[™] performed equally for naturally occurring SARS-CoV-2
- 3 Higher MHV recoveries were calculated using centrifugal ultrafiltration devices
- 4 Naturally occurring viruses complement concentration methods comparison
- 5 A 23% of detected SARS-CoV-2 adsorb to the solid fraction of wastewater
- 6 CP-Select[™] fits into a BSL-2 cabinet enabling to work under biosafety containment

1 **Evaluation of two rapid ultrafiltration-based methods for SARS-CoV-2**
2 **concentration from wastewater**

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18

19 **Abstract**

20 Quantitative measurements of the severe acute respiratory syndrome coronavirus 2
21 (SARS-CoV-2) in raw wastewater have been implemented worldwide since the beginning
22 of the pandemic. Recent efforts are being made to evaluate different viral concentration
23 methodologies to overcome supplier shortages during lockdowns. A set of 22-wastewater
24 samples seeded with murine hepatitis virus (MHV), a member of the *Coronaviridae*
25 family, and the bacteriophage MS2, were used to characterize and compare two
26 ultrafiltration-based methods: a centrifugal ultrafiltration device (Centricon[®] Plus-70)
27 and the automated concentrating pipette CP-Select[™]. Based on the recovery efficiencies,
28 significant differences were observed for MHV, with Centricon[®] Plus-70 (24%) being the
29 most efficient method. Nevertheless, concentrations of naturally occurring SARS-CoV-
30 2, Human adenoviruses and JC polyomaviruses in these samples did not result in
31 significant differences between methods suggesting that testing naturally occurring
32 viruses may complement the evaluation of viral concentration methodologies. Based on
33 the virus adsorption to solids and the necessity of a pre-centrifugation step to remove
34 larger particles and avoid clogging when using ultrafiltration methods, we assessed the
35 percentage of viruses not quantified after ultrafiltration. Around 23% of the detected
36 SARS-CoV-2 would be discarded during the debris removal step. The CP-Select[™]
37 provided the highest concentration factor (up to 333x) and the lowest LoD (6.19×10^3
38 GC/L) for MHV and proved to be fast, automatic, highly reproducible and suitable to
39 work under BSL-2 measures.

40

41 **Keywords:** SARS-CoV-2, wastewater, viral concentration method, ultrafiltration, viral
42 recovery

43

44 **1. Introduction**

45 There is increasing evidence that untreated wastewater is a promising unbiased indicator
46 of the presence of SARS-CoV-2 virus in the population as it has been reported by different
47 research groups as a possible way to monitor trends and the approximate overall
48 prevalence of COVID-19 in the population (Kitajima et al., 2020; Medema et al., 2020a).

49 Given the coronavirus pandemic impacts, the method to detect SARS-CoV-2 RNA in
50 wastewater had, by necessity, to be developed and implemented at warp-speed. One of
51 the major challenges in SARS-CoV-2 research in wastewater has been the lack of
52 standardized protocols for its detection. The way the virus is concentrated seems to be
53 crucial in order to avoid false negative results or inaccurate reported concentrations.

54 On the lack of much data regarding coronavirus recovery efficiency when using common
55 methods for viral concentration, we should rely on what it is known for other enveloped
56 viruses considering that every single virus will have a different behaviour during viral
57 concentration. Alone or combined, electropositive and electronegative filtration,
58 centrifugal ultrafiltration, organic flocculation and PEG/Al(OH)₃ precipitation methods
59 have been used in different studies targeting enveloped viruses' in environmental waters
60 as recently reviewed (Rusiñol et al., 2020).

61 Preliminary data obtained by our research group in a study evaluating different
62 concentration methods for the detection of SARS-CoV-2 in wastewater showed no
63 significant differences between skimmed milk organic flocculation and Centricon® Plus-
64 70 and CP-Select™ ultrafiltration devices (Rusiñol et al., 2020). Centricon® Plus-70
65 ultrafilters have been described as a useful method for SARS-CoV-2 concentration from
66 wastewater. Ultrafiltration is an interesting method since: i) samples do not need
67 preacidification, ii) nor a long time of precipitation, which could not favour the stability

68 of enveloped viruses, and iii) their concentration relies mainly on their size. However,
69 and due to COVID-19 pandemic, there has been a shortage of these ultrafiltration devices.
70 For this reason, this study was focused on the evaluation of two ultrafiltration methods
71 described as useful for SARS-CoV-2 concentration from wastewater. Centricon® Plus-
72 70 30kDa devices and the Concentrator Pipette CP-Select™ from Innovaprep were tested
73 to concentrate raw wastewater samples artificially spiked with MS2 bacteriophage and
74 Murine Hepatitis Coronavirus (MHV) and presenting also naturally occurring SARS-
75 CoV-2, Human adenoviruses (HAdV) and JC polyomaviruses (JCPyV). Centricon® of
76 different cut-off size (10, 30 and 100kDa) have been applied to concentrate SARS-CoV-
77 2 (Medema et al., 2020a; Rusiñol et al., 2020). In this issue 30kDa were the filters of
78 election, trying to favour viral retention while avoiding the retention of smaller molecules
79 that could act as enzymatic inhibitors. Regarding filter tips to be coupled to CP-Select™,
80 the smallest available pore size tips (150kDa) were used. The novelty of this method
81 resides in the use of a pressurized eluent in the form of wet foam.

82

83 **2. Material and methods**

84 *2.1. Viruses and cell lines*

85 Bacteriophage MS2 (ATCC 23631), a model for non-enveloped RNA viruses and Murine
86 Hepatitis Virus-A59 (MHV-A59), a model for enveloped betacoronaviruses (like SARS-
87 CoV-2), were propagated using the following protocols. Bacteriophage MS2 was cultured
88 in *Salmonella typhimurium* strain WG49 (NCTC 12484) following ISO 10705-1
89 indications. MHV-A59 and DBT murine cell line were kindly provided by Wigginton
90 Group Research, Michigan University, Michigan. MHV were propagated by infecting
91 confluent monolayers of DBT cells following previously described instructions

92 (Leibowitz et al., 2011). Viruses were clarified from the supernatant by centrifugation at
93 3,000xg for 15 min and the supernatants were kept at - 80°C.

94 2.2. *Sample collection*

95 A total of 22 24-hours-composite raw wastewater samples (500 ml) were collected
96 between March and September 2020 from 6 WWTPs, located in Catalonia (Spain) (Table
97 1). The selected WWTPs treat urban and industrial wastewater from approximately 20%
98 of the Catalan population. Samples were either shipped to the laboratory under cool
99 conditions or alternatively stored after collection at - 20°C.

100 Additionally, to determine the relation between the viral recovery and wastewater
101 physicochemical characteristics, the turbidity was measured using a turbidimeter
102 HI98703 (Hanna Instruments Inc.), the pH was measured using a pHmeter 902/4 (Nahita
103 Inc.) and the BOD₅ values were provided by WWTP managers.

104 2.3. *Viral concentration methods*

105 An aliquot of 200 ml of each wastewater sample was seeded with 10⁷ GC/ml of MS2 and
106 MHV (1:100, v/v). Samples were centrifuged at 4,750xg for 30 minutes in order to
107 remove suspended solids that may interfere with the ultrafiltration. The resulting
108 supernatant was divided into two aliquots of 100 ml and subjected to two different viral
109 concentration methods:

110 1) Concentration Pipette CP-Select™ using Hollow Fiber Polysulfone PVP high-flow
111 pipette ultrafilter tips (CPT) with a cut-off of 150 KDa (InnovaPrep) and 2) Centricon®
112 Plus-70 centrifugal ultrafiltration (CeUF) devices, with a cut-off of 30 KDa (Millipore).
113 CP-Select™ method began with filtration of 80 ml of supernatant through single-use
114 CPT. Viral particles were eluted with 0.075% Tween-20/Tris using *Wet Foam Elution*™
115 cans (Innovaprep) into a final volume of between 240 µl and 600 µl.

116 The CeUF devices were pre-rinsed before use, following manufacturer instructions, and
117 then 70 ml of supernatant was centrifuged at 3,000xg for 30 minutes. Viruses were eluted
118 inverting the CeUF device and centrifuged at 1,000xg for 3 minutes to obtain the final
119 concentrate of approximately 280-900 µl.

120 *2.4. Nucleic acid extraction and q(RT)PCR quantification*

121 Viral nucleic acids (NA) were extracted using the QIAmp Viral RNA Mini kit (Qiagen,
122 Inc., Valencia, CA) according to the manufacturer's protocol in an automated QIAcube
123 platform (Qiagen, Inc., Valencia, CA). The volume of the concentrates used for the
124 extraction were 140 µL and the elution volumes were 60 - 80 µL. A negative control of
125 the viral nucleic acid extraction was added per batch of samples.

126 Specific real-time qPCR and RT-qPCR assays previously described were used to quantify
127 SARS-CoV-2 N1 and N2 (probes, primers and cycling conditions described in the CDC-
128 006-00019 CDC/DDID/NCIRD/ Division of Viral Diseases protocol), MS2
129 bacteriophage (Pecson et al., 2009), MHV (Ahmed et al., 2020), HAdV (Hernroth et al.,
130 2002) and JCPyV (Pal et al., 2006) by using TaqManTM Environmental Master Mix 2.0
131 and RNA UltraSenseTM One-Step RT-qPCR System (Invitrogen) for DNA and RNA
132 viruses respectively. Quantification was performed in a StepOne plus Real-Time PCR
133 System (Applied Biosystems, USA). Undiluted and 10-fold dilutions of the nucleic acid
134 extracts were analysed in duplicate. All the qPCR and RT-qPCR assays included non-
135 template controls to demonstrate that the mix did not produce fluorescence and bovine
136 serum albumina (BSA) (1mg/ml), was added to RT-qPCR assays to reduce PCR
137 inhibitors. The standards for viruses were prepared using synthetic gBlocks[®] Gene
138 Fragments (IDT) and quantified with a Qubit[®] fluorometer (Thermo Fisher Scientific)
139 except for SARS-CoV-2 standard which was constructed using the EURM-019 single
140 stranded RNA fragments of SARS-CoV-2, provided by the European Commission Joint

141 Research Centre. For all the standards, ten-fold dilutions were prepared from 10^0 to 10^7
142 copies per reaction.

143 As for enzymatic inhibition we performed previous tests, when setting up qPCR for N1
144 and N2 assays for SARS-CoV-2 detection, by adding known amounts of target RNA into
145 wastewater. Inhibition was reduced when including BSA to the qPCR master mix. Every
146 tested sample was previously spiked with MS2 bacteriophages that were used as a process
147 control as well as for controlling inhibition by analysing tenfold dilutions of every nucleic
148 acid extraction.

149 *2.5.LOD/LOQ determination*

150 The limit of detection (LoD) of the whole method (including ultrafiltration, extraction
151 and RT-qPCR detection) was calculated by running six replicate tenfold dilutions of
152 target DNA/RNA suspensions around the detection end point (2.5, 5, 25 and 50
153 GC/reaction), for each analysed virus. The concentration that produced at least 95%
154 positive replicates was assumed to be the LoD of the qPCR assay, which was transformed
155 to LoD of the entire method using the sample volume tested in each of the methodologies.
156 The limit of quantification (LoQ) was estimated using the procedure described by
157 Forootan and colleagues (Forootan et al., 2017).

158 *2.6.Evaluation of viral recovery*

159 Viral recovery percentage was calculated according to experimental values obtained by
160 spiking samples with MS2 and MHV viral stocks, shaking for 10 min and using as input
161 viral concentration the direct quantification of the viral stock added:

$$162 \quad \text{Virus recovery (\%)} = \frac{\text{Concentrate Titer (GC/ml)} \times \text{Sample Volume (ml)}}{\text{Inoculum Titer (GC/ml)} \times (\text{Sample Volume (ml)}/100)} \times 100$$

163 To shed some light into the role that the matrix into which viral stock is embedded may
164 play when calculating viral recoveries, four different quantification strategies were
165 conducted: 1) direct quantification of the viral stocks; 2) quantification of raw wastewater
166 spiked with known concentrations of the viral stocks; 3) same as 2 but after debris
167 removal, and 4) quantification of the viral stocks in a concentrated wastewater sample.
168 All these quantifications were assayed in triplicate.

169 *2.7. Virus attachment to suspended solids*

170 To investigate the percentage of coronaviruses which could remain attached to suspended
171 material and not be properly quantified using ultrafiltration methods, viruses present in
172 pellets obtained after centrifugation of 9 raw wastewater samples were further eluted in
173 3.5 ml of glycine buffer pH9.5 for 30 minutes and after the addition of 3.5 ml of 2xPBS
174 centrifuged at 3000xg for 20 minutes. The resulting supernatant (6.5 - 7.5 ml) was filtered
175 using Amicon[®] Ultra-15 Centrifuge Filters Ultracell[®] 50KDa (Merck Millipore) and
176 eluted for further viral quantification. Simultaneously supernatants obtained after first
177 centrifugation were further concentrated as described in section 2.3 using Centricon[®]
178 Plus-70 devices.

179 *2.8. Tween-20 addition in the pre-concentration step before ultrafiltration with CP-* 180 *Select™*

181 CP-Select™ manufacturer recommends the addition of Tween-20 before ultrafiltration in
182 order to increase viral recovery. The appropriateness of including this step to the CP-
183 Select™ concentration protocol step was evaluated in 3 selected wastewater samples (100
184 ml). Prior to ultrafiltration, 5% Tween 20 (1:100, v/v) was added to raw wastewater and
185 processed as described above.

186 *2.9. Data visualization and statistical analysis*

187 Data visualization, plotting and statistical test was done using R version 4.0.2 (R Core
188 Team, 2020). For each virus, Wilcoxon signed rank tests for paired data were used to
189 evaluate whether there were statistically significant differences between both
190 ultrafiltration methods. To evaluate potential associations between viral recovery and raw
191 wastewater turbidity we run Pearson's correlation coefficient tests.

192

193 **3. Results**

194

195 *3.1. Comparison between CP-Select™ and Centricon® Plus-70 devices*

196 The MS2 phage, a non-enveloped RNA virus frequently used as a process control in
197 environmental studies (Coulliette et al., 2014; Ikner et al., 2011; Ye et al., 2016) and the
198 MHV, an enveloped RNA surrogate for human coronavirus (Ahmed et al., 2020;
199 Casanova et al., 2009; Ye et al., 2016), were seeded to calculate viral concentration
200 methods recovery efficiencies.

201 Mean recovery values for MS2 and MHV in wastewater are represented in Figure 1. No
202 statistically significant differences were observed between concentration methods
203 regarding MS2 recovery (p -value = 0.75) but CeUF provided significant highest mean
204 recoveries for MHV (p -value = 0.004). However, no statistical differences were observed
205 between the two methods when naturally occurring viruses were quantified (Figure 2):
206 SARS-CoV-2 (p -value of 0.27 and 0.73 for N1 and N2, respectively), HAdV, JCPyV (p -
207 values > 0.05). CeUF provided higher mean recovery percentages for MHV whereas CP-
208 Select™ provided higher recovery rates for MS2.

209 Table 2 summarizes equivalent sample volumes analyzed and the resulting concentration
210 factors by using CP-Select™ or CeUF methods as well as the limits of detection and

211 quantification (LoD_{95%} and LoQ), calculated mean recoveries, standard deviations and
212 coefficients of variation of the compared concentration methods based on MS2 and MHV
213 quantifications. By using the concentrating pipette, a higher concentration factor was
214 obtained, and a larger sample volume was analyzed in each RT-qPCR reaction.

215 After addition of Tween-20 into wastewater previously to concentration with CP-
216 Select™, no statistical differences were observed when adding Tween-20 (p -value =
217 0.105), obtaining mean values of 50.7 and 20.9 GC/ml SARS-CoV-2 with and without
218 Tween-20 addition respectively. However, the ultrafiltration time when adding Tween-
219 20 was reduced.

220 *3.2. Viral stock quantification*

221 When evaluating if calculation of viral recoveries could be biased by the effect of the
222 matrix in which viral stocks were embedded, no significant differences were observed
223 when quantifying MS2 stocks directly or within different wastewater matrices (p -values
224 >0.05) (Figure 3). On the other hand, MHV stock quantification showed a matrix effect
225 suggesting that the way the viral stock, used for spiking recovery assays, is quantified
226 may influence recovery values obtained. In this study, the recoveries represented in
227 Figure 1 were calculated according to the direct quantification of the MHV used for
228 spiking whereas MHV stock quantification in wastewater matrices would have showed
229 higher viral recoveries (data not shown).

230 *3.3. Virus attachment to suspended solids*

231 Seeded MS2 and naturally occurring SARS-CoV-2 (N1 gene) were quantified from
232 sample concentrates and in the generated pellets at the debris removal step (Figure 4).
233 For MS2, similar fractions were measured from the pellet (49%) and the supernatant
234 (51%). For the naturally occurring SARS-CoV-2 (N1 assay), those samples that could be

235 quantified showed more variability. In samples 1-9, most of the detectable SARS-CoV-2
236 fraction (mean values of 77%) was measured in the supernatant whereas the remaining
237 23% was detected in the pellets.

238 The turbidity of the wastewater samples was highly variable, ranging from 106 to 830
239 NTU (Nephelometric Turbidity Units). Weak correlations were observed between sample
240 turbidity and viral quantifications obtained by using the CP-Select™ method (Pearson's
241 correlation coefficients of 0.2 and 0.4 for MS2 and MHV, respectively) and inverse
242 relation with sample turbidity was observed when using CeUF (Pearson's correlation
243 coefficients of 0.2 and 0.1 for MS2 and MHV, respectively). No correlations between
244 viral concentrations and pH and BOD₅ were observed (<0.3).

245

246 4. Discussion

247 In the actual pandemic scenario, viral concentration methods showing acceptable
248 performance for both enveloped and non-enveloped viruses have received increased
249 attention. As recently reviewed, a wide variety of strategies are being used to study viral
250 presence in wastewater (Corpuz et al., 2020) but few of those concentration
251 methodologies has been implemented for SARS-CoV-2 surveillance (Rusiñol et al.,
252 2020). When comparing methodologies, ultrafiltration achieves higher MHV recoveries
253 (25%) than PEG precipitation (5%), but the ultrafiltration devices are less used than
254 flocculation/precipitation methods (Ye et al., 2016). This has been mainly caused by the
255 shortage of supplies and the lack of readily material in many countries during lockdowns.
256 Nevertheless, the one-step centrifugal ultrafiltration techniques enable the detection of
257 viruses from relatively small sample volumes (70 – 80 mL).

258 Three ultrafiltration devices: the Centricon® Plus-70 (Medema et al., 2020b), the
259 Amicon® Ultra-15 (Ahmed et al., 2020) and the new automatic Concentrating Pipette

260 (CP-Select™) from Innovaprep (Gonzalez et al., 2020; Rusiñol et al., 2020) have been
261 successfully used to detect SARS-CoV-2 from wastewater. The first two devices have
262 also been used to concentrate other human enteric viruses from water (Qiu et al., 2016;
263 Sidhu et al., 2018). Viruses are retained based on size exclusion and backwashed from
264 the ultrafilters. Both CeUF devices (Centricon® and Amicon®) contain an Ultracell®
265 regenerated cellulose membrane that results in 19 cm² and 7.6 cm² respectively, whereas
266 the CP-Select™ with Hollow Fiber Polysulfone ultrafiltration tips has a surface area of 98
267 cm², which is 5 to 13 times larger than those of the other CeUF devices. To our knowledge
268 this is the first study that compares the performance of the CP-Select™ with Centricon®
269 Plus-70 to concentrate SARS-CoV-2 and other viruses from wastewater samples. It
270 should be noticed that this system includes a wet foam elution step which according to
271 the manufacturer's improves viral elution from filter cells.

272 When applying ultrafiltration to wastewater, samples need to be pre-centrifuged to
273 remove larger particles and avoid clogging. The resulting supernatant (70 - 80ml) is then
274 passed in a single-step through the ultrafilter. Viruses have been reported to adsorb to the
275 solid fraction of wastewater (Ye et al., 2016). According to our results, 23% of total
276 detected SARS-CoV-2 would be discarded during the debris removal step while higher
277 percentage of the detectable MS2 (49%) would be retained in the pellet. Ye et al. (2016)
278 reported MHV to adsorb to the solid fraction of wastewater samples in higher percentages
279 (26%) than MS2 (6%) while Ahmed et al., reported similar loss for seeded MHV (30%)
280 at the pre-filtration step (Ahmed et al., 2020). According to our results and considering
281 the need of easy and fast method for SARS-CoV-2 detection in wastewater as an early
282 warning tool, a straightforward and routinely adopted method shouldn't consider
283 including viruses attached to the debris. This would imply an extra elution step, from the
284 debris, and addition to the wastewater sample, which would suppose an addition of only

285 a percentage of viruses attached to solid material. Thus, in our opinion, this step is not
286 worth doing for routine testing and only when very high sensitivities and accurate
287 quantifications are needed. Regarding the two ultrafiltration methods evaluated in this
288 study, significant differences were only observed for MHV for which CeUF devices
289 performed better than CP-Select™. In contrast, for naturally occurring SARS-CoV-2 both
290 methods provided similar results showing that, as expected, each single virus behave
291 differently under the same concentration procedure. Despite MHV is also a member of
292 the Genus *Betacoronavirus* (as SARS-CoV-2), it did not show equivalent recovery rates
293 to CeUF. Interestingly, however, the concentration of naturally occurring SARS-CoV-2
294 from wastewater using both concentration methods resulted in equivalent outcomes. This
295 suggest that the best way to compare concentration methods for SARS-CoV-2 could be
296 testing real environmental samples since, as observed for other viruses and other
297 concentration methods, each virus has a particular behaviour for each of the
298 methodologies applied. The way the MHV stock was quantified seemed to affect the
299 recovery value obtained thus pointing at a clear effect of the matrix into which the viral
300 stock is suspended. This could be probably due to different RNA protection/degradation
301 phenomena or to the presence/absence of enzymatic inhibition in the different matrices
302 assayed. This is another reason to consider when evaluating viral concentration methods
303 and another argument in favour of using naturally occurring virus to complement
304 concentration methods comparison studies, although this strategy does not allow the
305 estimation of recovery rates.

306 Overall, CeUF devices were confirmed as an efficient ultrafiltration procedure for SARS-
307 CoV-2 as it has been previously reported by others (Ahmed et al., 2020; Medema et al.,
308 2020b). Moreover, CP-Select™ with Hollow Fiber Polysulfone tips showed to be useful
309 for SARS-CoV-2 concentration from wastewater as well as for the concentration of other

310 wastewater occurring viruses independently of the turbidity of the samples. It is worth
311 mentioning that equipment fits into a BSL-2 cabinet which makes this procedure strongly
312 recommended for viruses requiring biosafety containment. In turn, CeUF devices should
313 be used in a superspeed centrifuge that is difficult to fit into BSL-2 facility especially in
314 routine laboratories that require extreme security measures to avoid spill overs.

315 Also, CP-Select™ provides with good concentration factor and equivalent LoD, LoQ and
316 variance than CeUF devices. The use of Tween-20, as it has been recommended by
317 manufacturers, has not proven to increase SARS-CoV-2 recovery although it has been
318 observed it may help to filtrate samples reducing the time required for ultrafiltration.

319 CP-Select™ is a handy equipment that can be applied without previous debris elimination
320 or by only using syringe filters or vacuum filtration devices. This device allows
321 concentration at the point-of-use by simply connecting the CP-Select™ equipment to a
322 power supply. The number of methods available for SARS-CoV-2 concentration from
323 wastewater is increasing, as well as data on their performance, which will be relevant for
324 researchers and routine laboratories in order to make a good election on their SARS-CoV-
325 2 testing strategies. Detection of other potential pandemic enveloped viruses, that could
326 emerge soon, would require optimized and well characterized viral concentration
327 methods.

328 **Conclusions:**

- 329 • Ultrafiltration devices (Centricon® and CP-Select™) performed equally for
330 different naturally occurring viruses, including SARS-CoV-2, whereas for the
331 spiked MHV, used as a model of enveloped viruses of the genus betacoronavirus,
332 the CeUF achieved higher recoveries.
- 333 • The way the viral stock is quantified may influence recovery values calculations.
- 334 • Up to 23% of detected SARS-CoV-2 adsorb to the solid fraction and is not
335 considered in the further detection by quantitative PCR.

- 336 • The CP-Select™ fits into a BSL-2 cabinet enabling to work under biosafety
337 containment

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422

Table 1. Characteristics of the selected wastewater treatment plants (WWTP). Mean values and standard deviations. BOD₅: biological organic demand.

WWTP	Number of samples	Design capacity (Hab. Eq.)	Turbidity (NTU)	pH	BOD ₅ (mgO ₂ /L)
1	10	2843750	816±17	7.39±0.13	364±72
2	2	451250	218±2.31	7.54±0.15	390±72
3	2	285666	113±8.14	8.17±0.21	69±30
4	3	196167	165±4.36	7.62±0.10	217±63
5	2	165450	106±1.15	7.55±0.20	316±126
6	3	99166	222±5.86	7.80±0.15	191±47

Table 2. Characterization of the concentration methods: volume of wastewater sample analyzed in each reaction, mean concentration factor, estimated 95% limit of detection (LoD_{95%}) and limit of quantification (LoQ) and mean recovery values for each of the seeded viruses.

	CP-Select™	CeUF
Sample volume analyzed per reaction	1,56-2,92 ml	0,91-2,19 ml
Concentration factor	133-333x	77-250x
LoD_{95%} (CI)*	MS2: 5,14 x10 ³ (3,02 x10 ³ -9,40 x10 ³) MHV: 6,19 x10 ³ (2,43 x10 ³ -1,58 x10 ⁴)	MS2: 5,67 x10 ³ (3,22 x10 ³ -1,03 x10 ⁴) MHV: 6,61 x10 ³ (2,59 x10 ³ -1,68 x10 ⁴)
LoQ*	MS2: 2,32 x10 ³ MHV: 2,35 x10 ⁴	MS2: 3,56 x10 ³ MHV: 2,51 x10 ⁴
Mean recovery ± SD (CV)	MS2: 27,72 ± 24,46% (0,65) MHV: 7,51 ± 6,14% (0,68)	MS2: 26,34 ± 22,71% (0,66) MHV: 24,07 ± 14,48% (0,58)

*LoD_{95%} and LoQ values are given in genome copies detected per liter of the original wastewater sample. CI: confidence interval; SD: Standard deviation; CV: coefficient of variation.

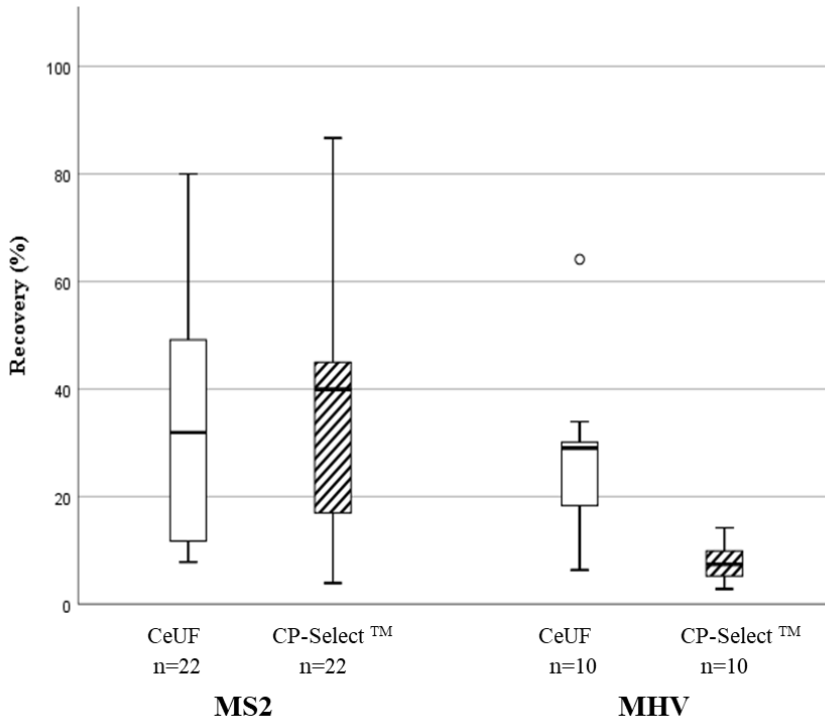


Figure 1. Barplots of the mean recovery values (%) of MS2 and MHV by using two different ultrafiltration methods: InnovaPrep concentrating pipette with single-use ultrafiltration tips 150KDa (CP Select™) and centrifugal ultrafiltration with Centricon® Plus-70 30KDa (CeUF).

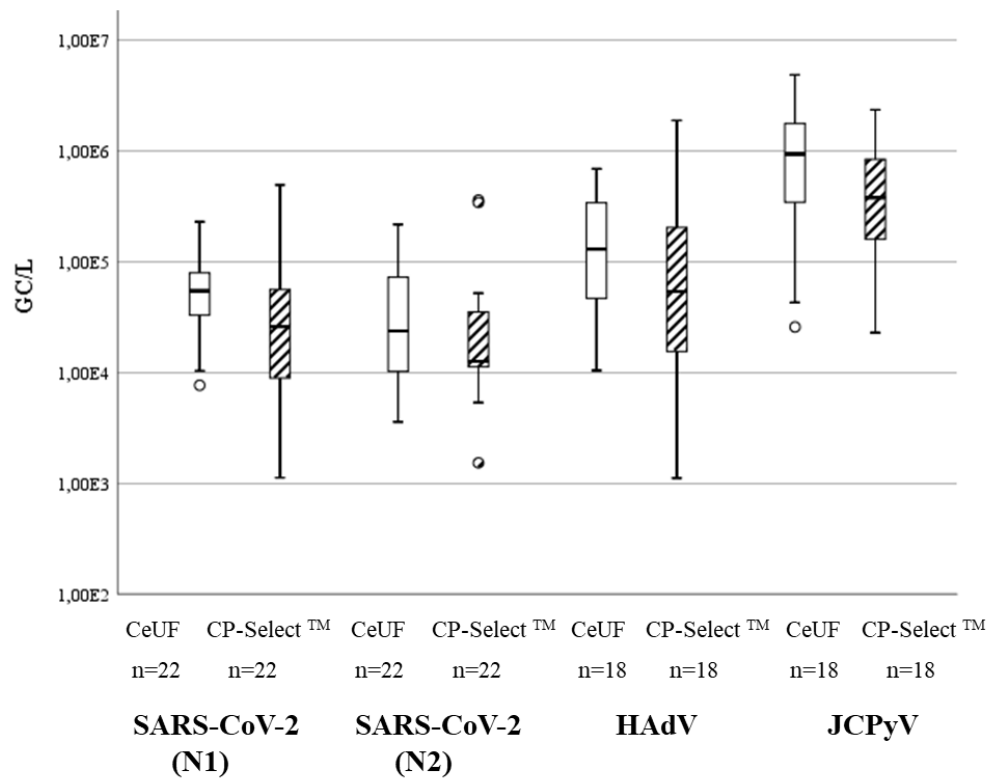
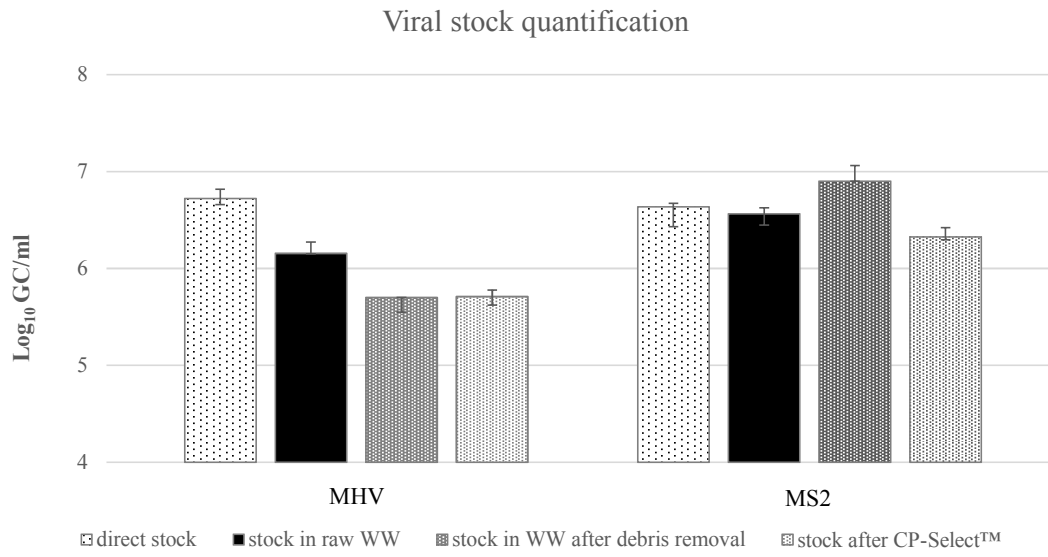


Figure 2. Barplots of the concentrations of naturally occurring SARS-CoV-2 (N1 and N2 assays), HAdV and JCPyV (expressed in genome copies per liter) by using two different ultrafiltration methods: InnovaPrep concentrating pipette with single-use ultrafiltration tips 150KDa (CP Select™) and centrifugal ultrafiltration with Centricon® Plus-70 30KDa (CeUF).

Figure 3. Mean concentration values of the viral stocks, using 4 different quantification strategies.



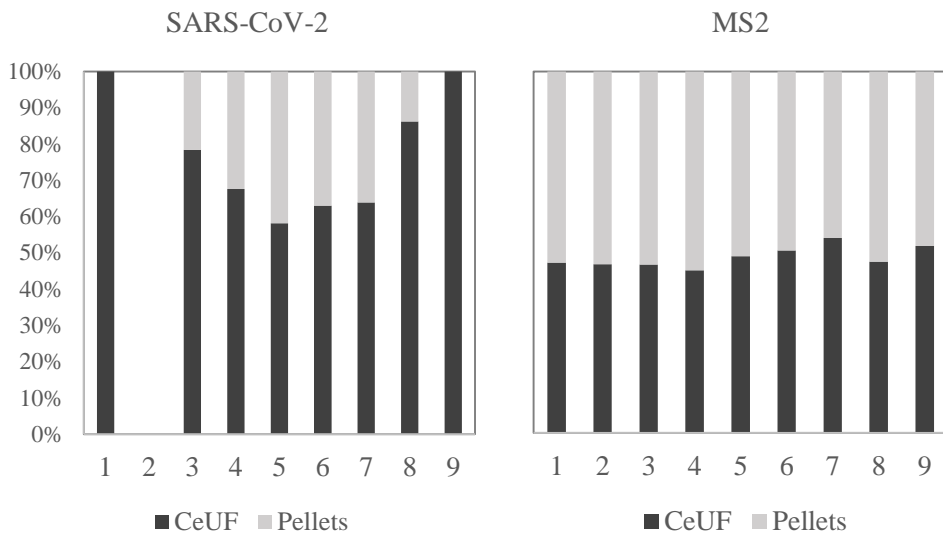


Figure 4. Detection of naturally occurring SARS-CoV-2 (N1 assay) and seeded MS2 in the pellet or supernatant fractions of nine wastewater samples after 4,700xg 30 minutes centrifugation expressed as the percentage of total viruses detected.

CRediT authorship contribution statement

Eva Forés: Sampling, Methodology, Formal analysis, Writing-Original draft preparation

Bofill-Mas S: Methodology, Formal analysis, Writing- Original draft preparation, Conceptualization, Writing- Reviewing and Editing

Itarte M: Methodology, Formal analysis.

Martínez-Puchol S: Methodology.

Hundesda A: Methodology.

Calvo M: Data statistical analysis

Borrego C.M.: Sampling, Reviewing and Editing

Corominas LL: Sampling, Reviewing and Editing

Girones R: Reviewing and Editing

Rusiñol M: Methodology, Formal analysis, Conceptualization, Writing-Original draft preparation, Supervision

Declaration of interests

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

The authors declare the following financial interests/personal relationships which may be considered as potential competing interests: