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Decoding the molecular heterogeneity of pediatric monomorphic post-solid organ transplant lymphoproliferative disorders

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Abstract:

Post-transplant lymphoproliferative disorders (PTLD) represent a broad spectrum of lymphoid proliferations, frequently associated with Epstein-Barr Virus (EBV) infection. The molecular profile of pediatric monomorphic PTLDs (mPTLD) has not been elucidated and it is unknown whether they display similar genetic features as their counterpart in adult and immunocompetent (IMC) pediatric patients. In this study, we investigated 31 pediatric mPTLD after solid organ transplantation, including 24 diffuse large B-cell lymphomas (DLBCL), mostly classified as activated B-cell, and seven Burkitt lymphoma (BL), 93% of which were EBV positive. We performed an integrated molecular approach, including fluorescence in situ hybridization, targeted gene sequencing and copy-number (CN) arrays. Overall, PTLD-BL carried mutations in MYC, ID3, DDX3X, ARID1A or CCND3 resembling IMC-BL, higher mutational burden than PTLD-DLBCL and less CN alterations than IMC-BL. PTLD-DLBCL showed a very heterogeneous genomic profile with fewer mutations and CN alterations than IMC-DLBCL. Epigenetic modifiers and genes of Notch pathway were the most recurrently mutated in PTLD-DLBCL (both 28%). Mutations in cell cycle and Notch pathways correlated with worse outcome. All seven PTLD-BL were alive after treatment with pediatric B-cell Non-Hodgkin Lymphoma protocols, whereas 54% of DLBCL patients were cured with immunosuppression reduction, rituximab and/or low-dose chemotherapy. These findings highlight the low complexity of pediatric PTLD-DLBCL, their good response to low intensity treatment and the shared pathogenesis between PTLD-BL and EBV+ IMC-BL. We also suggest new potential parameters that could help in the diagnosis and the design of better therapeutic strategies for these patients.

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Decoding the molecular heterogeneity of pediatric monomorphic post-solid organ transplant lymphoproliferative disorders

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Data Sharing Statement

Copy-number and sequencing data are available at GEO and ENA under accession numbers GSE198253 and ERP134862, respectively.

Key points

- The mutational profile of pediatric PTLD-BL resembles immunocompetent EBV+ BL, suggesting the need of an intensive therapy.
- Pediatric PTLD-DLBCL is genetically less complex than adult PTLD-DLBCL and pediatric immunocompetent DLBCL.

ABSTRACT

Post-transplant lymphoproliferative disorders (PTLD) represent a broad spectrum of lymphoid proliferations, frequently associated with Epstein-Barr Virus (EBV) infection. The molecular profile of pediatric monomorphic PTLDs (mPTLD) has not been elucidated and it is unknown whether they display similar genetic features as their counterpart in adult and immunocompetent (IMC) pediatric patients. In this study, we investigated 31 pediatric mPTLD after solid organ transplantation, including 24 diffuse large B-cell lymphomas (DLBCL), mostly classified as activated B-cell, and seven Burkitt lymphoma (BL), 93% of which were EBV positive. We performed an integrated molecular approach, including fluorescence in situ hybridization, targeted gene sequencing and copy-number (CN) arrays. Overall, PTLD-BL carried mutations in MYC, ID3, DDX3X, ARID1A or CCND3 resembling IMC-BL, higher mutational burden than PTLD-DLBCL and less CN alterations than IMC-BL. PTLD-DLBCL showed a very heterogeneous genomic profile with fewer mutations and CN alterations than IMC-DLBCL. Epigenetic modifiers and genes of Notch pathway were the most recurrently mutated in PTLD-DLBCL (both 28%). Mutations in cell cycle and Notch pathways correlated with worse outcome. All seven PTLD-BL were alive after treatment with pediatric B-cell Non-Hodgkin Lymphoma protocols, 54% of DLBCL patients whereas were cured with immunosuppression reduction, rituximab and/or low-dose chemotherapy. These findings highlight the low complexity of pediatric PTLD-DLBCL, their good response to low intensity treatment and the shared pathogenesis between PTLD-BL and EBV+ IMC-BL. We also suggest new potential parameters that

could help in the diagnosis and the design of better therapeutic strategies for these patients.

INTRODUCTION

Post-transplant lymphoproliferative disorders (PTLD) are relatively common complications after solid organ (SOT) or hematopoietic stem cell transplantations and represent a major cause of morbidity and mortality.¹ Pediatric transplant recipients are at increased risk of developing PTLD, in part because of Epstein-Barr virus (EBV) seronegativity at the time of transplantation.¹ Three main EBV latency patterns are described in B cells, through which the virus is able to transit, distinguished by different viral gene expression profiles.² Characteristically, PTLDs show a latency III pattern.³

Diffuse large B-cell lymphoma (DLBCL) is the most common subtype of monomorphic PTLD (mPTLD) in both, adult and pediatric patients, while Burkitt lymphoma (BL) is a less frequent form, with aggressive clinical presentation and different pathological and prognostic characteristics.^{4,5} These two entities are also the most frequent mature B-cell Non-Hodgkin Lymphoma (B-NHL) in immunocompetent (IMC) children and adolescents, and their genetic landscapes have been extensively studied.^{6–10} In detail, sporadic BL (sBL) is characterized by t(8;14)(q24;q32)/IGH::*MYC* translocation and recurrent mutations in *ID3*, *CCND3*, *MYC*, *TP53* and *TCF3*.^{6–8} Additionally, EBV-associated BL, including the endemic variant (eBL), displays less frequent mutations.^{11–15} On the other hand, pediatric and young adult DLBCL lacks *BCL2* and *BCL6* primary aberrations, but similarly to adults, has recurrent 2p16/*REL* gains, 19p13/*CD70* homozygous deletions, and mutations in *SOCS1* and *KMT2D*.^{9,10}

Despite the clear morphological similarities of mPTLD with lymphomas in IMC patients, few studies have systematically analyzed their biology and genetics beyond their relationship to EBV infection. The genetic profile of PTLD-BL has not been unraveled yet, whereas the few studies that describe the molecular profile of PTLD-DLBCL have been performed in adult patients.^{16–18} In this sense, adult PTLD-DLBCL carries recurrent alterations in the JAK-STAT pathway,¹⁸ lower genetic complexity of germinal center B-cell like (GCB) PTLD-DLBCL than IMC-DLBCL, and absence of mutations in key genes characteristic of IMC-DLBCL lymphomagenesis such as *SOCS1*, and genes involved in the NFKB signaling pathway as *CARD11*.^{16,17} Moreover, EBV-positive (EBV+) PTLD-DLBCL patients carry a lower mutational burden than the EBV-negative (EBV-).¹⁶ Additionally, gains of 3q, 5p, 8q and 11p, and 1p36, 12p and 17p13 losses have been described in adult PTLD-DLBCL.^{19–21}

The lack of information in pediatric mPTLD also raises the question whether large B-cell lymphoma/high grade with 11q aberration (LBCL/HG-11q) and large B-cell lymphoma with *IRF4* translocation (LBCL-*IRF4*), two recently described entities predominantly occurring in children, are represented in this setting.^{22,23} Of note, the mutational profile of these diseases has been well characterized in IMC population. LBCL/HG-11q lacks BL related mutations, but displays recurrent mutations in *BTG2*, *DDX3X*, *ETS1*, *EP300*, *GNA13* and *NFRKB*,^{24,25} whereas LBCL-*IRF4* carries frequent alterations in *IRF4* and *CARD11* genes.⁹

In terms of clinical management, there is no standard of care (SOC) for mPTLD in children, a significant gap sharpened by the fact that these patients are particularly vulnerable due to risk of toxicity and rejection, and susceptibility to

infections. Reduction of immunosuppression (RIS) is generally the first treatment step, followed by rituximab, combined or not, with chemotherapy.^{26,27}

The aim of this study was to delineate for the first time, the specific genetic, clinical and pathological features of pediatric mPTLD after SOT and to clarify whether the pathogenesis of these lymphomas is the same as in IMC patients. A thorough morphological review and molecular characterization of these cases, together with correlation between these aspects and clinical parameters, might help select better treatment strategies for this challenging population.

METHODS

Patients and samples

Thirty-one SOT-related pediatric mPTLD (≤18 years old) patients at diagnosis were recruited from pediatric transplantation Spanish national reference centers in the context of a centralized review supported by the *Sociedad Española de Hematología y Oncología Pediátricas* (SEHOP). All cases were centrally reviewed by four hematopathologists (N-CdA, B.G-F, O.B and E.C) and classified according to the revised 4th World Health Organization (WHO) classification 2017 criteria.²⁸ Available clinical data, details on transplant and mPTLD management were retrospectively collected (**Supplemental Table 1**).

All formalin-fixed and paraffin-embedded (FFPE) samples investigated contained more than 60% of neoplastic cells. DNA and RNA from FFPE materials were extracted using Qiagen AllPrep DNA/RNA FFPE kit (Qiagen, Hilden, Germany). Polymerase chain reaction (PCR) amplifications for clonal IGH chain gene rearrangements detection were performed according to

BIOMED-2 protocols.²⁹ This study was approved by the Hospital Clinic of Barcelona's Review Board (HCB/2018/0365) and was performed in accordance with the Declaration of Helsinki.

Immunohistochemical studies, EBV infection and cell of origin (COO) determination

Phenotypical profile was studied using standard immunohistochemistry (IHC) protocols on an automated platform (Ventana BenchmarkUltra, Roche, Switzerland). EBV infection, latency pattern and replicative phase were determined by *in situ* hybridization (EBER) and IHC (LMP-1, EBNA-2, ZEBRA) (**Supplemental Methods**; **Supplemental Table 2**). Cases were classified as GC and non-GC subtypes according to Hans algorithm.³⁰ Additionally, COO classification was performed using Lymph2Cx assay (NanoString Technologies, Seattle, WA).³¹

Fluorescence in situ hybridization (FISH)

FISH analyses were performed using standard protocols. Breaks at *BCL2*, *PAX5*, *BCL6*, *MYC*, IGK, *IRF4* loci, t(8;14) and t(2;8) fusions, and 11q alterations were analyzed by commercial (Metasystems, Altlußheim, Germany; Empire Genomics, Williamsville, NY; Agilent Technologies, Santa Clara, CA; Abbot, Chicago, IL; ZytoVision, Bremerhaven, Germany) or home-made FISH probes.³²

DNA CN alterations (CNAs) analysis

CNAs were examined in 16 PTLD-DLBCL cases and 7 PTLD-BL, using OncoScan array (Thermo Fisher Scientific, Waltham, MA) following standard

protocols (**Supplemental Methods**). Gains, losses and CN neutral-loss of heterozygosity (CNN-LOH) regions were evaluated using Nexus Biodiscovery v9.0 software (Biodiscovery, Hawthorne, CA). Previously published CN data of pediatric IMC-BL,³³ IMC-DLBCL,⁹ and adult PTLD-DLBCL^{21,34} were used for comparison.

Targeted NGS and mutational analysis

Samples from 25 mPTLD patients, with material available, were included in the mutational analysis. The custom NGS panel interrogated 167 B-cell lymphoma related genes (**Supplemental Table 3**), including those needed to perform *LymphGen* prediction algorithm.³⁵ SureSelectXT Target Enrichment System Capture NGS strategy library (Agilent Technologies, Santa Clara, CA) was used before sequencing in a MiSeq equipment (Illumina, San Diego, CA) (**Supplemental Methods**). Pathway enrichment analysis was performed defining the contribution of each gene based on previous literature (**Supplemental Methods**).³⁶ Previously published mutational profiles of adult PTLD-DLBCL,^{17,18} pediatric and young adult IMC-DLBCL,⁹ pediatric sporadic and endemic BL,^{6,11–13} were used for comparisons.

Statistical methods

Disease-free survival (DFS) was defined as time from diagnosis until lymphoma disease related death and event-free survival (EFS) was established as time to progression, death of any cause, organ loss or secondary malignancy. The Kaplan–Meier method was used to estimate the DFS and EFS distributions, whereas differences were assessed by the log-rank test.³⁷ Differences in the distribution of individual parameters among patient subsets were analyzed by

Fisher's exact test for categorized variables, and the Student's t-test for continuous variables. Non-parametric tests were applied when necessary. The *P*-values for multiple comparisons were adjusted using the Benjamini–Hochberg correction (false discovery rate, FDR). A *P*=0.05 cut-off was considered significant unless otherwise indicated. Statistical analyses were carried out using R software v4.1.2.

RESULTS

Clinical features

From the 31 recruited patients, eighteen were male and 13, female. The mean age at the time of lymphoma diagnosis was 8.7 years (range 2-17). Kidney and liver were the most frequently transplanted organs (11 and 10 patients, respectively), followed by heart (8 patients), intestine (2 cases) and lung (1 case). Twenty-four cases presented with extranodal disease, and the most frequently affected site was the gastrointestinal tract (21/29)(**Supplemental Figure 3**). Five patients had Stage IV disease (4 had bone marrow involvement, and one case had both central nervous system and bone marrow infiltration). The mean time from transplant to PTLD diagnosis was 35 months (range 2-170). Fifteen out of 31 patients (48%) had early-onset PTLD diagnosed in the first-year post-transplant, while only 6% (2/31) had very late-onset PTLD (≥10 years). Clinicopathological features are detailed in **Table 1** and **Supplemental Table 1**.

At PTLD diagnosis, EBV was detectable in blood in 23 out of 27 patients with available information, and PTLD was secondary to EBV primary infection in 47% (7/15) of the patients with known seronegativity before transplant.

Pathological characteristics

Individual description of clinicopathological features, COO and EBV latency pattern of mPTLD patients is detailed in **Supplemental Table 4**.

Twenty-four cases were classified as DLBCL, and seven as BL. Of note, two BL had less monotonous cytology (p28 and p29) (**Supplemental Figure 1**) and one BL (p38) (**Supplemental Figure 2**) showed DLBCL morphology. However, all 3 cases harbored typical phenotypic and molecular features and were, therefore, classified as BL. IGH gene rearrangements analysis identified a clonal peak in 23 out of 24 mPTLD. In case p27 IGH-FR3 was polyclonal, but no further analyses of IGH-FR1 and IGH-FR2 could be performed due to DNA quality.

Most DLBCL cases showed a non-GC phenotype (18/19, 95%), including four DLBCL with plasmacytic differentiation, in line with Lymph2Cx results, that showed an ABC signature in 71% (15/21), followed by 19% unclassified and 10% GCB. The seven BL were classified as GCB by either one of both methods.

EBER was positive in 28 out of 30 cases. A complete EBV latency pattern was determined in 26 cases. All seven BL were EBV+, and all 6 studied cases had a latency pattern I, whereas 21/23 DLBCL were EBV+, 60% of which had latency III. Viral replication was identified in six out of 16 analyzed cases, 5 of them among the latency pattern III group (**Table 1**).

FISH results

MYC rearrangements were detected in all seven BL. Six cases carried t(8;14)(q24;q32) translocation confirmed by IGH::*MYC* dual fusion FISH probe whereas t(2;8)(p11.2;q24) rearrangement was identified in case p28 using IGK::*MYC* dual fusion probe. Furthermore, no 11q alterations or chromosomal rearrangements involving *IRF4*, *PAX5*, *BCL2* or *BCL6* were observed in the 29, 29, 29, 13 and 11 analyzed cases, respectively (**Supplemental Figure 3**).

Copy number alteration profile

Eleven out of the 23 pediatric mPTLD studied displayed CNAs (mean 1.5 alt/case; range 0-7) (**Figure 2A**; **Supplemental Table 5**). Seven out of the 16 (44%) PTLD-DLBCL showed CNAs (mean 1.8 alt/case), with gains of 3p and 9q21.11-q34.3 (3 cases each; 19%) being the most frequent. Four out of seven (57%) PTLD-BL displayed CNAs (mean 0.9 alt/case) with no common CNA among those cases. None of the 25 investigated cases showed the characteristic 11g alteration found in LBCL/HG-11g.

Additionally, we compared those CN profiles to the ones observed in IMC patients. Of note, pediatric PTLD-BL had lower genetic complexity than IMC-sBL³³ (0.9 alt/case vs 6.3 alt/case; *P*<0.005) and lacked the 1q23.2-q25.3 gains characteristic of BL (**Figure 2B**). Similarly, PTLD-DLBCL had less CNA than IMC-DLBCL⁹ (1.8 alt/case vs 4.4 alt/case; *P*<0.05) (**Figure 2C** and **Figure 3A**) and adult PTLD-DLBCL^{21,34} (**Figure 3B**), with absence of CNAs frequently observed in IMC-DLBCL⁹ such as telomeric 1q31.3-q42.13 gains and losses of 6q21 and 19p13.3 (**Figure 2C**).

Identification of mutational profiles by targeted NGS

Eighteen DLBCL and 7 BL samples were analyzed by NGS (mean coverage 2586x, range 402-7256x). After filtering, a total of 187 somatic variants were detected in 23 out of 25 (range 0-28) mPTLD cases, 127 of them predicted as drivers (mean 5.5 driver mutation/case)(**Supplemental Methods; Supplemental Table 6**).

The mutational burden of PTLD-BL was higher than PTLD-DLBCL (7.8 vs 4.4 mutations, *P*=0.03). All 7 PTLD-BL cases carried *MYC* mutations. *ID3* and *DDX3X* variants were seen in four, *FOXO1* in three, *ARID1A* in two, and *CCND3* in one case (**Figure 4**). Four cases harbored exonic *MYC* driver mutations, three of which carried further multiple intronic mutations. Three cases had only multiple intronic mutations.

Comparison with previously published series of EBV+ and EBV- BL, both sporadic and endemic subtypes,^{11–13} revealed that PTLD-BL display a mutational frequency closer to EBV+ BL, with lower incidence of *TCF3*, *CCND3*, *TP53* and *SMARCA4* mutations and higher of *ARID1A* and *FOXO1* (**Supplemental Figure 4A**). Furthermore, the recently defined IC-BL (*ID3/CCND3*) and DGG-BL (*DDX3X/GNA13/GNAI2*) molecular genetic subgroups seem to be represented in our PTLD-BL cohort (4 and 3 cases, respectively) (**Supplemental Figure 4B**).¹⁵

Differently, the mutational profile of the 18 PTLD-DLBCL was very heterogeneous with lower complexity than their counterparts in IMC patients,⁹ when 94 commonly investigated genes were considered (2.4 vs 4.6 mutation/case, P=0.2) (**Figure 3C**; **Supplemental Table 7**). The most recurrently mutated gene was *KMT2C* in 3 cases (17%), followed by mutations

in *SPTNB5*, *TP53*, *NOTCH1*, *BCOR*, *CARD11*, *CD70*, *FAS*, *IL16* and *TP73* (2 cases each; 11%). The three *KMT2C* mutations were spread throughout the coding region of the gene (exons 36 and 43), while three *NOTCH1* mutations were detected in two cases, both carrying a mutation predicted to truncate the protein in its PEST domain. Of note, PTLD-DLBCL lacked the characteristic mutations of *SOCS1*, *MYD88*, *PIM1*, *BTG1*, *EZH2* and *PRDM1* genes observed in pediatric and young adult IMC-DLBCL (**Supplemental Table 7**).⁹

Pathway enrichment analysis showed that, in line with IMC-DLBCL, genes involved in epigenetic modifications were one of the most recurrently affected (5 cases). On the other hand, and differently to IMC-DLBCL, PTLD-DLBCL carried frequent mutations in Notch and MAPK pathways (5 and 2 cases, respectively), whereas lacked alterations affecting JAK-STAT pathway (**Supplemental Figure 5**). In order to discern whether these PTLD-DLBCL could belong to previously defined genetic subtypes of DLBCL,³⁵ *LymphGen* algorithm was applied to 19 PTLD-DLBCL. Only two cases (11%) were classified as N1, while the rest remained undetermined. These data suggest that this group of pediatric PTLD-DLBCL, mainly EBV+ (91%), do not fit in any of the established genetic DLBCL subtypes.

Moreover, comparison with adult PTLD-DLBCL^{17,18} showed that pediatric PTLD-DLBCL lack mutations in *KMT2D* (30% vs 0%, *P*=0.01), *PIK3CD*, *MYD88*, *PIM1*, *BTG1* and *EZH2* genes, and have lower incidence of *TP53* mutations (11% vs 32%, *P*=0.11)(**Figure 3D**; **Supplemental Table 7**).

Clinical outcome and prognostic value of clinical and molecular features

Four out of the 7 PTLD-BL (57%) presented with Stage IV. All of them underwent RIS and received chemotherapy according to the SOC for B-NHL. All seven patients were alive and in complete remission at last follow up (mean follow up 8.7 years; range 2-22 years) (**Table 1; Supplemental Table 1**).

Different to PTLD-BL, treatment strategy in PTLD-DLBCL was more heterogeneous. In detail, 7 out of 24 cases were initially treated following firstline B-NHL protocols. The remaining cases received a less aggressive regimen, mainly consisting of RIS (12 cases) and/or rituximab (12 cases). In five patients, LDCT was added (**Supplemental Table 1**).

The 5-year EFS (5y-EFS) rate was 73% in the whole cohort. Seven patients died, five of them related to lymphoma (5y-DFS 84%; **Figure 5A**). Of note, all deceased patients were PTLD-DLBCL and three of them with clinical information available, were at advanced stage (III-IV) at diagnosis. Only one out of 14 PTLD-DLBCL patients treated with less aggressive therapy (RIS, rituximab and/or LDCT) deceased because of lymphoma (5y-DFS 93%). This survival rate is in line with the one observed in patients who received intense chemotherapy (6/7 patients, 86%)(**Figure 5B**). Two of the patients died of F-PTLD before any therapy could be initiated, and there was no information available on the treatment received by the remaining case.

Of note, among PTLD-DLBCL patients, both, presence of mutations affecting cell cycle signaling (*CCND3* and *TP53*)(5y-DFS 33% vs 93%; *P*=0.008) (**Figure 5C**) and alterations of Notch pathway (*NOTCH1*, *FBXW7* and *SGK1*) (5y-DFS 50% vs 93%; *P*=0.043) (**Figure 5D**), were associated with a worse outcome.

DISCUSSION

PTLD is a major complication in recipients of both SOT and hematopoietic stem cell transplantations and occurs in up to 20% of patients depending on the series.^{5,38} In this study, we report for the first time an integrative genetic and molecular characterization of a large series of mPTLD in children and adolescents who received a SOT. Our results show relevant differences in the molecular landscape of these tumors with both PTLD in adults and DLBCL in IMC pediatric patients. Moreover, we show different molecular and clinical features between the 24 PTLD-DLBCL and 7 PTLD-BL included in the study, stressing the need for an adequate morphological and molecular diagnosis.

In detail, all seven PTLD-BL cases had a *MYC* rearrangement and, unlike PTLD-DLBCL, had a GCB phenotype and EBV latency pattern I, fully resembling IMC-BL. However, morphology ranged from typical BL cytology to less monotonous cytology or even DLBCL morphology. Nevertheless, all cases harbored typical phenotypic and molecular features of BL, highlighting the relevance of latency and COO determination in those cases. Additionally, all 7 cases depicted the mutational profile described in IMC-BL patients. Specifically, *MYC* mutations, either exonic or multiple intronic, were detected in all BL samples, followed by frequent mutations in *ID3* and *DDX3X* (4/7 cases each; 57%) and *ARID1A* (2/7 cases; 29%), with all 7 BL patients harboring, at least, one of these mutations. Our PTLD-BL also displayed lower genetic complexity than IMC-BL,³³ with absence of the characteristic 1q gains, which could be related to the EBV infection, as previously reported.¹⁵

Prior studies have observed differences in the mutational profile between EBVand EBV+ BL, the latter being associated with recurrent mutations in *ARID1A* and *RHOA* and lower incidence of mutations in *ID3*, *CCND3* or *SMARCA4*.^{11–}

^{13,15} Our 7 PTLD-BL were EBV positive, an expected higher frequency than that reported in sporadic and HIV related BL,³⁹ and similar to eBL.⁴⁰ All six investigated PTLD-BL had an EBV latency pattern I, typically seen in BL,^{12,41} unlike other types of PTLD, where latency type III is the most prevalent.^{3,42} We have also compared the mutational profile of previously published EBV+ and EBV- BL cohorts^{11–13} with our cases having a mutational distribution closer to EBV+ BL.^{4,5} Similarly, our cases seem to belong to the recently defined molecular subgroups highly represented in a large series of EBV+ BL tumors (DGG-BL and IC-BL) by whole-exome sequencing.¹⁵ Despite the limited number of our PTLD-BL series, our data also confirm the lower mutational burden in IC-BL-like cases than those with DGG-BL characteristics (6 vs 13 mutations/case) and the overrepresentation of male patients within DDG-BL.

Clinically, PTLD-BL showed a longer interval between transplant and lymphoma onset compared to PTLD-DLBCL (**Table 1**) and had a more aggressive presentation (4/7 were at Stage IV) than PTLD-DLBCL (1/24). All PTLD-BL were alive and in complete remission at last follow-up after treatment with RIS and chemotherapy following SOC for mature B-cell NHL (rituximab included in 4/7).

In this line, previous studies have highlighted that PTLD-BL required chemotherapy in most cases.^{26,27,43} In detail, a phase II trial proved the use of LDCT combined with rituximab to be effective in pediatric mPTLD after SOT, with a 2y-OS and EFS of 90% and 76%, respectively.²⁶ In this study from Gross *et al,* 4 out of 5 PTLD-BL were long-term survivors. However, *MYC* status was not known in all cases. Another non-randomized prospective multicentre trial studied a response-adapted sequential immuno-chemotherapy in PTLD after

SOT observing a 2y-OS and EFS of 86% and 67%, respectively. In half of the patients, chemotherapy could be avoided after responding to rituximab induction. Of note, in this study, 86% of PTLD-BL cases required chemotherapy.²⁷ All these observations claimed that PTLD-BL should be considered as an independent entity and would benefit from intensive therapeutic regimes, therefore, its distinction from other mPTLD is relevant.

On the other hand, our data show that the genetic features of pediatric PTLD-DLBCL are very heterogeneous. Latency pattern III was the most common among PTLD-DLBCL, though patterns II and I were also observed in 30% and 10% of the cases respectively, in line with previous literature on mPTLD.³ Seven out of the 16 samples evaluated showed CNAs, with the most common alterations being gains of 3p and 9q21.11-q34.3. This CN landscape lacks characteristic alterations specific of pediatric and young adult DLBCL in IMC cases such as 1g31.3-g42.13 gains and losses of 6g21 and 19p13.3;⁹ and it is also different from the one observed in adult PTLD-DLBCL, with absence of most CNAs reported in those cases.^{21,34} The mutational profile was very diverse as well, with *KMT2C* being the most recurrently mutated gene (3 cases; 17%), followed by mutations in NOTCH1 or TP53 genes (2 cases each; 11%), among others. In terms of global mutational burden, PTLD-DLBCL showed a lower mutational load than IMC counterparts, lacking typically found SOCS1 mutations, and the profile of affected pathways was enriched with alterations in epigenetic modifiers and Notch signaling. EBNA-2 has been described to use Notch pathway to immortalize B cells.⁴⁴ In our series, none of the ten EBNA-2 positive cases with mutational information harbored mutations in Notch pathway, while 40% (2/5) of the EBV+ PTLD-DLBCL, EBNA-2 negative had

mutations in Notch pathway, supporting an alternative role for EBNA-2 and Notch in the oncogenesis of PTLD-DLBCL.⁴⁵

The mutational landscape of pediatric PTLD-DLBCL was different to the one reported in adult PTLD-DLBCL, either EBV+ or EBV- cases, with lack of mutations in *KMT2D* and *PIK3CD* and lower frequency of *TP53* mutations. This lack of recurrent genomic alterations in pediatric PTLD-DLBCL sheds light into the biology of these tumors, although, in order to confirm these observations, analyses in independent series by, e.g., whole-exome or whole-genome approaches need to be performed.

Regarding clinical aspects, in our series, the PTLD-DLBCL patients that were treated with less intensive schemes (14/24; 58%) had a 5y-DFS of 93%, and there was only one lymphoma related death. Furthermore, among the seven patients treated with B-NHL specific strategies (5y-DFS 86%), there was one toxic death, stressing the vulnerability of this fragile population and the need of adapting treatments to reduce toxicities.

In summary, the different genetic background of pediatric PTLD-DLBCL in comparison with adult PTLD-DLBCL and IMC children and young adults, and the response to different treatment strategies, suggest that perhaps less intensive schemes with only RIS and rituximab or LDCT, could be a therapeutic option in selected patients. Additionally, correlation analyses between molecular features and clinical outcome revealed that alterations in the cell cycle signaling and Notch pathway were associated with a worse outcome. The real impact of these alterations as potential biomarkers needs to be further investigated.

Moreover, previous studies in adult PTLD have reported association of EBV latency pattern III and replication with a significantly shorter survival and early PTLD onset.³ In pediatric mPTLD, patients with EBV latency III and viral replication presented a shorter time from transplant to PTLD diagnosis (8.2 vs 47.6 months; P=0.004) and a tendency towards a worse prognosis (2y-DFS 60 vs 82%; P=0.33) (**Supplemental Figure 6B**). Although EBV latency pattern is currently not available for clinicians, its inclusion in the design of future therapeutic strategies should be considered.

The incidence of LBCL/HG-11q and LBCL-*IRF4* in pediatric mPTLD had not been previously investigated. Remarkably, Ferreiro *et al*⁴⁶ described an overrepresentation of LBCL/HG-11q in a subset of molecularly defined adult BL in the post-transplant setting. Interestingly, in our pediatric mPTLD, no 11q aberrations were identified by FISH and/or CN arrays. Similarly, although 20 mPTLD with DLBCL diagnosis expressed IRF4/MUM1, none of the 23 investigated cases displayed *IRF4* rearrangements. These results suggest the rarity of LBCL/HG-11q and LBCL-*IRF4* in pediatric mPTLD, although larger series of cases should validate these observations. Also, *PAX5* alterations, previously reported to occur in B-NHL in the post-transplant setting and associated with unfavorable outcome,^{3,47,48} were absent in pediatric mPTLD, suggesting that *PAX5* is not a key gene related to lymphomagenesis in these patients.

Altogether, we performed a multidisciplinary genome-wide analysis of a large series of pediatric SOT-associated mPTLD for the first time, observing distinct molecular profiles among PTLD-BL and PTLD-DLBCL. Additionally, PTLD-BL

showed similarities with EBV+ IMC-BL whereas PTLD-DLBCL was genetically different than adult PTLD and IMC-DLBCL counterpart.

Moreover, although the low number of patients included and the diverse therapeutic approaches in our series is a limitation, our analyses support previous evidence stating that PTLD-DLBCL in SOT recipients can respond to less intensive treatment.^{26,27} Further prospective studies should be undertaken to corroborate our findings and help in defining more personalized therapeutic strategies for these patients.

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AUTHOR CONTRIBUTION

J.S-V performed research, analyzed data and wrote the manuscript; N. C-dA reviewed and interpreted pathological data, analyzed data and wrote the manuscript. P.G-G reviewed and interpreted clinical data, analyzed data and wrote the manuscript. J-E. R-Z, M. L-G, S.M., D.C., N.G., and I.M-G performed research and analyzed data. F. D-C, J.M., M.G., E. G-F, M. LL, G.F., B.G-F, and E.C. reviewed and interpreted pathological data. M.A., C.G-C, I.A., A.F., J.V-A., S.G-M, B.G., V.C. reviewed and interpreted clinical data. O.B. performed morphological diagnosis, analyzed data and wrote the manuscript. I.S. performed research, analyzed data, designed research and wrote the manuscript. All authors approved the final manuscript.

CONFLICT OF INTEREST DISCLOSURES

The authors declare no conflict of interest.

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Table 1. Clinicopathological characteristics of 31 pediatric B-cell monomorphic

	PTLD-DLBCL, n= 24	PTLD-BL, n = 7	Total, n = 31
Mean age (range, years old)	8 (2-17)	12 (6-16)	9 (2-17)
Male	13/24 (54%)	5/7 (71%)	18/31 (58%)
Female	11/24 (46%)	2/7 (29%)	13/31 (42%)
Localization			
Extranodal involvement	17/23 (74%)	6/7 (86%)	23/30 (77%)
Gastrointestinal tract	17/23 (74%)	4/6 (67%)	21/29 (72%)
Stage			
Stage I	1/19 (5%)	1/7 (14%)	2/26 (8%)
Stage II	3/19 (16%)	0/7 (0%)	3/26 (11%)
Stage III	14/19 (74%)	2/7 (29%)	16/26 (62%)
Stage IV	1/19 (5%)	4/7 (57%)	5/26 (19%)
Cell of origin (NanoString and/or Hans algorithm) ^{&}			
GCB	2/22 (9%)	7/7 (100%)	9/29 (31%)
ABC/non-GCB	20/22 (91%)	0/7 (0%)	20/29 (69%)
EBER Hybridization positive	21/23 (91%)	7/7 (100%)	28/30 (93%)
EBV replication	6/11 (55%)	0/5 (0%)	6/16 (38%)
Latency pattern			
Latency I	2/20 (10%)	6/6 (100%)	8/26 (31%)
Latency II	6/20 (30%)	0/6 (0%)	6/26 (23%)
Latency III*	12/20 (60%)	0/6 (0%)	12/26 (46%)
Mean time from transplant to PTLD diagnosis (range, months)	24 (2-141)	74.4 (32-170)	35 (2-170)
Early-onset PTLD	15/24 (63%)	0/7 (0%)	15/31 (48%)
Alive with no evidence of disease	17/24 (71%)	7/7 (100%)	24/31 (77%)
Died of disease	5/24 (21%)	0/7 (0%)	5/31 (16%)
Fulminant-PTLD	3/5 (60%)	0/0 (0%)	3/5 (60%)
Follow up (median)	1.3 years	5.2 years	2.7 years
(range)	(10 days – 14.1 years)	(2.1 – 22.6 years)	(10 days – 22.6 years)
5y-DFS	78.9%	100%	83.7%

PTLD patients

ABC: activated B cell; GCB, germinal center B cell; EBER: Epstein-Barr virus-encoded small RNAs; EBV, Epstein-Barr Virus; DFS: disease-free survival. *In the absence of LMP-1 assessability, two cases with latency pattern IIb-III are included in latency pattern III group.

[&] Unclassified cases by NanoString analysis were classified according to Hans algorithm results.

FIGURE LEGENDS

Figure 1. Morphological, immunohistochemical and genetic features of two prototypical post-SOT mPTLD, Burkitt lymphoma (BL) (case p11) and diffuse large B-cell lymphoma (DLBCL) (case p10), respectively. Case p11: (A) H&E, original magnification at 400x. (B) Tumor cells are diffusely positive for CD10 (Immunostain, original magnification at 400x). (C) In situ hybridization for EBV RNA is positive in tumor cells (*In situ* hybridization, original magnification at 400x). (D-F) Neoplastic cells are negative for LMP1 (D), EBNA2 (E) and Zebra (F) immunostains, indicating a latency pattern I (Immunostains, original magnification at 400x). (G) FISH using MYC Break-Apart Probe (BAP) shows a signal constellation of one colocalized signal (yellow arrow) and two split signals (green and red arrows) in accordance with the gene rearrangement. **Case p10**: (H) DLBCL with focal plasmacytic differentiation, (H&E, original magnification at 400x). (I) Atypical cells are diffusely positive for MUM1 (Immunostain, original magnification at 400x). (J-L) LMP1 (J), EBNA2 (K) and ZEBRA (L) immunostains show positive tumor cells, reflecting a pattern III of latency (Immunostains, original magnification at 400x). (M-N) FISH using BAP for MYC (M) and *IRF4* (N) genes show a normal pattern with two colocalizations in each nucleus for both hybridizations (yellow arrows).

Figure 2. (A) Global CN profile of 23 B-cell post-SOT mPTLD. X-axis represents chromosomes from 1 to Y and p to q. The Y-axis indicates frequency of each genomic alteration among the analyzed cases; CN gains are represented in blue and CN losses are depicted in red. Chromosomal bands of regions altered in more than 10% cases are indicated in the plot. (B) Comparative plot of CN profile comparison with previously published data on

BL³³ and **(C)** DLBCL⁹ in IMC patients. Asterisks indicate significant differences between both groups according to Fisher's exact test raw P<0.1, considering a minimum number of altered cases n=3.

Figure 3. Comparison between pediatric PTLD-DLBCL, pediatric and young adult IMC-DLBCL⁹ and adult PTLD-DLBCL^{17,18,21,34} in terms of (A-B) copy number alterations (CNA) and (C-D) mutational frequencies. The vertical axis of (A) and (C) represents number of alterations and the two groups are separated in the X-axis (PTLD-DLBCL represented in purple and IMC-DLBCL represented in blue). Asterisks in (A) mark significant differences between both groups according to Wilcoxon rank-sum test *P*<0.05. The vertical axis of (B) and (D) represented in the horizontal axis. Asterisks in (B) mark significant differences between adult and pediatric PTLD-DLBCL, according to Fisher's exact test *P*-<0.1 and in (D) indicate significant differences between adult, either EBV+ or negative, and pediatric PTLD-DLBCL, according to Fisher's exact test * *P*<0.1 and ** *P*<0.05.

Figure 4. Molecular CN and mutational information on 25 B-cell post-SOT mPTLD. Each column corresponds to a case, the top histogram depicts number of CN alteration (CNA) and each row of the bottom plot represents a gene, where dark blue color marks driver mutation. Only genes with driver mutation in more than two cases are represented. Blue in *MYC*-R row indicates *MYC* rearrangement was detected by FISH using *MYC* BAP and, in cases p11, p15, p22, p29, p38 and p42, also using IGH::*MYC* DF probe whereas in case p28, IGK::*MYC* DF probe was used. NA: Not available; DLBCL: diffuse large B-cell lymphoma, BL: Burkitt lymphoma; COO: cell of origin; GCB: germinal center B-

cell, ABC: activated B-cell; RIS: reduction of immunosuppression; LDCT: lowdose chemotherapy: SOC-BNHL: standard-of-care for B-NHL.

Figure 5. (A) Event-free survival (EFS) and disease-free survival (DFS) probabilities within the complete cohort. (B) DFS probability of PTLD-DLBCL patients treated with reduction of immunosuppression (RIS) and/or rituximab (R) and/or low-dose chemotherapy (LDCT) compared to that of cases treated with standard-of-care for B-NHL (SOC-BNHL). (C) DFS probability of PTLD-DLBCL patients according to mutations affecting cell cycle signaling and (D) Notch pathway.




Figure 2



Figure 3







Supplemental Material

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Supplemental Methods

EBV latency pattern immunohistochemical evaluation

The latency pattern was assessed using EBNA-2 and LMP-1 antibodies available in our lab (Supplemental Table 2) and established according to the study from Murata T et al, Viruses 2021.¹

In detail, when EBER was positive, and both EBNA-2 and LMP-1 were negative, the case was considered to show a latency pattern I, whereas when both markers were positive it was regarded as a latency pattern III. A case showing EBNA-2 positivity and LMP-1 negativity was considered a latency phase IIb, conversely, EBNA-2 negativity and LMP-1 positivity phenotype was regarded as a latency pattern IIa.

COO determination according to Lymph2Cx assay (NanoString, Seattle, WA, USA)

Four hundred nanograms of total RNA from FFPE samples were processed using Gene Expression Lymph2Cx NanoString Assay. First, Probe Mixes A and B and hybridization master mix (including the TagSet) were prepared and split into the 12 tubes containing the samples to analyze. Samples were then incubated in a PCR machine, for 16 hours at 67°C. In the next morning, on the deck of the NanoString nCounter Prep Station, purified Target/Probe complexes are eluted off and are immobilized in the cartridge for data collection on the nCounter Digital Analyzer. This instrument collects data, yielding hundreds of thousands of target molecule counts, using a microscope objective and a CCD camera. Digital images are processed on the nCounter Digital Analyzer and the barcode counts are tabulated in a comma separated value (CSV) format, which can be downloaded via a memory stick.

Library preparation SureSelectXT and Targeted sequencing approach

A total of 25 formalin fixed paraffin embedded (FFPE) samples were processed using SureSelectXT (Agilent Technologies, Santa Clara, CA) a custom panel interrogating 167 genes, that was designed according to previous literature in DLBCL and other B-cell lymphomas in both adult and children population (**Supplemental Table 3**), including those needed to perform *LymphGen* prediction algorithm.² A total of 100ng of genomic DNA were sheared using the Covaris S220 focused-ultra sonicator (Covaris, Woburn, MA) to a target peak size of 150–200 bp. Library preparation was performed using SureSelectXT Custom Capture Library baits as described in SureSelectXT Target Enrichment System protocol (Agilent Technologies, Santa Clara, CA). For amplification of the post capture libraries, 10 to 13 cycles were performed depending on the initial sample quality. The libraries were qualified using the Bioanalyzer HS (Agilent Technologies, Santa Clara, CA), quantified with the KAPA Library Quantification Kit (Kapa Biosystems, Wilmington, Massachusetts) and sequenced in a MiSeq instrument (Illumina, San Diego, CA) in a paired-end run of 150 bp. The average sequencing coverage of the across regions was 2554x (range 255-7421x) and over 84% of the targeted regions were covered by at least 100x.

FASTQ files were generated by MiSeq control software and quality control of the raw data performed the FastQC tool was using (https://www.bioinformatics.babraham.ac.uk/projects/fastqc/). Sequencing reads were subsequently aligned to the human reference genome (GRCh37/hg19) using the Burrows-Wheeler Aligner–MEM algorithm.³ Variant calling was performed using two different variant callers, Somatic Variant Caller (Illumina, San Diego, CA) and Mutect2 (Genome Analysis Toolkit (GATK), version 4.0.3)⁴ and variants were annotated using the VariantStudio software v3.0 and ANNOVAR, respectively.⁵ For Somatic Variant Caller (Illumina, San Diego, CA), default settings were used to analyze sequencing results and to call the variants. Low quality or low coverage calls (total depth <20) were excluded. For Mutect2 variants, low quality variants were also excluded using FilterMutectCalls (GATK) with default thresholds. Only variants identified by both algorithms were considered. We excluded variants affecting non-interrogated regions and known polymorphisms described in the GnomAD, 1000 Genomes and/or ExAC03 database (latest version included in ANNOVAR) with more than 0.1% frequency in normal population. In order to exclude

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artifacts, each variant was also inspected with the Integrative Genomics Viewer (IGV, Broad Institute, version 2.3) software (**Supplemental Figure 2**). Variants detected in *BRCA2* were specifically consulted in ClinVar database (<u>https://www.ncbi.nlm.nih.gov/clinvar/</u>), which reports relationships between human variations and phenotypes, regarding their clinical significance and they were excluded after confirmation of their benign effect. Variant detected in case p27 was additionally detected in DNA extracted from blood sample (data not shown).

Driver prediction by mutation effect

Since no germline DNA was available and in order to select somatic variants, potential driver mutations were predicted according to previously published criteria⁶ in which the 90% of the mutations classified as functional were demonstrated to be somatic. Inclusion criteria were: 1) variants described previously as somatic or functional on previous reports or COSMIC, 2) all truncating variants (nonsense, frameshift, splice donor or acceptor mutations; and 3) the remaining missense variants that were predicted to be functionally deleterious using Mutation Assessor⁷ or SIFT predictor if a score was not provided by Mutation Assessor.⁸ Other functional predictors as Polyphen-2 (Polymorphism Phenotyping-2)⁹, CADD (Combined Annotation Dependent Depletion)¹⁰ and CHASM-3.1¹¹ were also applied. In detail, there were 96 non-synonymous missense driver SNVs, 12 frameshift indels, 3 in-frame deletions, 3 splicing variants and 9 nonsense mutations.

Copy number analysis

DNAs from 23 samples were hybridized on an OncoScan array (ThermoFisher Scientific inc.). Gains and losses and copy number neutral-loss of heterozygosity (CNN-LOH) regions were evaluated and visually inspected using Nexus Biodiscovery version 9.0 software (Biodiscovery, Hawthorne, CA). Human reference genome was GRCh37/hg19. The copy number alterations (CNAs) with minimum size of 100 kb and CNN-LOH larger than 5 Mb

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were considered informative. Published CN data from pediatric IMC-BL¹² and IMC-DLBCL,¹³ adult PTLD-DLBCL^{14,15} were used for comparison.

Supplemental Figures

Supplemental Figure 1. Cases p28 and p29 showing an intermediate morphology between DLBCL and a BL, with *MYC* gene rearrangement. (A) Case p28 shows a quite monotonous proliferation of medium-sized lymphocytes, with a round-oval irregular nucleus, finely clumped chromatin in most of them, some demonstrating a small nucleoli, abundant mitosis figures and intermingled scattered body tangible macrophages (H&E, original magnification at 400x). (B) FISH with IGK::*MYC* dual fusion probe demonstrates the presence of fusion signals, consistent with the t(2;8) translocation. (C) Case p29 demonstrates a rather monotonous proliferation of medium-sized lymphocytes, with round irregular nucleus and finely clumped chromatin with conspicuous nucleoli. (D) FISH with *MYC* break-apart probe exhibits split signals, in accordance to the presence of the *MYC* gene rearrangement.



Supplemental Figure 2. Histopathological features of case p38. (A) H&E staining (original magnification 40x), showing DLBCL morphology. **(B)** CD20 staining (original magnification 10x). **(C)** CD10 expression (original magnification 20x). **(D)** EBV *in situ* hybridization showing positivity in virtually all tumor cells (original magnification 20x). **(E)** Negative LMP1 immunostaining (original magnification 20x). **(F)** Negative EBNA2 immunostaining (original magnification 20x); **(G)** KI-67 immunostaining (original magnification 20x). **(H)** FISH with *MYC* break-apart probe, showing split signals demonstrating the rearrangement (original magnification 100x). **(I)** FISH with IGH::*MYC* dual color dual fusion probe, showing fusion signals demonstrating the t(8;14) rearrangement (original magnification 100x).



Supplemental Figure 3. Results from FISH analysis performed on 31 post-SOT mPTLD to investigate the status of *MYC*, *PAX5*, *IRF4*, *IGH*, *BCL2* and *BCL6* loci as well as 11q region. # Marks case p27, with IGH-FR3 was polyclonal, but no further analyses of IGH-FR1 and IGH-FR2 could be performed due to DNA quality.



Supplemental Figure 4. (A) Comparison of mutational profile between pediatric PTLD-BL and previously published BL series, stratified according to EBV status.^{16–18} To perform statistical analysis using comparable data, only exonic *MYC* mutations in PTLD-BL were considered. **(B) Genetic landscape of seven PTLD-BL** representing mutations and CNA frequently associated to previously defined genetic subgroups.¹⁹



Supplemental Figure 5. **Pathway enrichment analysis** on PTLD-DLBCL and comparison with pediatric IMC-DLBCL.¹³ The horizontal axis represents frequency of pathway alteration (%) and each pathway is represented in the vertical axis. Asterisks mark significant differences between the two groups, according to Fisher's exact test P < 0.1.



Supplemental Figure 6. Disease-free survival (DFS) probability of post-SOT mPTLD stratified according to EBV latency pattern and replication information.



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Case	Age at DX (years), gender	Morphological diagnosis	Allograft	TX-PTLD (months)	EBV in blood	LDH high	Extranodal	BM/CNS	Stage*	RIS	Rituximab	Chemotherapy
p1	2, M	PTLD-DLBCL	Kidney	10	Pos	Yes	Yes	NA	NA	NA	NA	NA
р5	2, M	PTLD-DLBCL	Liver	6	Pos	NA	Yes	No	Ш	Yes	Yes	No
р6	2, F	PTLD-DLBCL	Liver	8	Pos	No	Yes	No	≡	Yes	Yes	No
p10	5, F	PTLD-DLBCL	Kidney	18	Pos	yes	Yes	No	III	NA	Yes	No
p11	16, M	PTLD-BL	Heart	88	Pos	Yes	Yes	Both	IV	Yes	Yes	SOC-BNHL ¹
p12	9, F	PTLD-DLBCL	Liver	73	Neg	NA	NA	NA	NA	NA	NA	NA
p13	4, M	PTLD-DLBCL	Heart	5	Pos	No	Yes	No	Ш	Yes	Yes	SOC-BNHL ¹
p14	3, M	PTLD-DLBCL	Intestine	2	Pos	No	Yes	NA	NA	Yes	Yes	No
p15	15, F	PTLD-BL	Liver	170	Neg	No	No	No	I	Yes	Yes	SOC-BNHL ²
p17	12, F	PTLD-DLBCL	Kidney	7	Pos	Yes	No	No		Yes	Yes	No
p19	4, F	PTLD-DLBCL	Liver	16	Pos	No	Yes	No	II	Yes	Yes	LDCT
p20	4, M	PTLD-DLBCL	Kidney	10	Pos	No	Yes	No	III	Yes	Yes	LDCT
p21	6, F	PTLD-DLBCL	Liver	10	Pos	NA	Yes	NA	NA	Yes	No	No
p22	6, F	PTLD-BL	Liver	35	Pos	No	Yes	BM	IV	Yes	Yes	SOC-BNHL ¹
p23	14, F	PTLD-DLBCL	Lung	8	Pos	Yes	Yes	NA	NA	Yes	Yes	LDCT
p25	17, M	PTLD-DLBCL	Kidney	6	Pos	No	No	No	Ш	Yes	Yes	No
p26	3, M	PTLD-DLBCL	Kidney	9	Pos	Yes	Yes	No	≡	No	Yes	No
p27	12, M	PTLD-DLBCL	Heart	141	Neg	No	Yes	No		Yes	No	SOC-BNHL ¹
p28	14, M	PTLD-BL	Heart	73	Pos	No	Yes	No	III	Yes	No	SOC-BNHL ²
p29	13, M	PTLD-BL	Heart	88	Pos	Yes	Yes	BM	IV	Yes	No	SOC-BNHL ²
p31	9, F	PTLD-DLBCL	Heart	16	Pos	No	Yes	No		Yes	No	SOC-BNHL ¹
p32	4, M	PILD-DLBCL	Kidney	6	Pos	No	No	No	I	Yes	No	No
p33	6, F	PTLD-DLBCL	Intestine	2	NA	NA	Yes	No	II	Yes	Yes	SOC-BNHL
p35	14, M	PTLD-DLBCL	Kidney	54	Neg	No	Yes	No	III	Yes	No	SOC-BNHL ⁴
p36	7, M	PTLD-DLBCL	Heart	90	NA	NA	Yes	No		NA	NA	NA
p37	13, M	PTLD-DLBCL	Liver and kidney	53	NA	No	Yes	No		Yes	Yes	SOC-BNHL'
p38	5, M	PTLD-BL	Liver	32	NA	NA	Yes	No	III	Yes	No	SOC-BNHL⁵
p39	17, F	PTLD-DLBCL	Heart	4	Pos	No	No	BM	IV	Yes	Yes	SOC-BNHL ¹
p41	8, F	PTLD-DLBCL	Liver	14	Pos	No	Yes	No	III	Yes	Yes	LDCT
p42	13, M	PTLD-BL	Kidney	35	Pos	Yes	Yes	BM	IV	Yes	Yes	SOC-BNHL ¹
p43	12, M	PTLD-DLBCL	Kidney	8	Pos	No	No	No	III	Yes	Yes	LDCT

Supplemental Table 1. Clinical features of 31 pediatric post-SOT mPTLD detailed per case

M, male; F, female; DX, diagnosis; PTLD, post-transplant lymphoproliferative disorder; TX-PTLD, months from transplant to PTLD; EBV, Epstein-Barr Virus; Pos, positive; Neg, negative; LDH, lactate dehydrogenase; BM, bone marrow immunosuppression; LDCT, low dose chemotherapy; SOC-BNHL, standard of care for mature B-cell non-Hodgkin lymphomas; CR, complete remission; PD, progressive disease; F-PTLD: fulminant PTLD; NA: not assessable. The LDI the reference laboratory of each hospital.

*Stage was established according Murphy or International pediatric NHL staging system (IPNHLSS)

†Patient p7 received COP followed by dexamethasone and bortezomib administration; Patient p24 received COP and GEM/STEROID/CYS/ICE SOC-BNHL: 1, Inter-B-NHL-2010; 2, Inter-B-NHL 2004; 3, SIOP-98; 4, SIOP-94 Cellular therapy: a, specific T cells against EBV; b, memory T cells

	Other	Outcome,	Causo of
Antiviral	treatment	Follow-up	death
	strategy	(months)	ueath
NA	NA	PD, 0	F-PTLD
Pharmacologic therapy	Surgery	CR, 169	
Pharmacologic			
therapy		CR, 101	
No		CR, 30	
Pharmacologic		OD 05	
therapy		UR, 20	
NA	NA	PD, 8	PD
Pharmacologic		CR 1	Toxicity
therapy			TOXICITY
Pharmacologic		CR. 13	Transplant
therapy			related
No		CR, 49	
No		CR, 13	
Pharmacologic therapy		CR, 147	
No		CR, 13	
Pharmacologic			
therapy		CR, 15	
Pharmacologic	Surgery	CR 62	
therapy	ourgery	013, 02	
Pharmacologic		PD. 0	F-PTLD
therapy		,	
Pharmacologic		CR, 32	
Inerapy Pharmacologic			
therany		CR, 33	
No		CR 37	
No		CR 144	
No	Surgery	CR 128	
No	Surgery	CR, 120	
NU	Surgery		
No	Surgery	CR, 4	
therapy	Surgery	CR, 86	
Pharmacologic			
therapy		CR, 136	
NA		PD. 0	F-PTLD
No		, CR, 88	
Pharmacologic	0	00.074	
therapy	Surgery	GR, 271	
Pharmacologic	Surgen	CR 51	Other
therapy	Surgery	UN, 91	
No		CR, 8	
No		CR, 57	
No		CR, 6	

r; CNS, central nervous system; RIS, reduction of H high levels were defined as above twice the normal value of

Antibody	Clone	Source	Antigen retrieval/visualization	Dilution
CD20	L26	DAKO, (Copenhagen, Denmark)	EDTA 1 mM pH 9/ ENVISION FLEX (DAKO)	RTU
CD79a	JCB 117	DAKO	EDTA 1 mM pH 9/ ENVISION FLEX (DAKO)	RTU
PAX-5	SP34	Roche	CC1 solution / ultraView Universal DAB Detection Kit. Automated immunostainer (Benchmark XT; Ventana)	RTU
CD3	Polyclonal	DAKO	EDTA 1 mM pH 9/ ENVISION FLEX (DAKO)	RTU
CD5	4C7	DAKO	EDTA 1 mM pH 9/ ENVISION FLEX (DAKO)	RTU
CD10	56C6	DAKO	EDTA 1 mM pH 9/ ENVISION FLEX (DAKO)	RTU
BCL6	PG-B6p	DAKO	EDTA 1 mM pH 9/ ENVISION FLEX (DAKO)	RTU
BCL2	124	DAKO	EDTA 1 mM pH 9/ ENVISION FLEX (DAKO)	RTU
Ki67	Mib-1	DAKO	Citrate 10 mM pH 6/ ENVISION FLEX (DAKO	RTU
MUM1	MRQ-43	Ventana, Roche (Oro Walley, AR, USA)	CC1 solution / ultraView Universal DAB Detection Kit. Automated immunostainer (Benchmark XT; Ventana)	RTU
MYC	Y69	Ventana, Roche	CC1 solution / ultraView Universal DAB Detection Kit. Automated immunostainer (Benchmark XT; Ventana)	RTU
CD21	EP3093	Ventana, Roche	CC1 solution / ultraView Universal DAB Detection Kit. Automated immunostainer (Benchmark XT; Ventana)	RTU
EBNA-2	PE2	Abcam	CC1 solution / ultraView Universal DAB Detection Kit. Automated immunostainer (Benchmark XT; Ventana)	0.09375
LMP-1	CS1-4	Roche	CC1 solution / ultraView Universal DAB Detection Kit. Automated immunostainer (Benchmark XT; Ventana)	RTU
ZEBRA		Argene	CC1 solution / ultraView Universal DAB Detection Kit. Automated immunostainer (Benchmark XT; Ventana)	1:100

Supplemental Table 2. Details of all antibodies used for immunohistochemical characterization, including source and conditions of use.

RTU, ready to use.

According to previous reports, MYC, BCL2,²¹ BCL6 and MUM1²² were considered positive when \geq 40%, \geq 50%, \geq 30% or \geq 30% of the cells were positive, respectively. Staining for EBNA-2 or LMP-1 were assessed as positive when any neoplastic cell showed nuclear or cytoplasmic/membranous positivity, respectively.²²

Gene	Transcript	Position (GRCh37/hg19)	Exons Pathway		
ABCA7	NM_019112.3	chr19:1041361-1065424	All CDS E2-47		
ACTB	NM_001101.3	chr7:5567379-5569288	All CDS E2-6		
APC	NM_000038.5	chr5:112090588-112179823	All CDS E2-16		
ARID1A	NM_006015.4	chr1:27022895-27107247	All CDS E1-20	Epigenome/Chromatin modifier	
ATM	NM_000051.3	chr11:108098352-108236235	All CDS E2-63		
B2M	NM_004048.2	chr15:4500374545008540	All CDS E1-3	Immune Response	
BCL10	NM_003921.4	chr1:85733310-85742035	All CDS E1-3	BCR-TLR signaling pathway	
BCL2	NM_000633.2	chr18:60795858-60985899	All CDS E1-2		
BCL6	NM_001706.4	chr3:187440245-187451481	All CDS E3-10	B-cell differentiation	
BCL7A	NM_020993.4	chr12:122459997-122497069	All CDS E1-6		
BLM	NM_000057.2	chr15:91290623-91358509	All CDS E2-22		
BRAF	NM_004333.4	chr7:140453075-140481493	E11-15	MAP-kinase pathway	
BRCA2	NM_000059.3	chr13:32890598-32972907	All CDS E2-27		
BTG1	NM_001731.2	chr12:92537856-92539311	All CDS E1-2		
BTG2	NM_006763.2	chr1:203274735-203276566	All CDS E1-2		
BTK	NM_000061.2	chrX:100604873-100630272	All CDS E2-19	BCR-TLR signaling pathway	
CARD11	NM_032415.4	chr7:2946272-2998140	All CDS E2-25	BCR-TLR signaling pathway	
CCND1	NM_053056	chr11:69456082-69466050	All CDS E1-5	Cell cycle	
CCND3	NM_001760.3	chr6:41903678-41909387	All CDS E1-5	Cell cycle	
CD58	NM_001779.2	chr1:117057435-117113594	All CDS E1-6	Immune Response	
CD70	NM_001252.3	chr19:6586031-6591013	All CDS E1-3		
CD79A	NM_001783.3	chr19:42381375-42385047	All CDS E1-5	BCR-TLR signaling pathway	
CD79B	NM_001039933.2	chr17:62006585-62009621	All CDS E1-6	BCR-TLR signaling pathway	
CIITA	NM_000246.3	chr16:10971188-11017160	All CDS E1-19	Immune Response	
CREBBP	NM_004380.2	chr16:3777719-3929917	All CDS E1-31	Epigenome/Chromatin modifier	
DDX3X	NM_001356.3	chrX:41193506-41206972	All CDS E1-17		
DIS3	NM_014953.3	chr13:73333933-73355970	All CDS E1-21		
EGR1	NM_001964.2	chr5:137801451-137803770	All CDS E1-2		
EP300	NM_001429.3	chr22:41489009-41574960	All CDS E1-31	Epigenome/Chromatin modifier	
ETS1	NM_001143820.1	chr11:128332256-128443025	All CDS E2-10	MAP-kinase pathway	

Supplemental Table 3. One-hundred sixty-seven genes included in the SureSelectXT custom panel used for the NGS analysis of pediatric post-SOT mPTLD cases and pathway in which they are involved.

ETV6	NM_001987.4	chr12:11803062-12043980	All CDS E1-8	
EZH2	NM_004456.4	chr7:148504738-148508812	E16-20	Epigenome/Chromatin modifier
FAM46C	NM_017709.3	chr1:118165491-118166666	All CDS E2	
FBXW7	NM_033632.3	chr4:153244033-153332955	All CDS E2-12	Notch pathway
FOXO1	NM_002015.3	chr13:41133660-41240349	All CDS E1-2	
GNA13	NM_006572.4	chr17:63010375-63052711	All CDS E1-4	PI3K-AKT-mTOR pathway
HDAC7	NM_015401.3	chr12:48177624-48213568	All CDS E1-26	
HIST1H1D	NM_005320.2	chr6:26234496-26235161	All CDS E1	Epigenome/Chromatin modifier
HIST1H1E	NM_005321.2	chr6:26156619-26157278	All CDS E1	Epigenome/Chromatin modifier
HIST1H2BC	NM_003526.2	chr6:26123752-26124132	All CDS E1	Epigenome/Chromatin modifier
HIST1H3B	NM_003537.3	chr6:26031878-26032288	All CDS E1	Epigenome/Chromatin modifier
ID3	NM_002167.4	chr1:23885451-23885917	All CDS E1-2	PI3K-AKT-mTOR pathway
IRF4	NM_002460.3	chr6:393153-407598	All CDS E2-9	B-cell differentiation
IRF8	NM_002163.2	chr16:85936622-85954888	All CDS E2-9	B-cell differentiation
KLHL6	NM_130446.2	chr3:183209715-183273441	All CDS E1-7	
KMT2D	NM_003482.3	chr12:49415563-49449107	All CDS E1-54	Epigenome/Chromatin modifier
KMT2C	NM_170606	chr7:151833918-152132871	All CDS E1-59	Epigenome/Chromatin modifier
KRAS	NM_033360.2	chr12:25368375-25398318	All CDS E2-5	MAP-kinase pathway
MAP2K1	NM_002755.3	chr15:66679686-66782953	All CDS E1-11	MAP-kinase pathway
MAPK1	NM_002745.4	chr22:22123493-22221730	All CDS E1-8	MAP-kinase pathway
MEF2B	NM_001145785.1	chr19:19256606-19261544	All CDS E2-9	Epigenome/Chromatin modifier
MYC	NM_002467.4	chr8:128748840-128753674	All CDS E1-3 + intron1	Cell cycle
MYD88	NM_002468.4	chr3:38180153-38182777	All CDS E1-5	BCR-TLR signaling pathway
MYOM2	NM_003970.2	chr8:1998881-2092905	All CDS E2-37	
NFKB1	NM_003998.3	chr4:103446676-103537751	All CDS E2-24	BCR-TLR signaling pathway
NFKBIE	NM_004556.2	chr6:44226956-44233500	All CDS E1-6	BCR-TLR signaling pathway
NOTCH1	NM_017617.3	chr9:139390023-139399556	E26-27, 34 + 3'UTR	Notch pathway
NOTCH2	NM_024408.3	chr1:120457929-120459317	E34	Notch pathway
NRAS	NM_002524.4	chr1:115251156-115258781	All CDS E2-5	MAP-kinase pathway
PCBP1	NM_006196	chr2:70314876-70315946	All CDS E1	
PDGFRA	NM_006206.4	chr4:55124936-55161439	All CDS E2-23	
PIK3CD	NM_005026.3	chr1:9770513-9787104	All CDS E3-24	PI3K-AKT-mTOR pathway
PIK3R1	NM_181523.2	chr5:67522504-67593429	All CDS E2-16	PI3K-AKT-mTOR pathway

PIM1	NM_001243186.1	chr6:37138079-37141867	All CDS E1-6	
POU2F2	NM_001207025.2	chr19:42595704-42636563	All CDS E1-14	
PRDM1	NM_001198.3	chr6:106534429-106555361	All CDS E1-7	B-cell differentiation
PRKCB	NM_002738.6	chr16:23847497-24226137	All CDS E1-17	BCR-TLR signaling pathway
PRKDC	NM_006904.6	chr8:48686734-48872686	All CDS E1-87	
PTPN1	NM_002827	chr20:49127065-49199252	AI CDS E1-10	
RB1	NM_000321	chr13:48878049-49054207	All CDS E1-27	Cell cycle
RELN	NM_005045.3	chr7:103113259-103629803	All CDS E1-65	
SEMA5A	NM_003966.2	chr5:9043008-9380058	All CDS E3-23	
SETD2	NM_014159.6	chr3:47058583-47205414	All CDS E1-21	
SGK1	NM_001143676.1	chr6:134491406-134638598	All CDS E1-14	Notch pathway
SMARCA4	NM_001128844.1	chr19:11132400-11144541	E20-28	
SOCS1	NM_003745.1	chr16:11348699-11349335	All CDS E2	JAK-STAT pathway
SP140	NM_007237	chr2:231090560-231177399	All CDS E1-27	
SPTBN5	NM_016642.2	chr15:42140813-42185695	All CDS E2-68	
STAT3	NM_139276.2	chr17:40467763-40500534	All CDS E2-24	JAK-STAT pathway
STAT6	NM_001178078.1	chr12:57490355-57502061	All CDS E2-22	JAK-STAT pathway
TAF1	NM_004606.3	chrX:70586165-70683896	All CDS E1-38	Epigenome/Chromatin modifier
TBL1XR1	NM_024665.4	chr3:176743285-176782765	All CDS E3-16	
TCF3	NM_003200.3	chr19:1611706-1650247	All CDS E2-19	PI3K-AKT-mTOR pathway
TET2	NM_001127208.2	chr4:106155099-106197676	All CDS E3-11	Epigenome/Chromatin modifier
TMEM30A	NM_018247.3	chr6:75965818-75994354	All CDS E1-7	
TNFAIP3	NM_006290.3	chr6:138192365-138202456	All CDS E2-9	
TNFRSF14	NM_003820.2	chr1:2488104-2494712	All CDS E1-8	
TNFSF9	NM_003811.3	chr19:6531048-6535077	All CDS E1-3	
TNIP1	NM_001252390.1	chr5:150410274-150444656	All CDS E2-18	
TP53	NM_000546.5	chr17:7572927-7579912	All CDS E2-11	Cell cycle
TRAF3	NM_003300.3	chr14:103336539-103372121	All CDS E2-11	
UNC5D	NM_080872.2	chr8:35093303-35648081	All CDS E1-17	
WHSC1	NM_001042424.2	chr4:1902382-1980636	All CDS E2-22	
XBP1	NM_005080	chr22:29191534-29196512	All CDS E1-5	
NFKBIA	NM_020529.2	chr14:35871219-35873850	All CDS E1-6	BCR-TLR signaling pathway
TRAF2	NM_021138.4	chr9:139793193-139820353	All CDS E2-11	

NFKBIZ	NM_031419.4	chr3:101568473-101578400	All CDS E1-12	BCR-TLR signaling pathway
ACTG1	NM_001614	chr17:79477716-79479380	All CDS E2-6	
ALDH18A1	NM_002860	chr10:97366519-97413134	All CDS E2-18	
ARID5B	NM_032199	chr10:63661469-39911653	All CDS E1-10	
BCL11A	NM_022893	chr2:60687539-60780405	All CDS E1-4	
BCL2L1	NM_138578	chr20:30253752-30310021	All CDS E2-3	
BCOR	NM_001123385	chrX:39911362-39937182	All CDS E2-15	
C10orf12	NM_015652	chr10:98741148-98744891	All CDS E1	
CD83	NM_004233	chr6:14118043-14135467	All CDS E1-5	
CDKN2A	NM_000077	chr9:21968228-21974826	All CDS E1-3	Cell cycle
CHST2	NM_004267	chr3:142839659-142841251	All CDS E2	
CLTC	NM_004859	chr17:57697493-57771213	All CDS E1-32	
DAZAP1	NM_170711	chr19:1407773-1433826	All CDS E1-12	
DOCK8	NM_001190458	chr9:286509-464219	All CDS E2-46	
DTX1	NM_004416	chr12:113495998-113534744	All CDS E1-9	Notch pathway
DUSP2	NM_004418	chr2:96809562-96811093	All CDS E1-4	
EBF1	NM_024007	chr5:158126119-158526486	All CDS E1-16	
EDRF1	NM_001202438	chr10:127408377-127452041	All CDS E1-25	
EIF4A2	NM_001967	chr3:186501400-186507058	All CDS E1-11	
EPB41	NM_203343	chr1:29313950-29442309	All CDS E2-16	
FAS	NM_000043	chr10:90750634-90774207	All CDS E1-9	
FBXO11	NM_001190274	chr2:48035257-48132859	All CDS E1-23	
FOXC1	NM_001453	chr6:1610681-1612342	All CDS E1	
GNAI2	NM_002070	chr3:50273768-50295122	All CDS E1-8	
GRHPR	NM_012203	chr9:37422748-37436779	All CDS E1-9	
HIST1H2BK	NM_080593	chr6:27114197-27114577	All CDS E1	
HLA-A	NM_002116	chr6:29910331-29913232	All CDS E1-8	
HLA-B	NM_005514	chr6:31322260-31324935	All CDS E1-7	
HLA-C	NM_002117	chr6:31236946-31239848	All CDS E1-8	
HLA-DMB	NM_002118	chr6:32902748-32908584	All CDS E1-6	
IKBKB	NM_001556	chr8:42129619-42188497	All CDS E2-22	
IL10RA	NM_001558	chr11:117857183-117870356	All CDS E1-7	
IL16	NM_172217	chr15:81517741-81601139	All CDS E2-19	

IRF2BP2	NM_182972	chr1:234742883-234745240	All CDS E1-2	
ITPKB	NM_002221	chr1:226822372-226925159	All CDS E1-8	
JUNB	NM_002229	chr19:12902586-12903629	All CDS E1	
KLF2	NM_016270	chr19:16435735-16437842	All CDS E1-3	
KLHL14	NM_020805	chr18:30254620-30350554	All CDS E2-9	
KLHL21	NM_014851	chr1:6653425-6662877	All CDS E1-4	
KLHL42	NM_020782	chr12:27933264-27951099	All CDS E1-3	
MED16	NM_005481	chr19:868100-891131	All CDS E2-16	
MPEG1	NM_001039396	chr11:58978188-58980338	All CDS E1	
NOL9	NM_024654	chr1:6585914-6614562	All CDS E1-12	
OSBPL10	NM_017784	chr3:31703564-32022671	All CDS E1-12	
PABPC1	NM_002568	chr8:101716525-101733811	All CDS E1-14	
PIM2	NM_006875	chrX:48771408-48776111	All CDS E1-6	
PPP1R9B	NM_032595	chr17:48212696-48227874	All CDS E1-12	
PRRC2C	NM_015172	chr1:171481228-171560986	All CDS E2-34	
RAC2	NM_002872	chr22:37622711-37640188	All CDS E1-6	
RFTN1	NM_015150	chr3:16358335-16535376	All CDS E2-10	
S1PR2	NM_004230	chr19:10334520-10335581	All CDS E2	
SEC24C	NM_004922	chr10:75506591-75530853	All CDS E3-24	
SETD1B	NM_015048	chr12:122242644-122268145	All CDS E1-17	
SLC1A5	NM_005628	chr19:47278767-47291222	All CDS E1-8	
SPEN	NM_015001	chr1:16174563-16265922	All CDS E1-15	Notch pathway
TAP1	NM_000593	chr6:32813356-32821593	All CDS E1-11	
TNRC18	NM_001080495	chr7:5347737-5460877	All CDS E2-30	
ΤΟΧ	NM_014729	chr8:59720306-60031546	All CDS E1-9	
TP53BP1	NM_001141979	chr15:43699581-43785241	All CDS E1-28	
TP63	NM_003722	chr3:189349305-189612291	All CDS E13-14	
TP73	NM_005427	chr1:3598930-3649643	All CDS E2-14	
TRIP12	NM_001284214	chr2:230632270-230725247	All CDS E2-42	
TRRAP	NM_001244580	chr7:98478774-98609978	All CDS E2-72	
UBE2A	NM_003336	chrX:118708675-118717218	All CDS E1-6	
VMP1	NM_030938	chr17:57808808-57917272	All CDS E2-12	
WDR24	NM_032259	chr16:734734-739640	All CDS E1-9	

WEE1	NM_003390	chr11:9595481-9610149	All CDS E1-11	
ZC3H12D	NM_207360	chr6:149771819-149795679	All CDS E2-6	
ZFP36L1	NM_004926	chr14:69256250-69259655	All CDS E1-2	
ZNF516	NM_014643	chr18:74074453-74155010	All CDS E3-8	
RHOA	NM_014643	chr3:49397642-49413024	All CDS E2-5	

BCR-TLR: B-cell receptor/Toll-like receptor; CDS: coding DNA sequence

Sorios	Morphology	Bioney site			Immu	unophei	notype			Clonality	600 ^{&}	ERED	I MD_1	EBNA-2	Zebra-	Latoncy pattorn
Series	worphology	Biopsy site	CD20	CD79	BCL2	BCL6	CD10	MUM1	PAX5	Cionanty	000	LDLK		LDNA-2	EBV	
p1	DLBCL	Intestine	+	+	+		-	+			ABC	+	+	+	+	Type III with replication
р5	DLBCL*	Small bowel	+		+	-	-	+		Monoclonal	Non-GC /Unc		+			
р6	DLBCL	Stomach	+		+	-	-	+			Non-GC /ABC	+	+	-		Type IIa
p10	DLBCL*	Intestine	+		+	-	-	+	+	Monoclonal	Non-GC/ ABC	+	+	+	+	Type III with replication
p11	BL	Inguinal LN	+		-	+	+	-		Monoclonal	GCB	+	-	-	-	Type I
p12	DLBCL	LN	+	+	-	+ focal	I	+	+	Monoclonal	Non-GC /ABC	+	+	-	-	Type IIa
p13	DLBCL	Cervical LN	+	+	+	-	-	+	+	Monoclonal	Non-GC /ABC	+	+	+	+	Type III with replication
p14	DLBCL	LN	+	+	-	+	-	+	+	Monoclonal	Non-GC /ABC	+	+	+	+	Type III with replication
p15	BL	LN	+	+	-	+	+	-	+		GCB	+	-	-	-	Туре I
p17	DLBCL	LN	+		+ focal	-	-	+	+	Monoclonal	Non-GC /Unc	+				
p19	DLBCL	Intestine	+	+	+	-	-	+		Monoclonal	Non-GC /ABC	+	-	-	-	Туре I
p20	DLBCL	Intestinal LN	+	+	+	-		+	+	Monoclonal	ABC	+	+	+	-	Type III
p21	DLBCL	Mesenteric LN	+	+	+	+	-	+	+	Monoclonal	Non-GC /ABC	+	+	-	+	Type II with replication
p22	BL	Tonsil		+	-	focal	+		+	Monoclonal	GCB	+	-	-	-	Туре I
p23	DLBCL	Intestine	+			focal	-			Monoclonal		+	-	-	-	Туре I
p25	DLBCL	LN	-	+ focal	+	+ focal	-	+ focal	+ focal	Monoclonal	Non-GC /Unc	+	+	-	-	Type IIa
p26	DLBCL	Mesenteric LN	+	+	+	+	-	+	+	Polyclonal	Non-GC /ABC	+	+	+		Type III
p27	DLBCL	Intestine	+	+	-	+	NA	-	+	Polyclonal#	GCB	-	-			
p28	BL	Intestine	+	+	-	+	+	+	+	Monoclonal	GCB	+	-	-	-	Туре І
p29	BL	Intestine	+	+	-	+	-	_	NA	Monoclonal	GCB	+				

Supplemental Table 4. Morphologic features, COO, EBV and clonality sta	atus of 31 pediatric post-SOT mPTLD detailed per case.
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p31	DLBCL	Intestine	+	+	+	-	-	+	+	Monoclonal	Non-GC /ABC	+		+		Type IIb-III†
p32	DLBCL	Axilar LN	+		+	-	-	+		Monoclonal	Non-GC /Unc	+	+	+	+	Type III with replication
p33	DLBCL*	Mesenteric LN	-	+		-	-	+			Non- GC/ABC	+	+	+		Type III
p35	DLBCL	Cervical LN	+ focal	+		-	I	+		Monoclonal	Non- GC/ABC	+	+ focal	-		Type IIa
p36	DLBCL*	LN	+ focal	+		-	-	+	-	Monoclonal	Non-GC /ABC	+	+ focal	-		Type IIa
p37	DLBCL	Scalp	+			+	+	-		Monoclonal	GCB	-	-	-		
p38	BL	Intestine	+			+	+	-		Monoclonal	GCB	+	-	-		Туре I
p39	DLBCL	Tonsil	+	+		-	-	+			Non-GC	+	+	+		Type III
p41	DLBCL*	Abdominal mass	-	+				+	-	Monoclonal		+	+	+		Type III
p42	BL	Abdominal mass and intestine	+	+	-	+	+	+ focal	+		GCB	+	-	-	-	Туре I
p43	DLBCL	Abdominal adenopathy								Monoclonal	Non-GC /ABC	+	+	+		Type III

NA: not assessable

* cases with plasmacytic differentiation

† In the absence of LMP1 assessability, two cases with latency pattern IIb-III are included in latency pattern III group

[#]IGH clonality analysis was not complete

[&] Cell of origin determination according to Hans and/or NanoString analyses. GCB results derive from either one or both algorithms. Unc: Unclassified.

Case	Group	Chromosome Region (GRCh37/hg19)	Event	Length	Cytoband
p10	PTLD-DLBCL	No CNA			
p11	PTLD-BL	chr6:0-44,239,109	CNN-LOH	44239110	p25.3 - p21.1
p12	PTLD-DLBCL	chr7:92,977,364-105,256,711	CNN-LOH	12279348	q21.3 - q22.3
p12	PTLD-DLBCL	chr19:0-6,735,114	CNN-LOH	6735115	p13.3
p13	PTLD-DLBCL	No CNA			
p14	PTLD-DLBCL	chr1:14,244,382-23,427,526	CNN-LOH	9183145	p36.21 - p36.12
p14	PTLD-DLBCL	chr2:32,437,183-50,937,966	CNN-LOH	18500784	p22.3 - p16.3
p14	PTLD-DLBCL	chr3:59,350,609-64,821,813	CNN-LOH	5471205	p14.2 - p14.1
p14	PTLD-DLBCL	chr5:97,552,139-102,911,071	CNN-LOH	5358933	q15 - q21.2
p14	PTLD-DLBCL	chr8:139,752,488-146,364,022	CNN-LOH	6611535	q24.23 - q24.3
p14	PTLD-DLBCL	chr9:16,354,115-33,476,795	CNN-LOH	17122681	p22.3 - p13.3
p14	PTLD-DLBCL	chr11:123,273,988-128,279,122	CNN-LOH	5005135	q24.1 - q24.3
p14	PTLD-DLBCL	chr13:33,960,165-41,140,013	CNN-LOH	7179849	q13.1 - q14.11
p14	PTLD-DLBCL	chr17:7,241,535-12,838,706	CNN-LOH	5597172	p13.1 - p12
p15	PTLD-BL	chr11:12,030,761-13,113,696	CN Loss	1082936	p15.3 - p15.2
p15	PTLD-BL	chr11:45,916,091-51,575,951	CNN-LOH	5659861	p11.2 - p11.12
p15	PTLD-BL	chrX:0-155,270,560	CN Loss	155270561	p22.33 - q28
p19	PTLD-DLBCL	chr3:0-91,000,000	CN Gain	91000001	p26.3 - q11.1
p19	PTLD-DLBCL	chr8:0-7,004,147	CN Gain	7004148	p23.3 - p23.1
p19	PTLD-DLBCL	chr9:70,984,372-141,213,431	CN Gain	70229060	q21.11 - q34.3
p19	PTLD-DLBCL	chr20:29,519,156-63,025,520	CN Loss	33506365	q11.21 - q13.33
p19	PTLD-DLBCL	chr21:14,344,537-48,129,895	CN Gain	33785359	q11.2 - q22.3
p20	PTLD-DLBCL	chr9:204,738-141,054,761	CN Gain	140850024	p24.3 - q34.3
p20	PTLD-DLBCL	chr12:112,915,434-117,209,665	CN Loss	4294232	q24.13 - q24.22
p21	PTLD-DLBCL	chr3:187,566,343-189,106,752	CN Loss	1540410	q27.3 - q28
p22	PTLD-BL	chr11:45,916,586-51,575,951	CNN-LOH	5659366	p11.2 - p11.12
p22	PTLD-BL	chr20:14,819,848-14,972,461	CN Loss	152614	p12.1
p25	PTLD-DLBCL	chr8:110,835,796-116,086,438	CNN-LOH	5250643	q23.2 - q23.3
p26	PTLD-DLBCL	chr1:30,307,979-48,275,263	CNN-LOH	17967285	p35.2 - p33
p26	PTLD-DLBCL	chr2:21,494-10,020,168	CNN-LOH	9998675	p25.3 - p25.1

Supplemental Table 5. Copy number (CN) alterations and CN neutral loss of heterozygosity (CNN-LOH) detected in 23 pediatric post-SOT mPTLD.

p26	PTLD-DLBCL	chr2:147,262,273-173,642,237	CNN-LOH	26379965 q22.3 - q31.1
p26	PTLD-DLBCL	chr3:63,411-10,239,161	CNN-LOH	10175751 p26.3 - p25.3
p26	PTLD-DLBCL	chr3:47,120,640-52,638,564	CNN-LOH	5517925 p21.31 - p21.1
p26	PTLD-DLBCL	chr3:130,227,840-142,566,337	CNN-LOH	12338498 q22.1 - q23
p26	PTLD-DLBCL	chr4:105,732,804-114,022,691	CNN-LOH	8289888 q24 - q25
p26	PTLD-DLBCL	chr4:163,501,404-173,005,461	CNN-LOH	9504058 q32.2 - q34.1
p26	PTLD-DLBCL	chr4:184,294,510-190,915,650	CNN-LOH	6621141 q35.1 - q35.2
p26	PTLD-DLBCL	chr5:106,804,363-142,535,605	CNN-LOH	35731243 q21.3 - q31.3
p26	PTLD-DLBCL	chr6:142,331,397-155,742,113	CNN-LOH	13410717 q24.1 - q25.3
p26	PTLD-DLBCL	chr7:68,913,353-125,848,263	CNN-LOH	56934911 q11.22 - q31.33
p26	PTLD-DLBCL	chr9:204,738-7,370,794	CNN-LOH	7166057 p24.3 - p24.1
p26	PTLD-DLBCL	chr9:9,859,772-14,903,800	CNN-LOH	5044029 p23 - p22.3
p26	PTLD-DLBCL	chr15:38,731,045-45,598,335	CNN-LOH	6867291 q14 - q21.1
p26	PTLD-DLBCL	chr17:6,645,258-12,289,647	CNN-LOH	5644390 p13.1 - p12
p26	PTLD-DLBCL	chr17:55,932,988-62,982,717	CNN-LOH	7049730 q22 - q24.1
p28	PTLD-BL	No CNA		
p29	PTLD-BL	chr2:100,603,279-243,052,331	CNN-LOH	142449053 q11.2 - q37.3
p29	PTLD-BL	chr11:46,541,220-51,575,951	CNN-LOH	5034732 p11.2 - p11.12
p29	PTLD-BL	chr18:1-78,077,248	CN Gain	78077248 p11.32 - q23
p29	PTLD-BL	chrY:2,660,163-28,799,935	Homozygous Copy Loss	26139773 p11.31 - q11.23
p31	PTLD-DLBCL	chr3:1-198,022,430	CN Gain	198022430 p26.3 - q29
p31	PTLD-DLBCL	chr17:38,023,745-38,504,415	CN Loss	480671 q12 - q21.2
p31	PTLD-DLBCL	chrX:76,819,736-90,582,240	CN Gain	13762505 q21.1 - q21.31
p32	PTLD-DLBCL	chr16:29,710,682-35,271,725	CNN-LOH	5561044 p11.2 - p11.1
p33	PTLD-DLBCL	chr5:36,737,343-145,702,632	CNN-LOH	108965290 p13.2 - q32
p33	PTLD-DLBCL	chr15:30,591,713-55,631,810	CNN-LOH	25040098 q13.2 - q21.3
p36	PTLD-DLBCL	chr1:149,044,448-249,212,878	CNN-LOH	100168431 q21.2 - q44
p36	PTLD-DLBCL	chr4:84,099,324-88,072,085	CN Gain	3972762 q21.22 - q22.1
p36	PTLD-DLBCL	chr4:98,696,259-101,924,060	High Copy Gain	3227802 q22.3 - q24
p36	PTLD-DLBCL	chr4:101,947,930-103,635,183	High Copy Gain	1687254 q24
p36	PTLD-DLBCL	chr4:103,647,047-107,168,578	CN Gain	3521532 q24
p36	PTLD-DLBCL	chr4:107,185,824-190,915,650	CN Loss	83729827 q24 - q35.2
p36	PTLD-DLBCL	chr19:1-59,128,983	CN Loss	59128983 p13.3 - q13.43

p36	PTLD-DLBCL	chr20:1-63,025,520	CN Gain	63025520	p13 - q13.33
p37	PTLD-DLBCL	chr2:60,868,614-61,178,711	High Copy Gain	310098	p16.1
p37	PTLD-DLBCL	chr2:61,179,160-63,133,418	CN Gain	1954259	p16.1 - p15
p37	PTLD-DLBCL	chr10:43,581,501-57,966,196	CN Gain	14384696	q11.21 - q21.1
p37	PTLD-DLBCL	chr18:25,473,942-60,614,806	CN Gain	35140865	q12.1 - q21.33
p37	PTLD-DLBCL	chr18:60,616,440-70,727,724	CN Gain	10111285	q21.33 - q22.3
p38	PTLD-BL	No CNA			
p39	PTLD-DLBCL	No CNA			
p42	PTLD-BL	chr8:172,417-25,544,268	CN Loss	25371852	p23.3 - p21.2
p43	PTLD-DLBCL	chr1:144,009,053-167,484,888	CNN-LOH	23475836	q21.1 - q24.2
p43	PTLD-DLBCL	chr3:1-198,022,430	CN Gain	198022430	p26.3 - q29
p43	PTLD-DLBCL	chr5:1-180,915,260	CN Gain	180915260	p15.33 - q35.3
p43	PTLD-DLBCL	chr7:81,796,619-95,692,964	CNN-LOH	13896346	q21.11 - q21.3
p43	PTLD-DLBCL	chr9:1-141,213,431	CN Gain	141213431	p24.3 - q34.3
p43	PTLD-DLBCL	chr10:70,413,477-77,689,439	CNN-LOH	7275963	q21.3 - q22.2
p43	PTLD-DLBCL	chr12:1-133,851,895	CN Gain	133851895	p13.33 - q24.33
p43	PTLD-DLBCL	chr16:19,104,616-25,077,977	CNN-LOH	5973362	p12.3 - p12.1
p43	PTLD-DLBCL	chr17:20,124,289-20,517,074	CN Loss	392786	p11.2
p43	PTLD-DLBCL	chr17:20,517,075-21,911,243	CN Gain	1394169	p11.2
p43	PTLD-DLBCL	chr18:3,444,836-8,910,868	CNN-LOH	5466033	p11.31 - p11.22
p43	PTLD-DLBCL	chr20:34,101,821-40,679,500	CNN-LOH	6577680	q11.22 - q12

Start Case Group Chr End Ref Alt Func.refGene Gene Transcript PTLD-DLBCL 171486769 171486769 C PRRC2C NM 015172 p10 chr1 т exonic PTLD-DLBCL 2071443 2071443 C MYOM2 NM 003970 p10 chr8 Т exonic p10 PTLD-DLBCL chr8 128750345 128750345 C intronic-MYC MYC PTLD-DLBCL 42383204 42383204 C CD79A NM 001783 chr19 p10 exonic PTLD-BL chr1 23885829 23885829 T ID3 NM 002167 p11 exonic PTLD-BL 27100176 27100176 C ARID1A NM 006015 p11 chr1 Α exonic NM 001136125 PTLD-BL chr6 41903688 41903688 A G exonic CCND3 p11 101733638 G PTLD-BL 101733638 С PABPC1 NM 002568 chr8 exonic p11 С intronic-MYC PTLD-BL 128748998 128748998 A MYC p11 chr8 AGAGGAGAAG MYC PTLD-BL 128749066 128749066 GT p11 chr8 intronic-MYC PTLD-BL chr8 128749078 128749078 C intronic-MYC MYC p11 128749671 A MYC p11 PTLD-BL chr8 128749671 G intronic-MYC PTLD-BL 128749924 128749924 G С MYC chr8 intronic-MYC p11 PTLD-BL 117869503 117869503 C NM 001558 chr11 IL10RA p11 exonic 103342855 C TRAF3 PTLD-BL chr14 103342855 exonic NM 001199427 p11 PTLD-BL chr15 42145506 42145506 G A SPTBN5 NM 016642 p11 exonic PTLD-BL 41205627 41205627 C G DDX3X chrX exonic NM 001193417 p11 p12 PTLD-DLBCL chr1 2492096 2492096 С exonic TNFRSF14 NM 001297605 PTLD-DLBCL 115256530 115256530 G Т NRAS NM 002524 p12 chr1 exonic PTLD-DLBCL 101578250 101578250 C **UTR3-NFKBIZ** NFKBIZ c.*35C>T;NM 031419 p12 chr3 Т p12 PTLD-DLBCL chr3 101578251 101578251 T G UTR3-NFKBIZ NFKBIZ c.*36T>G;NM 031419 PTLD-DLBCL 101578368 101578372 AAAAG UTR3-NFKBIZ NFKBIZ p12 chr3 c.*153 *157delAAAAG;NI CCND3 p12 PTLD-DLBCL 41903682 41903682 A NM 001136125 Т exonic chr6 p12 PTLD-DLBCL chr7 5428547 5428547 C G TNRC18 NM 001080495 exonic PTLD-DLBCL 139390656 G NOTCH1 p12 chr9 139390656 A exonic NM 017617 NOTCH1 PTLD-DLBCL 139390861 139390861 G NM 017617 p12 chr9 А exonic p12 PTLD-DLBCL chr12 12037499 12037499 C ETV6 NM 001987 т exonic chr13 p12 PTLD-DLBCL 32906766 32906766 C BRCA2 NM 000059 Т exonic

Supplemental Table 6. List of somatic mutations identified in the analysis of 25 pediatric post-SOT mPTLD including prediction of amino acid chan prediction.⁶

p12	PTLD-DLBCL	chr15	42154384	42154384	С	Т	exonic	SPTBN5	NM_016642
p12	PTLD-DLBCL	chr19	6586078	6586078	G	A	exonic	CD70	NM_001252
p13	PTLD-DLBCL	chr11	117869676	117869676	С	Т	exonic	IL10RA	NM_001558
p14	PTLD-DLBCL	chr6	31323175	31323175	С	Т	exonic	HLA-B	NM_005514
p14	PTLD-DLBCL	chr9	139814840	139814840	А	С	exonic	TRAF2	NM_021138
p14	PTLD-DLBCL	chr12	48190875	48190875	G	A	exonic	HDAC7	NM_001308090
p14	PTLD-DLBCL	chr12	113532642	113532642	G	A	exonic	DTX1	NM_004416
p14	PTLD-DLBCL	chrX	39932564	39932564	С	Т	exonic	BCOR	NM_001123383
p15	PTLD-BL	chr1	23885664	23885666	GCC	-	exonic	ID3	NM_002167
p15	PTLD-BL	chr1	23885671	23885675	CTACC	-	exonic	ID3	NM_002167
p15	PTLD-BL	chr8	128748866	128748866	С	G	exonic	MYC	NM_002467
p15	PTLD-BL	chr8	128748893	128748893	Т	G	intronic-MYC	MYC	
p15	PTLD-BL	chr8	128748919	128748919	А	G	intronic-MYC	MYC	
p15	PTLD-BL	chr8	128748927	128748927	А	G	intronic-MYC	MYC	
p15	PTLD-BL	chr8	128748949	128748949	G	A	intronic-MYC	MYC	
p15	PTLD-BL	chr8	128748964	128748964	Т	A	intronic-MYC	MYC	
p15	PTLD-BL	chr8	128748982	128748982	Т	С	intronic-MYC	MYC	
p15	PTLD-BL	chr8	128749005	128749005	С	Т	intronic-MYC	MYC	
p15	PTLD-BL	chr8	128749047	128749047	-	С	intronic-MYC	MYC	
p15	PTLD-BL	chr8	128749087	128749087	А	G	intronic-MYC	MYC	
p15	PTLD-BL	chr8	128749186	128749186	А	G	intronic-MYC	MYC	
p15	PTLD-BL	chr8	128749250	128749250	с	т	intronic-MYC	МҮС	
p15	PTLD-BL	chr8	128749252	128749252	G	Т	intronic-MYC	MYC	
p15	PTLD-BL	chr8	128749266	128749266	А	Т	intronic-MYC	MYC	
p15	PTLD-BL	chr8	128749289	128749289	А	С	intronic-MYC	MYC	
p15	PTLD-BL	chr8	128749338	128749338	Т	С	intronic-MYC	MYC	
p15	PTLD-BL	chr8	128749356	128749356	G	С	intronic-MYC	MYC	
p15	PTLD-BL	chr8	128749422	128749422	А	С	intronic-MYC	MYC	
p15	PTLD-BL	chr8	128749436	128749436	Т	С	intronic-MYC	MYC	
p15	PTLD-BL	chr8	128749484	128749484	А	Т	intronic-MYC	MYC	
p15	PTLD-BL	chr8	128749514	128749514	G	A	intronic-MYC	MYC	
p15	PTLD-BL	chr8	128749515	128749515	G	С	intronic-MYC	MYC	
p15	PTLD-BL	chr8	128749564	128749564	Т	С	intronic-MYC	MYC	
p15	PTLD-BL	chr8	128749587	128749587	А	-	intronic-MYC	MYC	
p15	PTLD-BL	chr8	128749594	128749594	А	G	intronic-MYC	MYC	
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p15	PTLD-BL	chr8	128749661	128749661	G	С	intronic-MYC	MYC	
p15	PTLD-BL	chr8	128749790	128749790	А	G	intronic-MYC	MYC	
p15	PTLD-BL	chr8	128750135	128750135	Т	С	intronic-MYC	MYC	
p15	PTLD-BL	chr8	128750140	128750140	Т	A	intronic-MYC	MYC	
p15	PTLD-BL	chr8	128750609	128750609	А	G	exonic	MYC	NM_002467
p15	PTLD-BL	chr8	128750680	128750680	A	G	exonic	MYC	NM_002467
p15	PTLD-BL	chr8	128750698	128750698	A	G	exonic	MYC	NM_002467
p15	PTLD-BL	chr13	41240349	41240349	Т	С	exonic	FOXO1	NM_002015
p15	PTLD-BL	chr22	41525969	41525969	Т	С	exonic	EP300	NM_001429
p17	PTLD-DLBCL	chr1	6659354	6659354	G	A	exonic	KLHL21	NM_001324309
p17	PTLD-DLBCL	chr3	50273842	50273842	G	С	exonic	GNAI2	NM_002070
p17	PTLD-DLBCL	chr10	63852709	63852709	Т	С	exonic	ARID5B	NM_001244638
p20	PTLD-DLBCL	chr1	27106228	27106228	С	Т	exonic	ARID1A	NM_006015
p20	PTLD-DLBCL	chr3	16419320	16419320	С	A	exonic	RFTN1	NM_015150
p20	PTLD-DLBCL	chr3	47165327	47165327	С	Т	exonic	SETD2	NM_001349370
p20	PTLD-DLBCL	chr5	112175528	112175528	А	G	exonic	APC	NM_001127511
p20	PTLD-DLBCL	chr6	27114468	27114468	С	G	exonic	HIST1H2BK	NM_001312653
p20	PTLD-DLBCL	chr16	85953739	85953739	A	G	exonic	IRF8	NM_002163
p20	PTLD-DLBCL	chrX	39922007	39922007	С	Т	exonic	BCOR	NM_001123384
p21	PTLD-DLBCL	chr7	98609012	98609012	G	A	exonic	TRRAP	NM_003496
p21	PTLD-DLBCL	chr7	151878913	151878913	G	A	exonic	KMT2C	NM_170606
p22	PTLD-BL	chr1	23885727	23885727	А	С	exonic	ID3	NM_002167
p22	PTLD-BL	chr1	23885752	23885752	G	A	exonic	ID3	NM_002167
p22	PTLD-BL	chr1	27106133	27106133	Т	A	exonic	ARID1A	NM_006015
p22	PTLD-BL	chr3	142840130	142840130	G	A	exonic	CHST2	NM_004267
p22	PTLD-BL	chr8	128749703	128749703	G	А	intronic-MYC	MYC	
p22	PTLD-BL	chr8	128750381	128750381	С	G	intronic-MYC	MYC	
p22	PTLD-BL	chr8	128750677	128750677	С	Т	exonic	MYC	NM_002467
p22	PTLD-BL	chr11	128359281	128359281	С	Т	exonic	ETS1	NM_001330451
p22	PTLD-BL	chr12	57499332	57499332	G	A	exonic	STAT6	NM_001178081
p26	PTLD-DLBCL	chr3	142839722	142839722	С	Т	exonic	CHST2	NM_004267
p26	PTLD-DLBCL	chr19	1650232	1650232	Т	С	exonic	TCF3	NM_001136139
p27	PTLD-DLBCL	chr1	203274876	203274876	G	A	exonic	BTG2	NM_006763

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p27	PTLD-DLBCL	chr1	203274877	203274877	G	A	splicing	BTG2	exon1
p27	PTLD-DLBCL	chr2	96810513	96810513	G	A	exonic	DUSP2	NM_004418
p27	PTLD-DLBCL	chr2	230650559	230650559	G	-	exonic	TRIP12	NM_001284216
p27	PTLD-DLBCL	chr5	137801629	137801629	G	А	exonic	EGR1	NM_001964
p27	PTLD-DLBCL	chr5	137801737	137801737	С	Т	exonic	EGR1	NM_001964
p27	PTLD-DLBCL	chr6	26157154	26157154	С	Т	exonic	HIST1H1E	NM_005321
p27	PTLD-DLBCL	chr6	134494418	134494418	С	G	exonic	SGK1	NM_001143677
p27	PTLD-DLBCL	chr6	134494482	134494482	С	Т	exonic	SGK1	NM_001143677
p27	PTLD-DLBCL	chr6	134494647	134494647	А	С	exonic	SGK1	NM_001143677
p27	PTLD-DLBCL	chr6	134494697	134494697	G	С	exonic	SGK1	NM_001143677
p27	PTLD-DLBCL	chr6	134495155	134495155	G	С	exonic	SGK1	NM_001143677
p27	PTLD-DLBCL	chr6	134495196	134495196	Т	G	exonic	SGK1	NM_001143677
p27	PTLD-DLBCL	chr6	134495217	134495217	G	С	exonic	SGK1	NM_001143677
p27	PTLD-DLBCL	chr6	134495658	134495658	G	A	exonic	SGK1	NM_001143677
p27	PTLD-DLBCL	chr7	2983994	2983994	С	Т	exonic	CARD11	NM_032415
p27	PTLD-DLBCL	chr7	5353327	5353327	С	Т	exonic	TNRC18	NM_001080495
p27	PTLD-DLBCL	chr10	90750636	90750636	G	A	exonic	FAS	NM_000043
p27	PTLD-DLBCL	chr12	113496233	113496233	Т	A	exonic	DTX1	NM_004416
p27	PTLD-DLBCL	chr12	113496242	113496242	Т	G	exonic	DTX1	NM_004416
p27	PTLD-DLBCL	chr12	122460005	122460005	G	С	exonic	BCL7A	NM 001024808
p27		chr13	32911295	32911295	G	А	exonic	BRCA2	NM 000059
p27	PTI D-DI BCI	chr15	66727441	66727441	Т	C	exonic	MAP2K1	NM_002755
p27	PTI D-DI BCI	chr15	81517990	81517990	G	Т	exonic	II 16	NM 172217
p27	PTI D-DI BCI	chr16	10996027	10996027	C	A	exonic	CIITA	NM_000246
p27	PTI D-DI BCI	chr17	63014369	63014369	A	Т	splicing	GNA13	exon3
p27	PTLD-DI BCI	chr18	74154731	74154731	C	T	exonic	ZNF516	NM 014643
p27	PTI D-DI BCI	chr19	12902770	12902770	G	A	exonic	JUNB	NM_002229
p27	PTLD-DI BCI	chr19	19256795	19256795	G	A	exonic	MEF2B	NM 005919
p28	PTI D-BI	chr1	23885618	23885618	C	G	exonic	ID3	NM_002167
L			2000010	2000010	. ~				

p28	PTLD-BL	chr1	23885710	23885710	G	С	exonic	ID3	NM_002167
p28	PTLD-BL	chr1	23885755	23885755	-	CA	exonic	ID3	NM_002167
p28	PTLD-BL	chr2	70315818	70315818	А	С	exonic	PCBP1	NM_006196
p28	PTLD-BL	chr2	70315897	70315897	А	С	exonic	PCBP1	NM_006196
p28	PTLD-BL	chr3	38180208	38180208	С	Т	exonic	MYD88	NM_001172566
p28	PTLD-BL	chr4	103514671	103514671	G	A	exonic	NFKB1	NM_001165412
p28	PTLD-BL	chr5	112176402	112176402	А	G	exonic	APC	NM_001127511
p28	PTLD-BL	chr5	112177130	112177130	А	G	exonic	APC	NM_001127511
p28	PTLD-BL	chr6	26031949	26031949	G	A	exonic	HIST1H3B	NM_003537
p28	PTLD-BL	chr6	149795667	149795667	Т	G	exonic	ZC3H12D	NM_207360
p28	PTLD-BL	chr7	5352564	5352564	G	A	exonic	TNRC18	NM_001080495
p28	PTLD-BL	chr7	98507794	98507794	С	G	exonic	TRRAP	NM_001244580
p28	PTLD-BL	chr8	2007287	2007287	А	G	exonic	MYOM2	NM_003970
p28	PTLD-BL	chr8	128748873	128748873	А	G	intronic-MYC	MYC	
p28	PTLD-BL	chr8	128748874	128748874	G	A	intronic-MYC	MYC	
p28	PTLD-BL	chr8	128748895	128748895	Т	G	intronic-MYC	MYC	
p28	PTLD-BL	chr8	128748973	128748973	Т	G	intronic-MYC	MYC	
p28	PTLD-BL	chr8	128748975	128748975	Т	С	intronic-MYC	MYC	
p28	PTLD-BL	chr8	128748995	128748995	G	С	intronic-MYC	MYC	
p28	PTLD-BL	chr8	128749013	128749013	G	Т	intronic-MYC	MYC	
p28	PTLD-BL	chr8	128749035	128749035	Т	С	intronic-MYC	MYC	
p28	PTLD-BL	chr8	128749036	128749036	G	A	intronic-MYC	MYC	
p28	PTLD-BL	chr8	128749037	128749037	G	С	intronic-MYC	MYC	
p28	PTLD-BL	chr8	128749124	128749124	С	Т	intronic-MYC	MYC	
p28	PTLD-BL	chr8	128749135	128749135	А	G	intronic-MYC	MYC	
p28	PTLD-BL	chr8	128749174	128749174	С	G	intronic-MYC	MYC	
p28	PTLD-BL	chr8	128749212	128749212	G	-	intronic-MYC	MYC	
p28	PTLD-BL	chr8	128749267	128749267	_	GGACCGCATAT CGCCTGT	intronic-MYC	мүс	
p28	PTLD-BL	chr8	128749338	128749338	Т	С	intronic-MYC	МҮС	
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p28		cnr8	128749488	128749488	C		Intronic-IVIYC	MYC	
p28	PILD-BL	cnr8	128749504	128749504	G	A	Intronic-IVIYC	MYC	
						TTTTTTCTTTCA GATGCTGGCTC			
p28	PTLD-BL	chrX	41204672	41204672	-	GTGATC	exonic	DDX3X	NM 001193417
p29	PTLD-BL	chr1	118166170	118166170	А	G	exonic	FAM46C	NM_017709
p29	PTLD-BL	chr6	26031910	26031910	G	A	exonic	HIST1H3B	NM_003537
p29	PTLD-BL	chr6	26234965	26234965	G	A	exonic	HIST1H1D	NM_005320
p29	PTLD-BL	chr8	128749108	128749108	Т	С	intronic-MYC	MYC	
p29	PTLD-BL	chr8	128749144	128749144	С	A	intronic-MYC	MYC	
p29	PTLD-BL	chr8	128749431	128749431	G	С	intronic-MYC	MYC	
p29	PTLD-BL	chr8	128749444	128749444	А	С	intronic-MYC	MYC	
p29	PTLD-BL	chr8	128749478	128749478	А	Т	intronic-MYC	MYC	
p29	PTLD-BL	chr8	128749763	128749763	G	A	intronic-MYC	MYC	
p29	PTLD-BL	chr8	128749810	128749810	А	Т	intronic-MYC	MYC	
p29	PTLD-BL	chr8	128750169	128750169	А	Т	intronic-MYC	MYC	
p29	PTLD-BL	chr8	128750284	128750284	А	Т	intronic-MYC	MYC	
p29	PTLD-BL	chr8	128750292	128750292	С	G	intronic-MYC	MYC	
p29	PTLD-BL	chr8	128750512	128750512	С	G	exonic	MYC	NM_002467
p29	PTLD-BL	chr8	128750522	128750522	Т	С	exonic	MYC	NM_002467
p29	PTLD-BL	chr8	128750632	128750632	С	G	exonic	MYC	NM_002467
p29	PTLD-BL	chr8	128750663	128750663	А	С	exonic	MYC	NM_002467
p29	PTLD-BL	chr8	128750671	128750671	С	G	exonic	MYC	NM_002467
p29	PTLD-BL	chr8	128750743	128750743	С	G	exonic	MYC	NM_002467
p29	PTLD-BL	chr8	128750771	128750771	G	А	exonic	MYC	NM_002467
p29	PTLD-BL	chr8	128750921	128750921	Т	G	exonic	MYC	NM_002467
p29	PTLD-BL	chr8	128750953	128750953	С	G	exonic	MYC	NM_002467
p29	PTLD-BL	chr8	128751210	128751210	С	A	exonic	MYC	NM_002467
p29	PTLD-BL	chr8	128751251	128751251	С	G	exonic	MYC	NM_002467
p29	PTLD-BL	chr13	41240288	41240288	С	Т	exonic	FOXO1	NM_002015
p29	PTLD-BL	chrX	41205590	41205590	G	A	exonic	DDX3X	NM_001193417
p32	PTLD-DLBCL	chr1	3644716	3644716	G	A	exonic	TP73	NM_001126240
p32	PTLD-DLBCL	chr4	55161387	55161387	Т	-	exonic	PDGFRA	NM_001347829

p33	PTLD-DLBCL	chr1	6585940	6585940	G	A	exonic	NOL9	NM_024654
p33	PTLD-DLBCL	chr6	138200098	138200098	G	А	exonic	TNFAIP3	NM_001270507
p33	PTLD-DLBCL	chr8	48739308	48739308	G	A	exonic	PRKDC	
p33	PTLD-DLBCL	chr8	128749023	128749023	G	Т	intronic-MYC	MYC	
p33	PTLD-DLBCL	chr8	128750260	128750260	С	Т	intronic-MYC	MYC	
p33	PTLD-DLBCL	chr9	382626	382626	G	A	exonic	DOCK8	NM_001190458
p33	PTLD-DLBCL	chr15	42147805	42147805	G	A	exonic	SPTBN5	NM_016642
p33	PTLD-DLBCL	chr16	736078	736078	G	Т	exonic	WDR24	NM_032259
p35	PTLD-DLBCL	chr1	3648092	3648092	Т	С	exonic	TP73	NM_001204191
p35	PTLD-DLBCL	chr1	29379717	29379717	А	-	exonic	EPB41	NM_203343
p35	PTLD-DLBCL	chr1	171509892	171509892	С	Т	exonic	PRRC2C	NM_015172
p35	PTLD-DLBCL	chr1	171527028	171527028	С	А	exonic	PRRC2C	NM_015172
p35	PTLD-DLBCL	chr2	96811047	96811047	G	Т	exonic	DUSP2	NM_004418
p35	PTLD-DLBCL	chr3	187442733	187442733	Т	С	exonic	BCL6	NM_001134738
p35	PTLD-DLBCL	chr4	153249384	153249384	С	Т	exonic	FBXW7	NM_001013415
p35	PTLD-DLBCL	chr6	32820185	32820185	С	Т	exonic	TAP1	NM_000593
p35	PTLD-DLBCL	chr7	2956965	2956965	G	-	exonic	CARD11	NM_032415
p35	PTLD-DLBCL	chr7	2983905	2983905	С	Т	exonic	CARD11	NM_032415
p35	PTLD-DLBCL	chr7	151860427	151860427	С	Т	exonic	KMT2C	NM_170606
p35	PTLD-DLBCL	chr7	5567923	5567923	G	A	exonic	ACTB	NM_001101
p35	PTLD-DLBCL	chr8	128748903	128748903	Т	-	intronic-MYC	MYC	
p35	PTLD-DLBCL	chr10	90768708	90768708	Т	-	exonic	FAS	NM_000043
p35	PTLD-DLBCL	chr10	90773948	90773948	G	A	exonic	FAS	NM_152871
p35	PTLD-DLBCL	chr11	108206646	108206646	С	G	exonic	ATM	NM 000051
p35	PTLD-DLBCL	chr13	32910767	32910767	С	А	exonic	BRCA2	NM_000059
p35	PTLD-DLBCL	chr15	42167199	42167199	С	Т	exonic	SPTBN5	NM_016642
p35	PTLD-DLBCL	chr15	81595890	81595890	С	Т	exonic	IL16	NM_004513
p35	PTLD-DLBCL	chr16	735089	735089	G	A	exonic	WDR24	NM 032259
p35	PTLD-DLBCL	chr16	3801727	3801727	G	А	exonic	CREBBP	NM_001079846
p35	PTLD-DLBCL	chr19	6586201	6586201	G	А	exonic	CD70	NM_001252

p35	PTLD-DLBCL	chr19	6586333	6586333	С	А	exonic	CD70	NM_001252
p35	PTLD-DLBCL	chr19	42596336	42596336	G	-	exonic	POU2F2	NM_001207025
p36	PTLD-DLBCL	chr6	44233425	44233425	A	G	exonic	NFKBIE	NM_004556
p36	PTLD-DLBCL	chr9	139390684	139390684	G	А	exonic	NOTCH1	NM_017617
p36	PTLD-DLBCL	chr17	7577538	7577538	С	Т	exonic	TP53	NM_001126115
p37	PTLD-DLBCL	chr1	117087113	117087113	Т	А	exonic	CD58	NM_001144822
p37	PTLD-DLBCL	chr7	151878676	151878676	G	Т	exonic	KMT2C	NM_170606
p37	PTLD-DLBCL	chr16	11016049	11016049	G	А	exonic	CIITA	NM_001286403
p37	PTLD-DLBCL	chr17	7577556	7577556	С	A	exonic	TP53	NM_001126115
p37	PTLD-DLBCL	chrX	48772479	48772479	С	Т	exonic	PIM2	NM_006875
p38	PTLD-BL	chr2	48050381	48050381	Т	С	exonic	FBXO11	NM_001190274
p38	PTLD-BL	chr3	50294456	50294458	AAG	-	exonic	GNAI2	NM_001282618
p38	PTLD-BL	chr8	128748922	128748922	С	G	intronic-MYC	MYC	
p38	PTLD-BL	chr8	128749034	128749034	С	G	intronic-MYC	MYC	
p38	PTLD-BL	chr8	128749434	128749434	С	G	intronic-MYC	MYC	
p38	PTLD-BL	chr8	128749779	128749779	А	С	intronic-MYC	MYC	
p38	PTLD-BL	chrX	41205620	41205620	А	С	exonic	DDX3X	NM_001193417
p38	PTLD-BL	chrX	41205658	41205658	A	G	exonic	DDX3X	NM_001193417
p39	PTLD-DLBCL	chr1	16259009	16259009	А	G	exonic	SPEN	NM_015001
p39	PTLD-DLBCL	chr3	187447651	187447651	G	А	exonic	BCL6	NM_001134738
p39	PTLD-DLBCL	chr6	1611960	1611960	С	A	exonic	FOXC1	NM_001453
p39	PTLD-DLBCL	chr7	103244814	103244814	G	А	exonic	RELN	NM_005045
p39	PTLD-DLBCL	chr7	151945442	151945442	Т	С	exonic	KMT2C	NM_170606
p39	PTLD-DLBCL	chr8	2054325	2054325	С	A	exonic	MYOM2	NM_003970
p39	PTLD-DLBCL	chr8	35624487	35624487	G	А	exonic	UNC5D	NM_001322818
					GCGGCT				
p39	PTLD-DLBCL	chr14	69257012	69257022	GTCTC	-	exonic	ZFP36L1	NM_001244698
p42	PTLD-BL	chr1	203274864	203274864	G	С	exonic	BTG2	NM_006763
					TGATATA				
					AATCATC				
p42	PTLD-BL	chr3	47163511	47163528	AAAA	-	exonic	SETD2	NM 001349370

p42	PTLD-BL	chr3	187443367	187443367	С	Т	exonic	BCL6	NM_001134738
p42	PTLD-BL	chr5	112174464	112174464	A	G	exonic	APC	NM_001127511
p42	PTLD-BL	chr8	128749034	128749034	С	G	intronic-MYC	MYC	
p42	PTLD-BL	chr8	128749254	128749254	G	С	intronic-MYC	MYC	
p42	PTLD-BL	chr8	128749993	128749993	G	A	intronic-MYC	MYC	
p42	PTLD-BL	chr8	128750483	128750483	С	Т	intronic-MYC	MYC	
p42	PTLD-BL	chr8	128750530	128750530	А	G	exonic	MYC	NM_001354870
p42	PTLD-BL	chr8	128750625	128750625	G	С	exonic	MYC	NM_001354870
p42	PTLD-BL	chr8	128750680	128750680	А	G	exonic	MYC	NM_001354870
p42	PTLD-BL	chr8	128750705	128750705	G	A	exonic	MYC	NM_001354870
p42	PTLD-BL	chr8	128750802	128750802	G	С	exonic	MYC	NM_001354870
p42	PTLD-BL	chr8	128751144	128751144	С	G	exonic	MYC	NM_001354870
p42	PTLD-BL	chr8	128751210	128751210	С	A	exonic	MYC	NM_001354870
p42	PTLD-BL	chr8	128751221	128751221	Т	С	exonic	MYC	NM_001354870
p42	PTLD-BL	chr8	128751239	128751239	С	Т	exonic	MYC	NM_001354870
p42	PTLD-BL	chr13	41240273	41240273	G	С	exonic	FOXO1	NM_002015
p42	PTLD-BL	chr15	42149184	42149184	С	Т	exonic	SPTBN5	NM_016642
p42	PTLD-BL	chr17	7578265	7578265	A	G	exonic	TP53	NM_001126115
p42	PTLD-BL	chr17	57697512	57697512	Т	G	exonic	CLTC	NM_001288653
p42	PTLD-BL	chr19	1615444	1615444	А	С	exonic	TCF3	NM_001351778
p42	PTLD-BL	chrX	41201908	41201908	т	с	splicing	DDX3X	exon5
p43	PTLD-DLBCL	chr12	122460041	122460041	А	С	exonic	BCL7A	NM_001024808

ges that affect the protein function (MA, SIFT, Polyphen2, CADD) and driver variants

HGVS c	HGVS n	ExonicEurc refGene	AF		SIFT scor	SIFT prod
C560T	n S187l		0.462			
C.C.5001	p.5167E		0.402		0.009	
0.000001	p.111951		0.493	COSN25520744	0.049	D
o C224T			0.494	0031123320744	. 0.001	D
0.02241	p.175M,CD79A	framashift dalation	0.493	•	0.001	D
			0.42	COSM007720	· ·	•
C.C3972A			0.451	COSM907720	. 0.004	
C. 1653C	p.12181,CCND3		0.84	COSM5948876	0.001	
c.C174G	p.N58K	nonsynonymous SNV	0.402		0	D
			0.383	COSN4876650	•	•
			0.476			
			0.459	COSN25178966	· ·	•
			0.400			•
			0.521			•
c C884T	n P295I	nonsynonymous SNV	0.021		0.45	T
c C563T	n A188V TRAF3	nonsynonymous SNV	0 459	COSM6032802	0 155	Т
c C10120T	n R3374W	nonsynonymous SNV	0.479	COSM5786167	0 115	Т
c C1413G	p F471L DDX3X	nonsynonymous SNV	0.821		0	D
c 495dupC	p C165fs TNERSE14	frameshift insertion	0.271			2
c C181A	p.Q61K	nonsynonymous SNV	0.276	COSM580	0.009	D
		UTR3-NFKBIZ	0.331			_
		UTR3-NFKBIZ	0.322		· .	
M 031419		UTR3-NFKBIZ	0.529			
c.T659A	p.L220Q.CCND3	nonsynonymous SNV	0.249		0.002	D
c.G908C	p.G303A	nonsynonymous SNV	0.554		0.779	Т
c.C7535T	p.P2512L	nonsynonymous SNV	0.22	COSM27928	0	D
c.C7330T	p.Q2444X	stopgain	0.323	COSM27925.COSM4775092		-
c.C1130T	p.A377V	nonsynonymous SNV	0.285	COSM4384671	0	D
c.C1151T	p.S384F	nonsynonymous SNV	0.35	COSM5757636	0.012	D

c.G7492A	p.A2498T	nonsynonymous SNV	0.424		1	Т
c.C535T	p.R179X	stopgain	0.536			
c.C1057T	p.P353S	nonsynonymous SNV	0.464		0.351	Т
c.G814A	p.V272M	nonsynonymous SNV	0.068		0	D
c.A833C	p.K278T	nonsynonymous SNV	0.499		0.003	D
c.C577T	p.R193W,HDAC7	nonsynonymous SNV	0.493		0	D
c.G1276A	p.G426S	nonsynonymous SNV	0.477		0.058	Т
c.G2035A	p.V679I,BCOR	nonsynonymous SNV	0.963		0	D
c.252_254del	p.84_85del	nonframeshift deletion	0.12			
c.243_247del	p.Q81fs	frameshift deletion	0.127			-
c.C27G	p.N9K	nonsynonymous SNV	0.451	COSM6205982	0.005	D
			0.464			
			0.406			
			0.394			
			0.416			
			0.411			-
			0.408			-
			0.394	COSN23137013,COSN4876655		-
			0.427			-
			0.4			-
			0.403			-
			0.4	COSN23365627,COSN25205836		
			0.398			
			0.383			
			0.387			
			0.362			
			0.373			
			0.366	COSN23353412	-	-
			0.374		-	-
			0.374			
			0.371			
			0.37	COSN8574900		
		-	0.384			
			0.383	•		

		•	0.384			-
		•	0.384			
		·	0.42			
		·	0.416			
		•	0.421			
c.A146G	p.Q49R	nonsynonymous SNV	0.417		0.041	D
c.A217G	p.T73A	nonsynonymous SNV	0.395	COSM1163490,COSM1163491	0.037	D
c.A235G	p.S79G	nonsynonymous SNV	0.404		0.091	Т
c.A1G	p.M1V	nonsynonymous SNV	0.412	COSM220645,COSM220646	0.006	D
c.T1244C	p.L415P	nonsynonymous SNV	0.381	COSM221269	0.004	D
c.C1180T	p.R394C,KLHL21	nonsynonymous SNV	0.512	COSM4009497	0.065	Т
c.G75C	p.E25D	nonsynonymous SNV	0.469		0.074	Т
c.T2758C	p.Y920H,ARID5B	nonsynonymous SNV	0.49		0	D
c.C5839T	p.Q1947X,ARID1A	stopgain	0.273	COSM2235552		
c.G731T	p.G244V	nonsynonymous SNV	0.28		0.246	Т
c.G667A	p.V223I,SETD2	nonsynonymous SNV	0.511		1	Т
c.A4183G	p.M1395V,APC	nonsynonymous SNV	0.479	COSM4169481	0.01	D
c.G110C	p.S37T,HIST1H2BK	nonsynonymous SNV	0.229		0.035	D
c.A1013G	p.Q338R	nonsynonymous SNV	0.493		0.587	Т
c.G4009A	p.D1337N,BCOR	nonsynonymous SNV	0.427	COSM4434915,COSM4434916,CO SM4434917	0.043	D
c.G11062A	p.A3688T,TRRAP	nonsynonymous SNV	0.51	COSM6810121,COSM6810122	0.614	Т
c.C6032T	p.S2011F	nonsynonymous SNV	0.477		0.029	D
c.T191G	p.L64R	nonsynonymous SNV	0.277		0.001	D
c.C166T	p.P56S	nonsynonymous SNV	0.285	COSM1159770	0	D
c.T5744A	p.L1915X,ARID1A	stopgain	0.273			
c.G472A	p.G158R	nonsynonymous SNV	0.504		0.486	Т
		•	0.372		-	
		•	0.385		-	
c.C214T	p.P72S	nonsynonymous SNV	0.382	COSM3316879,COSM3316880	0.029	D
c.G307A	p.A103T,ETS1	nonsynonymous SNV	0.478		0.167	Т
c.C401T	p.A134V,STAT6	nonsynonymous SNV	0.498	COSM4043769	0.273	Т
c.C64T	p.P22S	nonsynonymous SNV	0.484		0	D
c.A16G	p.R6G,TCF3	nonsynonymous SNV	0.462	•	0	D
c.G142A	p.E48K	nonsynonymous SNV	0.35	COSM5947100	0.045	D

		splicing	0.338	COSM4799443		•
c.C497T	p.P166L	nonsynonymous SNV	0.37	COSM3799278	0.001	D
c.3973delC	p.R1325fs,TRIP12	frameshift deletion	0.094			
c.G179A	p.S60N	nonsynonymous SNV	0.241		0.161	Т
c.C287T	p.P96L	nonsynonymous SNV	0.484		0.005	D
c.C536T	p.A179V	nonsynonymous SNV	0.323		0.099	Т
				COSM1487239,COSM1487240,CO SM1487241,COSM1487242,COSM		
c.G495C	p.K165N,SGK1	nonsynonymous SNV	0.328	1487243	0.019	D
c.G431A	p.R144K,SGK1	nonsynonymous SNV	0.08		0.508	Т
c.T370G	p.S124A,SGK1	nonsynonymous SNV	0.085		0.085	Т
c.C320G	p.P107R,SGK1	nonsynonymous SNV	0.304		0.043	D
c.C300G	p.N100K,SGK1	nonsynonymous SNV	0.273		0.164	Т
c.A259C	p.K87Q,SGK1	nonsynonymous SNV	0.308		0.27	Т
c.C238G	p.P80A,SGK1	nonsynonymous SNV	0.238		0.121	Т
c.C227T	p.A76V,SGK1	nonsynonymous SNV	0.247		0.257	Т
c.G536A	p.R179Q,CARD11	nonsynonymous SNV	0.457	COSM6620089	0.101	Т
c.G7195A	p.A2399T	nonsynonymous SNV	0.28		0.051	Т
c.G3A	p.M1I,FAS	nonsynonymous SNV	0.453	COSM6660632	0	D
c.T236A	p.M79K	nonsynonymous SNV	0.247		0.018	D
c.T245G	p.F82C	nonsynonymous SNV	0.245		0.009	D
c.G8C	p.G3A,BCL7A	nonsynonymous SNV	0.238		0.035	D
c.G2803A	p.D935N	nonsynonymous SNV	0.454		0.043	D
c.T157C	p.F53L	nonsynonymous SNV	0.335	COSM555604	0.013	D
c.G250T	p.A84S,IL16	nonsynonymous SNV	0.491		0.016	D
c.C614A	p.T205N,CIITA	nonsynonymous SNV	0.457		0.023	D
exon3	c.276+2T>A	splicing	0.291	COSM4170815		-
c.G280A	p.E94K	nonsynonymous SNV	0.292		0.048	D
c.G185A	p.G62D	nonsynonymous SNV	0.098		0.647	Т
c.C806T	p.P269L	nonsynonymous SNV	0.488		0	D
c.G300C	p.Q100H	nonsynonymous SNV	0.071		0.056	Т

c.C208G	p.L70V	nonsynonymous SNV	0.071	COSM2049209	0	D
c.162_163insTG	p.V55fs	frameshift insertion	0.624			•
c.A943C	p.I315L	nonsynonymous SNV	0.308		0.004	D
c.A1022C	p.Y341S	nonsynonymous SNV	0.291		0	D
c.C56T	p.P19L,MYD88	nonsynonymous SNV	0.414		0.011	D
c.G1153A	p.G385R,NFKB1	nonsynonymous SNV	0.296		0.006	D
c.A5057G	p.K1686R,APC	nonsynonymous SNV	0.38		0.229	Т
c.A5785G	p.T1929A,APC	nonsynonymous SNV	0.4		0.174	Т
c.C340T	p.H114Y	nonsynonymous SNV	0.136		•	
c.A13C	p.S5R	nonsynonymous SNV	0.424		0.176	Т
c.C7958T	p.S2653F	nonsynonymous SNV	0.367		0	D
c.C1466G	p.T489S,TRRAP	nonsynonymous SNV	0.43		0.214	Т
c.A574G	p.S192G	nonsynonymous SNV	0.449		0.054	Т
			0.301		•	
		-	0.297		•	
			0.278		•	
			0.265			•
			0.257			•
				COSN23179746,COSN23224107,C		
				OSN23262474,COSN25184301,C		
				OSN25264478,COSN25522845,C		
				OSN4876647,COSN4876648,COS		
			0.278	N4876649		
			0.274			•
		-	0.278		•	
			0.294		•	
			0.289			•
			0.326			•
			0.32			
			0.324	COSN23136724	•	
		•	0.102			
			0.141		•	
		•	0.342			•

			0.237	COSN25179444,COSN25519226		
			0.235			
c.1138_1139insT						
TTTTTCTTTCAG						
ATGCTGGCTCG						
TGATC	p.F380fs,DDX3X	frameshift insertion	0.35			
c.A680G	p.Q227R	nonsynonymous SNV	0.425		0.796	Т
c.C379T	p.L127F	nonsynonymous SNV	0.34	COSM4171535	-	•
c.C197T	p.A66V	nonsynonymous SNV	0.416		0.081	Т
			0.399			-
			0.394		-	•
			0.067		-	•
			0.354		-	•
			0.37		-	
			0.382	•	-	
			0.368		-	•
			0.433			
			0.414			
			0.409			
c.C49G	p.P17A	nonsynonymous SNV	0.42		0.002	D
c.T59C	p.V20A	nonsynonymous SNV	0.419	COSM1159785,COSM1159786	0.547	Т
c.C169G	p.P57A	nonsynonymous SNV	0.059		0.002	D
c.A200C	p.K67T	nonsynonymous SNV	0.408		0	D
c.C208G	p.L70V	nonsynonymous SNV	0.42		0	D
c.C280G	p.P94A	nonsynonymous SNV	0.433		0.18	Т
c.G308A	p.G103D	nonsynonymous SNV	0.427	COSM1454788,COSM1454789	0.138	Т
c.T458G	p.F153C	nonsynonymous SNV	0.448	COSM1161732,COSM1161733	0.008	D
c.C490G	p.L164V	nonsynonymous SNV	0.442	COSM3316931,COSM3316932	0.056	Т
c.C747A	p.S249R	nonsynonymous SNV	0.407		0.011	D
c.C788G	p.T263S	nonsynonymous SNV	0.398		0	D
c.G62A	p.R21H	nonsynonymous SNV	0.386	COSM5948479,COSM5948480	0	D
c.G1376A	p.R459H,DDX3X	nonsynonymous SNV	0.746	COSM1161825	0.001	D
c.G862A	p.V288I,TP73	nonsynonymous SNV	0.486	COSM3934695,COSM908805	1	Т
c.3218delT	p.M1073fs,PDGFRA	frameshift deletion	0.47		•	•

c.C2083T	p.P695S	nonsynonymous SNV	0.34		0.255	Т
c.G1516A	p.A506T,TNFAIP3	nonsynonymous SNV	0.343		0.272	Т
		unknown	0.354			
			0.409	COSN17149849,COSN17151752		-
			0.372			-
c.G2515A	p.D839N,DOCK8	nonsynonymous SNV	0.098	COSM3657440,COSM3657441	0.102	Т
c.C9160T	p.R3054C	nonsynonymous SNV	0.126	COSM2186557	0.003	D
c.C1441A	p.L481I	nonsynonymous SNV	0.36		0.553	Т
c.T1115C	p.I372T,TP73	nonsynonymous SNV	0.179		0.002	D
				COSM1341625,COSM1341626,CO		
c.1633delA	p.K545fs,EPB41	frameshift deletion	0.18	SM6654552	-	
c.C3281T	p.T1094I	nonsynonymous SNV	0.136		0.004	D
c.C5771A	p.P1924H	nonsynonymous SNV	0.141		0.002	D
c.C47A	p.T16K	nonsynonymous SNV	0.418		1	Т
c.A1805G	p.Y602C,BCL6	nonsynonymous SNV	0.169		0	D
				COSM1149856,COSM117308,COS		
				M117309,COSM117310,COSM229		
c.G1040A	p.R347H,FBXW7	nonsynonymous SNV	0.123	65	0.001	D
c.G873A	p.M291I,TAP1	nonsynonymous SNV	0.153		0.002	D
c.2662delC	p.R888fs,CARD11	frameshift deletion	0.131		•	
c.G625A	p.A209T,CARD11	nonsynonymous SNV	0.113	COSM5138977	0.071	Т
c.G10235A	p.R3412Q	nonsynonymous SNV	0.137		0	D
c.C791T	p.P264L	nonsynonymous SNV	0.142			
			0.2			
c.397delT	p.F133fs,FAS	frameshift deletion	0.17	COSM1349688	•	
c.G686A	p.R229Q,FAS	nonsynonymous SNV	0.144	COSM5703782	0	D
c.C8226G	p.N2742K	nonsynonymous SNV	0.113		0.011	D
c.C2275A	p.L759I	nonsynonymous SNV	0.108	COSM946771,COSM946772	0.444	Т
c.G4343A	p.G1448E	nonsynonymous SNV	0.439		0.008	D
c.C1216T	p.Q406X,IL16	stopgain	0.138		•	
c.C2107T	p.R703C	nonsynonymous SNV	0.092		0.021	D
c.C3665T	p.T1222M,CREBBP	nonsynonymous SNV	0.133		0.009	D
c.C412T	p.R138C,CD70	nonsynonymous SNV	0.141	COSM1200315	0.011	D

c.G280T	p.G94W,CD70	nonsynonymous SNV	0.117		0	D
c.1285delC	p.L429fs,POU2F2	frameshift deletion	0.214	COSM4611039,COSM4611040	-	-
c.T76C	p.S26P	nonsynonymous SNV	0.364		0	D
c.C7507T	p.Q2503X	stopgain	0.15	COSM28669,COSM4775114		
				COSM10662,COSM1640830,COS		
				M3356964,COSM99020,COSM990		
c.G347A	p.R116Q,TP53	nonsynonymous SNV	0.181	21,COSM99602	0.005	D
c.A184T	p.K62X,CD58	stopgain	0.153			
c.C6269A	p.S2090X	stopgain	0.066			
c.G1423A	p.V475M,CIITA	nonsynonymous SNV	0.425	COSM966958	0.164	Т
				COSM10810,COSM129834,COSM		
				129835,COSM129836,COSM1646		
c.G329T	p.C110F,TP53	nonsynonymous SNV	0.136	852,COSM3378347	0	D
c.G413A	p.G138D	nonsynonymous SNV	0.901		0.466	Т
c.A1517G	p.Y506C,FBXO11	nonsynonymous SNV	0.498		0.078	Т
c.568_570del	p.190_190del,GNAI2	nonframeshift deletion	0.203			-
			0.214	COSN25450465		-
			0.144			-
			0.229			-
			0.223			-
c.A1406C	p.H469P,DDX3X	nonsynonymous SNV	0.163		0.109	Т
c.A1444G	p.T482A,DDX3X	nonsynonymous SNV	0.114	COSM1203253	0	D
c.A6274G	p.S2092G	nonsynonymous SNV	0.422		0.346	Т
c.C542T	p.A181V,BCL6	nonsynonymous SNV	0.385		0.19	Т
c.C1280A	p.P427H	nonsynonymous SNV	0.4		0.354	Т
c.C3125T	p.S1042L,RELN	nonsynonymous SNV	0.482		0.651	Т
c.A2077G	p.T693A	nonsynonymous SNV	0.472		0.159	Т
c.C2936A	p.T979N	nonsynonymous SNV	0.415		0.015	D
c.G2366A	p.R789H,UNC5D	nonsynonymous SNV	0.183	COSM1231806	0.001	D
c.245_255del	p.R82fs,ZFP36L1	frameshift deletion	0.142			
c.G130C	p.E44Q	nonsynonymous SNV	0.439		0.128	Т
c.2466_2483del	p.822_828del,SETD2	nonframeshift deletion	0.456			

c.G1591A	p.A531T,BCL6	nonsynonymous SNV	0.363	COSM220321	0.013	D
c.A3119G	p.D1040G,APC	nonsynonymous SNV	0.495	COSM6503097	0.005	D
			0.485			
			0.521			
			0.592	COSN23136164		
			0.496			
c.A64G	p.T22A,MYC	nonsynonymous SNV	0.48		1	Т
c.G159C	p.E53D,MYC	nonsynonymous SNV	0.537	COSM1159795,COSM1159796	0.152	Т
c.A214G	p.T72A,MYC	nonsynonymous SNV	0.56	COSM1163490,COSM1163491	0.037	D
c.G239A	p.R80H,MYC	nonsynonymous SNV	0.114		0	D
c.G336C	p.Q112H,MYC	nonsynonymous SNV	0.511	COSM1159799,COSM1159800	0.004	D
c.C678G	p.D226E,MYC	nonsynonymous SNV	0.51		0.115	Т
c.C744A	p.S248R,MYC	nonsynonymous SNV	0.457		0.011	D
c.T755C	p.L252P,MYC	nonsynonymous SNV	0.468		0.001	D
c.C773T	p.T258I,MYC	nonsynonymous SNV	0.163		0.012	D
c.C77G	p.P26R	nonsynonymous SNV	0.376		0	D
c.G8675A	p.R2892Q	nonsynonymous SNV	0.446		0.523	Т
				COSM11089,COSM116921,COSM 116922,COSM116923,COSM1169		
c.T188C	p.I63T,TP53	nonsynonymous SNV	0.092	24,COSM1645297,COSM3421936	0	D
c.T20G	p.I7S,CLTC	nonsynonymous SNV	0.093		0	D
c.T1659G	p.N553K,TCF3	nonsynonymous SNV	0.416		0.001	D
	c.395+2T>C;NM_00119341					
exon4	6	splicing	0.675			
c.A44C	p.K15T,BCL7A	nonsynonymous SNV	0.238		0.001	D

unhan? coord	Polyphon2 prod	Mutation Accessor, coor	MutationAccosor prod	CADD phrod	Driver Production
0.013	R	0 955		19 41	Passenger
0.010	B	0.345	N	18.6	Passenger
0.110	5			1010	Intronic
1	D	2.545	M	23.8	Driver
					Driver
				38	Driver
0.361	В	2.265	М	23.5	Driver
1	D	0.45	N	27	Passenger
					Intronic
		•			Intronic
					Intronic
			•		Intronic
			•		Intronic
0.054	В	0.945	L	4.476	Passenger
0.01	В	0.625	Ν	19.46	Driver
0.004	В	0.255	Ν	13.72	Driver
1	D	1.62	L	32	Passenger
					Driver
0.948	Р	3.42	Μ	24.1	Driver
					Driver
					Driver
					Driver
0.998	D	2.435	Μ	26.7	Driver
0	В	0.345	Ν	0.004	Passenger
1	D	2.88	Μ	31	Driver
				40	Driver
1	D	3.475	Μ	34	Driver
0.994	D	1.87	L	23.4	Benign variant (ClinVar ID 41541)

0	В	-0.59	Ν	0.003	Passenger
				26.4	Driver
0.047	В	1.245	L	0.229	Passenger
0.938	Р	2.225	М	25.7	Driver
0.999	D	3.02	М	25.2	Driver
1	D	3.11	М	32	Driver
0.05	В	1.53	L	24.6	Passenger
0.999	D	2.095	Μ	23.9	Driver
					Driver
					Driver
				21.5	Driver
					Intronic
		•	•		Intronic
		•			Intronic
					Intronic
		•			Intronic
		•	-		Intronic
			-		Intronic
					Intronic
		•			Intronic
			•		Intronic
					Intronic
•	•	•	•	-	Intronic

•					Intronic
					Intronic
0.145	В	1.59	L	12.37	Passenger
0.994	D	3.195	Μ	25.7	Driver
0.961	D	2.385	Μ	23	Driver
0	В		•	12.55	Driver
1	D	2.665	Μ	24.1	Driver
0.138	В	0.765	Ν	24.5	Driver
0	В	1.68	L	15.52	Passenger
0.999	D	1.845	L	25.8	Passenger
			•	42	Driver
0.977	D	2.08	Μ	21.7	Driver
0.001	В	-0.255	Ν	0.001	Passenger
0.03	В	0.975	L	19.87	Driver
0.131	В	0.125	Ν	23.8	Passenger
0	В	2.03	Μ	7.112	Driver
0.839	Р	1.355	L	25.2	Driver
0.493	Р	-0.28	Ν	21.2	Driver
0.976	D	2.48	Μ	24.7	Driver
0.998	D	2.445	Μ	26.7	Driver
1	D	3.57	Н	28.1	Driver
				37	Driver
0	В	0	Ν	18.37	Passenger
					Intronic
			-		Intronic
0.999	D	2.975	Μ	27.1	Driver
0.987	D	1.59	L	26.1	Passenger
0.252	В	1.87	L	22.1	Driver
0.98	D	0	N	15.19	Passenger
1	D	2.695	Μ	22.3	Driver
0.02	В	2.005	Μ	22.7	Driver

				25.1	Driver
0.804	P	3.06	M	24.8	Driver
					Driver
0.003	В	-0.69	N	11.84	Passenger
0.022	В	1.445	L	14.72	Passenger
0.056	В	1.095	L	23.3	Passenger
0.203	В	-0.035	N	22.8	Driver
0	В	0.835	L	21.3	Passenger
0	В	1.87	L	17.98	Passenger
0.265	В	1.955	М	23.5	Driver
0.004	В	1.67	L	22.7	Passenger
0.004	В	1.39	L	21.3	Passenger
0.009	В	0.83	L	22.6	Passenger
0.001	В	2.08	М	23.6	Driver
0.898	Р	2.085	М	28.7	Driver
0.002	В	2.125	М	14.68	Driver
0.225	В			23.4	Driver
0.968	D	2.465	М	23.9	Driver
0.992	D	2.4	М	23.7	Driver
0.714	Р	1.79	L	24.5	Passenger
					Benign variant
					detected in germline
0.055	В	2.67	М	8.869	(ClinVar ID 51349)
0.989	D	2.7	М	28	Driver
0.267	В	2.25	М	2.046	Driver
0.95	Р	0	N	9.318	Passenger
				25.2	Driver
0.976	D			23.9	Driver
0.005	В	0.345	Ν	10.29	Passenger
0	В	0	Ν	21	Passenger
0.115	В	0.805	L	12.86	Passenger

1	D	3.515	Н	25	Driver
					Driver
0.948	Р	2.465	М	24.1	Driver
0.949	Р	2.08	М	24.7	Driver
0	В	2.2	М	23.3	Driver
0.997	D	2.005	М	25.4	Driver
0	В	1.65	L	8.353	Passenger
0	В	1.845	L	8.716	Passenger
				24.6	Driver
0.031	В	1.845	L	1.131	Passenger
0.985	D	0	N	6.005	Passenger
0.006	В	0.09	N	10.26	Passenger
0.109	В	0.84	L	9.801	Passenger
					Intronic
					Intronic
					Intronic
					Intronic
			-		Intronic
			-		Intronic
			-		Intronic
					Intronic
		•			Intronic

					Intronic
					Intronic
					_ .
					Driver
0	В	-1.2	N	1.232	Passenger
			•	24.6	Driver
0.954	P	2.685	M	27.3	Driver
					Intronic
	•				Intronic
•			-		Intronic
					Intronic
1	D	2.9	Μ	25.7	Driver
0	В	-0.41	Ν	4.024	Driver
0.999	D	2.395	Μ	24.7	Driver
0.997	D	3.3	Μ	25.9	Driver
0.994	D	3.305	М	26	Driver
0.015	В	0.49	Ν	0.005	Passenger
0.807	Р	0.69	Ν	17.6	Driver
1	D	3.21	М	27.2	Driver
0.509	Р	2.31	Μ	23.5	Driver
0.582	Р	2.25	М	23.6	Driver
0.979	D	0.35	Ν	21.3	Passenger
1	D	2.89	М	31	Driver
1	D	1.93	L	34	Driver
0.001	B	-0.9	N	0.002	Driver
					Driver

0.002	В	0.55	Ν	0.434	Passenger
0.002	В	0	N	10.33	Passenger
0.956	Р	2.84	М	26.2	Driver
					Intronic
					Intronic
0.661	Р	2.215	М	24.4	Driver
0.998	D	2.565	Μ	26.1	Driver
0.982	D	2.08	Μ	18.23	Driver
0.911	Ρ	-		22.8	Driver
					Driver
0.078	В	1.245	L	5.347	Passenger
0.688	Р	1.15	L	0.984	Passenger
0	В	0	Ν	8.384	Passenger
1	D	2.39	Μ	27.1	Driver
1	D	1.255	L	35	Driver
0.998	D	2.705	М	32	Driver
					Driver
0.89	Р	1.52	L	28.6	Driver
1	D	2.395	М	27.6	Driver
1	D	4.805	Н	24.2	Driver
					Intronic
					Driver
1	D	3.005	М	33	Driver
0.933	Ρ	1.93	L	24.1	Passenger
					Benign variant
0	В	0.225	Ν	0.001	(ClinVar ID 51264)
0.983	D	2.135	Μ	24.1	Driver
	•	•		41	Driver
0.69	Ρ	1.65	L	21.5	Passenger
1	D	2.3	Μ	24.3	Driver
0.672	Р	1.24	L	24.5	Driver

1	D	1.7	L	27.8	Passenger
					Driver
0.994	D	0	Ν	11.98	Passenger
				47	Driver
1	D	2.935	Μ	34	Driver
-				35	Driver
-				37	Driver
0.631	Р	1.18	L	0.045	Driver
1	D	3.325	Μ	33	Driver
0	В	-0.75	N	0.483	Passenger
0.991	D	1.945	M	24.3	Driver
•					Driver
•					Intronic
•					Intronic
-					Intronic
-		-			Intronic
0.002	В	1.61	L	15.94	Passenger
1	D	4.01	Н	27.2	Driver
0	В	0.805	L	8.424	Passenger
0.267	В	1.265	L	16.5	Passenger
0.214	В	1.04	L	17.67	Passenger
0	В	0.45	Ν	6.076	Passenger
0.022	В	1.245	L	0.001	Passenger
0.995	D	3.015	Μ	23.7	Driver
1	D	2.81	Μ	34	Driver
					Driver
0.012	В	1.065	L	22.4	Passenger
					Driver

1	D	0.775	Ν	26.4	Driver
0.376	В	0.895	L	22.1	Driver
					Intronic
		•			Intronic
					Intronic
					Intronic
0	В	-1.125	N	0.002	Passenger
0.219	В	1.065	L	14.18	Driver
0.994	D	3.195	Μ	25.7	Driver
1	D	2.74	Μ	32	Driver
0.545	Р	2.21	М	23.4	Driver
0.224	В	-0.06	N	3.49	Passenger
0.582	Р	2.25	Μ	23.6	Driver
0.739	Р	0.925	L	12.24	Passenger
0.827	Р	2.05	Μ	23.5	Driver
0.123	В	2.9	Μ	23.8	Driver
0.018	В	0.805	L	9.692	Passenger
1	D	3 12	м	23.9	Driver
0.007	B	3.41	M	20.0	Driver
0.999	D	3.2	M	27.2	Driver
0.000	-	0.2			2
				24.7	Driver
0.643	Р	2.075	Μ	13.11	Driver

Supplemental Table 7. Frequency of driver mutations in pediatric PTLD-DLBCL. IC-DLBCL¹³ and in adult EBV-positive and negative PTLD-DLBCL.^{23,24} Frequencies marked in bold represent statistically significant differences with pediatric PTLD-DLBCL, according to Fisher's exact test * P<0.1..

Gene	Frequency of driver mutations(%)						
	Ped IC-DLBCL (n=22)	Ped PTLD-DLBCL (n=18)	Ped EBV+ PTLD- DLBCL (n=16)	Adult EBV+ PTLD DLBCL (n=16)	Adult EVB- PTLD- DLBCL (n=28)		
SOCS1	27.3	0	0	6.25	3.57		
KMT2D	22.7	0	0	18.75	35.71		
KMT2C	9.1	16.7	12.5	6.25	7.14		
CARD11	0	11.1	6.2	0	7.14		
CD70	4.5	11.1	12.5				
NOTCH1	0	11.1	12.5	6.25	14.29		
TP53	9.1	11.1	6.2	12.5	42.86		
MYD88	13.6	0	0	6.25	10.71		
GNA13	13.6	5.6	0	0	7.14		
PIM1	13.6	0	0	6.25	10.71		
BTG1	13.6	0	0	6.25	10.71		
EZH2	13.6	0	0	6.25	14.29		
PRDM1	4.5	0	0	0	10.71		
PIK3CD		0	0	18.75	3.57		
DDX3X	4.5	5.6	6.2				
SGK1	0	5.6	0	0	7.14		
SPTBN5	0	11.1	12.5				