1 Harnessing the immune-mediated cancer field as chemopreventive strategy for

- 2 hepatocellular carcinoma
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4 **Short-title:** Immune-mediated field cancerization as target for HCC prevention

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35 **Abbreviations:** ALT: alanine aminotransferase; α-SMA: α-smooth muscle actin; AKT: 36 protein kinase B; AST: aspartate aminotransferase; DEN: diethylnitrosamine; CCl4: 37 carbon tetrachloride; EMT: epithelial-mesenchymal transition, ERK: extracellular signal-38 regulated kinase; FC: Fold change, FDR: false discovery rate; FFPE: formalin-fixed 39 paraffin-embedded; FGFR: fibroblast growth factor receptor; GSEA: gene set 40 enrichment analysis; HCC: hepatocellular carcinoma; HCV: hepatitis C virus; HSC: 41 hepatic stellate cell; ICF: immune-mediated cancer field; IPA: ingenuity pathway 42 analysis; mo: months; NTP: nearest template prediction; PDGFR: platelet-derived 43 growth factor receptor; ssGSEA: single sample gene set enrichment analysis; TKIs: 44 tyrosine kinase inhibitors; Treq: regulatory T cells; gRT-PCR: guantitative real-time 45 polymerase chain reaction; VEGFR: vascular endothelial growth factor receptor.

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#### 91 ABSTRACT

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Background & aims: Cirrhosis and chronic inflammation precede hepatocellular
carcinoma (HCC) development in ~80% of cases. We sought to understand the
molecular and immune-related features governing the chronic inflammation from which
HCC arises, as a prelude to optimizing chemoprevention strategies.

97 **Methods:** Gene expression profiling of non-tumor liver tissues from 392 early HCC 98 patients [training (n=167); validation (n=225)] was analyzed to characterize the immune 99 features of the surrounding field. A 172-gene signature capturing the immune-mediated 100 cancer field (ICF) was then tested in a cohort of cirrhotic patients (n=216; median 101 follow-up 10 years) to predict risk of HCC development and outcome. The preventive 102 efficacy of targeting this ICF was assessed in a chemically-induced murine model of 103 chronic liver damage and hepatocarcinogenesis.

104 Results: An immune-mediated cancer field of the non-tumoral liver was identified in 105 60% of early HCC cases and in 50% of cirrhotic patients without HCC. This immune 106 field effect comprised three distinct subtypes: 1) High Infiltrate-ICF with increased 107 effector T cells; 2) Immunosuppressive-ICF with stromal activation and TGF-B 108 signaling, and; 3) Pro-inflammatory-ICF with up-regulation of IFN- $\gamma$  signaling. The 109 Immunosuppressive-ICF (10% of cirrhotic patients)(HR:2.41; 95% 1.21-4.80) – as well 110 as platelet count <100,000/mm<sup>3</sup>- were independent predictors of HCC development in 111 cirrhotic patients, Nintedanib, an anti-inflammatory and anti-angiogenic kinase inhibitor. 112 reverted this pro-carcinogenic field and significantly delayed HCC onset in preclinical 113 models.

114 **Conclusions:** An immunosuppressive *milieu* in non-tumor liver heightens the risk of 115 HCC development. Targeted therapies can revert this pro-carcinogenic field and 116 prevent HCC development in preclinical models. These data establish a rationale for 117 exploring chemopreventive strategies in patients at-risk of HCC development.

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### 119 Keywords:

120 Cancer field, immunosuppression, hepatocellular carcinoma, chemoprevention

#### 122 INTRODUCTION

Liver cancer is the fourth leading cause of cancer-related mortality worldwide<sup>1</sup>. 123 124 Hepatocellular carcinoma (HCC) accounts for more than 90% of liver cancers and is the main cause of death in patients with cirrhosis<sup>2,3</sup>. HCC cases arise from chronic liver 125 126 inflammation, fibrosis and eventually cirrhosis in 70-80% of cases<sup>2</sup>. In developed 127 countries, curative treatments are feasible in 30-40% of cases, but recurrence is high and no effective adjuvant therapies are available<sup>2,4</sup>. In addition, ~40-50% of patients 128 129 are diagnosed at advanced stages when currently approved molecular therapies yield 130 limited survival benefits (~1 year)<sup>5-8</sup>. Despite recent advances in the management and 131 clearance of HCV infection, there is an unmet need for early detection and application 132 of chemopreventive approaches in patients at high-risk of HCC development.

133 To date, there are no established preventive strategies for HCC in patients at risk beyond prevention with anti-viral therapies<sup>9,10</sup>. Once cirrhosis is established, anti-viral 134 therapies reduce but do not eliminate the risk of HCC<sup>4,11,12</sup>. Individual risk assessment 135 136 is a key first step in the successful development of any chemopreventive strategy. In 137 this regard, increasing evidence suggests the existence of the so-called "cancer field-138 effect" or field cancerization which consists of predisposing oncogenic and 139 inflammatory signals occurring during chronic liver injury and ultimately leading to malignant transformation<sup>13–15</sup>. Gene signatures derived from the cirrhotic tissue 140 141 adjacent to HCC tumors have been designed to predict poor outcome, particularly in HCV-infected cirrhotic patients at higher risk of HCC development<sup>13,16–19</sup>. Overall, these 142 143 studies support the feasibility of using molecular scores of the carcinogenic field to 144 identify patients at high risk of HCC development. However, the pro-carcinogenic roles 145 of inflammation and immune response in the context of the field cancerization have 146 been poorly explored. Understanding the immune features governing the unresolved 147 cancer field-effect is crucial for identifying potential therapeutic targets in patients at 148 high risk of HCC development.

149 In this study, the analysis of the inflammatory *milieu* that characterizes the underlying 150 liver disease in which HCC tumors arise has led to the identification of an immune-151 mediated cancer field (ICF) in 60% of early HCC patients and 50% of cirrhotic patients 152 without HCC. This ICF comprises three distinct molecular subtypes including the High 153 Infiltrate-ICF subtype with increased infiltration of effector T cells, the 154 *Immunosuppressive-ICF* subtype with activation of stroma and TGF- $\beta$  signaling, and 155 the *Pro-inflammatory-ICF* subtype with up-regulation of IFN-γ signaling. These immune 156 profiles, particularly the presence of the *Immunosuppressive-ICF* cancer field, predicts 157 increased risk of HCC development in cirrhotic patients. Inhibition of this procarcinogenic field with nintedanib, an anti-inflammatory and anti-angiogenic kinase
 inhibitor approved in pulmonary fibrosis, significantly delays HCC onset in a mouse
 model of chronic liver damage and hepatocarcinogenesis. Overall, our study provides
 the rationale to explore chemopreventive strategies in cirrhotic patients at high-risk of
 HCC development.

163

## 164 MATERIALS AND METHODS

#### 165 Human cohort

166 For the purpose of the study, gene expression data from a cohort of 167 surgically 167 resected fresh-frozen samples (Heptromic dataset, GSE63898) with matched tumor 168 and adjacent non-tumor tissue were analyzed. All samples were previously collected 169 (1998-2008) in the setting of the HCC Genomic Consortium upon institutional review 170 board approval. Full description of the cohort and RNA profiling data are available in 171 previous publications<sup>20,21</sup>. Supplementary Table 1 provides a summary of the clinical-172 pathological variables of the samples used in the current study (referred as training 173 cohort, n=167). Validation of the identified molecular profiles was then performed in an 174 independent set of 225 adjacent non-tumor liver tissues previously characterized by 175 our group (GSE10143)<sup>13</sup>. Finally, for the purpose of identifying those non-neoplastic 176 patients at higher risk of HCC development and most likely to benefit from 177 chemopreventive strategies, our findings were evaluated in a previously characterized cohort of patients with early cirrhosis (n=216, GSE15654)<sup>16</sup> and a publicly available 178 179 dataset of fibrotic liver tissues (n=124, GSE84044)<sup>22</sup>.

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#### 181 Modeling the immune-mediated cancer field

182 Enrichment scores of 4872 gene sets that represent cell states and perturbations of the 183 immune system (Collection C7 of MSigDB, Broad Institute)<sup>23</sup> were calculated by Single-184 sample Gene Set Enrichment Analysis (ssGSEA) in the non-tumor liver tissue of the 185 study cohort. Next, unsupervised clustering analysis by non-negative matrix factorization (NMF consensus)<sup>24</sup> method was performed to identify the presence of an 186 187 immune-mediated cancer field. To characterize the molecular features of those 188 samples presenting an ICF and to identify different immune-mediated field subtypes, a second unsupervised clustering was performed using ssGSEA scores obtained for a 189 190 curated set of gene signatures representative of individual cell types<sup>25,26</sup> (lymphocytes, 191 macrophages, dendritic cells, mast cells, neutrophils, and eosinophils), cancer 192 immune-related signaling pathways<sup>27</sup> (infiltrating lymphocyte, TGF-β response, IFN-γ response, macrophage activation, and wound healing), and inflammation- or immune specific biological processes (Hallmark collection of MSigDB, Broad Institute).

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### 196 Generation of an immune-mediated field gene signature

An ICF field gene signature was generated using top differentially expressed genes in each molecular group (FDR<0.05; Fold-change  $\geq$ 2). The ability of the gene signature to capture the key molecular features of the immune-mediated cancer field subtypes was then validated in an independent dataset using Nearest Template Prediction (NTP) analysis using p-value<0.05 for defining significant prediction (Gene Pattern modules)<sup>28</sup>.

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# 204 Molecular characterization of the ICF subtypes and identification of candidate205 therapies

206 Enrichment of additional molecular pathways and gene expression signatures in the ICF subtypes was evaluated using GSEA, ssGSEA, NTP and Ingenuity Pathway 207 208 Analyses (IPA). All gene signatures used are available in Molecular Signature 209 Database (MSigDB, Broad Institute) or were previously reported (Supplementary 210 Table 2). CIBERSORT<sup>25</sup> (cell-type identification by estimating relative subsets of RNA 211 transcripts) was used to estimate the relative fraction of 22 immune cell types within the 212 leukocyte compartment of non-tumor liver tissues. The Immunophenoscore (IPS) 213 algorithm<sup>29</sup> was used to analyze the major immunogenic determinants. An *in silico* 214 analysis based on ssGSEA scores of ~1230 gene sets (DSigDB) recapitulating targets 215 of approved therapies was also performed for the screening of candidate targeted 216 therapies. This approach provided the rationale for testing nintedanib in a preclinical 217 model of HCC development.

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### 219 Histological evaluation of infiltrating inflammation

Histopathological analysis was performed in 98 out of 167 cases. Specifically, hematoxylin and eosin (H&E) staining of formalin-fixed paraffin embedded (FFPE) tissue section of HCCs and their matched adjacent non-tumor livers were evaluated by two expert pathologists (CM and MS). The presence of inflammation (portal/septal, interface, pericentral and lobular) as well as the lymphoid aggregates were assessed in the non-tumor liver tissue sections. More details on the histological evaluation of the samples have been included in **Supplementary material**.

227

228 Animal model

229 For the purpose of this study, we generated a chemically-induced model of HCC and 230 fibrosis in male C57BL/6J mice (Harlan Laboratories, n=58) by a single injection of N-231 nitrosodiethylamine (DEN) followed by weekly dosing with carbon tetrachloride (CCl<sub>4</sub>), 232 as previously described<sup>30</sup>. Once fibrosis was established, mice were randomized to 233 receive vehicle or nintedanib (50 mg/kg, Boehringer Ingelheim). Mice were then 234 sacrificed at different time-points and liver and tumor tissue samples were collected 235 and processed for molecular characterization of histology, the transcriptome and 236 protein expression. All experimental procedures were carried out following the approval 237 of the institutional ethical committee of the University of Barcelona and Hospital Clinic 238 of Barcelona. For specific details, see **Supplementary material**.

239

### 240 Statistical analysis

241 All analyses were performed using SPSS software version 23 (IBM) or GraphPad 242 Prism version 5.00 (San Diego, CA). Correlations for categorical and continuous 243 variables were analyzed by Fisher's exact test and Wilcoxon rank-sum test, 244 respectively. The prognostic value of the signatures was assessed using Kaplan-Meier 245 estimates, log-rank test, and Cox regression models. In in vivo studies, the Mann-246 Whitney U test was used to compare differences in body weights, liver function, tumor 247 number, and tumor size. Fisher exact test was performed for analysis of HCC 248 incidence and pERK staining. Student T-Test was used to compare the differences in 249 Sirius Red guantification, CD31 staining, CD4/CD8 staining proportion of immune cell 250 infiltrate and relative gene expression.

251

#### 252 **RESULTS**

# 253 Identification of a novel immune-mediated cancer field effect in non-tumor liver 254 tissue of patients with early HCC.

255 In order to characterize the immune features governing the unresolved cancer-field in 256 which new cancers arise, transcriptome-based analysis of a compendium of ~5,000 257 annotated immunology-specific gene-sets<sup>23</sup> was performed in the non-tumor liver tissue 258 of patients with early stage HCC. This analysis revealed the presence of an immune-259 mediated cancer field (ICF) in ~60% (98/167) of samples (Figure 1A and 260 Supplementary Figure 1). Specifically, these samples were characterized by 261 enrichment of several gene-sets recapitulating the presence of activated immune cells, 262 up-regulation of core signaling pathways involved in immune response (both innate and 263 adaptive) as well as those involved in the modulation of inflammatory response (i.e. 264 IL2-STAT5, IL6-STAT3, IL17, IFN-γ, CSF, TNF-α, and TGF-β signaling) (Figure 1A-B

265 and Supplementary Figure 1). Moreover, histological evaluation confirmed that liver 266 tissues with ICF contained a higher frequency of moderate to marked inflammatory 267 infiltrate (74% in ICF vs. 52% in non-ICF, p=0.034) and lymphoid aggregates (80% in 268 ICF and vs. 55% in non-ICF, p=0.009) (Figure 1C-1D and Supplementary Table 3). 269 In contrast, histological evaluation of the tumor showed no significant correlation 270 between the presence of the ICF and the detection of intratumoral or peritumoral 271 infiltration (Supplementary Table 3). This is in accordance with our recent publication<sup>20</sup>, where the tumor immune-based profile did not correlate with presence or 272 273 absence of immune gene signatures in the surrounding non-tumor tissue.

274 While characterizing the ICF we detected that, in addition to immunogenic features, 275 several well-known pro-carcinogenic signals such as epithelial-to-mesenchymal 276 transition, KRAS, EGFR, and VEGF signaling were also significantly enriched in liver 277 tissues containing the ICF (Supplementary Table 4). In line with these oncogenic 278 signals, a significant enrichment of previously reported prognostic signatures derived 279 from the adjacent non-tumoral liver were also detected. These signatures included the 280 186-gene cancer-field signature<sup>13</sup>, activated hepatic stellate cells (HSCs)<sup>17</sup>, hepatic injury and regeneration (HIR)<sup>19</sup>, and multicentric occurrence of HCCs<sup>31</sup> (Figure 1A). 281 282 The presence of the ICF significantly correlated with HCV infection, poor survival 283 [median OS 43.4 mo in the ICF group vs 94.8 mo in non-ICF; p=0.001] and features 284 indicative of liver dysfunction such as high bilirubin, low platelet count and albumin 285 levels (Supplementary Table 5 and Supplementary Figure 1B). Altogether, our data 286 highlight the presence of an immune-mediated cancer field in 60% of early HCC patients. This ICF is characterized by activation of immunomodulatory signaling 287 288 cascades (i.e. IFN-y, TNF- $\alpha$ , TGF- $\beta$ , IL6) along with cancer promoting signaling 289 pathways (i.e. EMT, EGFR and VEGFR), and is associated with HCV infection and 290 poor prognosis.

291

#### 292 The immune-mediated cancer field contains 3 distinct molecular subtypes.

293 Further dissection of the key immune-modulating signaling pathways and immune-cell 294 infiltrates in those samples harboring the immune-mediated cancer field revealed the 295 existence of three distinct molecular subtypes. The first molecular subtype, henceforth called the "High Infiltrate-ICF" subtype (23% of the ICF), showed a significant 296 297 enrichment of several previously established gene signatures mirroring the presence 298 and/or activation of immune cell infiltrates such as lymphocytes (T and B cells)<sup>27,32</sup>, 299 macrophages<sup>33</sup> and ectopic lymphoid structures<sup>34</sup> (Figure 2A-2B). Consistently, 300 immunogenicity, herein captured either by the cytolytic activity score (Figure 2A)<sup>35</sup> or

301 using the immunophenoscore algorithm<sup>29</sup> (Figure 2C), was also significantly higher in 302 these samples (p<0.001). Specifically, non-tumor liver samples belonging to the High 303 Infiltrate-ICF subtype showed significant infiltration of effector cells, including activated 304 and memory CD8+/CD4+ T cells (p<0.01) together with up-regulation of the main 305 components of the major histocompatibility complex (MHC) class I and class II (Figure 306 **2C**, p≤0.001). The second subtype, the so-called "Immunosuppressive-ICF" (36% of 307 the ICF), was characterized primarily by activation of stroma and HSCs, increased 308 TGF- $\beta$  signaling and T cell exhaustion (Figure 2A). Moreover, several immune-309 checkpoint inhibitors (i.e. CTLA-4, TIGIT, LAG3) were significantly over-expressed 310 (p<0.01) in this class, along with higher levels of M2 macrophages (p<0.05), activated 311 mast cells (p < 0.05), and neutrophils (p < 0.01) which are among the main mediators of 312 immune tolerance and inhibition (Figure 2B-2C). The third subtype (41% of the ICF) 313 showed a clear predominance of IFN-y signaling (p<0.001) and enrichment of the 314 inflammatory M1 macrophages (p<0.001) and follicular helper T cells (p<0.05), and 315 was called the "Pro-inflammatory-ICF" subtype (Figure 2). Interestingly, the High 316 Infiltrate and Immunosuppressive subtypes shared several molecular features including 317 the enrichment of key signaling pathways involved in modulating the immune response 318 (i.e. IL2 and TNF signaling), proliferation (i.e. KRAS signaling) and angiogenesis 319 (**Figure 2A**, p<0.001).

320 In order to further confirm the presence and molecular traits of the identified ICF, we 321 generated a transcriptome-based gene signature able to capture the three immune-322 mediated cancer field subtypes. Interestingly, this signature only showed minimal 323 overlap (0-5%) with previously reported gene signatures of field cancerization in HCC (Supplementary Table 6) <sup>13,16,18,36</sup>. The predictive capacity of the resulting 172-gene 324 325 signature was then validated in the adjacent non-tumor tissue of 225 patients with early 326 HCC, previously characterized by our group<sup>13,37</sup> (Supplementary Figure 2, 327 Supplementary Table 7). Similar to what previously observed in the training cohort, 328 58% (130/225) of patients belonged to the immune-mediated field. Within this group, 329 ~31% (40/130) presented the High Infiltrate-ICF profile, ~27% (35/130) the 330 Immunosuppressive-ICF and ~42% (55/130) the Pro-inflammatory-ICF subtype 331 (Supplementary Figure 2). Subsequent characterization further confirmed the ability 332 of the signature to capture the main molecular traits defining each subtype, such as 333 increased infiltration of effector T cells in High Infiltrate subtype, activation of stroma 334 and TGF- $\beta$  signaling in *Immunosuppressive* subtype and up-regulation of IFN- $\gamma$ 335 signaling in *Pro-inflammatory* subtype (**Supplementary Figure 2**). Overall, our results 336 highlight the presence of 3 molecular subtypes within the immune-mediated cancer field with high degree of lymphocyte infiltration (overall 16% of HCC patients) or either
immunosuppressive (overall 20% of HCC patients) or pro-inflammatory (24% of HCC
patients) signaling cascades.

340

# The immunosuppressive-ICF subtype predicts high risk of HCC development in cirrhotic patients

343 Following the identification of an immune-mediated cancer field in the livers of 60% of 344 patients with early HCC, we next sought to assess its role in liver disease progression 345 and HCC primary occurrence. To this end, the 172-gene signature was analyzed in a 346 cohort of 216 non-malignant cirrhotic patients with a median follow-up of 10 years in the context of an HCC surveillance program<sup>16</sup>. Overall, 51% (110/216) of cirrhotic 347 348 patients harbored the ICF, including the High Infiltrate-ICF subtype in 28% (31/110), 349 the Immunosuppressive-ICF subtype in 19% (21/110), and the Pro-inflammatory-ICF 350 subtype in 53% (58/110) of cirrhotic patients harboring the ICF. Next, we tested the 351 capacity of the ICF subtypes to predict the risk of HCC development in cirrhotic 352 patients. Interestingly, the presence of the Immunosuppressive-ICF subtype (10% of all 353 cirrhotic patients) was significantly associated with a higher risk of HCC development 354 [median time to HCC development of 7.4 years (95% CI: 3.2-11.7) vs 17.1 years (95% 355 CI: 10.6-23.7) in Rest, p<0.0001] and was found to be an independent predictor of 356 HCC occurrence in cirrhotic patients in a multivariate analysis [HR 2.41 (95% CI: 1.2-357 4.8), p=0.012] (Figure 3A and Table 1). In addition, the *Immunosuppressive-ICF* was 358 also significantly associated with poor survival [median overall survival of 7.1 years 359 (95% CI: 4.5-9.6) vs 16.3 years (95% CI: 9.1-23.5) in Rest, p<0.0001] and higher risk 360 of hepatic decompensation [median time to hepatic decompensation of 6.5 years (95% CI: 4.3-8.6) vs >15 years in Rest, p<0.0001] (Figure 3B-3C). Cirrhotic patients 361 362 harboring the other two ICF subtypes (*High Infiltrate* and *Pro-inflammatory* subtypes) 363 also showed a non-significant trend towards a higher risk of HCC development 364 compared to those patients lacking the ICF [mean time to HCC development of 12.8 365 years (95% CI:11.5-14.2) in Other-ICF subtypes vs 16.3 years (95% CI: 14.2-18.5) in 366 non-ICF, p=0.06] (Supplementary Figure 3A).

Moreover, the analysis of an additional cohort of 124 non-neoplastic patients with liver fibrosis<sup>22</sup> revealed that the immune-mediated cancer field may occur as a progressive event, as it significantly correlated with increasing levels of fibrosis stage and degree of inflammation (**Supplementary Figure 3B**). Particularly, the presence of the *Immunosuppressive-ICF* significantly correlated with the presence of advanced liver
 fibrosis (Scheuer fibrosis S3-4 score<sup>22</sup>, p=0.034) (Supplementary Figure 3B).

In conclusion, the immune-mediated cancer field detected in patients with early HCC is also present in the livers of ~50% of cirrhotic patients and captures the presence of a damaging and continuous inflammatory response in the underlying liver disease. Furthermore, our results underscore the critical role of an *Immunosuppressive-ICF* cancer field (overall, 10% of cirrhotic patients) in defining a 2.4 risk of HCC development vs the rest of patients, and to a smaller extent of the *High Infiltrate* and *Pro-inflammatory* subtypes.

380

# The immune-mediated field as target for chemoprevention in a mouse model recapitulating chronic liver inflammation and HCC development

383 Based on the compelling results described above, we hypothesized that the immune-384 mediated cancer field, and particularly the Immunosuppressive-ICF subtype, may 385 represent an ideal target for chemopreventive strategies in cirrhotic patients at high risk 386 of HCC development. To this purpose, an *in silico*-based analysis was performed using 387 our training cohort to identify those candidate therapies most likely to modulate the 388 identified ICF. This analysis was based on the enrichment of a compendium of ~1230 389 gene sets (DSigDB collections D1 and D2)<sup>38</sup> recapitulating the main targets of 1202 390 approved drugs. Among the top 10 most significantly enriched drugs (Supplementary 391 Figure 4), nintedanib was the only FDA-approved therapy indicated for a non-cancer 392 condition. Specifically, nintedanib is the first molecular targeted therapy with clinical 393 efficacy in patients with idiopathic pulmonary fibrosis as both an anti-fibrogenic and 394 anti-inflammatory agent<sup>39</sup>. Given these considerations, the efficacy of nintedanib in 395 reverting the pro-tumorigenic immune-mediated cancer field was tested in a mouse 396 model of HCC development in the setting of chronic inflammation and liver fibrosis 397 (Supplementary Figure 5A). In this model, the macroscopic evaluation of explanted 398 livers in DEN/CCl<sub>4</sub> treated mice sacrificed at the age of 15, 17 and 18 weeks confirmed 399 the development of numerous hepatic tumors in the context of chronic inflammation 400 and liver fibrosis (Figure 4A). Tumor penetrance and number of tumors progressively 401 increased, ultimately reaching a 100% incidence at 18 weeks of age (Figure 4A and 402 **4B**). At all-time points, histological evaluation of the liver sections showed that a portion 403 of the tumors were pre-neoplastic (dysplastic) nodules (Figure 4C). After only 3 weeks 404 of treatment (mice sacrificed at 15 weeks of age, Supplementary Figure 5A), 405 nintedanib showed a clear trend towards reducing HCC incidence, number and size of

406 tumors (Figure 4B, D and E). These differences reached significance at 17 weeks of 407 age (Figure 4B), having a marked decrease in both overall number of tumors (30% in 408 nintedanib vs 89% in vehicle group, p=0.019) and specifically in HCC incidence (7% in 409 nintedanib vs 33% in vehicle group, p=0.04). Similarly, at 18 weeks of age, HCC 410 incidence was significantly reduced in treated mice (Figure 4B, 22% in nintedanib vs 411 77% in vehicle group, p<0.001). In addition, nintedanib significantly reduced the overall 412 tumor number and size both at 17 and 18 weeks of age (Figure 4D-E). Overall, 413 nintedanib was well tolerated with no significant induction of body weight loss or 414 hepatotoxicity measured by serum ALT and AST levels (Supplementary Figure 5B-415 C). Taken together, our data suggest that nintedanib is safe and efficacious in 416 preventing HCC development in our experimental model.

417

#### 418 Nintedanib treatment reverts the immunosuppressive-ICF effect

419 Next, we sought to assess the impact of nintedanib treatment on the immune-mediated 420 cancer field. For this purpose, we analyzed gene expression profiling of non-tumor liver 421 samples from 17 weeks-old DEN/CCl<sub>4</sub> mice treated with nintedanib (n= 10) or vehicle 422 (n=9), and 3 healthy control mice. First, the comparison between the healthy control 423 group and vehicle treated mice revealed a profile of activated pathways compatible 424 with HCC development within a fibrotic and inflammatory background. In this regard, 425 functional analysis of differentially expressed genes (Supplementary Table 8) 426 highlighted the activation of hepatic stellate cells and fibrogenesis, as well as immune 427 system activation (inflammatory response, chemotaxis, binding of myeloid and 428 leukocytes) in vehicle treated DEN/CCl<sub>4</sub> livers (Supplementary Table 9). Notably, our 429 model faithfully recapitulated the human immune-mediated field subtypes described 430 above (Figure 5A). The comparison of the gene expression profiles of adjacent non-431 tumor liver from mice treated with vehicle or nintedanib demonstrated that nintedanib 432 significantly reverted the different ICF subtypes, more specifically the 433 Immunosuppressive-ICF phenotype, which predicts risk of HCC development in 434 cirrhotic patients (Figure 5A, p=0.005). A non-significant trend was also observed for 435 the other two types of ICF (Figure 5A). Treatment with nintedanib led to a significant 436 down-regulation of inflammatory cues (IL-6/STAT3, interferon- $\alpha$ , interferon- $\gamma$ ) and 437 immune-related signaling (IL-2/STAT5 activation, allograft rejection) (Figure 5A). 438 Among the infiltrating immune cells, nintedanib significantly reduced the presence of B 439 and T cells, activated macrophages, helper T cells and Tregs (Figure 5A). 440 Furthermore, nintedanib significantly decreased the expression levels of immune 441 response modulators characteristic of the Immunosuppressive-ICF subtype (including

*IL1, CCL5, PDL1,* Figure 5A). Despite exhibiting similar global levels of inflammatory
infiltrates, quantification of CD4 and CD8 positive infiltrating lymphocytes by IHC
revealed a significant decrease of CD4+ T cells in nintedanib-treated mice compared to
controls (Figure 5B, p<0.05). These data are in accordance with the down-regulation</li>
of gene sets representing helper and regulatory T cells (Figure 5A).

447 Next, in order to further characterize the chemopreventive effects of nintedanib we 448 assessed the activation status of the main nintedanib targets (i.e. VEGFR2 and 449 PDGFR-β). Western blot of non-tumor liver tissue confirmed that nintedanib blocked 450 the activation of VEGFR2 (Figure 5C) and its downstream effectors AKT and ERK 451 (Supplementary Figure 6A). Consistently, both liver parenchyma and liver tumors 452 were pERK positive in vehicle-treated mice and pERK negative in nintedanib treated 453 mice (Supplementary Figure 6B, p<0.05), indicating an anti-proliferative effect of 454 nintedanib as well. Given the strong inhibition of VEGFR signaling observed, we next 455 assessed the anti-angiogenic effect of nintedanib in DEN/CCl<sub>4</sub> mice. In this model, 456 reduced CD31 staining was associated with diminished blood vessel area in both liver 457 parenchyma and liver tumors of nintedanib-treated mice (Figure 5D). Altogether, these 458 data suggest that nintedanib exhibits its chemopreventive effects in part by inducing 459 vascular normalization and inhibiting hepatic proliferation.

460 In parallel, we evaluated the potential of nintedanib to ameliorate liver fibrosis. No 461 significant inhibition of the main pro-fibrogenic signaling pathway, PDGFR- $\beta$ , was 462 detected in the livers of nintedanib-treated mice (Supplementary Figure 6C). 463 Consistently, similar levels of fibrosis degree were observed between mice treated with 464 nintedanib or vehicle (Supplementary Figure 6D). Furthermore, the gene expression 465 levels of collagen markers (i.e. Col1a1 and Col1a2) and the pro-fibrogenic ligand Pdgfb 466 were similar in nintedanib- and vehicle-treated mice (Supplementary Figure 6E). In 467 contrast, α-SMA (Acta2), a marker of extracellular matrix (ECM) producing cells, was 468 significantly decreased in nintedanib-treated mice at 17 weeks of age, with a non-469 significant trend towards reduction at 18 weeks (Supplementary Figure 6E), in 470 accordance with the reduced liver angiogenesis.

471 Overall, our data confirm that our mouse model faithfully recapitulates the immune-472 mediated cancer field effect observed in human samples and that therapeutic targeting 473 of the *Immunosuppressive-ICF* subtype, accompanied by liver vascular normalization 474 and suppression of hepatic proliferation, can prevent the development of HCC 475 associated with advanced chronic liver disease.

#### 477 **DISCUSSION**

This study represents an in-depth analysis of the *inflammatory milieu* associated with the "field cancerization" in the chronically injured liver, and investigates its clinical implications in the prediction and prevention of HCC occurrence in cirrhotic patients.

481 The role of the "cancer field effect" in promoting neoplastic transformation has gained 482 much interest in recent years and currently an altered microenvironment is considered 483 a promoter of cancer<sup>40,41</sup>. Activation of HSC as well as certain pathways, such as 484 nuclear factor-KB and TGF-B signaling, have been previously associated with liver fibrogenesis, and eventually carcinogenesis<sup>13,18</sup>. With this study, we move beyond the 485 486 limits of current knowledge and provide a detailed description of the immune 487 microenvironment underlying the *field cancerization* in the non-tumor liver. To this end, 488 we first characterized the immune profile of the non-tumor liver parenchyma of 392 489 early HCCs and then investigated its role in predicting HCC development in 214 490 cirrhotic patients with long-term surveillance for HCC (median of 10 years)<sup>16</sup>. The 491 analysis revealed that up to 60% of HCCs and 50% of cirrhotic patients showed a 492 deleterious immune-mediated response in the surrounding tissue, which was 493 associated with impaired liver function, activation of specific oncogenic loops and poor 494 survival. Further dissection of the immunological portrait of this molecular group 495 identified three distinct subtypes with different levels of lymphocyte infiltration and 496 activation of either immunosuppressive or pro-inflammatory traits were identified. 497 Interestingly, the *Immunosuppressive-ICF* subtype (~10% of cirrhotic patients), mostly 498 characterized by stromal activation and TGF- $\beta$  signaling, was an independent predictor 499 of HCC development, increasing 2.4 the risk of cancer development on top of the other 500 cirrhotic molecular subtypes. These observations support the hypothesis that an 501 immunosuppressive microenvironment favors HCC development and provide insights 502 into the immune-mediated mechanisms underlying the cancerization field. Perhaps of 503 greater clinical relevance, the Immunosuppressive-ICF subtype may provide a novel 504 companion biomarker to enrich at-risk patients in chemoprevention clinical trials.

505 Reducing the incidence and mortality of HCC requires advances in chemopreventive 506 approaches at pre-neoplastic stages, in addition to curative treatment options for early 507 lesions. Universal immunization against HBV and antiviral therapies against HBV and HCV have been associated with very reduced HCC risk<sup>2,42-44</sup>. Once cirrhosis is 508 509 established, the risk of HCC development remains despite achieving a sustained 510 virologic response in HCV patients<sup>11,12</sup>. In addition, the incidence of other risk factors, 511 such as non-alcoholic steatohepatitis (NASH), is dramatically increasing<sup>45</sup>. Thus, 512 alternative HCC preventive strategies capable of interfering with molecular 513 hepatocarcinogenesis are an unmet need. Furthermore, identifying those patients at 514 high risk of HCC development should enable a cost-effective selection of patients most 515 likely to benefit from chemopreventive approaches. Since our results specifically 516 showed that the Immunosuppressive-ICF class identifies 10% of cirrhotic patients at 517 higher risk of HCC development (20% in patients that already developed an HCC), we 518 then sought to investigate if the molecular forces driving such cancer field could serve 519 as target for chemopreventive strategies. To test this hypothesis in the preclinical 520 setting, we first verified that the molecular profiles observed in human cirrhosis were 521 faithfully reproduced in animal models of chronic liver injury. The DEN/CCl<sub>4</sub> chemically-522 induced mouse model reliably reproduced the presence of a pro-carcinogenic 523 phenotype with increased inflammation, angiogenesis and immune response. In 524 particular, characterization of the immune-mediated field recreated in those animal 525 livers confirmed the recruitment of T cells, regulatory and helper T cells observed in 526 liver tissues from patients belonging to the immune-mediated cancer field.

527 In order to identify the most promising candidate therapies for novel chemopreventive 528 strategies, we conducted an *in silico* analysis using a large compendium of gene sets<sup>38</sup> 529 recapitulating the main targets of 1202 approved drugs. Among the top ten most 530 significantly enriched drugs, we selected nintedanib, the only FDA therapy approved for 531 non-neoplastic conditions. In our animal model, oral administration of nintedanib 532 reverted the immune-mediated cancer field, and particularly the Immunosuppressive-533 ICF subtype, ultimately reducing HCC incidence and growth. Reversion of the 534 *Immunosuppressive* profile induced by treatment with nintedanib was accompanied by 535 reduction of CD4+ lymphocytes. These findings are in line with previous reports 536 suggesting that CD4+, but not CD8+ T cells, propagate immune-mediated liver injury in 537 models of chronic liver inflammation or autoimmune liver disease<sup>46,47</sup>. Pretreatment 538 with T cell-specific Abs or immunosuppressive agents, such as anti-CD4 mAb, FK506 539 (Tacrolimus), or cyclosporine A, have shown to ameliorate hepatitis in these models, 540 further supporting the role of CD4+ T cells in inducing liver damage<sup>48</sup>. The damage is 541 thought to be partially mediated by T-cell-derived IFN-y and Kupffer-cell-derived TNF, 542 which lead to hepatocyte cell death<sup>49</sup>. Consistently, elevated IFN-y and TNF signaling 543 were among the key features up-regulated in our immune-mediated cancer field and 544 were both significantly inhibited upon treatment. Overall, our study identifies a novel 545 promising chemopreventive strategy for HCC and confirms the validity of using the 546 reversion of the *Immunosuppressive-ICF signature* as reliable read-out of efficacy. This 547 is of great clinical importance since there is currently no effective method to monitor the 548 short-term effects of chemopreventive drugs<sup>10</sup>.

549 Angiogenesis and inflammation are interdependent and, as demonstrated in different 550 studies conducted in animal models and human tissues, are active partners at onset of 551 cancer<sup>50</sup>. Thus, blockage of angiogenesis can also represent an attractive 552 chemoprevention strategy<sup>51</sup>. Nintedanib belongs to a new generation of TKIs that, in 553 addition to exerting immune modulating by inhibiting src family of kinases (i.e. LCK, 554 FLT3, and SRC), blocks the activation of main angiogenic receptors<sup>52</sup>. Many cytokines 555 and growth factors are involved in modulating the formation of new vessels. Expression 556 of VEGF and its receptors is elevated in HCC cell lines and tissues, as well as in the blood circulation of patients with HCC<sup>37,53-55</sup>. In our model, nintedanib exerted its 557 558 chemopreventive mechanisms in part through the inhibition of VEGF signaling, a major 559 driver of angiogenesis<sup>56</sup>. Consistently, nintedanib treatment led to significant decrease 560 of  $\alpha$ -SMA and CD31, markers highly expressed on activated endothelial cells. Thus far, 561 independent studies had described that HCC prevention can be achieved in animal 562 models by attenuating liver fibrosis through the inhibition of epidermal growth factor receptor (EGFR)<sup>57,58</sup> or lysophosphatidic acid (LPA)<sup>36</sup> signaling. With the current study, 563 564 we demonstrate that modulation of the liver microenvironment by molecular targeted 565 drugs, which simultaneously block liver inflammation and angiogenesis, might 566 represent a powerful alternative strategy.

567 We recently defined the immune class of HCC<sup>20</sup> and the Immune exclusion class (characterized by active Wnt/CTNNB1)<sup>3,20,59</sup>, which might predict response and primary 568 resistance to checkpoint inhibitors, respectively <sup>3,59</sup>. We herein explore the immune-569 570 mediated mechanisms underlying HCC occurrence by defining an immunosuppressive 571 field effect in ~10% of cirrhotic patients that conforms a cancer-permissive milieu, thus 572 posing them at the highest risk of HCC development. In addition, our pre-clinical data 573 with a drug approved in pulmonology and in non-small cell lung cancer treatment 574 suggest that the permissive microenvironment can be reverted leading to a delay in 575 HCC occurrence. These data provide the rationale for testing this strategy in early 576 chemoprevention trials targeting cirrhotic patients at high risk of HCC development. In 577 addition, this strategy could also be further explored in the adjuvant setting considering 578 that 20% of HCC undergoing resection also present this permissive *milieu* in the 579 adjacent non-tumoral tissue.

#### 581 **FIGURE LEGEND**

582 Figure 1. Identification of an immune-mediated cancer field effect in non-tumoral 583 liver tissue adjacent to early HCCs. A) Heatmap representation of the immune-584 mediate cancer field present in 60% of HCC patients. High and low single sample 585 gene-set enrichment (ssGSEA) analysis scores are represented in red and blue, 586 respectively. Positivity for previously reported gene signatures was evaluated by 587 nearest template (NTP) method. B) Top predicted upstream cytokine and transcription 588 factors activated in liver tissues belonging to the immune-mediated field effects. C) 589 Representative images of degree of Portal/Septal infiltrating inflammation. Moderate-to-590 marked inflammatory infiltrate was more prevalent in samples with immune-mediated 591 field effect. D) Representative images depicting presence or absence of lymphoid 592 aggregates. The presence of lymphoid aggregates was significantly associated with the 593 immune-mediated cancer field effect.

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595 Figure 2. The immune-mediated cancer field contains 3 distinct molecular 596 subtypes. A) Heatmap representation of the three ICF subtypes within the samples 597 presenting the immune-mediated cancer field. Statistical significance is highlighted 598 comparing the different subtypes within the ICF (Purple in High Infiltrate-ICF, Orange in 599 Immunosupressive-ICF, Green in Pro-inflammatory-ICF, and Black in both High 600 Infiltrate and Immunosuppressive). B) Comparison of estimated proportion of immune 601 cells (CIBERSORT method) between the immune-mediated cancer field subtypes. C) 602 Immunophenogram representing the enrichment of immunogenic determinants in the 603 distinct immune-mediated cancer field subtypes. Significant statistical differences are 604 defined as follows: \*=p<0.05, \*\*=p<0.01 and \*\*\*=p<0.001.

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Figure 3. Association of the presence of the *Immunnosuppressive-ICF* with HCC occurrence and prognostic variables in cirrhotic patients. (A) Kaplan-Meier estimates of HCC development, (B) overall survival, (C) hepatic decompensation, according to the presence of the *Immunosuppressive* -ICF subtype (orange), and the Rest of the cohort.

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Figure 4. Nintedanib prevents and delays the development of HCC. A)
Representative pictures of macroscopic evaluation of hepatic tumors in mice treated
with vehicle or nintedanib sacrificed at 15, 17 and 18 weeks of age. Arrows indicate
macroscopically visible tumors. B) Evaluation of overall tumor burden and HCC

616 incidence. (#) denotes statistical significance for overall tumor burden while (\*) for HCC 617 incidence. C) Microscopic evaluation of the number of tumors per mouse in each 618 experimental group. Ill-defined hepatic tumors were diagnosed as dysplastic nodules, 619 while circular well-defined lesions with pushing margins and/or vascular invasion were 620 diagnosed as HCCs. D) Number of macroscopic tumors per mouse treated with vehicle 621 or nintedanib at the three different time-points. E) Diameter size of the largest tumor 622 per mouse treated with vehicle or nintedanib at the three different time-points. A total of 623 7-11 mice were evaluated per group in each approach. Significant statistical differences are defined as follows: # or \*=p<0.05, \*\*=p<0.01 and \*\*\*=p<0.001. 624

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626 Figure 5. Nintedanib reverts the Immunosuppressive-ICF recapitulated in the 627 DEN/CCl<sub>4</sub> mice and induces vascular normalization. A) Single sample GSEA 628 analysis of the 172-gene signature and gene-sets recapitulating the immune-mediated 629 cancer field subtypes. In the heatmap, high and low ssGSEA analysis scores or gene 630 expression levels are represented in red and blue, respectively. B) Representative 631 images and quantification of CD4+ and CD8+ positive infiltrating lymphocytes in the 632 liver parenchyma of 17 weeks old mice treated with vehicle (n=5) or nintedanib (n=5). 633 C) Western-blot analysis of VEGFR2 activation in the non-tumor liver parenchyma of 634 17 weeks old mice treated with vehicle (n=6) or nintedanib (n=6). D) Morphometric 635 quantification of blood vessel area by CD31 immunostaining in 5 randomly selected low 636 magnification fields in mice treated with vehicle (n=5) or nintedanib (n=5). Significant 637 statistical differences are defined as follows: \*\*=p<0.01 and \*\*\*=p<0.001

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## 641 **TABLES**

Variable	Un <u>ivariate analy</u> sis p-value	Multivariate analysis (cox's regression)		
		HR	CI(95% low-high limits)	p-values
Non-tumoral liver tissue-based transcriptomic profiles				
Immunosuppressive-ICF	0.03	2.41	1.21-4.80	0.01
186-gene Poor prognosis signature	<0.0001	1.56	0.89-2.7	0.12
<u>Clinicopathological variables</u>				
Age (>median)	0.87			
Gender	0.22			
Diabetes	0.48			
HCV genotype 1b	0.18			
Alcohol consumption (>80 g/day)	0.68			
HCVetiology plus alcohol consumption	0.68			
History of antiviral treatment (interferon-based)	0.65			
Varices	0.02	1.49	0.85-2.6	0.17
Spleen	0.13			
Ishaak score 6 vs 5	0.24			
Platelet count (<100,000/mm3)	0.02	1.51	0.91-2.64	0.15
Bilirubin (> 1 mg/dL)	0.00	1.85	1.07-3.2	0.03
AFP (> 10 ng/mL)	0.87			
Prothrombin time (international normalized ratio >1.2)	0.38			

Table 1. Uni- and Multivariate Analysis of risk of HCC development in cirrhotic patients
 including gene signatures and clinico-pathological variables (n=216).

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#### 647 **REFERENCES**

- Bray F, Ferlay J, Soerjomataram I, et al. Global cancer statistics 2018:
   GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in
   185 countries. CA Cancer J Clin 2018;68:394–424.
- 652 2. Llovet JM, Zucman-Rossi J, Pikarsky E, et al. Hepatocellular carcinoma. Nat
  653 Rev Dis Prim 2016;2:16018.
- Llovet JM, Montal R, Sia D, et al. Molecular therapies and precision medicine for
  hepatocellular carcinoma. Nat Rev Clin Oncol 2018;15:599–616.
- 656 4. Galle PR, Forner A, Llovet JM, et al. EASL Clinical Practice Guidelines:
  657 Management of hepatocellular carcinoma. J Hepatol 2018;69:182–236.
- 5. Llovet JM, Ricci S, Mazzaferro V, et al. Sorafenib in advanced hepatocellular
  carcinoma. N Engl J Med 2008;359:378–390.
- 6. Cheng A-L, Finn RS, Qin S, et al. Phase III trial of lenvatinib (LEN) vs sorafenib
  (SOR) in first-line treatment of patients (pts) with unresectable hepatocellular
  carcinoma (uHCC). | 2017 ASCO Annual Meeting Abstracts. J Clin Oncol
  2017;35.

- 664 7. Bruix J, Qin S, Merle P, et al. Regorafenib for patients with hepatocellular
  665 carcinoma who progressed on sorafenib treatment (RESORCE): a randomised,
  666 double-blind, placebo-controlled, phase 3 trial. Lancet 2017;389:56–66.
- Abou-Alfa GK, Meyer T, Cheng A-L, et al. Cabozantinib (C) versus placebo (P)
  in patients (pts) with advanced hepatocellular carcinoma (HCC) who have
  received prior sorafenib: Results from the randomized phase III CELESTIAL trial.
  J Clin Oncol 2018;36:suppl 4S; abstr 207.
- 671 9. Singh S, Singh PP, Roberts LR, et al. Chemopreventive strategies in
  672 hepatocellular carcinoma. Nat Rev Gastroenterol Hepatol 2014;11:45–54.
- Fujiwara N, Friedman SL, Goossens N, et al. Risk factors and prevention of
  hepatocellular carcinoma in the era of precision medicine. J Hepatol
  2018;68:526–549.
- 676 11. Calvaruso V, Cabibbo G, Cacciola I, et al. Incidence of Hepatocellular
  677 Carcinoma in Patients With HCV-Associated Cirrhosis Treated With Direct678 Acting Antiviral Agents. Gastroenterology 2018;155:411–421.e4.
- Kanwal F, Kramer J, Asch SM, et al. Risk of Hepatocellular Cancer in HCV
  Patients Treated With Direct-Acting Antiviral Agents. Gastroenterology
  2017;153:996–1005.e1.
- Hoshida Y, Villanueva A, Kobayashi M, et al. Gene expression in fixed tissues
  and outcome in hepatocellular carcinoma. N Engl J Med 2008;359:1995–2004.
- Even 14. Zucman-Rossi J, Villanueva A, Nault J-C, et al. Genetic Landscape and
  Biomarkers of Hepatocellular Carcinoma. Gastroenterology 2015;149:1226–
  1239.e4.
- Farazi PA, DePinho RA. Hepatocellular carcinoma pathogenesis: from genes to
  environment. Nat Rev Cancer 2006;6:674–687.
- 689 16. Hoshida Y, Villanueva A, Sangiovanni A, et al. Prognostic gene expression
  690 signature for patients with hepatitis C-related early-stage cirrhosis.
  691 Gastroenterology 2013;144:1024–1030.
- 692 17. Ji J, Eggert T, Budhu A, et al. Hepatic stellate cell and monocyte interaction
  693 contributes to poor prognosis in hepatocellular carcinoma. Hepatology
  694 2015;62:481–95.
- 695 18. Zhang DY, Goossens N, Guo J, et al. A hepatic stellate cell gene expression
  696 signature associated with outcomes in hepatitis C cirrhosis and hepatocellular

- 697 carcinoma after curative resection. Gut 2016;65:1754–64.
- Kim JH, Sohn BH, Lee H-S, et al. Genomic predictors for recurrence patterns of
  hepatocellular carcinoma: model derivation and validation. Beck AH, ed. PLoS
  Med 2014;11:e1001770.
- Sia D, Jiao Y, Martinez-Quetglas I, et al. Identification of an Immune-specific
  Class of Hepatocellular Carcinoma, Based on Molecular Features.
  Gastroenterology 2017;153:812–826.
- Villanueva A, Portela A, Sayols S, et al. DNA Methylation-based prognosis and
  epidrivers in hepatocellular carcinoma. Hepatology 2015:1–12.
- Wang M, Gong Q, Zhang J, et al. Characterization of gene expression profiles in
  HBV-related liver fibrosis patients and identification of ITGBL1 as a key regulator
  of fibrogenesis. Sci Rep 2017;7:43446.
- Godec J, Tan Y, Liberzon A, et al. Compendium of Immune Signatures Identifies
  Conserved and Species-Specific Biology in Response to Inflammation. Immunity
  2016;44:194–206.
- 712 24. Brunet J-P, Tamayo P, Golub TR, et al. Metagenes and molecular pattern
  713 discovery using matrix factorization. Proc Natl Acad Sci U S A 2004;101:4164–9.
- 714 25. Newman AM, Liu CL, Green MR, et al. Robust enumeration of cell subsets from
  715 tissue expression profiles. Nat Methods 2015;12:453–7.
- Pindea G, Mlecnik B, Tosolini M, et al. Spatiotemporal dynamics of intratumoral immune cells reveal the immune landscape in human cancer. Immunity 2013;39:782–95.
- 719 27. Thorsson V, Gibbs DL, Brown SD, et al. The Immune Landscape of Cancer.
  720 Immunity 2018;48:812–830.e14.
- 721 28. Reich M, Liefeld T, Gould J, et al. GenePattern 2.0. Nat Genet 2006;38:500–
  722 501.
- Charoentong P, Finotello F, Angelova M, et al. Pan-cancer Immunogenomic
  Analyses Reveal Genotype-Immunophenotype Relationships and Predictors of
  Response to Checkpoint Blockade. Cell Rep 2017;18:248–262.
- 30. Dapito DH, Mencin A, Gwak G-Y, et al. Promotion of hepatocellular carcinoma
  by the intestinal microbiota and TLR4. Cancer Cell 2012;21:504–16.
- 31. Okamoto M, Utsunomiya T, Wakiyama S, et al. Specific gene-expression profiles
  of noncancerous liver tissue predict the risk for multicentric occurrence of

- hepatocellular carcinoma in hepatitis C virus-positive patients. Ann Surg Oncol2006;13:947–54.
- 32. Wolf DM, Lenburg ME, Yau C, et al. Gene co-expression modules as clinically
  relevant hallmarks of breast cancer diversity. Haibe-Kains B, ed. PLoS One
  2014;9:e88309.
- 33. Beck AH, Espinosa I, Edris B, et al. The macrophage colony-stimulating factor 1
  response signature in breast carcinoma. Clin Cancer Res 2009;15:778–87.
- Messina JL, Fenstermacher DA, Eschrich S, et al. 12-Chemokine gene signature
  identifies lymph node-like structures in melanoma: potential for patient selection
  for immunotherapy? Sci Rep 2012;2:765.
- 740 35. Rooney MS, Shukla SA, Wu CJ, et al. Molecular and genetic properties of
  741 tumors associated with local immune cytolytic activity. Cell 2015;160:48–61.
- 742 36. Nakagawa S, Wei L, Song WM, et al. Molecular Liver Cancer Prevention in
  743 Cirrhosis by Organ Transcriptome Analysis and Lysophosphatidic Acid Pathway
  744 Inhibition. Cancer Cell 2016;30:879–890.
- 745 37. Chiang DYY, Villanueva A, Hoshida Y, et al. Focal gains of VEGFA and
  746 molecular classification of hepatocellular carcinoma. Cancer Res 2008;68:6779–
  747 6788.
- 748 38. Yoo M, Shin J, Kim J, et al. DSigDB: drug signatures database for gene set
  749 analysis: Fig. 1. Bioinformatics 2015;31:3069–3071.
- Richeldi L, Bois RM du, Raghu G, et al. Efficacy and safety of nintedanib in
  idiopathic pulmonary fibrosis. N Engl J Med 2014;370:2071–82.
- 40. Lochhead P, Chan AT, Nishihara R, et al. Etiologic field effect: reappraisal of the
  field effect concept in cancer predisposition and progression. Mod Pathol
  2015;28:14–29.
- Hernandez-Gea V, Toffanin S, Friedman SL, et al. Role of the microenvironment
  in the pathogenesis and treatment of hepatocellular carcinoma.
  Gastroenterology 2013;144:512–27.
- Chang M-H, You S-L, Chen C-J, et al. Decreased incidence of hepatocellular
  carcinoma in hepatitis B vaccinees: a 20-year follow-up study. J Natl Cancer Inst
  2009;101:1348–55.
- 761 43. Papatheodoridis G V, Dalekos GN, Yurdaydin C, et al. Incidence and predictors
  762 of hepatocellular carcinoma in Caucasian chronic hepatitis B patients receiving

- 763 entecavir or tenofovir. J Hepatol 2015;62:363–70.
- Morgan RL, Baack B, Smith BD, et al. Eradication of hepatitis C virus infection
  and the development of hepatocellular carcinoma: a meta-analysis of
  observational studies. Ann Intern Med 2013;158:329–37.
- Mittal S, Sada YH, El-Serag HB, et al. Temporal trends of nonalcoholic fatty liver
  disease-related hepatocellular carcinoma in the veteran affairs population. Clin
  Gastroenterol Hepatol 2015;13:594–601.e1.
- Tiegs G, Hentschel J, Wendel A. A T cell-dependent experimental liver injury in
  mice inducible by concanavalin A. J Clin Invest 1992;90:196–203.
- 772 47. Omenetti S, Brogi M, Goodman WA, et al. Dysregulated intrahepatic CD4+ T773 cell activation drives liver inflammation in ileitis-prone SAMP1/YitFc mice. Cell
  774 Mol Gastroenterol Hepatol 2015;1:406–419.
- Tiegs G, Hentschel J, Wendel A. A T cell-dependent experimental liver injury in
  mice inducible by concanavalin A. J Clin Invest 1992;90:196–203.
- 49. Heymann F, Peusquens J, Ludwig-Portugall I, et al. Liver inflammation
  abrogates immunological tolerance induced by Kupffer cells. Hepatology
  2015;62:279–91.
- 50. Hanahan D, Weinberg RA. Hallmarks of cancer: The next generation. Cell2011;144:646–674.
- 782 51. Albini A, Tosetti F, Li VW, et al. Cancer prevention by targeting angiogenesis.
  783 Nat Rev Clin Oncol 2012;9:498–509.
- 52. Hilberg F, Roth GJ, Krssak M, et al. BIBF 1120: triple angiokinase inhibitor with
  sustained receptor blockade and good antitumor efficacy. Cancer Res
  2008;68:4774–82.
- 53. Mas VR, Maluf DG, Archer KJ, et al. Angiogenesis soluble factors as
  hepatocellular carcinoma noninvasive markers for monitoring hepatitis C virus
  cirrhotic patients awaiting liver transplantation. Transplantation 2007;84:1262–
  790
  71.
- Foon RTP, Ho JWY, Tong CSW, et al. Prognostic significance of serum vascular
  endothelial growth factor and endostatin in patients with hepatocellular
  carcinoma. Br J Surg 2004;91:1354–60.
- Forwitz E, Stein I, Andreozzi M, et al. Human and mouse VEGFA-amplified
  hepatocellular carcinomas are highly sensitive to sorafenib treatment. Cancer

796 Discov 2014.

- 797 56. Carmeliet P. VEGF as a key mediator of angiogenesis in cancer. Oncology
  798 2005;69 Suppl 3:4–10.
- 57. Schiffer E, Housset C, Cacheux W, et al. Gefitinib, an EGFR inhibitor, prevents
  hepatocellular carcinoma development in the rat liver with cirrhosis. Hepatology
  2005;41:307–14.
- 58. Fuchs BC, Hoshida Y, Fujii T, et al. Epidermal growth factor receptor inhibition
  attenuates liver fibrosis and development of hepatocellular carcinoma.
  Hepatology 2014;59:1577–90.
- 805 59. Pinyol R, Sia D, Llovet JM. Immune exclusion-Wnt/CTNNB1 class predicts
  806 resistance to immunotherapies in HCC. Clin Cancer Res
  807 2019:clincanres.3778.2018.