- 1 MS 08-486 2 Pharmaceuticals and Personal Care Products in the Environment 3 4 Running title: Pharmaceuticals and biological community in the Llobregat River 5 **Corresponding author:** 6 Isabel Muñoz Gracia 7 Department of Ecology, University of Barcelona 8 Av. Diagonal, 645, 08028 Barcelona, Spain 9 Tel: 34 934021512
- 10 Fax: 34 934111438
- 11 e-mail: imunoz@ub.edu
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This ET&C Paper in Press manuscript is in its original unedited form and has not been copyedited or formatted for final production. This manuscript is fully citable. ©2009 Society of Environmental Toxicology and Chemistry (SETAC). 17 Pharmaceuticals and Personal Care Products in the Environment 18 19 BRIDGING LEVELS OF PHARMACEUTICALS IN RIVER WATER WITH BIOLOGICAL 20 COMMUNITY STRUCTURE IN THE LLOBREGAT RIVER BASIN (NE SPAIN) 21 Isabel Muñoz\*<sup>†</sup>, Julio C. López-Doval<sup>†</sup>, Marta Ricart<sup>‡</sup>, Marta Villagrasa<sup>§</sup>, Rikke Brix<sup>§</sup>, Anita 22 23 Geiszinger<sup>‡</sup>, Antoni Ginebreda<sup>§</sup>, Helena Guasch<sup>‡</sup>, M. José López de Alda<sup>§</sup>, Anna M. Romaní<sup>‡</sup>, 24 Sergi Sabater, ‡ || and Damià Barceló§ || 25 26 <sup>†</sup> Department of Ecology, University of Barcelona, Av. Diagonal, 645, 08028 Barcelona, Spain 27 <sup>‡</sup> Department of Environmental Sciences, University of Girona, Campus Montilivi, 17071 28 Girona, Spain 29 § Department of Environmental Chemistry, Institute of Environmental Assessment and Water 30 Research (IDAEA), Spanish Council for Scientific Research (CSIC), C/ Jordi Girona, 18-26, 31 08034 Barcelona, Spain 32 || Catalan Institute for Water Research (ICRA), Scientific and Technologic Park of the University 33 of Girona, 17003 Girona, Spain 34 (Received 29 September 2008; Accepted 28 January 2009) 35 36 37 38 39

40 \*To whom correspondence may be addressed (imunoz@ub.edu).

42 Abstract-A wide range of human pharmaceuticals are present at low concentrations, in 43 freshwater systems, particularly in sections of polluted river. These compounds show high 44 biological activity, often associated with a high stability. These characteristics imply a potential 45 impact of these substances on aquatic biota even when present at low environmental 46 concentrations. Low flow conditions in Mediterranean rivers, most of which flow through 47 densely populated areas and are subjected to intensive water use, increase the environmental risk 48 of these emergent compounds. Here we studied whether pharmaceuticals in river water affect the 49 local benthic community structure (diatoms and invertebrates). For this purpose, we analyzed the 50 occurrence of pharmaceuticals along the Llobregat River and examined the benthic community 51 structure (diatoms and invertebrates) of this system. Some pharmaceutical products in the 52 Llobregat River registered concentrations greater than those cited in the literature. Multivariate 53 analyses revealed a potential causal association between the concentration of some anti-54 inflammatories and Beta-blockers and the abundance and biomass of several benthic 55 invertebrates (*Chironomus* spp. and *Tubifex tubifex*). Further interpretation in terms of cause-56 effect relationships is discussed, however it must be always taken with caution since other 57 pollutants may also have significant contributions. Combined with further community 58 experiments in the laboratory, our approach could be a desirable way to go on in future risk 59 management decisions. 60

61

Keywords–Llobregat River, Pharmaceuticals, Invertebrates, Diatoms, Multivariate analysis
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64

## 65 Introduction

66

67	Pharmaceuticals comprise an array of products, including a variety of chemical formulations and
68	multiple biological targets. These drugs exert specific biological effects; and are administered for
69	human and veterinary health care. Although a variety of pharmaceutical compounds have been
70	detected in the environment, their potential ecological significance remains unknown and few
71	studies have addressed their impact on non-target species [1].
72	
73	As a result of human population density and more intensive animal farming techniques,
74	water catchments are highly susceptible to be at risk for potential contamination by various
75	pharmaceutical products. A wide range of human pharmaceuticals, including analgesics,
76	antibiotics, steroids, cardiovascular drugs, and various drugs used to treat mental illness, are
77	present in effluents from sewage treatment plants that continuously enter freshwater systems [2-
78	4].
79	
80	Although concentrations of pharmaceuticals in the aquatic environment are generally in
81	the nanogram-per-liter and low microgram-per-liter range, these compounds show high reactivity
82	with biological systems; and are often highly stable. These characteristics may cause potential
83	toxic effects to aquatic organisms even at low environmental concentrations [5]. Most studies in
84	this field have addressed the effects of pharmaceuticals on aquatic vertebrates, namely fish.
85	Chronic toxicity data for algae and invertebrate have become more available in recent years,
86	although they are usually obtained from aqueous-exposure experiments [6] rather than from field

87	studies. Although pharmaceuticals are often moderately lipophilic [7], few studies have
88	examined their potential impact on benthic communities and sediment organisms.
89	
90	The rationale for the present study was to focus on algae and invertebrate fauna as
91	representatives of aquatic benthic communities inhabiting the sediment interphase, with the aim
92	to determine the potential effect of pharmaceuticals on the abundance and community structure
93	of these organisms. These chemicals, transported by water, are adsorbed by particulate matter
94	and accumulated in sediments. After adsorption, chemicals can be remobilized by resuspension
95	or desorption, being a primary source of contamination for benthic organisms.
96	Analyses of chronic toxicity of pharmaceuticals on organisms are essential to obtain a
97	realistic environmental risk assessment, since these substances were designed to exert distinct
98	molecular modes of actions. However, studies in this field usually provide data only for target
99	species and work with concentrations far above those found in nature. Multi-species tests, even
100	in standardized bioassays, allow realism in terms of ascribing biological endpoints to
101	contaminant impact [8]. Data on the responses of natural communities in field conditions to
102	pharmaceuticals may provide the information required to define the ecological relevance of these

103 substances.

104

Given that flowing waters are exposed to multiple stressors, the effects of which can be cumulative and interact across spatial and temporal scales, defining causal relationships can be complex. To establish some relationships between environmental stressors and biological response variables, it is necessary to integrate laboratory, field and other experimental approaches such as mesocosms [9]. Field observations are essential for the detection of emerging

- consequences of stressors on communities, and for allowing the generation of hypotheses toidentify potential cause-and-effect relationships.
- 112

113	The management of Mediterranean river basins implies considering their particular
114	hydrology (low winter and summer discharges and periodical floods in spring and autumn) as
115	well as continuing human pressure on resources and on the ecosystem. Characterized by low
116	flows during normal conditions (~5 m3/s) and extraordinary peak events (maximum recorded of
117	2500 m3/s in 1971) which periodically resets the system, the Llobregat River (NE Spain) is a
118	good example of a Mediterranean basin. The middle and lower sections of the river drain densely
119	populated and industrialized (tannery, textile, pulp, and paper) areas. The major land use types in
120	this area are: 38% urban and industrial activities and 13% farmlands. Furthermore, the Llobregat
121	provides drinking water to the large conurbation of Barcelona.
122	
123	Several works have investigated the presence of various synthetic and natural estrogens
124	and progestogens [10, 11] and other pharmaceuticals [12, 13] in scattered locations in the
125	Llobregat river basin. The invertebrate community of this river has been used as indicator of its
126	ecological quality since the early 1980s [14, 15]. Diatom data from this river have also been
127	available since the 1980s [16-18] and a surveillance program including macroinvertebrates and
128	diatoms is currently implemented by the Catalan Water Authority.
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the water in the Llobregat River basin; and to find potential relationships between the presenceof pharmaceuticals and the structural composition (changes in abundance and biomass) of the

133	biological community (benthic algae and invertebrates) in this river. A total of 29
134	pharmaceuticals, including analgesics, anti-inflammatories, lipid regulators, antibiotics, lipid
135	regulators, psychiatric drugs, anti-histamines and Beta blockers, were analyzed in river water.
136	Other physical and chemical variables related to eutrophication and other environmental
137	characteristics were also analyzed to delineate their influence on the natural community
138	structure. Thus, to the best of our knowledge, the present study is the first to provide extensive
139	data on the occurrence of a large number of pharmaceuticals in the aqueous phase of the
140	Llobregat basin, and in a range of locations and time periods. On the basis of our findings, spatial
141	and temporal variations in pharmaceutical load in the river can be deduced, together with the
142	potential effects of these substances on biota.
143	
144	Material and Methods
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	Study site
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- middle part of the basin receives natural salt slurries from salt formations, which have caused anincrease in water salinity downstream.
- 158

159 Four sampling sites were selected from the middle and lower part of the Llobregat main 160 channel and another three were selected in the Anoia tributary (Fig. 1). These sites were part of a 161 pollution gradient: sites LL1 and A1 were the least polluted but received some industrial 162 effluents and surface runoff from agricultural areas. Sites A3 and LL4 were located in the last 163 section of the two rivers, and were the most polluted sites. Site A2 was located in a highly 164 polluted area receiving wastewaters from tannery, textile and paper industries. Sites LL2 and 165 LL3 were located in a densely inhabited area and received urban and industrial wastewaters 166 inputs. 167 168 Water samples for chemical (nutrients, cations and anions) and pharmaceutical analysis 169 were collected simultaneously with the biological samples from the sediments. Sampling was 170 performed in early June 2005, November 2005, and late May 2006. These three periods covered 171 two of the most relevant periods (spring and autumn) in the system in terms of its hydrology. 172 Samples for grain size characterization were taken in the two first samplings. Ninety-eight 173 percent of the sediment was gravel and sand in all sites. Sites A1, A2, LL1, and LL2 had a 174 lightly more proportion of gravel and A3 a higher proportion of fine sand respect to the others 175 sites.

176

177 Water quality parameters

179	The pH, water temperature, conductivity and oxygen concentration were measured in situ with
180	appropriate probes. Water samples for nutrient analysis were collected in triplicate in each
181	sampling period. The samples were filtered immediately (Nylon Membrane Filters 0.2 mm,
182	Whatman) and frozen until analysis. Nitrate, sulphate, chloride, sodium, potassium, and calcium
183	were determined by ion chromatography (761 Compact IC, Metrohm, Herisau Switzerland).
184	Soluble reactive phosphate and ammonium were measured following standard procedures.
185	
186	Analysis of pharmaceuticals

187 A total of 29 pharmaceuticals, belonging to the classes of analgesics and non-steroidal anti-188 inflammatories, lipid regulators, psychiatric drugs, anti-histamines, anti-ulcer agents, antibiotics 189 and Beta-blockers were measured in water by means of off-line solid-phase extraction (SPE) 190 followed by liquid chromatography-tandem mass spectrometry. Briefly, water samples (400 ml), 191 previously passed through 0.45 µm nylon membrane filters, were preconcentrated on Oasis ® 192 HLB (hydrophilic-lipophilic balance; 60 mg, from Waters (Milford, USA) cartridges, which 193 were further rinsed with 5 ml of high-performance liquid chromatography-grade water, dried 194 under vacuum for 15 to 20 min, and eluted with 2x4 ml of methanol. The extracts were then 195 evaporated under a gentle nitrogen stream, reconstituted with 1ml of methanol-water (25:75, 196 v/v), and added to 10µl of an internal standard mixture. Analysis of the extracts was performed 197 by liquid chromatography-triple quadrupole-tandem mass spectrometry, equipped with an 198 electrospray interface, operating in the SRM mode. Two SRM transitions were monitored per 199 compound. Nine of the 29 compounds were measured in negative ion mode and the remaining 20 200 in positive ion mode. This method and its validation (linearity, detection and quantification 201 limits, repeatability, etc.) are described in detail in Gros et al. [19].

202

203 Diatom analysis

204	The sampling of biofilms for diatom analysis included those growing on large sediment particles
205	(cobbles and gravel; epilithic biofilms) and sand (in the site LL4). Sand samples (2-5cm depth)
206	were collected with a polyvinyl corer, and top sub-samples were taken by an untapped syringe.
207	Epilithic biofilms samples were collected by scrapping a known surface (1cm2) of gravel or
208	cobbles with a knife. Samples were fixed with 4% formaldehyde for further identification and
209	counting. One replicate from each sampling site was cleaned by means of acid oxidation. The
210	cleaned frustules were mounted on resin (Naphrax, refraction index 1.74; Brunel Microscope) on
211	permanent slides. The diatom community was observed and identified under a light microscope
212	(Nikon Eclipse 600W) using phase-contrast and Nomarski differential interference contrast
213	optics at a magnification of
214	x1,000. At least 400 valves were counted and identified to establish the relative abundance of the
215	diatom species present in each sample.
216	
217	Invertebrate analysis
210	

218

Five sediment samples (10-15 cm depth) were randomly collected with a polyvinyl sand corer in each sampling site. The animals retained when sieving the sediment through a 500 mm mesh (the benthic macrofauna) were fixed with 4% formaldehyde. The invertebrates were sorted, counted and identified in the laboratory under a dissecting stereoscopic microscope. Identification was at family level for Oligochaeta and at the genus or species level for the rest of the groups present (mainly Chironomidae and Ephemeroptera). Biomass was calculated as a dry weight from length

dimension using exponential equations [20]. Abundance and biomass were referred to thesediment surface area.

227

228 Data analysis

229 Diatom and invertebrate taxa accounting for more than 1% of total abundance in at least two of 230 the samples were included in the analyses. Taxa abundances were square root transformed prior 231 to analysis. A preliminary analysis based on the Bray-Curtis similarity index for abundance data 232 was conducted to examine the extent to which macroinvertebrate and diatom assemblages varied 233 among samples. This index calculates the similarity between two sites on the basis of species 234 composition. A non-metric multi-dimensional scaling (MDS) procedure was used to ordinate 235 communities (and hence sites) on the basis of the similarity between each pair of samples 236 estimated with the Bray-Curtis similarity index for abundances and Euclidean distances for 237 invertebrate biomass data [8]. The MDS draws the sample distribution in a space delimited by a 238 maximum of three axes. Short distances indicate high similarity in community composition 239 between sites. The stress index measures the goodness of fit between the rank order of 240 similarities and the rank order of distances [8]. Stress values higher than 0.3 indicates that the 241 points are closed to being arbitrarily placed in the two dimensional ordination space.

242

243 Physicochemical variables, except pH were transformed to reduce skewed distributions. 244 Values of environmental and pharmaceutical variables were normalized by subtracting them 245 from the mean and dividing this number by the standard deviation before their inclusion in the 246 analyses. To avoid co-linearity, the variables Ca+ (correlated with Mg+, r=0.95) and ranitidine 247 concentration (correlated with sotalol, r=0.96) were unselected. When the pharmaceutical

- concentrations were below method detection limits, a value equal to one-half the methoddetection limit was assigned to these data in the statistical analyses.
- 250

251	The optimal match between the community patterns and the environmental variables
252	associated with those samples was explored with the BEST (Bio-Env-Stepwise) procedure. This
253	procedure searches for the highest rank correlation (Spearman correlation) between the species
254	similarity matrix (calculated with abundances and biomass for invertebrates) and the
255	environmental matrix (based on Euclidean distances). Spearman rank correlations were
256	calculated by matching element to element [21]. The significance of the rank was assessed using
257	the Monte Carlo permutation test (999 unrestricted permutations). These analyses were
258	performed with the PRIMER® 6 statistical package (Plymouth Marine Laboratory, UK, 2001).
259	
260	In addition, invertebrate data were analysed by detrended correspondence analysis to
261	explore the species responses along an ordination axis. The maximum length of the gradient
262	obtained with the detrended correspondence analysis was 2.44, indicating that linear methods
263	were appropriate [22]. Consequently, we carried out a redundancy analysis, in which species data
264	were constrained by environmental variables. This ordination assumes a linear combination of
265	the species along the environmental gradients preserving the Euclidean distances. The
266	redundancy analysis was performed with CANOCO® software, version 4.5 [22].
267	
268	
269	

270 **Results and Discussion** 

272	Water quality parameters
273	Average values of the water quality parameters in the studied sites are shown in Table 1. The
274	Llobregat and Anoia waters were characterized with high conductivity values and elevated ion
275	concentrations. The upstream Anoia site had waters with high sulphate concentrations because of
276	its gypsum bedrock. Salt (mainly KCl and NaCl) outcrops in the Llobregat basin caused the high
277	concentration of these ions and the corresponding high water conductivity.
278	
279	The sites A2, LL3 and LL4 showed the highest nutrient concentrations and lowest
280	dissolved oxygen. Water flow was low in the two spring seasons (2005, mean discharge in June
281	of 4.79 m3/s, and in 2006, 5.67 m3/s, at LL2 site) and nutrient loads and conductivity increased
282	substantially from upper to downstream reaches. In autumn 2005 the river discharge was higher
283	(mean discharge in October: 8.71 m3/s) and most of the concentrations reached their lowest
284	values.
285	
286	Pharmaceuticals in the Llobregat River
287	
288	Twenty-one of the 29 compounds studied were present in at least one of the samples analyzed
289	(Table 2). Twelve of these compounds presented maximum concentrations above $1 \mu g/L$ and
290	three (the analgesic diclofenac, the lipid regulator bezafribate, and the antibiotic

- sulfamethoxazole) exceeded 10  $\mu$ g/L. Mean concentrations higher than 1 $\mu$ g/L were found for
- 292 eight analytes. The highest mean concentrations were observed for ibuprofen (in sites A3, LL4),

diclofenac (A2, LL4), clofibric acid (LL4), and ofloxacin (LL4), mainly as a result of punctual
 maximum concentrations in specific sites. Analgesics, anti-inflammatories, lipid regulators and
 antibiotics were the families with the highest concentrations.

296

- No significant seasonal variations in pharmaceutical occurrence were detected (analysis of
  variance, time as a factor, p>0.05) but a clear spatial pattern was observed in sites A2, A3, and
  especially LL4, which registered the highest concentrations (analysis of variance, site as a factor
- and post-hoc comparisons with Tukey test, p < 0.05) (Fig. 2).

301

- 302 The pharmaceutical products observed in the Llobregat closely matched those identified by the
- 303 Spanish National Health System as those most consumed. These are mostly analgesics, drugs to
- 304 treat ulcer, anti-histamines, antibiotics, and antidepressants. The concentrations of
- 305 pharmaceuticals that we recorded in the Llobregat River were higher than those reported by Gros
- 306 et al [3] in the Ebro River, where concentrations ranged from 0.1 to 0.6  $\mu$ g/L. In low flow
- 307 periods in a Mediterranean river in France, Comoretto and Chiron [23] reported similar
- 308 concentrations for carbamezapine and bezafribrate to those monitored in our present study. The
- 309 concentration of pharmaceuticals in the Llobregat River were in general in the range described
- 310 by Fent et al. [2], except for ketoprofen, diclofenac, gemfibrazil, bezafibrate, and ranitidine
- 311 which were detected at higher mean concentrations in our present study.

312

313 Biological community structure in the Llobregat River

The ordination of the diatom assemblages (**Fig. 3A**) was derived from the two-dimensional MDS based on Bray-Curtis similarities calculated from root-transformed diatom abundances. Diatom ordination was highly similar between the least polluted sites (A1 and LL1), which showed the highest species richness. The community was dominated by the taxa *Navicula cryptocephala* in A1 and *Nitzschia inconspicua* in LL1. The community composition of site A2, one of the most polluted, clearly differed from the other sites.

321

322 The plot carried out with the data on invertebrate abundance (Fig. 3B) separated one 323 group with the most polluted sites A2 and LL4. Most of the sites (except A1 and A3) in the 324 autumn sampling formed a separate group related to the general low abundance found in these 325 sites as a result of the higher discharge in that period. The rest of the sites presented a more 326 diverse community characterized by the presence of mayflies and several families of worms. The 327 ordination carried out with the data on invertebrate biomass (Fig. 3C) showed sites A2 and LL4 328 to be separated from the others. These sites were characterized by the dominance of the non-329 biting midge Chironomus spp (mainly bernensis and plumosus) both in terms of abundance and 330 biomass (Fig. 4A). Site A3 was characterized in the first spring sampling by the dominance of 331 Oligochaeta (mainly *Tubifex tubifex*, Fig. 4B). This taxon was also present in the other sampling 332 periods but with moderate values.

333

Communities of diatoms and invertebrates found in this study are characteristics of a perturbed fluvial system. There was a general decrease of species richness downstream and prevalent abundance of the most tolerant species to organic and chemical pollution. Salinity,

- high nutrient concentration and low flow are considered to be responsible for biologically poorcommunities, made up of tolerant taxa [15, 17].
- 339

340 *Relationships between chemical and biological parameters* 

341 The results of the BEST procedure showed no significant correlation between the diatom

342 community and the physicochemical and pharmaceutical variables. However, a significant rank

343 correlation (0.492, p=0.001) was observed between invertebrate abundance and the

344 concentrations of indomethacine and propranolol, as well as with water temperature. Invertebrate

biomass also showed a significant rank correlation (0.810, p=0.002) with the concentration of

ibuprofen, atenolol and propranolol. On the basis of the results from the Best analysis, if the true

347 driving abiotic variables are selected, and two sites have very similar suites of values for these,

348 then the assemblages would also be expected to be similar (and vice-versa). This was the case in

349 sites A3 and LL4, where the reduced invertebrate community, mostly made up of midge larvae

350 (*Chironomus* spp.) and Oligocaheta showed higher abundances and biomass when the river

351 carried higher concentrations of anti-inflammatories and Beta-blockers (Fig. 4D). Remarkably,

352 temperature was the only selected environmental parameter related to invertebrate community

abundance. This observation probably reflects the spatial arrangement of the polluted sites.

354 Neither nutrients nor conductivity were selected as significant variables correlated with

invertebrate community in these sites, regardless of their relevance as by-products of human

activities in the Llobregat River and their established effect on the biological communities [15].

357 The finding that diatom distribution was not affected by the pharmaceutical products is also

358 relevant, however, this observation may be attributed to the mode of action of these products,

359 which does not directly affect the primary producers at the observed concentrations [2].

360

361	The RDA determined that the concentration of some anti-inflammatories and
362	propranolol, as well as temperature, explained 71% of the taxonomic variance in invertebrate
363	density (Fig. 5). The first RDA axis reflected the distribution of sites along the presence of
364	indomethacine and propranolol but also on the increase of temperature. Ibuprofen was the
365	variable most correlated with axis 2; temperature was also correlated with this axis (Table 3).
366	Sites A2 and LL4 were associated with high concentrations of these pharmaceuticals, in contrast
367	to the remaining sites. The position of the samples in the second axis was associated with the
368	high concentration of ibuprofen in site A3, especially in the first sampling period. Chironomus
369	spp. abundance was closely associated with the highest concentrations of propranolol and
370	indomethacine and colder waters that were characteristic of site LL4 (Fig. 5B). Instead, the
371	abundance of families of Oligochaeta (Naididae, Enchytraeidae, and Tubificidae) was found to
372	be related to higher ibuprofen concentrations (site 3). The remaining taxa were related to low
373	concentrations of pharmaceuticals (Fig. 5B).
374	
375	The RDA analysis for diatoms showed a slight correlation of the environmental variables
376	with the axis, and no significance of the Monte Carlo test was found.
377	
378	The results of both multivariate analyses for invertebrate assemblage were similar, sites
379	with higher concentrations of anti-inflammatories and Beta blockers and higher temperatures
380	were characterized by a greater abundance and biomass of Chironomus spp. and Tubifex tubifex.
381	Several studies on the effects of chronic exposure indicate that ibuprofen has very little impact

382 on aquatic environments [24-26]. However, an increase in the somatic growth of *Daphnia magna* 

383	population when exposed to ibuprofen (20 mg/L) and a reduction in reproduction has been
384	described [27]. These authors suggested that the negative impact on reproduction would release
385	energy to be invested in growth. Preliminary results (López-Doval, unpublished results) of a
386	chronic laboratory test with Chironomus riparius exposed to sediments spiked with
387	indomethacine (120 $\mu$ g/g sediment), show an increase of nearly 60% of individual growth
388	respect to the control treatment.
389	
390	Other authors listed numerous effects of anti-inflammatories in freshwater organisms, for
391	example, Schwaiger et al. [28] and Triebskorn et al. [29] detected bioaccumulation and
392	hystopathological alterations in kidney and gills in rainbow trout and common carp exposed to
393	diclofenac (lowest-observed-effect concentrations: 1-5 mg/L) at a concentration range found in
394	our present study. Pharmaceuticals into the environment may affect the same pathways in
395	animals with similar target organs, tissues, cells or biomolecules. Knowledge about these targets
396	exists primarily for fish but less is known in invertebrates or other phyla [2].
397	
398	The toxicity of the Beta-blockers is difficult to ascertain in invertebrates since most
399	studies have analyzed the effects only in D. magna [30]. Propranolol shows the highest acute
400	toxicity and highest log $K_{OW}$ among beta-blockers. Stanley et al. [31] found 48-h propranolol
401	median effective concentration value for D. magna.of 1.67 mg/L. These experimental
402	concentrations were extremely higher than those we registered in the Llobregat water. Beta
403	blockers may have several effects on fish, such as cardiovascular dysfunction and impairment of
404	fitness [32] or reproduction [33]. Propranolol swells fish erythrocytes, thereby affecting oxygen
405	uptake.

406	Some psychiatric drugs, like carbamazepine, show low acute toxicity in aquatic
407	organisms, particularly invertebrates [34]. However, in response to chronic sediment-exposure,
408	the midge Chironomus riparius shows a blockage of pupation and emergence (10% effective
409	concentration values 70 to 210 mg/kg dry wt). Although information on the concentrations of
410	pharmaceuticals in sediment is limited, it is feasible that these compounds accumulate in this
411	compartment [35], thereby posing a risk for the survival of populations of benthic organisms.
412	

413 Some of the mechanisms of action of pharmaceuticals described in the literature could 414 explain the association between these substances and the invertebrates found in the Llobregat 415 River. However, the correlational findings could also be the result of cumulative or synergistic 416 effects caused by several stressors that co-occur in the system. Pharmaceuticals do not appear as 417 isolated compounds in river water, but as a mixture, and data on the responses of aquatic 418 organisms to a mixture of pharmaceuticals is very limited. Escher et al. [36] reported higher 419 toxicity of a pharmaceutical mixture than separated products. Flaherty et al. [37] indicated 420 unpredictable and complex effects of a pharmaceutical mixture on Daphnia survival, growth and 421 reproduction. Moreover, other indirect effects can be influential factors, such as habitat and food 422 availability, species competition or predator-prey interactions. Although habitat characterization 423 (sediment grain size) showed slight differences in the studied sites, little is known about the other 424 parameters. Therefore, our results should be taken as indicative and require further experimental 425 testing in controlled conditions such as mesocosms [38-40].

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- 427
- 428

## 429 Conclusion

430 Several pharmaceutical products in the Llobregat River were found at concentrations higher than 431 those cited in the literature. The low water flow of Mediterranean rivers increases the potential environmental risk of these emerging water contaminants. Thus, much of the aquatic biota will 432 433 be exposed, throughout their lives, to complex mixtures of these compounds. Like other 434 contaminants, the best way to reduce the ecological impact of pharmaceuticals in rivers may be 435 to prevent input by improving sewage treatment procedures. 436 One of the major objectives for environmental scientists is to establish causal links 437 between stressors and the quality of ecological systems. Procedures that identify causality allow 438 corrective action measures for habitat recovery and control. Identifying the cause of biological 439 impairment in freshwater systems is also an essential objective of the European Water 440 Framework Directive. Our results reveal a potential causal relationship between the 441 concentration of a number of pharmaceutical products and the abundance and biomass of several 442 key benthic invertebrates. Although our assessment has been based on field data, it provides 443 evidence on how pharmaceuticals disrupt biological communities. Although strict guidelines for 444 developing cause and effect relationships are not well established, some criteria support causality 445 in environmental impact studies [40]. Experiments that include evidence from multiple field and 446 laboratory studies are stronger than those based alone on only one kind of data. Knowledge of 447 field patterns will help us to focus subsequent monitoring efforts and to identify key taxa for use 448 as ecological indicators. This approach might be useful to find spatial and temporal correlation of 449 stressor and effects along gradients, although it should be combined with community 450 experiments in the laboratory to examine hypotheses generated from field studies. This could be 451 a desirable way to go on in future risk management decisions for these emerging water

- 452 contaminants taking into account both the scarcity of experimental studies with durations longer
- 453 than a few weeks and the few works that used multi-species in standardized bioassays.
- 454
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## 463 **References**

- 1. Sanderson H, Johnson DJ, Reitsma T, Brain RA, Wilson CJ, Solomon KR. 2004. Ranking
- 465 and prioritization of environmental risks of pharmaceuticals in surface waters. *Regul Toxicol*
- 466 *Pharmacol* 39:158–83.
- 467 2. Fent K, Weston AA, Caminada D. 2006. Ecotoxicology of human pharmaceuticals. *Aquat*468 *Toxicol* 76:122-159.
- 469 3. Gros M, Petrovic M, Barceló D. 2007. Wastewater treatment plants as a pathway for aquatic
- 470 contamination by pharmaceuticals in the Ebro river basin (NE Spain). Environ Toxicol Chem
- 471 26:1553-1562.
- 472 4. Martínez Bueno MJ, Aguera A, Gómez MJ, Hernando MD, García Reyes JF, Fernández
- 473 Alba AR. 2007. Application of liquid chromatography/quadrupole-linear ion trp mass
- 474 spectrmetry and time-of-flight mass spectrometry to the determination of pharmaceuticals and
- 475 related contaminants in wastewater. *Anal Chem* 79:9372-9384.
- 476 5. Hernando MD, Mezcua M, Fernandez-Alba AR, Barceló D. 2006. Environmental risk
- 477 assessment of pharmaceutical residues in wastewater effluents, surface waters and sediments.

478 *Talanta* 69: 334-342.

- 479 6. Crane M, Watts C, Boucard T. 2006. Chronic aquatic environmental risks from exposure to
  480 human pharmaceuticals. *Sci Total Environ* 367: 23-41.
- 481 7. Beausse J. 2004. Selected drugs in solid matices: A review of environmental determination,
- 482 occurrence and properties of principal substances. *Trends Anal Chem* 23:753-761.

- 484 8. Clarke KR. 1999. Nonmetric multivariate analysis in community level ecotoxicology.
- 485 Environ Toxicol Chem 18:118-127.

- 486 9. Adams SM. 2003. Establishing causality between environmental stressors and effects on
- 487 aquatic ecosystems. *Hum Ecol Risk Assess* 9:17-35.
- 488 10. Solé M, López de Alda MJ, Castillo M, Porte C, Ladegaard-Pedersen K, Barceló D. 2000.
- 489 Estrogenicity determination in sewage treatment plants and surface waters from the Catalonian
- 490 area (NE Spain). *Environ Sci Technol* 34:5076–5083.
- 491 11. Petrovic M, Solé M, López de Alda MJ, Barceló D. 2002. Endocrine disruptors in sewage
- 492 treatment plants, receiving river waters, and sediments: Integration of chemical analysis and
- 493 biological effects on feral carp. *Environ Toxicol Chem* 21:2146-2156.
- 494 12. Farré M, Ferrer I, Ginebreda A, Figueras M, Olivella L, Tirapu LL, Vilanova M, Barceló D.
- 495 2001. Determination of drugs in surface water and wastewater samples by liquid
- 496 chromatography-mass spectrometry: Methods and preliminary results including toxicity studies
- 497 with vibrio fischeri. J Chromatogr A 938:187-197.
- 498 13. Kuster M, López de Alda MJ, Hernando MD, Petrovic M, Martín-Alonso J, Barceló D.
- 499 2008. Analysis and occurrence of pharmaceuticals, estrogens, progestogens and polar pesticides
- 500 in sewage treatment plant effluents, river water and drinking water in the Llobregat river basin
- 501 (Barcelona, Spain). *J Hydrol* 358:112-123.
- 502 14. Prat N, Puig MA, Gonzalez G, Tort MF, Estrada M. 1984. *Llobregat*. In Whitton BA, ed,
- 503 Ecology of European Rivers. Blackwell Scientific Publications, Oxford, UK, pp 527–551.
- 504 15. Muñoz I, Prat N. 1994. A comparison between different biological water quality indexes in
- the Llobregat basin (NE Spain). Verhandlungen International Vereinigung Limnologie 25:19451949.
- 507 16. Sabater S, Sabater F, Tomàs X .1987. Water quality and diatom communities in two Catalan
  508 rivers (N.E. Spain). *Water Res* 21:901-911.

- 509 17. Leira M, Sabater S. 2005 Diatom assemblages destribution in catalan rivers, NE Spain, in
- 510 relation to chemical and physiographical factors. *Water Res* 39:73-82.
- 511 18. Tornes E, Cambra J, Goma J, Leira M, Ortiz R, Sabater S. 2007. Indicator taxa of benthic
- 512 diatom communities: a case study in Mediterranean streams. *Int J Limnol* 43:1-11.
- 513 19. Gros M, Petrovic M, Barcelo D. 2006. Development of a multi-residue analytical
- 514 methodology based on liquid chromatography-tandem mass spectrometry (LC-MS/MS) for
- 515 screening and trace level determination of pharmaceuticals in surface and wastewaters. *Talanta*
- 516 70:678-690.
- 517 20. Benke AC, Huryn AD, Smock LA, Wallace JB. 1999. Length-mass relationships for
- 518 freshwater macroinvertebrates in North America with particular reference to the southeastern
- 519 United States. J North Am Benthol Soc 18:308–343.
- 520 21. Clarke KR, Warwick RM. 2001. Change in Marine Communities: An Approach to
- 521 Statistical Analysis and Interpretation, 2nd ed. Primer-E. Plymouth, UK.
- 522 22. ter Braak CJF, Smilauer P. 1998. Software for Canonical Corommunity Ordination
- 523 CANOCO version 4. Microcomputer Power, Ithaca, NY, USA.
- 524 23. Comoretto L, Chiron S. 2005. Comparing pharmaceutical and pesticide loads into a small
- 525 Mediterranean river. *Sci Total Environ* 349:201-210.
- 526 24. Pascoe D, Karntanut W, Muller CT. 2003. Do pharmaceuticals affect freshwater
- 527 invertebrates? A study with the cnidarian *Hydra vulgaris*. *Chemosphere* 51:521-528.
- 528 25. Pomati F, Netting AG, Calamari D, Neilan BA. 2004. Effects of erythromycin, tetracicline
- and ibuprofen on the growth of *Synechocystis* sp. and *Lemna minor*. *Aquat Toxicol* 67:387-396.

- 530 26. Richards SM, Wilson CJ, Johnson DJ, Castle DM, Lam M, Mabury SA, Sibley PK,
- 531 Solomon KR. 2004. Effects of pharmaceutical mixtures in aquatic microcosms. *Environ Toxicol*
- 532 *Chem* 23:1035-1042.
- 533 27. Heckmann LH, Callaghan A, Hooper HL, Connon R, Hutchinson TH, Maund SJ, Sibly RM.
- 534 2007. Chronic toxicity of ibuprofen to Daphnia magna: Effects on life history traits and
- 535 population dynamics. *Toxicol Lett* 172:137-145.
- 536 28. Schwaiger JH, Mallowa U, Wintermayr H, Negele RD. 2004. Toxic effects of the non-
- 537 steroidal anti-inflammatory drug diclofenac Part I: histopathological alterations and
- 538 bioaccumulation in rainbow trout. *Aquat Toxicol* 68:141-150.
- 539 29. Triebskorn R, Casper H, Scheil V, Schwaiger J. 2007. Ultrastructural effects of
- 540 pharmaceuticals (carbamazepine, clofibric acid, metoprolol, diclofenac) in rainbow trout
- 541 (Oncorhynchus mykiss) and common carp (Cyprinus carpio). Anal Bioanal Chem 387:1405-
- 542 1416.
- 543 30. Hernando MD, Petrovic M, Fernández-Alba AR, Barceló D. 2004. Analysis by liquid
- 544 chromatography–electrospray ionization tandem mass spectrometry and acute toxicity evaluation
- 545 for Beta-blockers and lipid regulating agents in wastewater samples. J Chromatrogr A 1046:133-
- 546 140.
- 547 31. Stanley JK, Ramírez AJ, Mottaleb M, Chambliss CK, Brooks BW. 2006. Enantiospecific
- 548 toxicity of the Beta-blocker propranolol to *Daphnia magna* and *Primephales promelas*. *Environ*
- 549 *Toxicol Chem* 25:1780-1786.
- 550 32. Owen SF, Giltrow E, Huggett DB, Hutchinson TH, Saye J, Whter MJ, Sumpter JP. 2007.
- 551 Comparative physiology, pharmacology and toxicology of Beta-blockers: Mammals versus fish.
- 552 Aquat Toxicol 82:145-162.

- 553 33. Hugget DB, Brooks BW, Peterson B, Foran CM, Schlenk D. 2002. Toxicity of select beta
- adrenergic receptor blocking pharmaceuticals (Beta-blockers) on aquatic organisms. Arch

555 Environ Contam Toxicol 43:229-235.

- 556 34. Oetken M, Nentwig G, Löffler D, Ternes T, Oehlmann J. 2005. Effects of pharmaceuticals
- 557 on aquatic invertebrates. Part I. The antiepileptic drug carbamazepine. Arch Environ Contam
- 558 *Toxicol* 49:353–361.
- 559 35. Kinney CA, Furlong ET, Kolpin DW, Burkhardt MR, Zaugg SD, Werner SL, Bossio JP,
- 560 Benotti MJ. 2008. Bioaccumulation of pharmaceuticals and other anthropogenic waste indicators
- 561 in earthworms from agricultural soil emended with biosolid or swine manure. *Environ Sci*
- 562 *Technol* 42:1863-1870.
- 563 36. Escher BI, Bramaz N, Eggen RIL, Richter M. 2005. In vitro assessment of models toxic
- action of pharmaceuticals in aquatic life. *Environ Sci Technol* 39:3090-3100.
- 565 37. Flaherty CM, Dodson SI. 2005. Effects of pharmaceuticals on *Daphnia* survival, growth,
- and reproduction. *Chemosphere* 61:200-207.
- 567 38. Muñoz I, Real M, Guash H, Navarro, E, Sabater S. 2001. Effects of atrazine on periphyton
- 568 under grazing pressure. *Aquat Toxicol* 55:239-249.
- 569 39. Sabater S, Guasch H, Ricart M, Romaní A, Vidal G, Klünder C, Schmitt-Jansen M. 2007.
- 570 Monitoring the effect of chemicals on biological communities. The biofilm as an interface. *Anal*
- 571 Bioanal Chem 387:14425-1434.
- 572 40. Culp JM, Lowell RB, Cash KJ. 2000. Integrating mesocosm experiments with field and
- 573 laboratory studies to generate weight-of-evidence risk assessments for large rivers. *Environ*
- 574 *Toxicol Chem* 19:1167-1173.

## 576 Figure legends

- 578 Figure 1. Map of the sampling sites (Spain).
- 579 Figure 2. Concentrations (µg/L) of the families of pharmaceuticals in the sites studied for the
- 580 three sampling campaigns.
- 581 Figure 3. (A) Non metric multidimensional scaling (MDS) ordination of the sites using a Bray-
- 582 Curtis similarities on root-transformed diatom species relative abundances. Lines delimit the
- 583 groups formed by a cluster analysis (group average linked) at 40 % of similarity, (**B**) MDS site
- 584 ordination for invertebrate abundance (individual/cm2) using the same similarity index, (C)
- 585 MDS site ordination using Euclidean distances for invertebrate biomass (mg/cm2), lines delimit
- the groups formed by cluster analysis at 1.1 distance index. Site codes: (A) spring 2005; (B)
- 587 autumn 2005; (C) spring 2006.
- 588 Figure 4. (A-B) Invertebrate abundance multidimensional scaling (MDS) plot from Figure 3,
- 589 with values of *Chironomus* spp. and Tubificidae biomass (mg/m2) at distinct sites, superimposed
- 590 as circles of different sizes on the basis and proportional to biomass values, (**C-D**) the same
- 591 MDS plot on the basis and proportional to values of ibuprofen and propranolol concentrations
- 592 (É g/L) in water samples.
- 593 Figure 5. (A) Redundancy analysis (RDA) ordination plot for sites and environmental
- relationships based on the abundance of invertebrate taxa. The environmental variables more
- 595 significantly correlated with the RDA axes are represented in the byplot by arrows, which point
- 596 was in the direction of maximum change in the value of associated variable. (**B**) Ordination of
- 597 invertebrate species on the first two environmental axes of the RDA.
- 598

	A1		A2		A3		LL1		LL2		LL3		LL4	
	mean	SD												
pН	7.89	0.08	7.61	0.17	8.20	0.23	8.32	0.11	7.83	0.03	8.08	0.07	7.73	0.12
Temperature (°C)	14.67	1.12	23.07	3.49	21.87	4.88	20.33	4.65	20.77	3.36	21.30	3.41	24.47	2.76
Conductivity														
(µS/cm)	3163.33	271.54	3863.33	443.77	2177.67	294.31	1457.33	65.74	1624.33	145.11	1833.00	40.71	2766.67	457.86
Oxygen (mg/L)	8.31	1.72	6.74	2.01	10.25	5.44	11.00	1.08	7.90	0.13	7.63	1.64	6.96	2.04
NO3 (mg/L)	6.92	5.99	24.05	20.09	4.84	4.10	6.19	6.40	6.64	4.85	7.98	5.34	4.52	3.31
SO4 (mg/L)	837.90	237.97	528.89	162.85	372.90	111.20	140.38	8.64	125.92	34.99	239.01	85.04	282.42	71.42
Cl (mg/L)	300.03	78.50	521.43	254.10	278.05	95.93	308.69	121.42	259.47	104.78	315.74	47.12	479.31	131.38
NO2 (mg/L)	0.10	0.07	0.59	0.73	0.43	0.35	0.08	0.08	0.14	0.06	-	-	0.99	0.64
SRP (µgPO4/L)	32.09	18.05	598.98	162.33	313.35	65.26	200.62	43.80	117.11	57.78	350.33	236.19	635.14	164.71
Na (mg/L)	195.38	30.02	349.87	202.38	209.85	76.08	131.26	13.45	146.53	43.10	168.80	42.68	284.65	40.95
K (mg/L)	12.12	9.23	23.65	13.33	28.42	24.97	45.27	25.11	46.42	29.36	23.06	19.39	72.47	50.23
Ca (mg/L)	207.64	21.70	119.34	37.85	113.72	24.87	68.24	4.09	59.98	7.07	71.84	10.41	77.37	6.67
Mg (mg/L)	79.16	19.46	45.68	13.83	43.45	13.13	24.09	6.17	23.15	7.52	29.25	9.66	27.24	13.30
NH4 (mg/L)	0.33	0.32	0.95	0.06	0.47	0.41	0.24	0.34	0.67	0.54	2.31	1.32	0.65	0.18

**Table 1**. Physicochemical parameters and nutrients measured in the water samples collected at the selected sites. Average values and standard deviation (SD) are showed (n=3). SRP: Soluble reactive phosphate

Table 2. Average, minimum and maximum concentrations ( $\mu$ g/L) of pharmaceuticals in the water samples analysed, and concentration at the 75th percentile (*n*=21).

MDL=method detection level

		Mean	Min	Max	75th	MDL (ng/L)
Analgesics and anti-	Ketoprofen	0.79	0.16	2.71	0.71	30
inflamatories	Naproxen	0.53	0.02	2.06	0.65	7
	Ibuprofen	1.37	0.16	9.89	1.51	8
	Indomethacine	0.16	0.05	0.38	0.26	6
	Diclofenac	2.20	0.08	18.74	1.49	2
	Mefenamic acid	0.02	0.01	0.04	0.03	0.5
	Acetaminophen	0.42	0.06	2.42	0.45	17
	Propyphenazone	0.09	0.03	0.18	0.15	3
Lipid regulators and cholesterol lowering	Clofibric acid	2.28	0.01	7.91	3.29	1
statin drugs	Gemfibrozil	1.42	0.04	7.78	1.42	1
	Bezafibrate Pravastatin	1.02 <mdl< td=""><td>0.03</td><td>15.06</td><td>0.35</td><td>1 47</td></mdl<>	0.03	15.06	0.35	1 47
	Mevastatin	<mdl< td=""><td></td><td></td><td></td><td>7</td></mdl<>				7
Psychiatric drugs	Carbamazepine	1.07	0.08	3.09	1.93	2
	Fluoxetine	<mdl< td=""><td></td><td></td><td></td><td>20</td></mdl<>				20
Antiulcer agent	Paroxetine Lansoprazole	<mdl <mdl< td=""><td></td><td></td><td></td><td>8 5</td></mdl<></mdl 				8 5
Histamine H1 and H2	Loratadine	<mdl< td=""><td></td><td></td><td></td><td>2</td></mdl<>				2
receptor antagonists	Famotidine	<mdl< td=""><td></td><td></td><td></td><td>5</td></mdl<>				5
	Ranitidine	0.11	0.01	0.57	0.09	2
Antibiotics	Erythromycin	0.03	0.01	0.07	0.06	4
	Azythromycin	<mdl< td=""><td></td><td></td><td></td><td>1</td></mdl<>				1
	Sulfamethoxazole	1.11	0.03	11.92	0.44	5
	Trimethoprim	0.14	0.02	0.47	0.21	1
<b>D</b> 11 1	Ofloxacin	2.11	0.19	8.77	1.70	16
Beta blockers	Atenolol	0.22	0.05	0.67	0.32	9
	Sotalol	0.57	0.11	1.82	0.63	18
	Metoprolol	0.05	0.01	0.18	0.06	3
	Propranolol	0.03	0.01	0.06	0.06	2

**Table 3.** Correlation between axes and environmental variables following redundancy

 analysis of invertebrate species abundance data from Llobregat River

	Axis 1	Axis 2
Propranolol	0.90	-0.13
Ibuprofen	0.11	0.63
Indomethacine	0.72	-0.12
Temperature	0.52	0.47
Species-environment	0.92	0.81
Eigenvalues	0.44	0.22





Figure 2



DENSITY (ind/cm2)



BIOMASS (mg/cm2)







Figure 4





Figure 5