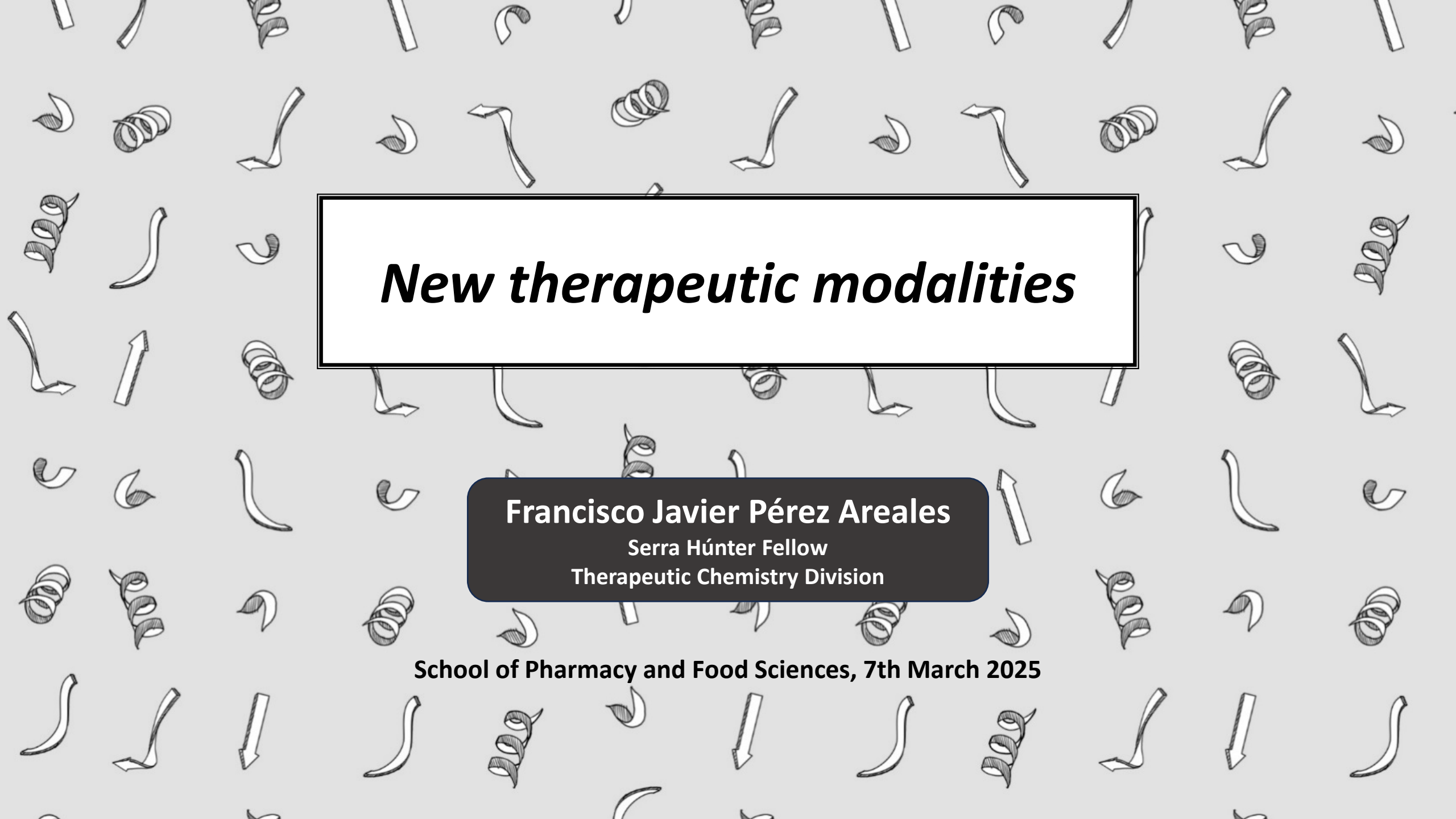


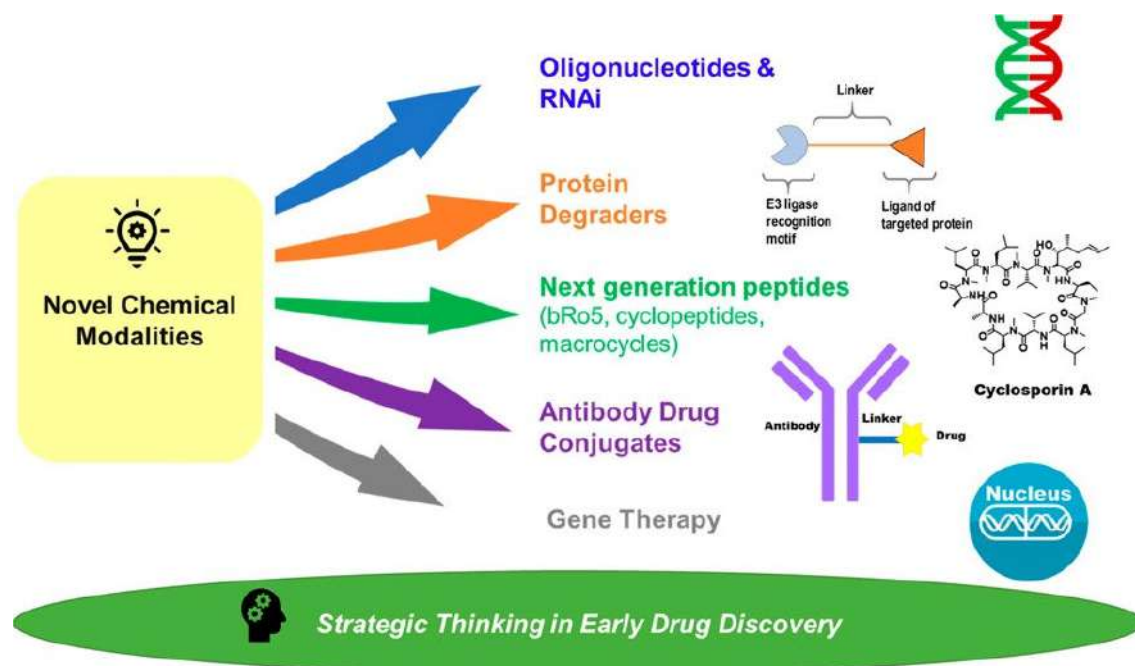
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New therapeutic modalities

Francisco Javier Pérez Areales

Serra Húnter Fellow
Therapeutic Chemistry Division

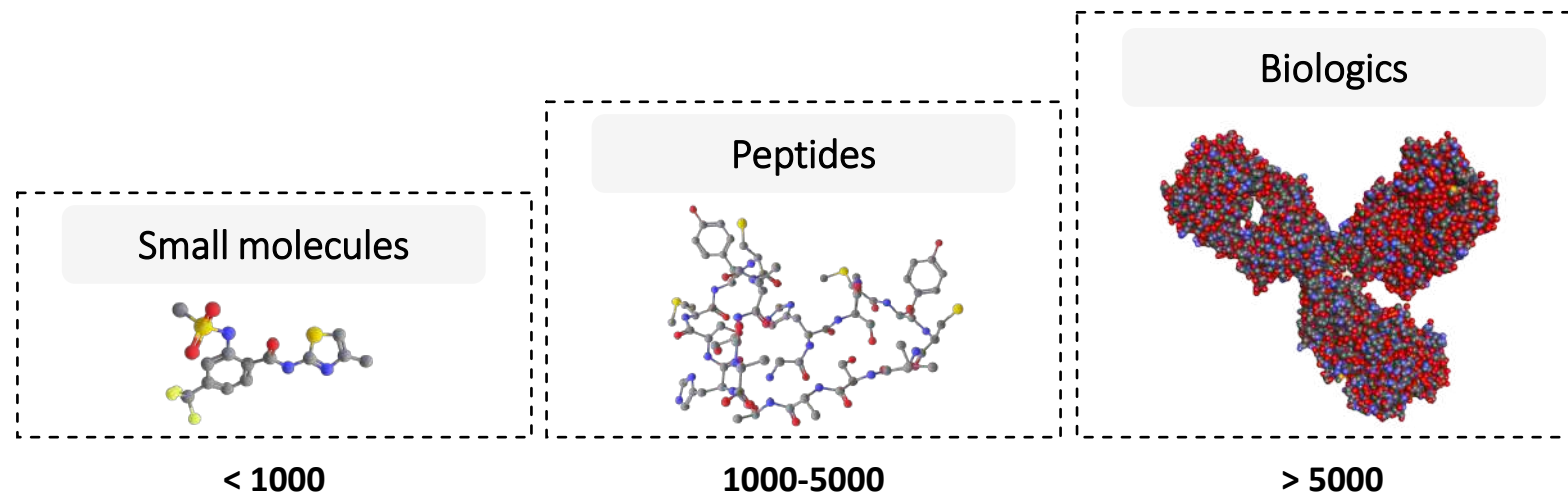
School of Pharmacy and Food Sciences, 7th March 2025



- Therapeutic peptides
- Antibody-drug conjugates (ADCs)
- Targeted protein degradation (TPD)

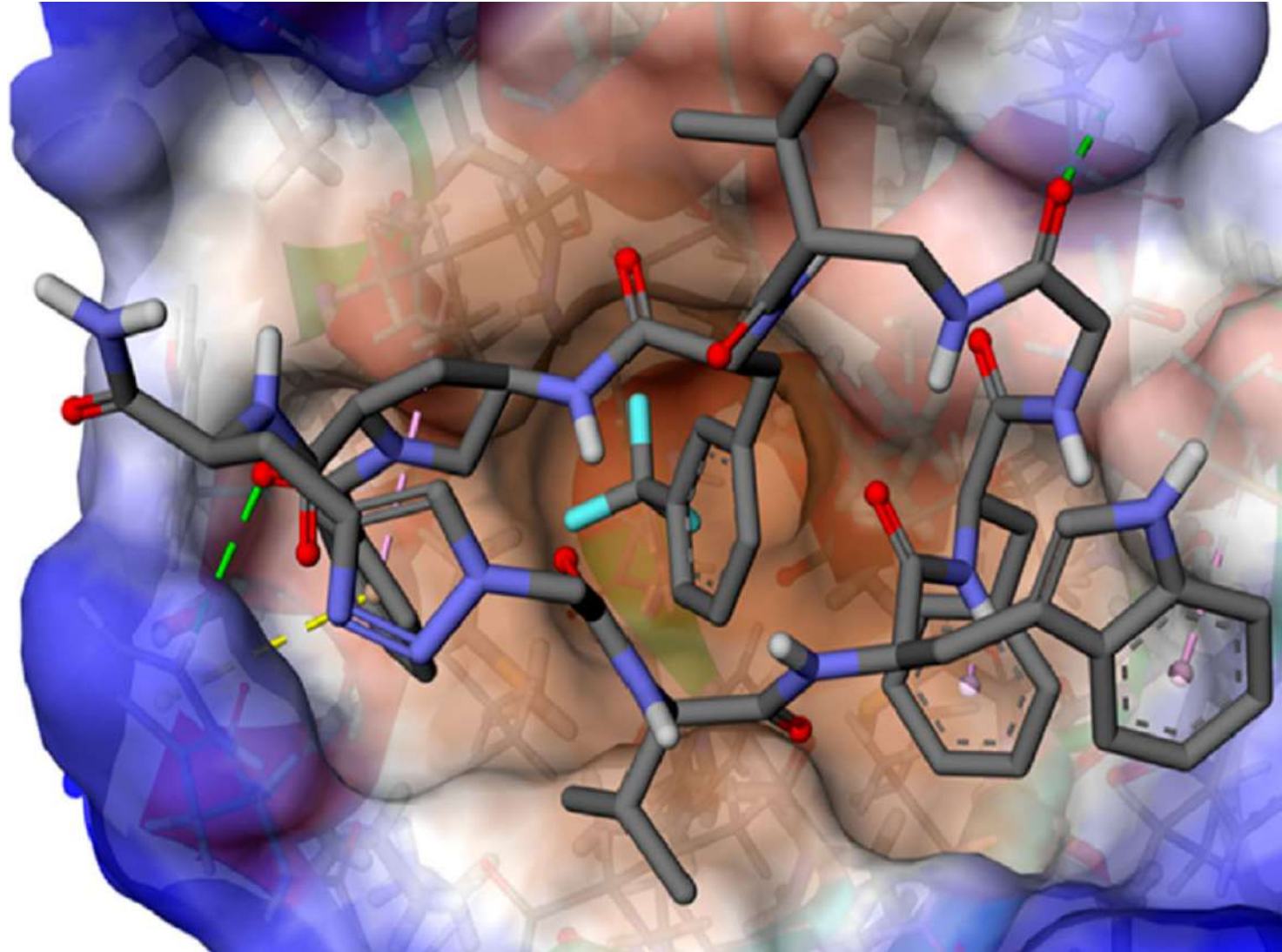
M.-J. Blanco *et al.* *ACS Med. Chem. Lett.* **2020**, 11, 228

Classification of therapeutics

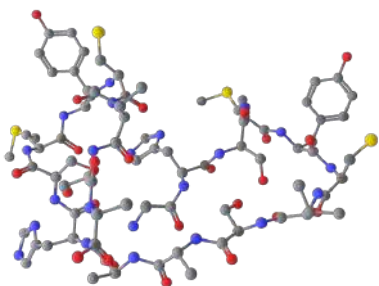


Targets	Small, well-defined pockets	Large, shallow surfaces	Extracellular proteins
Binding affinity	Moderate	High	High
Target specificity	Moderate	High	High
Oral bioavailability	Often	Rarely	No
Membrane permeability	Often	Rarely	No
Plasma half-life	Hours	Minutes to hours	Weeks
Production cost	Low	Moderate	High
Off-target toxicity	Yes	No	No
Immunogenicity	No	No	Yes

Photoswitches for peptide stapling



Peptides



Major limitations:

- Poor enzymatic stability
- Poor membrane permeability



MACROCYCLISATION

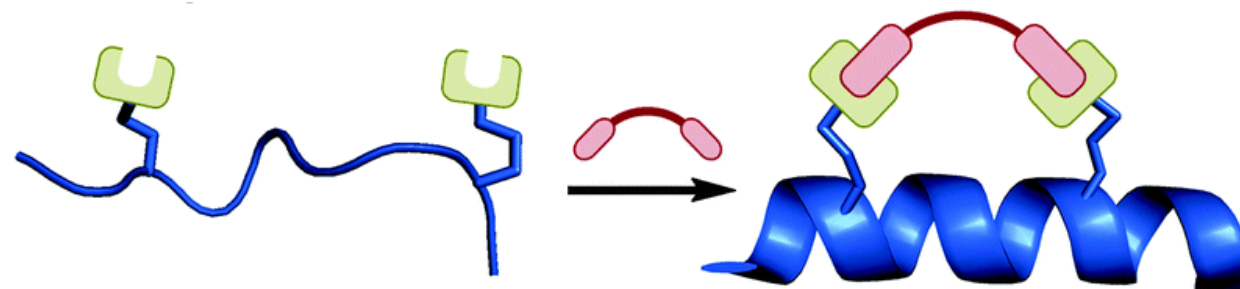
Peptide stapling

Reinforcing the alpha helical conformation:

↑ stability

↑ potency

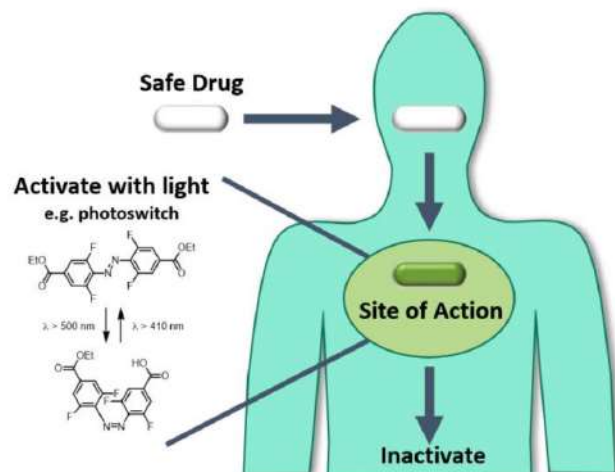
↑ permeability



Photopharmacology

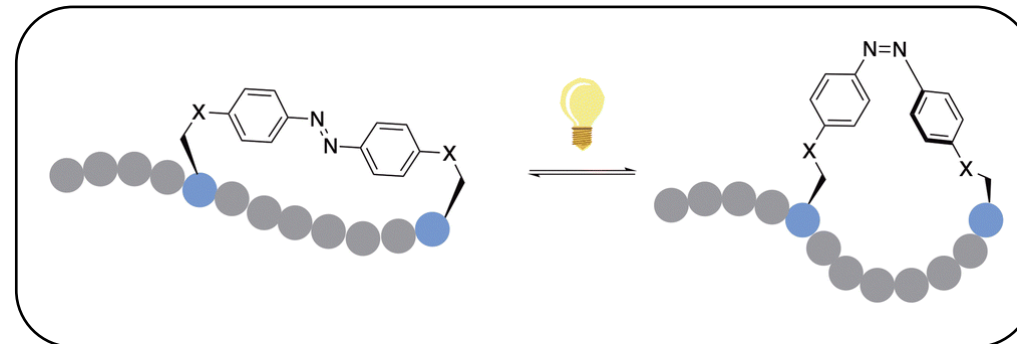
Staple peptides represent an ideal 'substrate' for **photopharmacology**, given the importance of structural changes for their PK profile and biological activity.

The incorporation a platform able to undergo *cis-trans* isomerization upon irradiation with light enables the formation of **photoswitchable peptides**.

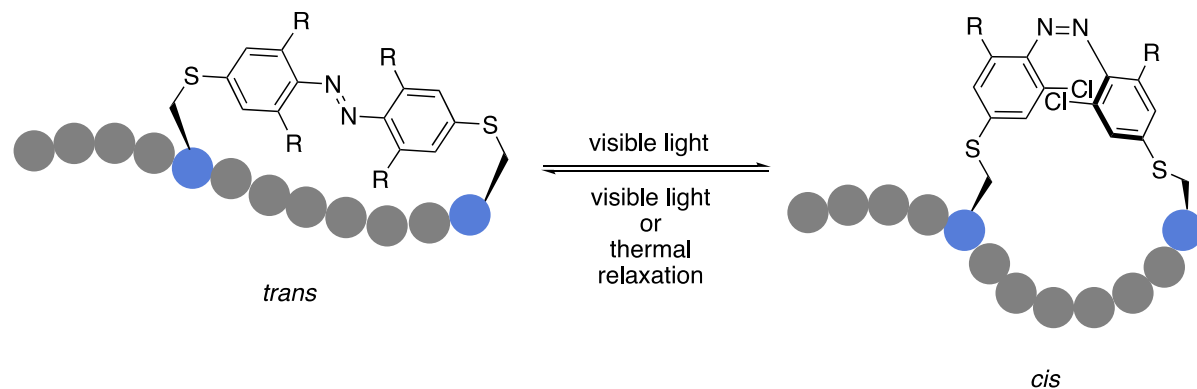


Azobenzene photoswitches

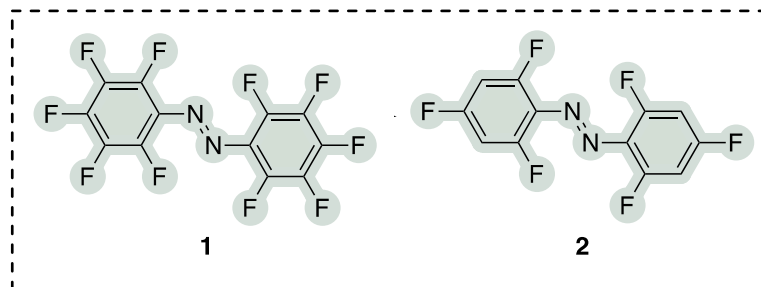
- ✓ Easy synthesis
- ✓ Highly photostable
- ✓ Readily undergo *cis-trans* isomerisation



Objectives

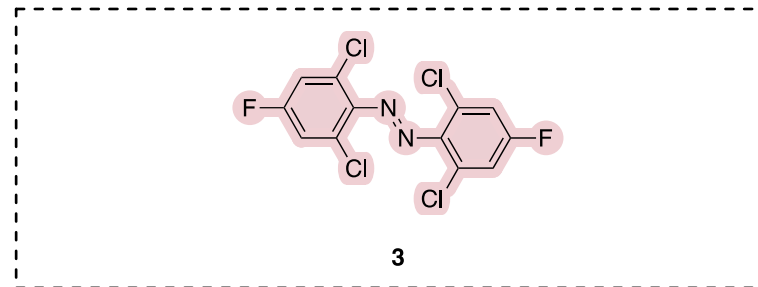


- Visible-light azobenzene platform
- Cysteine selective peptide stapling
- Stapling *via* S_NAr



✓ Green-light isomerisation

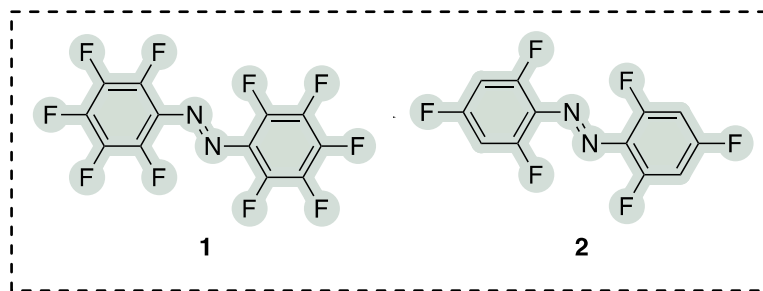
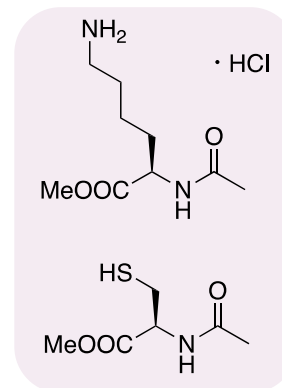
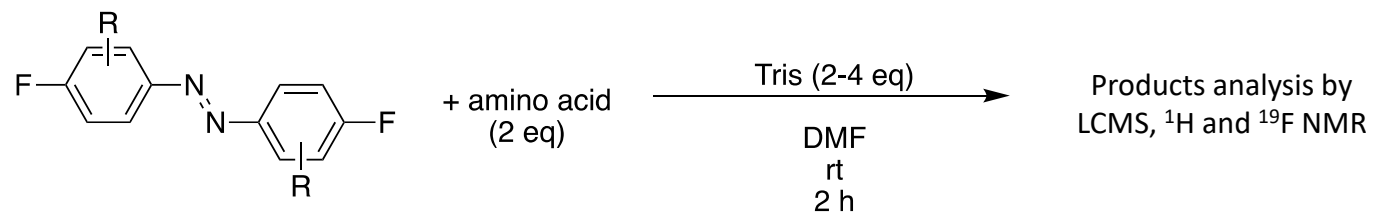
✓ One-step synthesis



✓ Red-light isomerisation

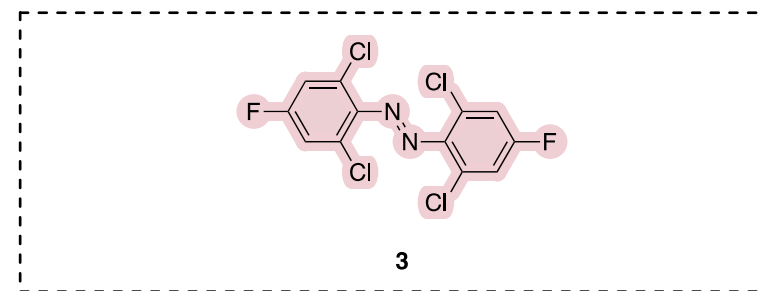
✓ One-step synthesis

Reactivity assessment



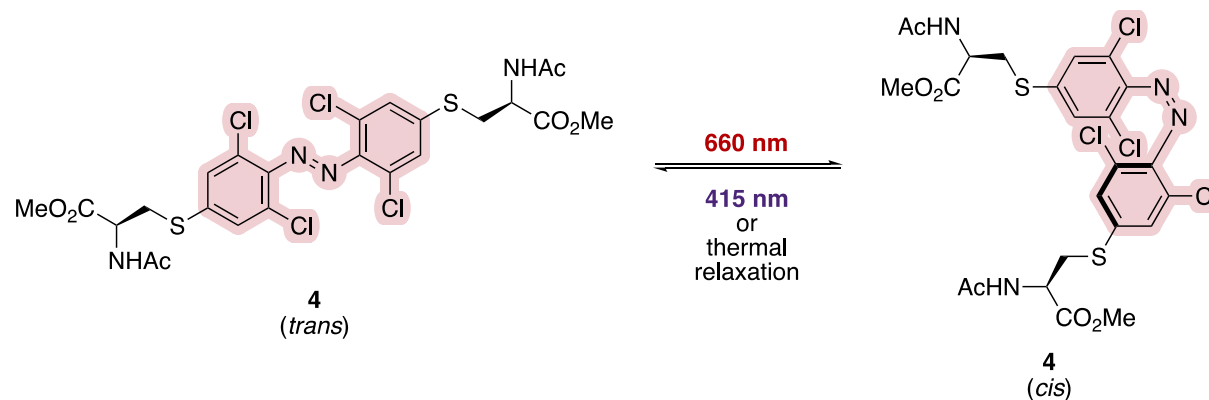
✗ Poly-substitution at *ortho* position

✗ Lysine reactivity

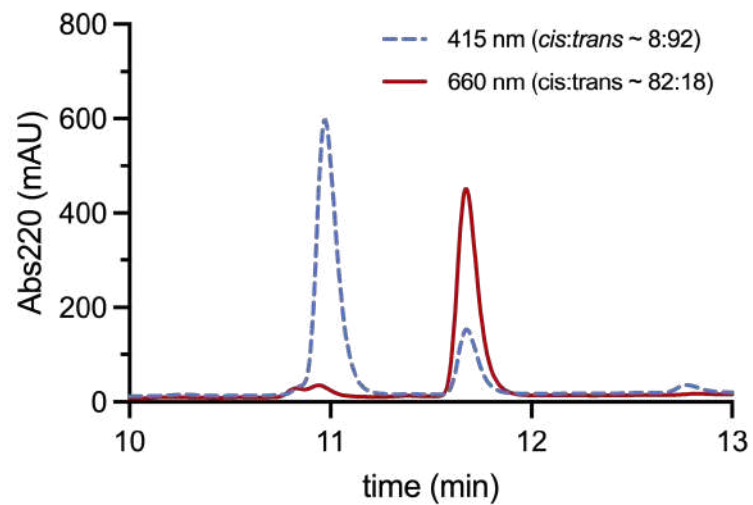


✓ Highly selective for *para* position

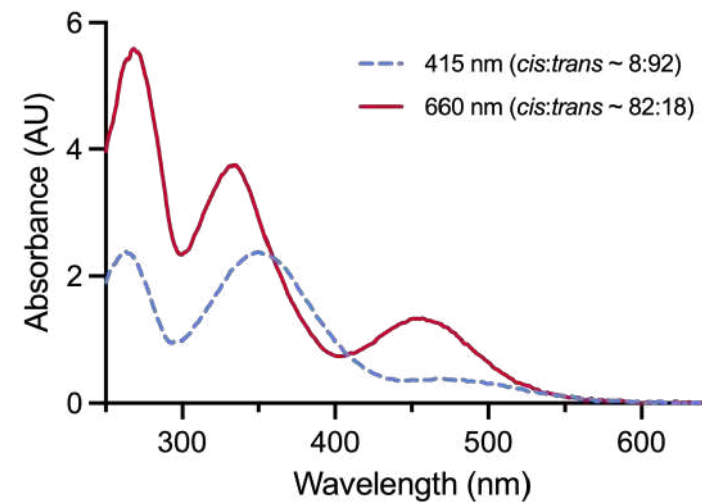
✓ Cysteine selective



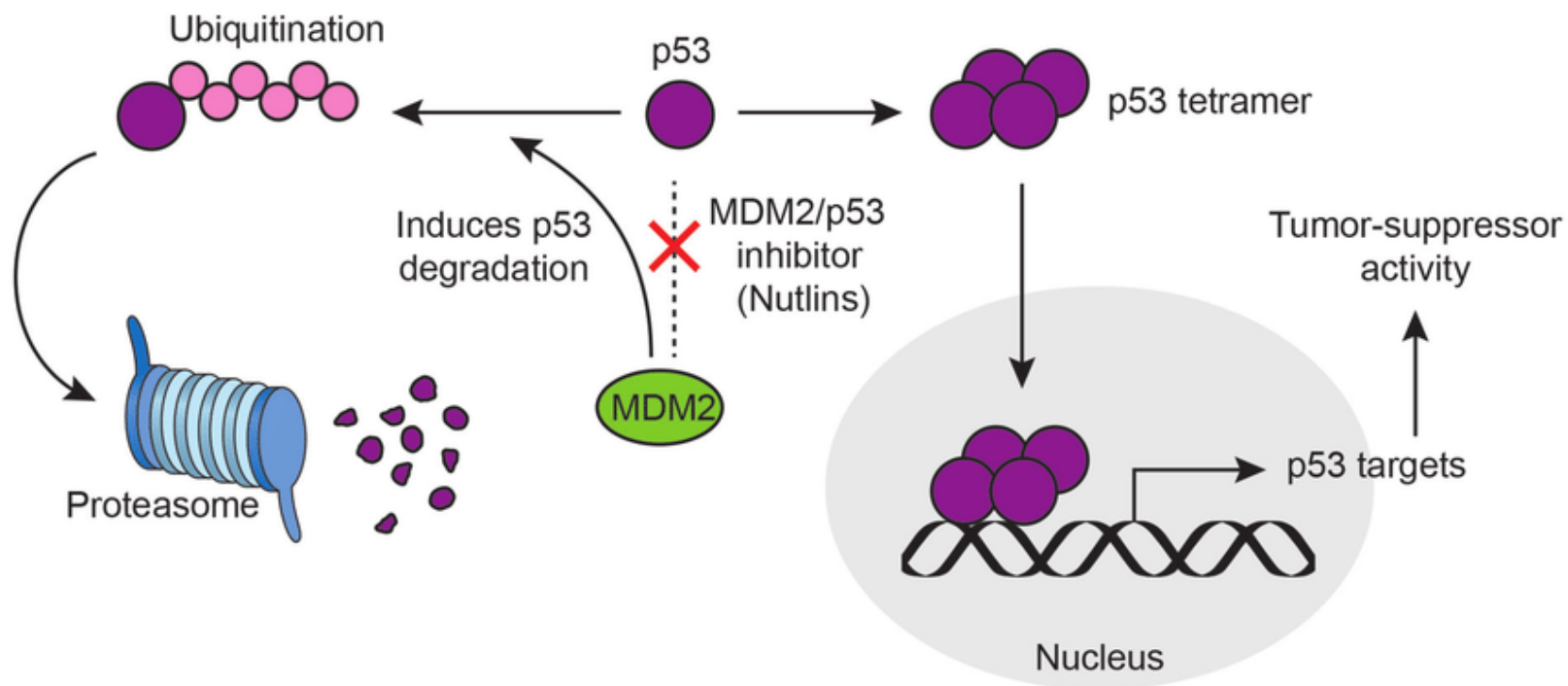
HPLC analysis



UV-vis analysis

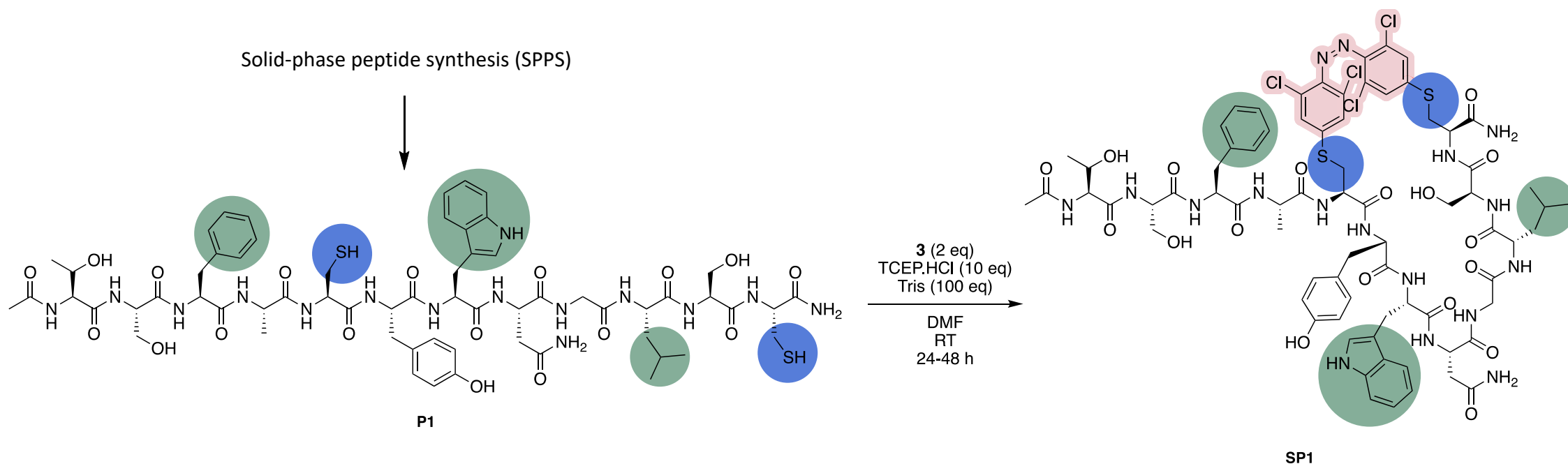


500 μM in $\text{H}_2\text{O}/\text{MeCN}$ 50:50. The spectra were recorded upon 30 min and 90 min irradiation with 415 nm and 660 nm LED lights, respectively.



Inhibition of the p53/MDM2 interaction has been shown to efficiently rescue p53 from degradation, thus recovering its tumour suppressor activity

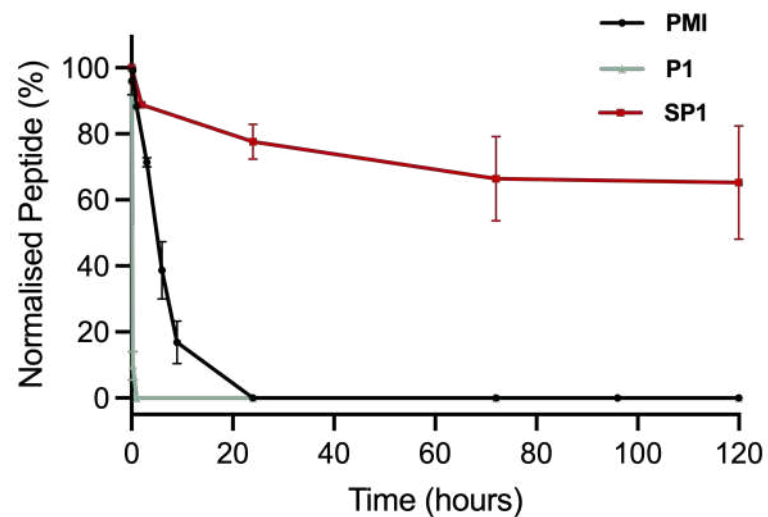
The photoswitchable *ortho*-chloro AB was applied to a derivative of **PMI** (Ac-TSFAEYWNNLSP-NH₂), a potent inhibitor p53–MDM2 interaction



Substitutions at positions 5 and 12 to facilitate (*i*, *i*+7) peptide stapling, allowing for the incorporation of the staple without disrupting the interaction with MDM2, driven by the 'hot spot' residues (Phe3, Trp7 and Leu10)

Peptide	Sequence
PMI	Ac-TSFAEYWNNLSP-NH ₂
P1 (Cys derivative)	Ac-TSFACYWNGLSLSC-NH ₂
SP1	Ac-TSFAXYWNGLSX-NH ₂

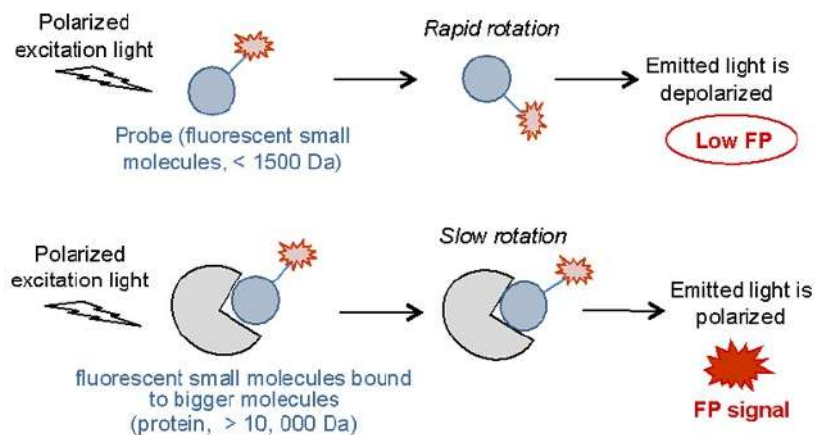
Stability in human serum



Incubated at 37 °C and monitored throughout 5 days by HPLC

Competitive fluorescence polarisation (FP) assay

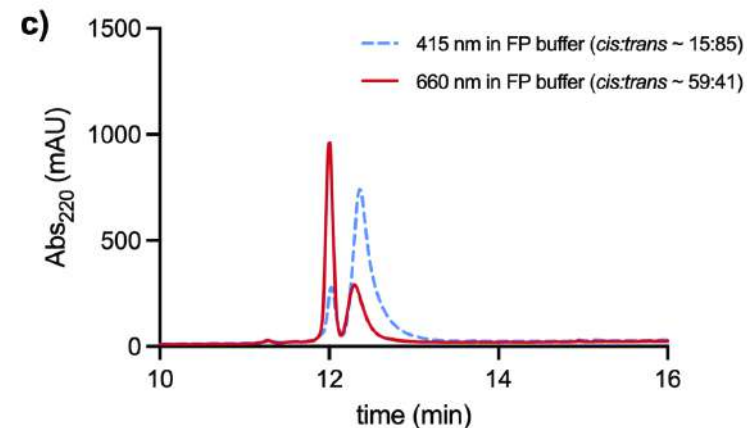
Determination of binding affinity of peptides for MDM2 direct FP, followed by competitive FP assay



Arkin, M.R. *et al.* *Assay Guidance Manual* Eli Lilly & Company and the National Center for Advancing Translational Sciences **2004**

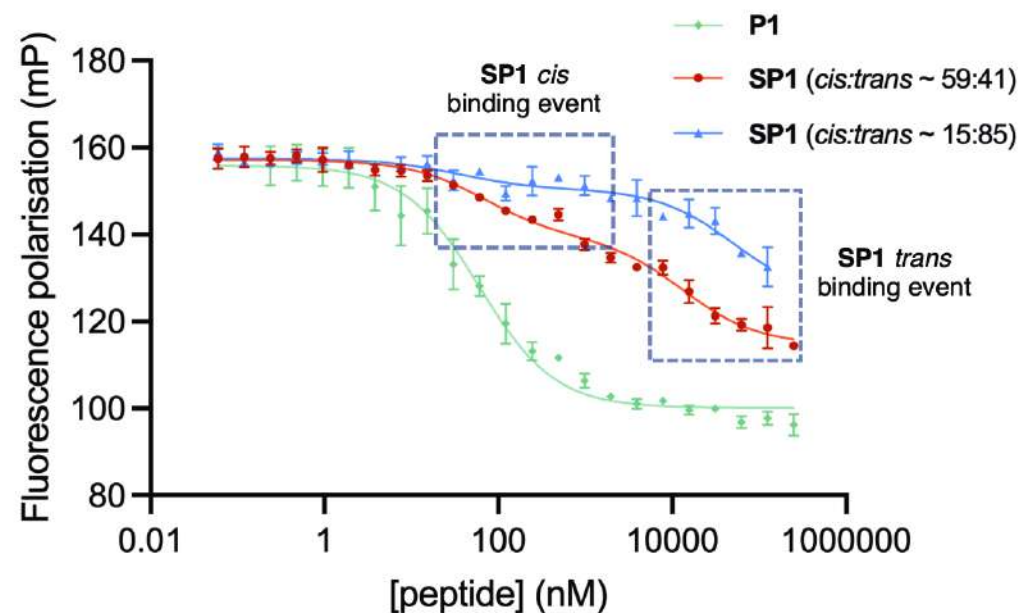
Peptide	Sequence
PMI	Ac-TSFAEYWNNLSP-NH ₂
FP tracer ($K_d = 7.6$ nM)	TAMRA-RFMDYWEGL-NH ₂
P1	Ac-TSFACYWNGLSLSC-NH ₂
SP1	Ac-TSFAXYWNGLSX-NH ₂

HPLC analysis in FP buffer

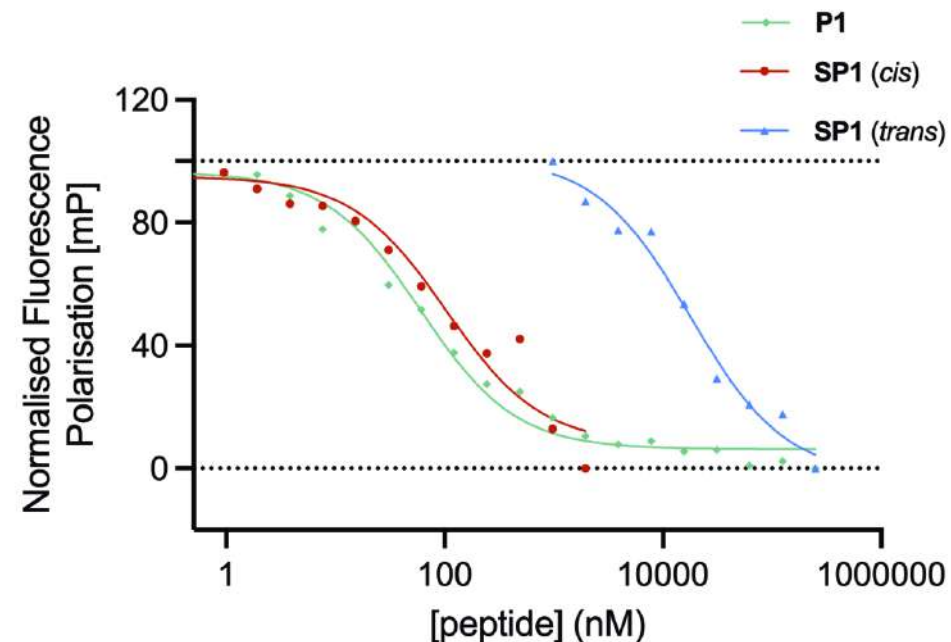


FP buffer (PBS, 0.05% (v/v) Tween-20, 3% DMSO)

Competitive FP assay



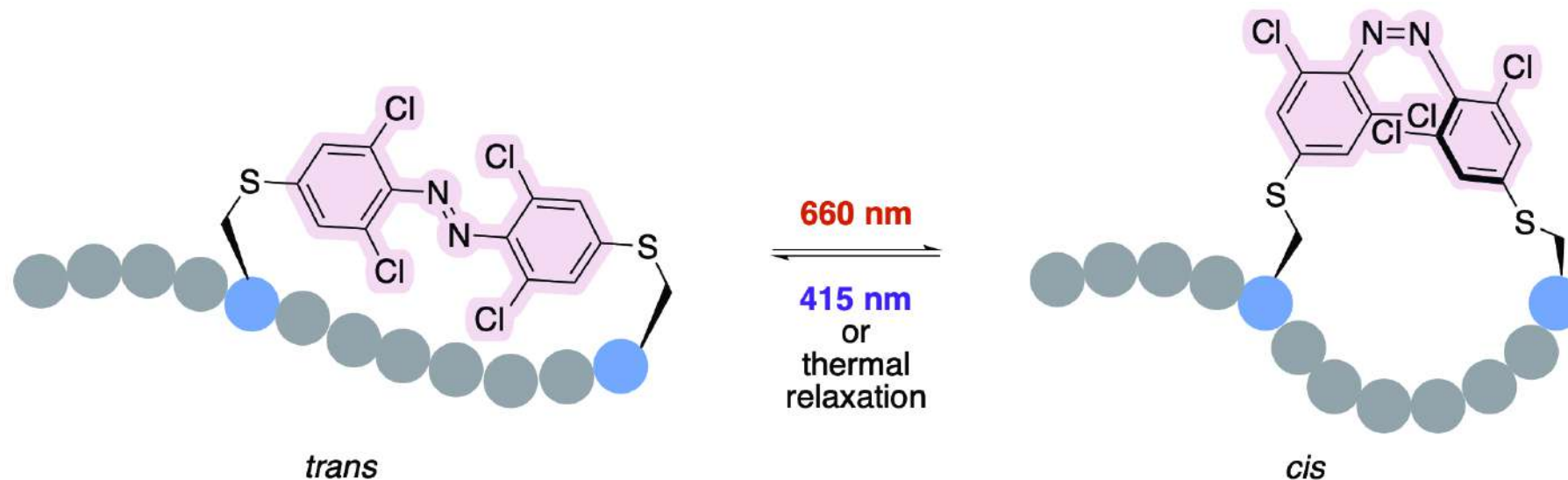
Normalised FP curves



Peptide	PMI	P1	SP1 (<i>trans</i>)	SP1 (<i>cis</i>)
K_i (nM)	0.78 ± 0.36	7.93 ± 3.83	> 13000	54 ± 4.9

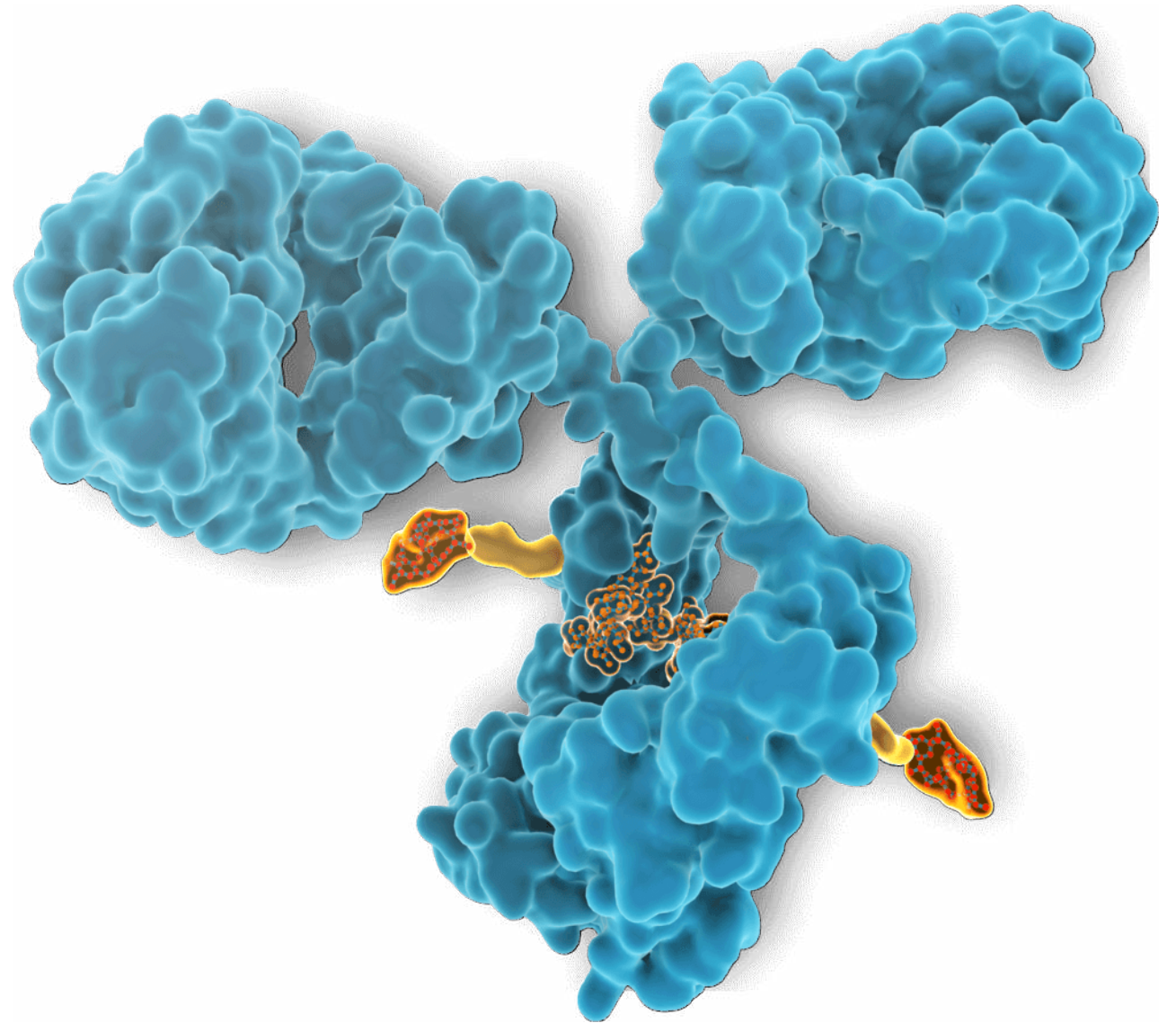
>240-fold higher affinity by *cis* isomer

ortho-chloro azobenzene photoswitch for peptide stapling

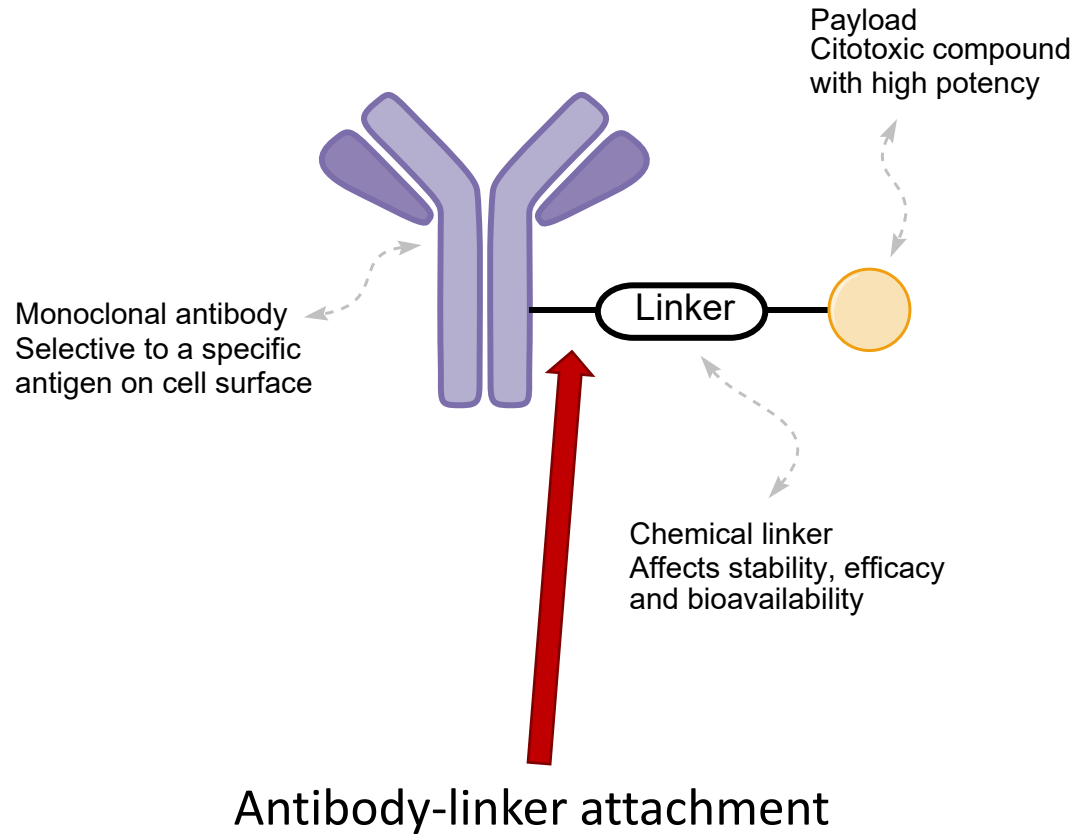


- ✓ Use of red light
- ✓ Cysteine-selective peptide stapling via S_NAr
- ✓ Light induced changes in peptide conformation
- ✓ >240-fold difference in binding affinity for MDM2 upon *trans-cis* isomerisation

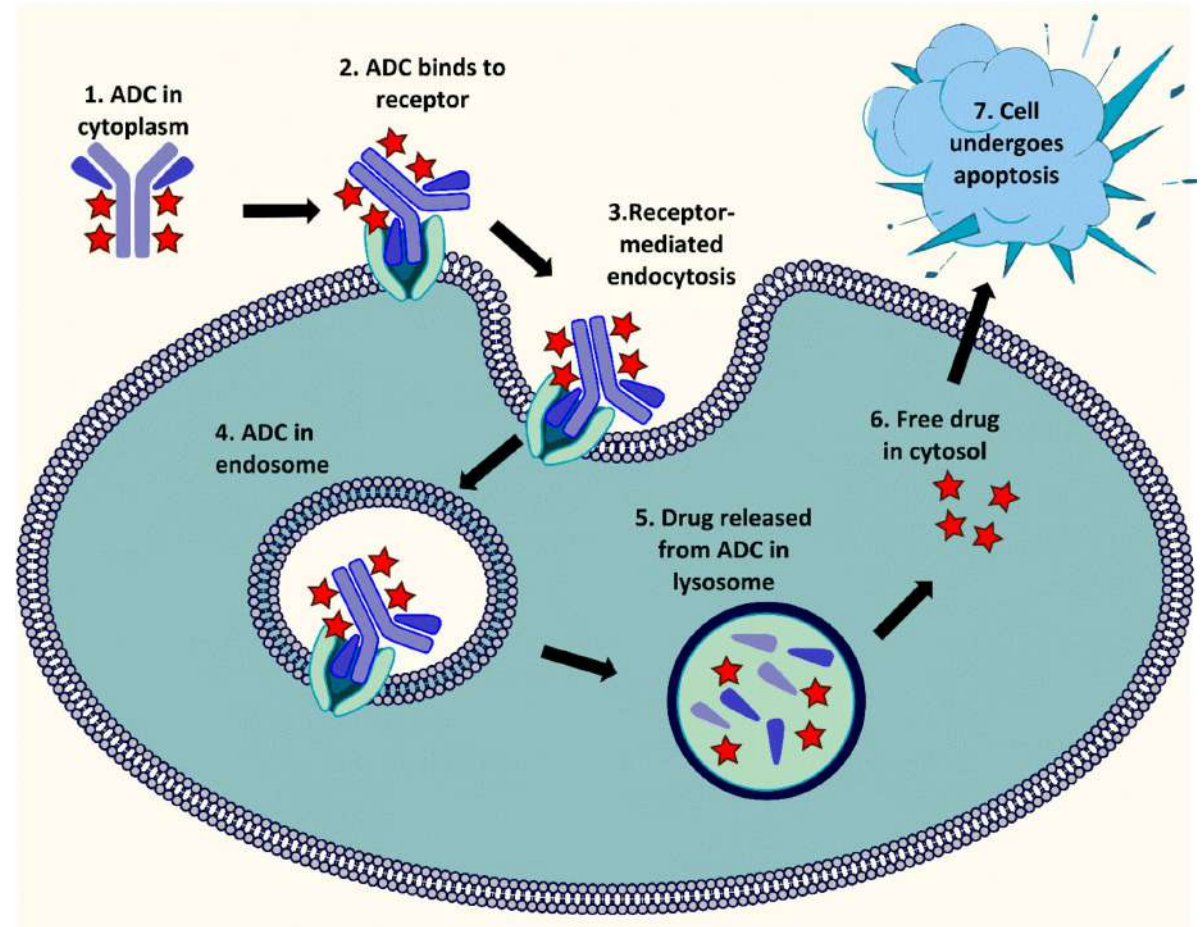
Tetra DVP



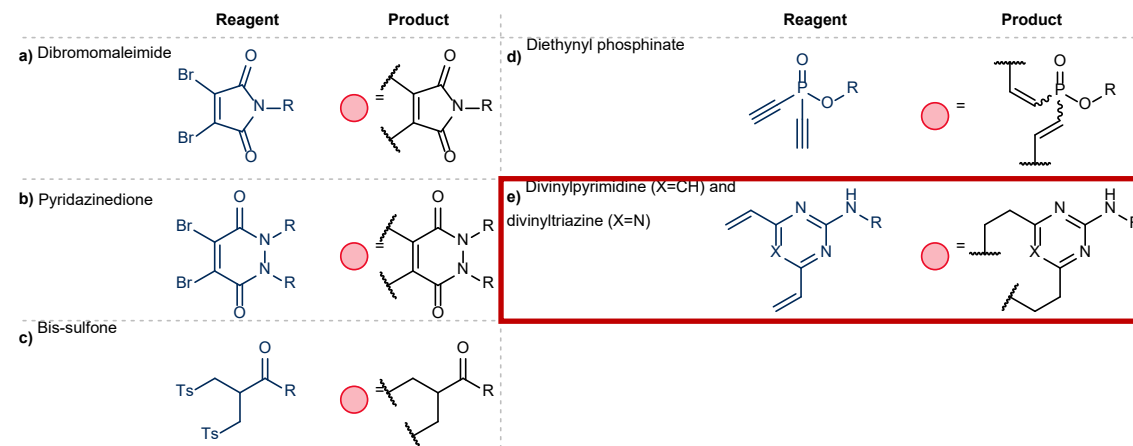
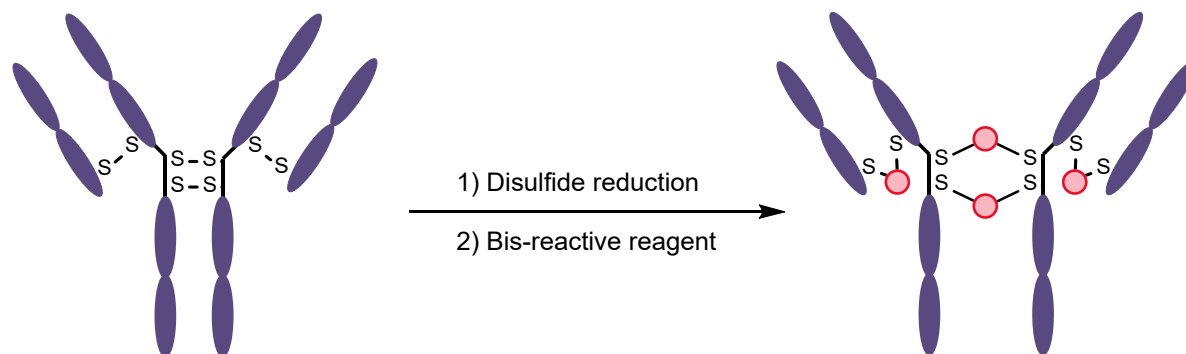
What is an ADC?



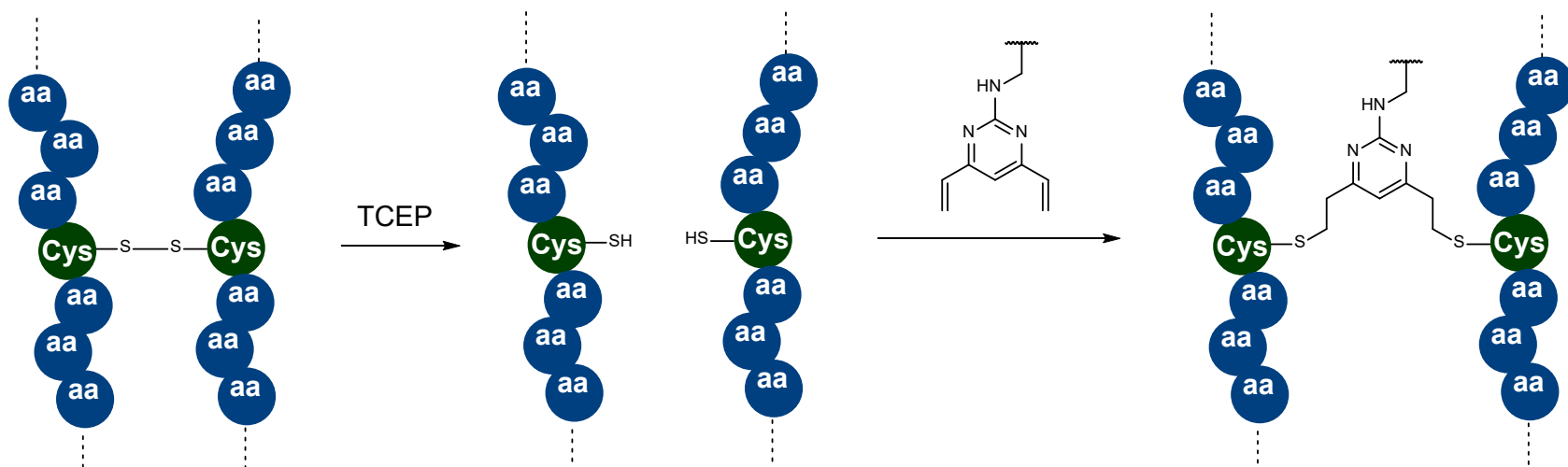
How ADCs work?



Disulfide re-bridging (interchain cross-linking)



Divinylpyrimidine (DVP)



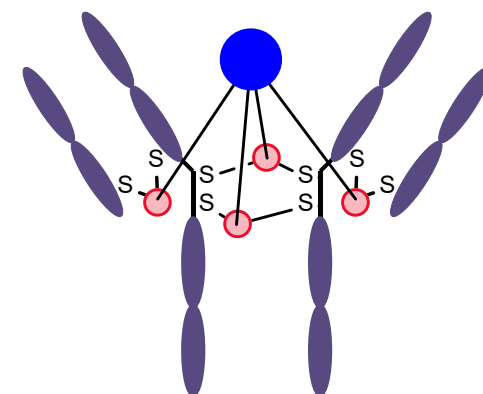
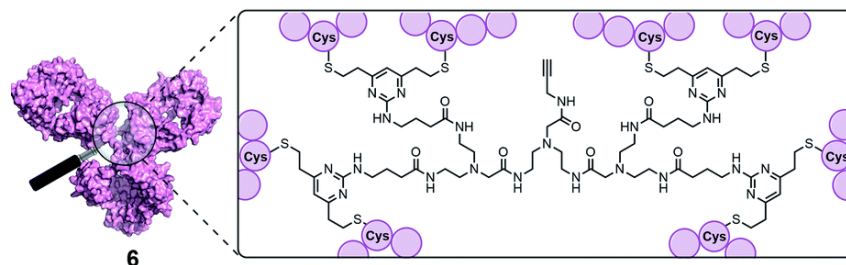
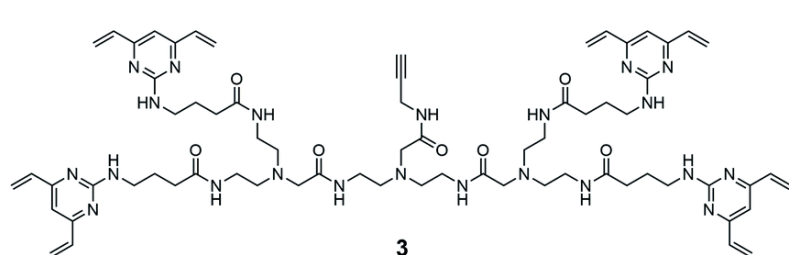
DVP conjugation Properties:

- Fast
- Irreversible
- Stable
- Chemoselective
- **Consistent DAR**
- Facile synthesis

DAR = drug-antibody ratio → Described as an average of cytotoxic units per antibody

DAR consistency is a major limitation of ADCs

Tetra-divinylpyrimidine (tetra-DVP)



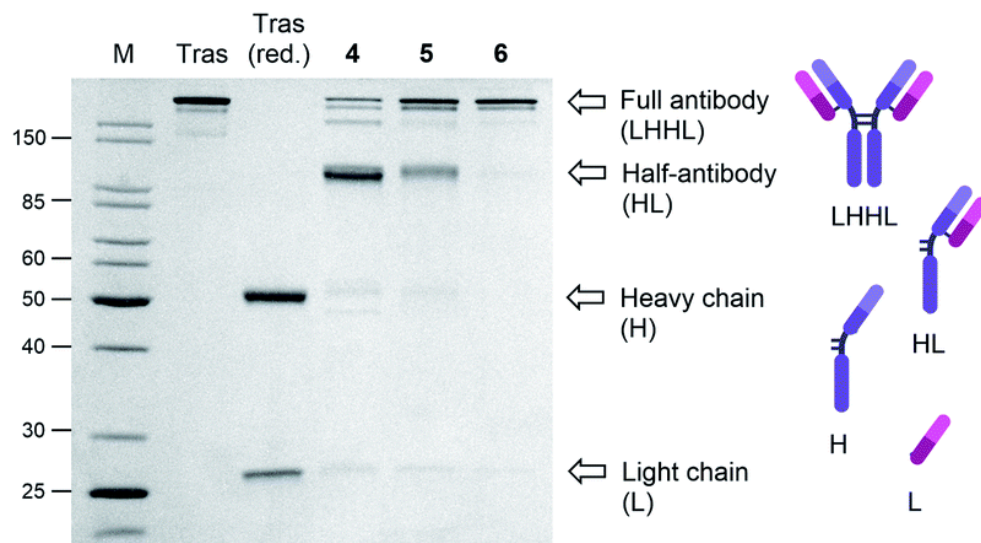
○ = DVP ● = functionalisable group

Tras = Trastuzumab

4 from DVP

5 from bis-DVP

6 from tetra-DVP

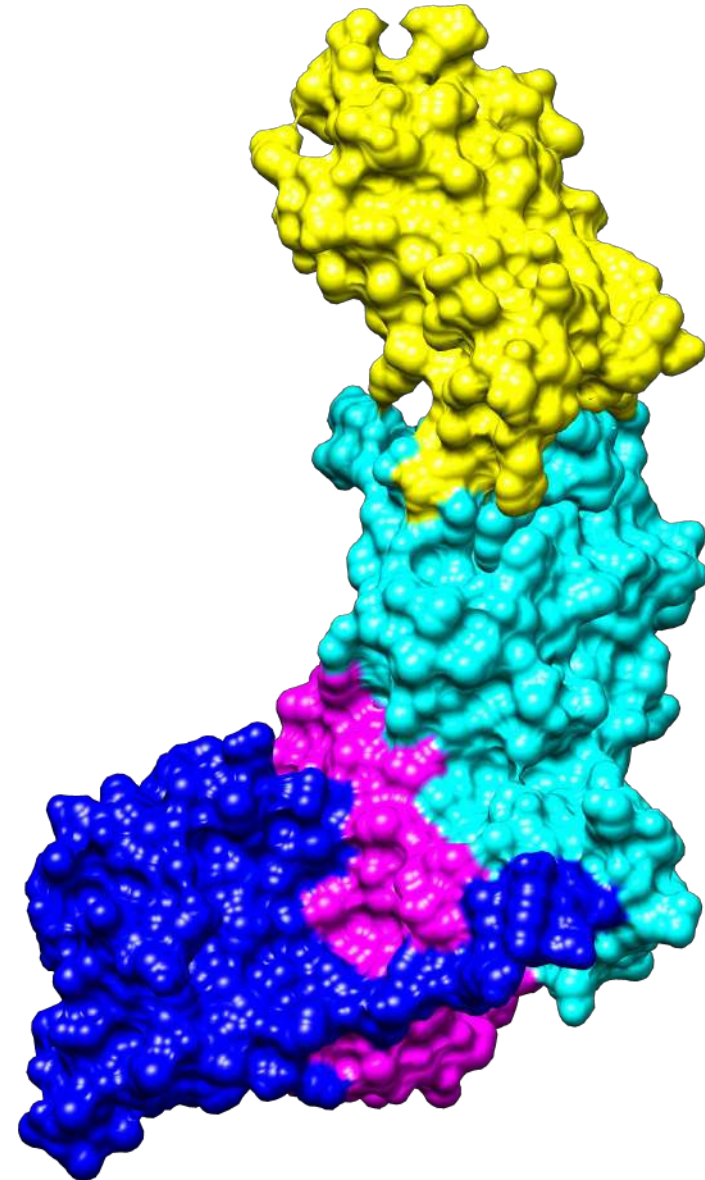


Functionalisable group

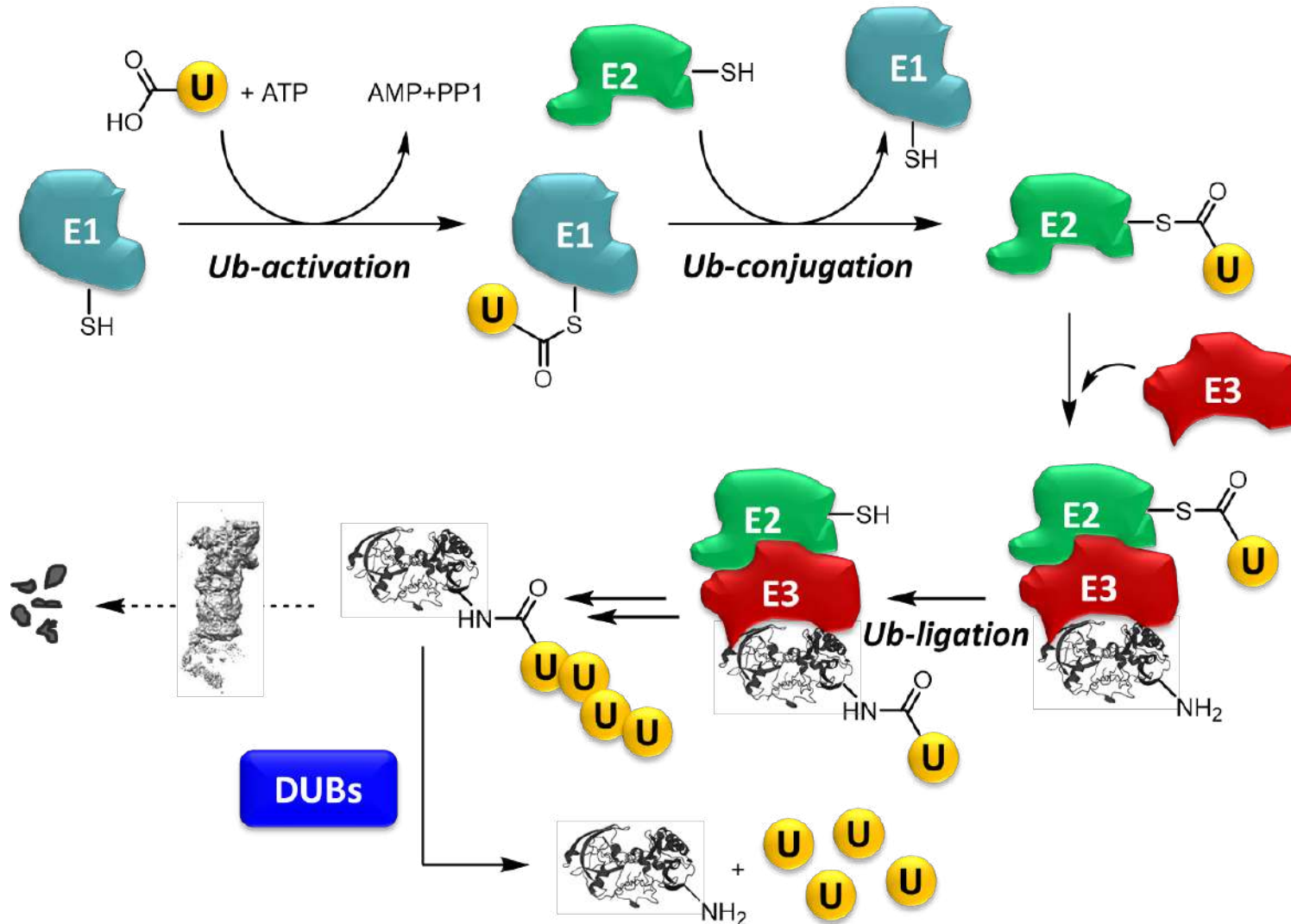


**Modular cargo loading
(consistent DAR)**

Ubiquitin-specific protease 2 targeted degraders (USP2-TD)



Ubiquitin proteasome system



Targeting **deubiquitinating enzymes (DUBs)** offers the opportunity to modulate the UPS with increased specificity, reduced toxicity profiles.

Target validation of DUBs currently represents a priority in the field since we are lacking appropriate **chemical probes** for most of them.



Ubiquitin-specific protease 2 (USP2)

Why USP2?

USP2 is an oncoprotein responsible for the stabilisation of different cell-cycle modulators and signalling pathway factors that lead to **cancer cell survival, proliferation and metastasis.**

www.impactjournals.com/oncotarget/

Oncotarget, Vol. 7, No. 30

Research Paper

Ubiquitin-specific protease 2 decreases p53-dependent apoptosis in cutaneous T-cell lymphoma

THE
EMBO
JOURNAL

The deubiquitinating enzyme USP2a regulates the p53 pathway by targeting Mdm2

Ubiquitin-specific Cysteine Protease 2a (USP2a) Regulates the Stability of Aurora-A^S

Cell
PRESS

Suppression of Cancer Cell Growth by Promoting Cyclin D1 Degradation

Article

Cell Chemical Biology

Lithocholic Acid Hydroxyamide Destabilizes Cyclin D1 and Induces G₀/G₁ Arrest by Inhibiting Deubiquitinase USP2a

J. BIOL. CHEM. VOL. 281, NO. 47, PP. 34608–34612, NOVEMBER 18, 2006
Biochemistry and Molecular Biology, Inc. Published in the U.S.A.

Small Molecule Inhibition of the Ubiquitin-specific Protease USP2 Accelerates cyclin D1 Degradation and Leads to Cell Cycle Arrest in Colorectal Cancer and Mantle Cell Lymphoma Models*

Oncogenic substrates of USP2

MDM2
Aurora A
Cyclin A1/D1
MDM4
FAS
c-Myc
Twist
AIF
TRAF2
EGFR

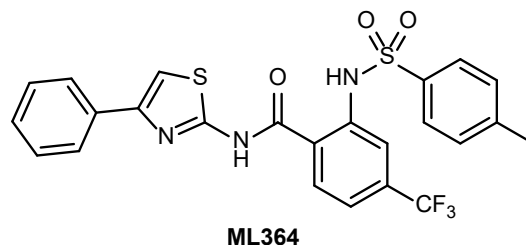
↑ USP2 or
↑ substrate

USP2-associated cancers:

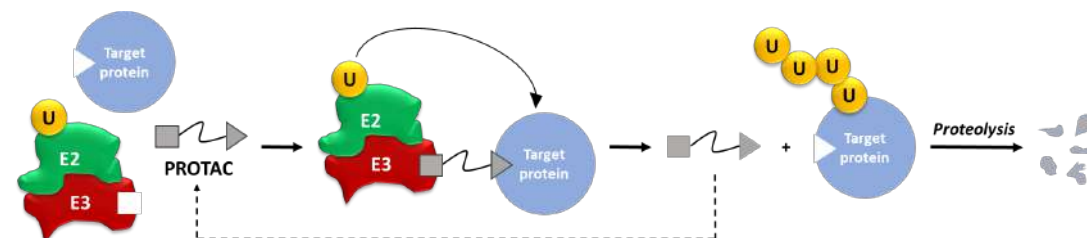
Triple-negative breast
Prostate
Ovarian
Cutaneous T-cell lymphoma
Colon
Breast
Mantle cell lymphoma
Lung
Glioblastoma

ML364, a “chemical probe”?

USP2 chemical inhibition has been barely explored while it has been extensively studied by biologists during the last decade, using ML364 as chemical probe.



Proximity-induced approach



ML364-based USP2 degraders

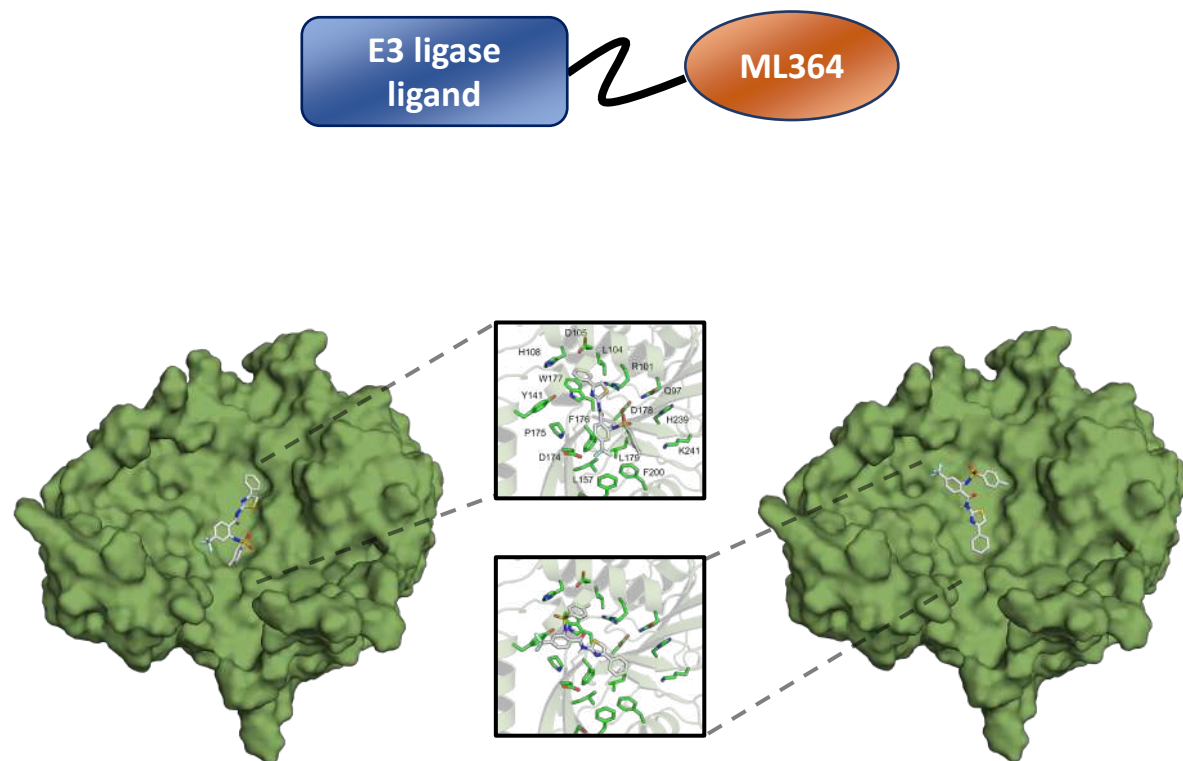
↑ **potency**: catalytic function and depletion of the target

↑ **selectivity**: ternary complex formation

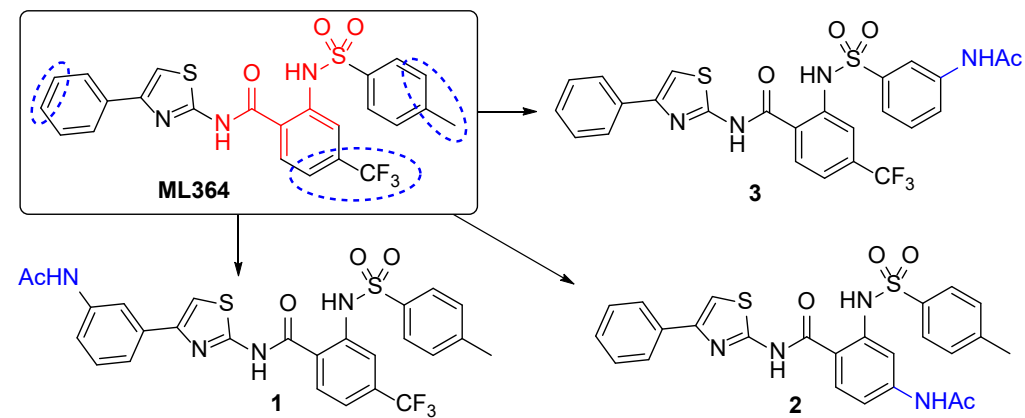
The discovery of an appropriate chemical probe that can be used for a confident interrogation and further understanding of USP2 biology and disease.

Poor quality chemical probe

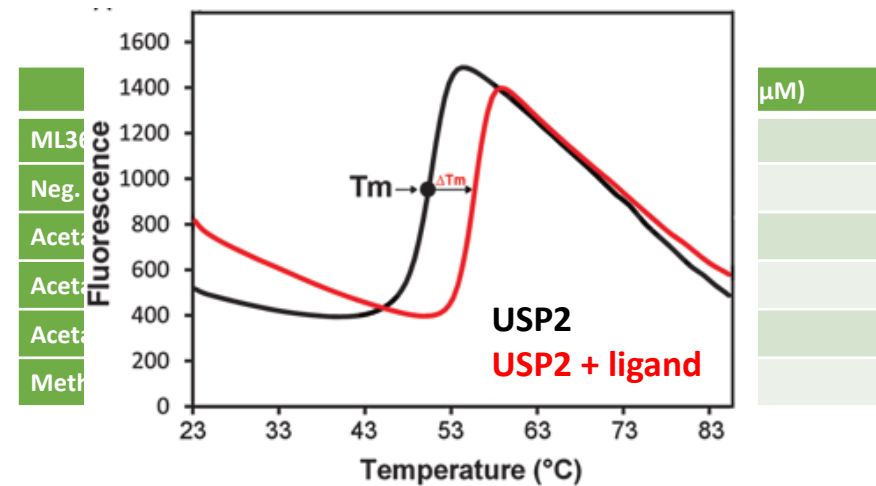
Exit vector investigation



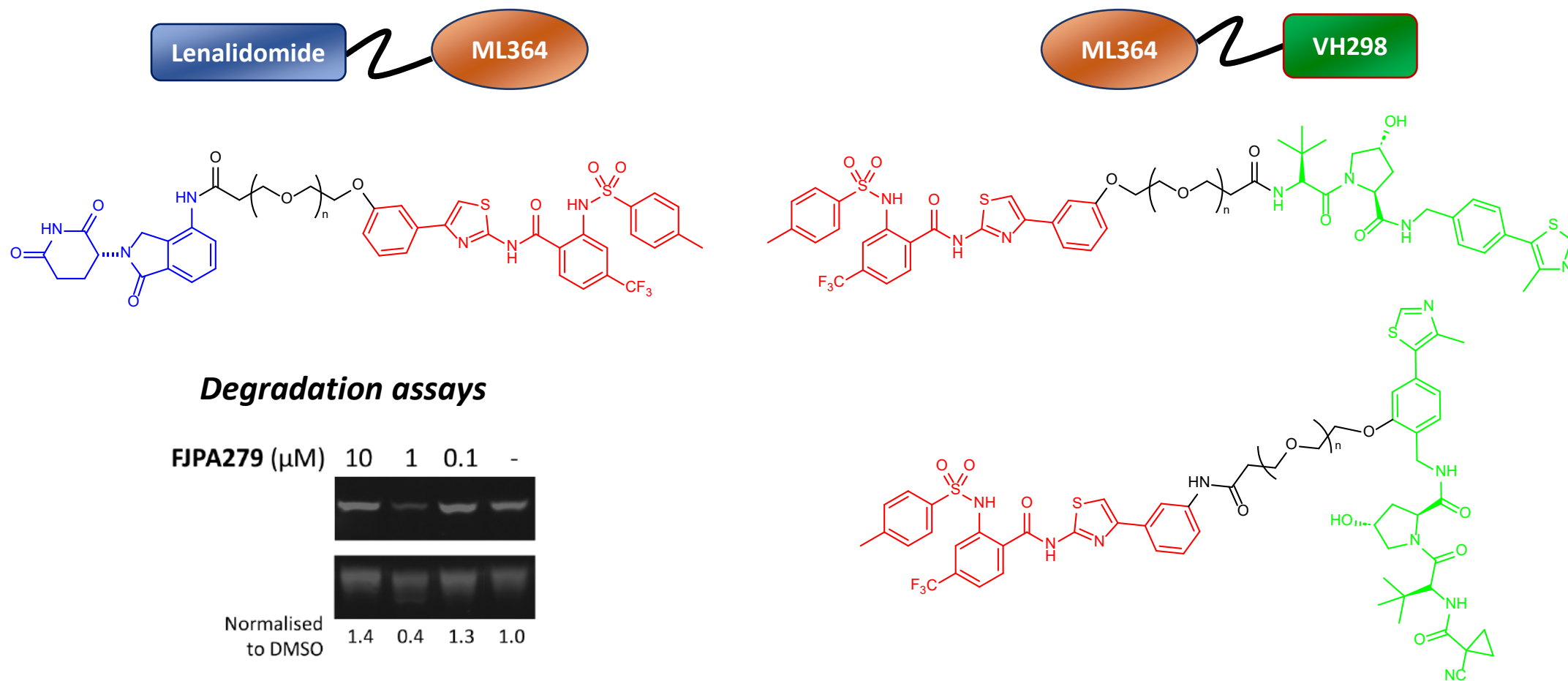
Docking calculations predicted two preferential binding modes at the PPI surface with Ub.



Differential Scanning Fluorimetry (DSF)



Heterobifunctional degrader candidates



FJPA279, a VH032-based compound, was identified as a partial degrader of USP2 at 1 μM, after 24 h treatment in MCF-7 cells.

Acknowledgements



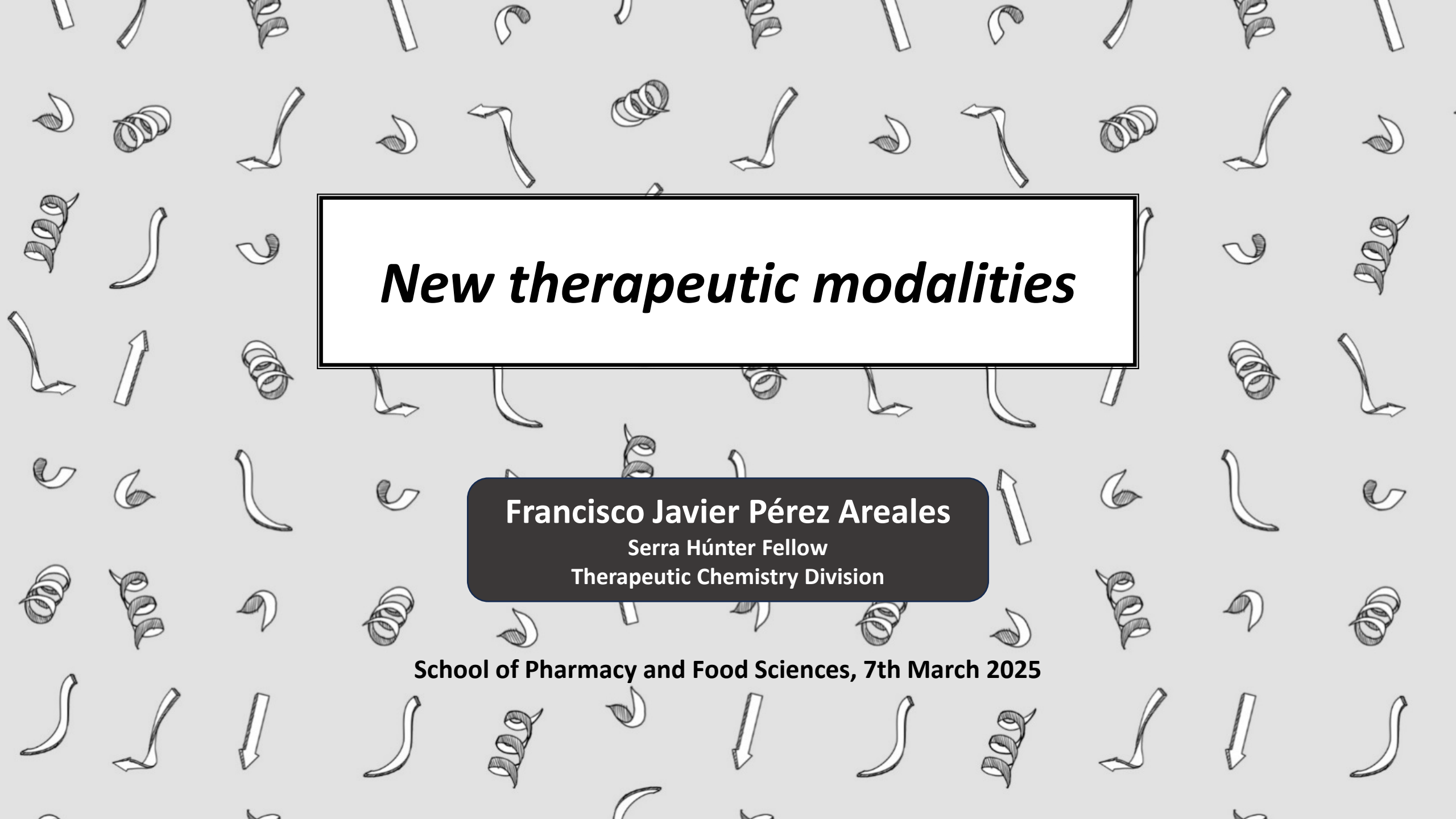
Prof. David Spring
Dr. Mia Kapun
Dr. Nicola Ashman
Dr. Mahri Park
Dr. Steve Walsh
Dr. Raoul Walter
Dr. Tim Schober



Prof. Alessio Ciulli
Aitana de la Cuadra

Dr. Mark Nakasome
Diane Cassidy



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School of Pharmacy and Food Sciences, 7th March 2025