

Contents lists available at [ScienceDirect](https://www.sciencedirect.com)

## Critical Reviews in Oncology / Hematology

journal homepage: [www.elsevier.com/locate/critrevonc](http://www.elsevier.com/locate/critrevonc)

## Autophagy and oxidative stress in solid tumors: Mechanisms and therapeutic opportunities

María Carretero-Fernández<sup>a,b,c,1</sup>, Antonio José Cabrera-Serrano<sup>a,b,1</sup>,  
 José Manuel Sánchez-Maldonado<sup>a,b</sup>, Lucía Ruiz-Durán<sup>a,b</sup>, Francisco Jiménez-Romera<sup>a,b</sup>,  
 Francisco José García-Verdejo<sup>a,c</sup>, Carmen González-Olmedo<sup>a,c</sup>, Aina Cardús<sup>d</sup>,  
 Leticia Díaz-Beltrán<sup>a,c</sup>, Juan Francisco Gutiérrez-Bautista<sup>a,b,e,f</sup>, Yolanda Benavente<sup>g,h,m</sup>,  
 Fernando Gálvez-Montosa<sup>a,i</sup>, José Antonio López-López<sup>a,c</sup>, Paloma García-Martín<sup>b,j</sup>,  
 Eva María Pérez<sup>a,b,j</sup>, Juan José Rodríguez-Sevilla<sup>k</sup>,  
 Delphine Casabonne<sup>g,h</sup>, Pedro Sánchez-Rovira<sup>a,c</sup>,  
 Fernando Jesús Reyes-Zurita<sup>l,\*</sup>, Juan Sainz<sup>a,b,h,l,\*\*</sup>

<sup>a</sup> Genomic Oncology Area, GENYO, Centre for Genomics and Oncological Research: Pfizer/University of Granada/Andalusian Regional Government, PTS, Granada 18016, Spain

<sup>b</sup> Instituto de Investigación Biosanitaria IBS.Granada, Granada 18012, Spain

<sup>c</sup> Medical Oncology Unit, University Hospital of Jaén, Jaén 23007, Spain

<sup>d</sup> Institut d'Investigacions Biomèdiques August Pi i Sunyer, Department of Hematology, Hospital Clínic de Barcelona, Barcelona, Spain

<sup>e</sup> Servicio de Análisis Clínicos e Inmunología, University Hospital Virgen de las Nieves, Granada 18014, Spain

<sup>f</sup> Department of Biochemistry, Molecular Biology and Immunology III, University of Granada, Granada 18016, Spain

<sup>g</sup> Unit of Infections and Cancer, Cancer Epidemiology Research Programme, IDIBELL, Catalan Institute of Oncology, L'Hospitalet de Llobregat, Spain

<sup>h</sup> Centro de Investigación Biomédica en Red de Epidemiología y Salud Pública (CIBERESP), Madrid 28029, Spain

<sup>i</sup> Oncology Department, Virgen de las Nieves University Hospital, Granada, Spain

<sup>j</sup> Campus de la Salud Hospital, PTS, Granada 18016, Spain

<sup>k</sup> Department of Hematology, Hospital Clínic, Barcelona, Spain

<sup>l</sup> Department of Biochemistry and Molecular Biology I, Faculty of Sciences, University of Granada, Granada 18012, Spain

<sup>m</sup> Department of Public Health, Mental Health and Maternal and Child Health Nursing, Faculty of nursing, University of Barcelona, Spain

**Abbreviations:** ADT, Androgen deprivation therapy; ADSL, Adenylosuccinate lyase; AKR1C1, Aldo-keto reductase family 1 member C1; AMPK, AMP-dependent protein kinase; AOPP, Advanced oxidative protein products; AORGs, Autophagy-related genes in gastric cancer; AR, Androgen receptor; ATO, Arsenic trioxide; ATG, Autophagy genes expression; BC, Breast cancer; BC HR+, Breast cancer with positive hormonal receptors; CAFs, Cancer-associated fibroblasts; Cav-1, Caveolin-1; CCS, Copper Chaperone for Superoxide Dismutase; CPE, Calotropis procera extract; CRC, Colorectal cancer; DCA, Dichloroacetate; DHM, Dihydropyridin; DPP4, Dipeptidyl peptidase-4; ENOS, Endothelial nitric oxide synthase; ER, Endoplasmic reticulum; ER+, Estrogen receptor positive; ET, Endocrine therapy; GA, Gingerol acid; Gln, Glutamine; GLS1, Glutaminase 1; GSH, Glutathione; HCC, Hepatocellular carcinoma; HMG-CoA, 3-hydroxy-3-methylglutaryl-CoA; HMGR, 3-hydroxy-3-methylglutaryl-CoA reductase; HNK, Hidroxil-Nor-Kavain; HO-1, Heme oxygenase-1 expression; ICD, Immunogenic cell death by damage-associated molecular patterns DAMPs; ICIs, Immune checkpoint inhibitors; IBD, Inflammatory bowel disease; IL-6, Interleukin-6; INOS, Inducible nitric oxide synthase; IRFA, Incomplete radiofrequency ablation; LC, Lung cancer; LRG1, Leucine-rich alpha-2-glycoprotein 1; MAOA, Monoamine oxidase A; MAPK, Mitogen-activated protein kinase; MDR, Multidrug resistance; MHC-I, Major histocompatibility complex class I; MMP-3, Metalloproteinase-3; MSI-H, Microsatellite instability-high; MtDNA, Mitochondrial DNA; NAC, N-acetylcysteine; NAFLD, Non-alcoholic fatty liver disease; NDUFS1, NADH, ubiquinone oxidoreductase subunit 1; NED, Neuroendocrine differentiation; NF-κB, Nuclear factor kappa B; NO, Nitric oxide; NOX4, NADPH oxidase 4; NSCLC, Non-small cell lung cancer; O<sub>2</sub><sup>-</sup>, Superoxide anion; OH•, Hydroxyl radical; PAHs, Polycyclic aromatic hydrocarbons; PCa, Prostate cancer; PCSC, Prostate cancer stem cells; PR+, Progesterone receptor positive; REST, Repressor element 1-silencing transcription factor; RNS, Reactive nitrogen species; ROS, Reactive oxygen species; RPRD1A, Regulation of Polymerase II Transcription DNA Repair and Damage 1 A; SCLC, Small cell lung cancer; SFA, Sophflarine A; SESN1/2, Sestrin proteins 1 and 2; SOD, Superoxide dismutase; STL, Sertraline; TFRC, Transferrin receptor 1; TIPRL, TOR Signaling Pathway Regulator-Like Protein; TNBC, Triple negative breast cancer; TNF-α, Tumor necrosis factor-alpha; TOR, Target of rapamycin; 5-FU, 5-fluorouracil; ΔΨ<sub>m</sub>, Mitochondrial membrane potential.

\* Correspondence to: Department of Biochemistry and Molecular Biology, Faculty of Sciences, University of Granada, Avenida Fuente Nueva, s/n, Granada 18071, Spain.

\*\* Correspondence to: Department of Biochemistry and Molecular Biology, Faculty of Sciences, University of Granada, Avenida Fuente Nueva, s/n, Granada 18071, Spain.

E-mail addresses: [ferjes@ugr.es](mailto:ferjes@ugr.es) (F.J. Reyes-Zurita), [jsainz@ugr.es](mailto:jsainz@ugr.es) (J. Sainz).

<sup>1</sup> These authors contributed equally to this work

<https://doi.org/10.1016/j.critrevonc.2025.104820>

Received 12 May 2025; Received in revised form 16 June 2025; Accepted 25 June 2025

Available online 26 June 2025

1040-8428/© 2025 The Author(s). Published by Elsevier B.V. This is an open access article under the CC BY-NC license (<http://creativecommons.org/licenses/by-nc/4.0/>).

## ARTICLE INFO

## Keywords:

Autophagy  
Solid tumors  
Oxidative stress  
Reactive oxygen species  
Resistance to therapy

## ABSTRACT

Cancer remains a leading cause of mortality worldwide, with solid tumors representing most cases. Autophagy and oxidative stress are two interconnected cellular mechanisms that influence tumor initiation, therapeutic response and disease progression. Autophagy plays a context-dependent role, functioning as a tumor suppressor by eliminating damaged organelles in early stages, while later supporting tumor survival under metabolic and therapeutic stress. Similarly, oxidative stress, characterized by an imbalance in reactive oxygen species (ROS), can drive tumorigenesis by promoting genomic instability and resistance to therapy but can also induce apoptosis in cancer cells. The crosstalk between autophagy and oxidative stress plays a pivotal role in shaping the tumor microenvironment, affecting immune evasion, drug resistance, and metabolic adaptation. Targeting these processes through pharmacological modulation presents both challenges and opportunities in cancer therapy. While autophagy inhibition can enhance chemotherapy efficacy by preventing tumor cell survival mechanisms, excessive oxidative stress induction may lead to cellular damage and systemic toxicity. This review explores the complex interplay between autophagy and oxidative stress in solid tumors, emphasizing their implications for cancer progression and treatment strategies. By understanding these mechanisms, novel therapeutic approaches, including combination therapies and precision medicine strategies, may be developed to improve patient outcomes.

## 1. Introduction

Nowadays, cancer is one of the leading causes of death in the world. According to GLOBOCAN 2022 estimates, approximately 20 million new cancer cases were diagnosed globally in 2022, with cancer-related deaths reaching around 9.7 million (Bray et al., 2024). These alarming statistics underscore the urgent need for continued research in oncology to gain a deeper understanding of the mechanisms driving their growth and progression (Bray et al., 2024). Solid tumors account for roughly 90 % of adult cancers and 60 % of childhood cancers (<https://seer.cancer.gov/statfacts/html/aya.html>, accessed 26/02/2025; <https://seer.cancer.gov/statfacts/html/all.html>, accessed 26/02/2025). Their high prevalence highlights the importance of gaining a deeper understanding of the mechanisms driving their growth and progression. Despite significant advancements, many aspects of tumor biology remain complex and incompletely understood. Recent studies have pointed to specific cellular processes that play pivotal roles in both the development and treatment resistance of these tumors. One such process is autophagy, which, along with oxidative stress, has emerged as a crucial factor influencing tumor behavior and response to therapies. However, many aspects of tumor biology remain complex and not fully understood while significant advancements have been made in cancer research.

Although numerous reviews have addressed the relationship between autophagy and oxidative stress in solid tumors, many focus on a specific cancer type, isolated molecular pathways, or partial aspects of this complex interaction. In contrast, this work provides an integrative and up-to-date perspective encompassing molecular mechanisms, their dual role in tumor promotion and suppression, and their impact on the tumor microenvironment across multiple cancer types, including breast, lung, prostate, colorectal cancers, and hepatocellular carcinoma. Furthermore, it thoroughly explores emerging therapeutic implications, emphasizing advances in modulators of autophagy and oxidative stress as strategies to overcome resistance to conventional treatments. This comprehensive approach facilitates the identification of unresolved questions within the literature and proposes an innovative conceptual framework that integrates molecular biology with clinical applications, thereby enhancing the rigor and translational relevance of this review.

Autophagy is a catabolic process in which the cell degrades its own components such as organelles, proteins and other molecules, particularly under conditions of nutrient deprivation or cellular stress (Hasan et al., 2022). This mechanism is crucial for maintaining the internal cellular homeostasis, as it also contributes to the elimination of the pathogens regulated by AMP-dependent protein kinase (AMPK), target of rapamycin (TOR) and growth factors such as insulin (Behrends et al., 2010). During this process, the substrates to be degraded are sequestered

into double membrane vesicles called autophagosomes, which then fuse with lysosomes producing autolysosomes. In the autolysosomes, the substrates are degraded, generating products that could be used as energy sources for multiple cellular processes (Onorati et al., 2018). Autophagy plays a dual role in solid tumors, acting as a tumor suppressor in early stages by eliminating damaged components and reducing genomic instability, while promoting tumor progression in advanced stages by providing energy and nutrients under stress conditions like hypoxia and chemotherapy. Key signaling pathways, including AMPK and mTOR, regulate this process, influencing tumor cell survival. Autophagy also impacts immune responses, either enhancing antigen presentation or aiding immune evasion. In particular, secretory autophagy has emerged as a relevant mechanism in solid tumors, influencing tumor-immune interactions and offering novel clinical insights (Li and Zhao, 2025). Given its context-dependent effects, targeting autophagy selectively could improve cancer therapies (Jalali et al., 2025).

Oxidative stress, defined as an imbalance between the production of reactive oxygen species (ROS) and the efficacy of antioxidant defenses, is another hallmark of cancer (Đuračková, 2010). During the endogenous metabolic reaction, aerobic cells produce ROS such as superoxide anion ( $O_2^-$ ), hydrogen peroxide ( $H_2O_2$ ), hydroxyl radical ( $OH\bullet$ ), and organic peroxides. The transfer of electrons to oxygen occurs in the mitochondrial membrane during the respiratory chain. Under hypoxic conditions, the respiratory chain could generate nitric oxide (NO) (Li et al., 2016). Oxidative stress plays a key role in cancer by promoting DNA damage, tumor growth, angiogenesis, and metastasis. It activates signaling pathways like PI3K/AKT and NF- $\kappa$ B, enhancing cell survival and therapy resistance (Iqbal, 2024). Additionally, it alters the tumor microenvironment, weakening immune responses (Iqbal, 2024). Targeting oxidative stress is a potential therapeutic strategy, either by increasing ROS to induce cancer cell death or reducing it to prevent tumor progression (Hayes et al., 2020; Reuter et al., 2010; Zhao et al., 2023).

Importantly, autophagy and oxidative stress are not independent phenomena but are intricately linked through a bidirectional regulatory relationship. Various stressors, including hypoxia or starvation, activate autophagy pathways that can raise ROS generation including superoxide and hydrogen peroxide (Redza-Dutordoir and Averill-Bates, 2021). This interplay has been extensively explored in hematological malignancies, offering mechanistic insights that may also be relevant to solid tumors (Cabrera-Serrano et al., 2025). Tumor initiation, growth, and possible therapeutic treatments all depend on the interaction between these processes, which each have a vital part to play in the development and progression of cancer (Chen et al., 2008). This review explores the intricate relationship between these processes, examining their dual

roles in tumorigenesis and their implications for therapeutic intervention. Notably, ongoing clinical trials targeting autophagy and ROS pathways highlight promising future strategies for cancer treatment (Pandey et al., 2024). In cancer biology, autophagy and oxidative stress are deeply interconnected, with their roles oscillating between protective and pathological depending on the tumor microenvironment.

## 2. Crosstalk between autophagy and oxidative stress in solid tumors

Autophagy and oxidative stress are two fundamental cellular processes that play a pivotal role in the development and progression of solid tumors. Autophagy, a tightly regulated lysosomal degradation pathway, is essential for maintaining cellular homeostasis by recycling damaged organelles and misfolded proteins (Hasan et al., 2022; Li et al., 2023; Taucher et al., 2022). This process maintains metabolic balance and prevents the accumulation of cytotoxic elements in cells. Conversely, oxidative stress results from an imbalance between the production of (ROS) and the efficacy of cellular antioxidant defenses, often culminating in DNA damage, genomic instability and cellular dysfunction (El Hout et al., 2020; Sharifi-Rad et al., 2020). The interaction between autophagy and oxidative stress can either suppress or promote tumor growth, depending on the context and stage of cancer development (Dong et al., 2023; Hasan et al., 2022; Taucher et al., 2022). Elevated ROS levels also activate various oncogenic pathways, such as the mitogen-activated protein kinase (MAPK) signaling pathway, and nuclear factor kappa B (NF- $\kappa$ B), promoting cancer development (Hasan et al., 2022). However, excessive ROS may induce cell death. This complex interaction contributes to the preservation of cellular stability and prevents the accumulation of alterations that could produce tumor development (Hasan et al., 2022).

### 2.1. Regulation of autophagy by oxidative stress

It is widely known that autophagy can be regulated by ROS through mTOR dependent pathways. When ROS levels are high it could produce the inactivation of PTEN by direct oxidation (Leslie, 2006), which increases the PIP3 levels, activating AKT and inhibiting mTORC1 signaling (Kma and Baruah, 2022). Other studies have shown that MAPK molecules such as JNK, p38 and ERK can also be regulated by ROS. As a result, these molecules play an important role in the activation of mTOR (Zhang et al., 2020; Zhou et al., 2015). MAPK/JNK is involved in the autophagy process through the direct phosphorylation of ULK1 or by promoting the autophagy key genes transcription like *BECN1* (He et al., 2018). In conclusion the accumulation of ROS could affect the autophagy pathway through MAPK/JNK/mTOR. In addition, ROS levels can also activate AMPK, which in turn induces autophagy. This process occurs when AMPK phosphorylates and activates ULK1 or inhibits mTORC1. It has been demonstrated that ROS can inhibit mTOR or activate the AMPK pathways, promoting autophagy in different tumor cell types, which directly impacts tumor progression (Zhao et al., 2017).

Additionally, transcription factors such as TFEB, HIF-1 $\alpha$ , p53, NF- $\kappa$ B, FOXO3, ATF4 and NRF2, which are essentials for the regulation of the autophagy genes expression (ATG), are activated by higher ROS levels (Dong et al., 2023). Furthermore, Sarkar et al., 2011 investigated the role of nitric oxide (NO) in S-nitrosyl IKK $\beta$  and reducing its phosphorylation, preventing the phosphorylation of AMPK which affects the initiation of autophagy (Sarkar et al., 2011). Whereas reactive nitrogen species (RNS) have traditionally been considered to inhibit autophagy, some studies suggest that NO could induce this process. In breast cancer cells for example, nitro oxidative stress triggered autophagy rapid activation in response to DNA damage mediated by ATM, activating LKB1. As a result, AMPK/TSC1/2/mTOR pathway was inhibited, promoting autophagy induction (Tripathi et al., 2013).

### 2.2. Autophagy and oxidative stress as tumor suppressor

Autophagy as a tumor suppressor in early cancer development plays a protective role by mitigating oxidative damage and maintaining cellular integrity. Removal of damaged mitochondria by mitophagy reduces ROS generation, thereby limiting oxidative damage to DNA and proteins (Choi et al., 2012; Li et al., 2020; Yan and Li, 2018). Loss or downregulation of core autophagy genes, such as *BECN1* and *LC3*, has been linked to increased ROS accumulation and heightened tumor initiation risk (Nurdinov, 2020; Poillet-Perez et al., 2015). This suggests that autophagy may act as a barrier to early tumorigenesis by preventing ROS-induced mutations (Fig. 1).

Mitophagy is regulated by two principal pathways, NIX/BNIP3L and PARKIN (PARK2/PINK1). The NIX/BNIP3L pathway interacts with proteins such as GABARAP and GABARAPL1 to label defective mitochondria and directs it to its degradation in the autophagosome (Poillet-Perez et al., 2015). Furthermore, the PARKIN/PINK1 pathway facilitates the removal of dysfunctional mitochondria in response to membrane depolarization, induced by the increase of ROS levels (Youle and Narendra, 2011). It has been demonstrated that the elimination of damaged mitochondria through autophagy reduces ROS production, thereby limiting its impact on tumor progression (Poillet-Perez et al., 2015).

ATG proteins modulate inflammatory and immune signaling through both autophagy dependent and independent mechanisms. In the former, mitophagy reduces mitochondrial ROS, indirectly modulating immune sensors such as RIG-I and the NLRP3 inflammasome. In the latter, ATG proteins interact directly with immune signaling pathways, independent of their role in autophagosome formation (Levine and Kroemer, 2019). Additionally, redox homeostasis is supported by enzymatic (e.g., superoxide dismutase (SOD), catalase, peroxidases) and non-enzymatic (e.g., glutathione (GSH), thioredoxin) antioxidant systems, which detoxify species like superoxide (O $_2^-$ ) and hydrogen peroxide (H $_2$ O $_2$ ) (Dorval and Hontela, 2003; Kern and Kehrer, 2005).

Furthermore, the p53 protein plays a role in the modulation of autophagy through the regulation of ROS levels (Maiuri et al., 2010; Rahman et al., 2022; Tang et al., 2015). Under oxidative stress conditions, basal p53 levels activate the expression of various antioxidants, such as, GPX1, MnSOD, ALDH4 and TPP53INP1, to mitigate oxidative damage (Budanov et al., 2010; Hu et al., 2010; Pani and Galeotti, 2011; Suzuki et al., 2010). This antioxidant function contributes to mTORC1 inhibition and autophagy induction (Budanov and Karin, 2008). Sestrin proteins (SESN1/2), transcriptional targets of p53, mediate this effect by activating AMPK and the TSC1/TSC2 complex, reinforcing autophagic flux and ROS detoxification (Budanov, 2011). In contrast, p53 can increase ROS levels. Duan et al. (2011), have shown that the silibinin can induce the autophagy cell death mediated by p53 through the activation of ROS-p38 and JNK pathways, as well as the inhibition of MEK/ERK and PI3K/AKT pathways (Duan et al., 2011). Thus, p53 plays a dual role in cancer by either preserving cellular homeostasis or initiating programmed cell death depending on the oxidative context.

### 2.3. Autophagy and oxidative stress as tumor promoter

Autophagy often switches to a pro-survival mechanism in established tumors. Cancer cells use autophagy to adapt to stressful microenvironmental conditions such as hypoxia, nutrient deprivation, and elevated ROS levels (Choi, 2012; Debnath et al., 2023; Yang et al., 2011). For example, hypoxia-induced autophagy enables tumor cells to withstand oxidative stress by degrading damaged components and providing metabolic substrates for survival. This paradoxical role highlights the adaptability of autophagy in cancer biology, where its modulation is context-dependent (Hasan et al., 2022; Hu et al., 2023).

One key molecule as a tumor promoter is p-62 (also known as SQSTM1) (Katsuragi et al., 2015), which plays a key role in protein accumulation and the formation of inclusion bodies, which are

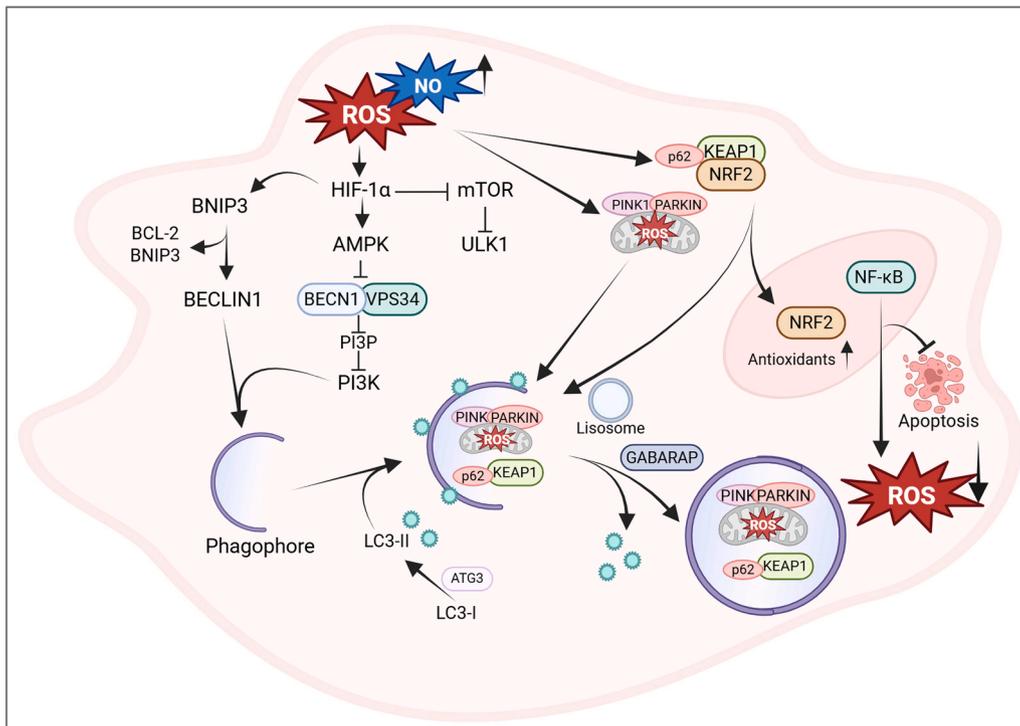


Fig. 1. Autophagy prevents tumorigenesis via BECN1 and LG3.

associated with diverse diseases (Stumptner et al., 1999; Zatloukal et al., 2002). For instance, in models lacking *ATG5* or *ATG7*, p62 aggregation promotes ROS accumulation and tumorigenesis (Takamura et al., 2011). This accumulation leads to increased ROS levels and promotes tumorigenesis, especially in cells with a defective autophagy. Specifically, p62 activates non canonical antioxidant pathway Nrf2, by competing with Keap1 and producing the translocation of Nrf2 to the nucleus, where it activates antioxidant genes (Jiang et al., 2015). However, excessive p62 accumulation also increases ROS levels, favoring tumor development. In addition, p62 can activate the NF- $\kappa$ B pathway, contributing to ROS production. Even so, when autophagy is impaired, p62 struggles to effectively activate the NF- $\kappa$ B pathway. While this might seem beneficial by reducing inflammation, the buildup of p62 also leads to increased oxidative stress, which can fuel tumor growth (Mathew et al., 2009).

Autophagy loss also disrupts mitochondrial homeostasis. Wei et al. (2011) have shown that in murine models with a mutation in FIP200, present abnormal mitochondria and an accumulation of healthy and dysfunctional mitochondria, in comparison with normal mice (Wei et al., 2011). The elimination of FIP200 in hematopoietic stem cells and neurons causes an accumulation of mitochondrial and ROS levels (Liu et al., 2010). Additionally, the accumulation of mitochondria with abnormal morphology has been identified in *ATG7*<sup>-/-</sup> murine livers, in  $\beta$  pancreatic cells, in *ATG5*<sup>-/-</sup> murine cardiac cells and Paneth cells with a mutation in *ATG16L1* (reviewed in (Chavez-Dominguez et al., 2020)). This investigation shows that the dysfunction of autophagy produces an increase of intracellular ROS levels that could be associated with tumor development. Another study describes the association between autophagy and endoplasmic reticulum (ER) stress, highlighting their role in tumor development and treatment response, specifically in melanoma. In melanomas with *BRAF*<sup>V600E</sup> mutation, the oncogenic alteration induces chronic ER stress, increasing basal autophagy levels (Corazzari et al., 2015). *BRAF*<sup>V600E</sup> activates p38 which in turn stimulates the pathways IRE1/ASK1/JNK and TRB3 pathways. The phosphorylation of Bcl-XL/Bcl-2 by JNK releases Beclin1, a key regulator of autophagy, while TRB3 inhibits Akt/mTOR, increasing basal autophagy (Corazzari et al., 2015). Furthermore, *ATG7* is essential for maintaining metabolic fitness in *BRAF*<sup>V600E+</sup> tumors and suggests that targeting autophagy may

have therapeutic potential in this context (Strohecker et al., 2013). Autophagy also regulates ferroptosis—a form of iron-dependent oxidative cell death, by degrading protective factors such as lipid droplets, GPX4, ferritin, SLC40A1, CDH2 and BMAL1 (An et al., 2024). Reticulophagy, a subtype of autophagy directed toward the ER, similarly enhances ferroptotic susceptibility (Liu et al., 2022).

Liu (2023) have investigated the role of genes related to oxidative stress and autophagy in gastric cancer (OARGs) (Liu, 2023). The study analyzed 17 OARGs with prognostic value, identifying three different patterns, each with unique biological and immunological characteristics. Finally, they found a significant negative correlation between the OARG score (a method to evaluate the patterns in specific tumors) and the immune cells infiltration. These results provide a new point of view to predict the prognostic and immunotherapy response in gastric patients, moreover, to highlight oxidative stress and autophagy in its progression (Liu, 2023).

#### 2.4. Autophagy and oxidative stress in tumor microenvironment

In the tumor microenvironment oxidative stress can mimic the effects of hypoxia, even in the presence of oxygen, leading to elevated ROS levels that damage DNA and drive genomic instability. This “bystander effect” also promotes aneuploidy in adjacent tumor cells ROS in the stroma trigger autophagy and mitophagy, generating recycled nutrients such as lactate and ketones that support tumor growth (Lisanti et al., 2010). Martinez-Outschoorn (2010) demonstrated that, in breast cancer, tumor cells can induce oxidative stress in neighboring stromal fibroblasts, triggering the autophagic degradation of caveolin-1 (Cav-1). This process, regulated by HIF-1 $\alpha$  and NF- $\kappa$ B, promotes nutrient recycling in the microenvironment and confers resistance to apoptosis in adjacent cancer cells, thereby supporting tumor progression (Martinez-Outschoorn, 2010).

### 3. The role of autophagy and oxidative stress in selected solid tumors

#### 3.1. Breast cancer

Breast cancer (BC) is the most common cancer among women globally, with incidence rates highest in North America, Northern Europe, and Oceania (Kelsey and Horn-Ross, 1993; Shimizu et al., 2012). Risk factors include demographic factors (age, race, socioeconomic status, and geographic location) and reproductive factors (early menarche, late menopause, nulliparity), as well as hormonal factors, genetics, and lifestyle. While BC is a multifactorial and complex disease, studies suggest that interactions between autophagy and oxidative stress may contribute to its initiation (Hasan et al., 2022; Poillet-Perez et al., 2015). In addition, the relationship between autophagy and ROS in BC appears to be context-dependent, with evidence indicating that these processes can also be involved in tumor suppression and disease progression (Table 1) (Hasan et al., 2022; Poillet-Perez et al., 2015).

In early-stage BC, autophagy functions as a tumor suppressor by removing ROS-damaged organelles and proteins, thereby maintaining genomic stability. Key mechanisms include Beclin-1 regulation, where monoallelic deletion (observed in ~50 % of BC) impairs autophagy, leading to ROS accumulation and mitochondrial dysfunction (Aita et al., 1999; Liang et al., 2006); mitophagy, whose defects (e.g., ATG5 or ATG7 deficiency) result in damaged mitochondrial accumulation, exacerbating ROS levels and promoting mutagenesis (Hasan et al., 2022; Kongara and Karantza, 2012). Prolonged NRF2 activation can lead to reductive stress and proteotoxicity, highlighting the importance of maintaining redox balance (Dodson et al., 2015). In fact, in established tumors, autophagy shifts towards a pro-survival role, enabling metabolic adaptation by maintaining mitochondrial health in response to increased ROS and metabolic demands driven by oncogenic signaling (e.g., mutant Ras) (Lock et al., 2014). Autophagy also facilitates metastasis, as detached mammary tumor cells activate ROS-dependent PERK signaling to induce autophagy, enhancing survival during matrix detachment and lung colonization (Avivar-Valderas et al., 2011). ROS-activated pathways like TGF- $\beta$ 1 and NF- $\kappa$ B promote EMT, while autophagy meets the energy demands of invasion. Under hypoxia, ROS triggers autophagy, creating a feedback loop that sustains tumor growth (Galluzzi et al., 2015). For example, ROS induce the oxidation of ATG4 and ATM, which triggers (Kim et al., 2011; Li et al., 2010) autophagosome formation. In addition, ROS activate AMPK, which inhibits mTORC1 in a H<sub>2</sub>O<sub>2</sub>-dependent manner, subsequently leading to ULK1 and Raptor phosphorylation and further promoting autophagy. On the other hand, HIF-1 $\alpha$ , which is stabilized under oxidative stress, has been shown to activate autophagy pathways, including the AMPK/mTOR signaling axis, highlighting the crucial role of ROS in modulating autophagic activity (Daskalaki et al., 2018; J. Li et al., 2022; Zhou et al., 2018). Therefore, understanding these mechanisms is vital for developing therapeutic strategies targeting the Keap1-Nrf2 and AMPK/mTOR pathways, but also TGF- $\beta$ 1- and NF- $\kappa$ B-induced pathways in various malignant diseases, including BC (Shan et al., 2024) (Fig. 2).

In advanced BC, autophagy shifts from a suppressive to a tumor supportive role, allowing cancer cells to adapt to harsh conditions such as hypoxia, nutrient deprivation, and oxidative stress (Fig. 1). Upregulation of autophagy-related proteins, including LC3B and ATG5, has been observed in BC cells resistant to therapies such as tamoxifen and taxanes, highlighting the role of autophagy in therapy evasion (Aqbi et al., 2018; Dwyer et al., 2024; Han et al., 2017). These findings emphasize the need for precision therapies that consider autophagy and oxidative stress, targeting these pathways in a context-dependent manner. This approach has emerged as a promising therapeutic strategy, with over 30 clinical trials investigating autophagy inhibition in combination with conventional cancer treatments.

In the metastatic scenario the interplay between autophagy and oxidative stress is also important. Autophagy supports the survival of

**Table 1**  
Therapeutic strategies in BC.

Therapeutic strategies	Molecular mechanism	References
<b>Preclinical study:</b> Simultaneous inhibition of the estrogen receptor, CDK4/6 and PI3K	Favors cell cycle arrest, apoptosis induction and tumoral regression <i>in vitro</i> and <i>in vivo</i>	(Jhaveri et al., 2024)
<b>Phase I/II study:</b> Inavolisib (PI3K inhibitor) + palbociclib (inhibitor of CDK4/6) + letrozole or fulvestrant (endocrine therapy)	Reduction of PI3K/AKT biomarker expression in tumoral tissue	(Jhaveri et al., 2024)
<b>Phase III study:</b> Inavolisib + palbociclib + fulvestrant in HR+, HER2- metastatic BC patients with PIK3CA mutation	Significant improvement in progression-free survival	(Juric et al., 2024)
<b>Preclinical study:</b> Autophagy inhibitors could enhance the effectiveness of lapatinib (HER2-targeted drugs)	Targeted treatments to HER2 can induce autophagy as a resistance mechanism	(M. Wang et al., 2024)
<b>Preclinical study:</b> IC87114 drug (inhibitor of the p110 $\delta$ PI3K isoform) in MCF-7 (BR HR+ cell line) and MDA-MB-231 (TNBC cell line)	Decreases cell migration in spheroids.	(Di Donato et al., 2022)
Vps34-IN1 (inhibitor of Vps34, class III PI3K isoform)	Mediates autophagy phenomena and increases sunitinib and erlotinib drugs' efficacy	(Di Donato et al., 2022)
MitoQ, a derivative of Coenzyme Q10 that specifically targets mitochondria	Generates the degradation of Keap1 through autophagy, increasing the transcriptional activity of Nrf2 leading to the expression of antioxidant genes such as p62. The p62 protein is associated with Keap1 and activates Nrf2, producing a positive feedback loop that protects cells from oxidative damage	(Gonzalez et al., 2014)
<b>REDD1 Modulation</b>	REDD1 inhibits mTORC1, reducing cancer cell proliferation and linking to chemotherapy-induced reduced BC cell survival.	(Lan et al., 2013; Tirado-Hurtado et al., 2018; Zhidkova et al., 2022, 2020)
<b>Pro-oxidants (Elesclomol, ATO)</b>	Pro-oxidants such as elesclomol and arsenic trioxide (ATO) exploit BC cells' redox vulnerabilities by increasing ROS, overwhelming antioxidant defenses, and inducing apoptosis. Their efficacy is enhanced when combined with autophagy inhibitors. Elesclomol induces oxidative stress by disrupting mitochondrial	(Alli and Ford, 2012; Alzuwaidi and Khalil, 2019; Buccarelli et al., 2021; Chow et al., 2004; Kirshner et al., 2023; Li et al., 2023; Moomivand et al., 2024; Nagai et al., 2012; Nasrollahzadeh, 2020; Nooshinfar et al., 2016; Qu et al., 2010; Rajaram et al., 2024; Wang et al., 2023; Wang et al., 2011; Xia et al., 2014; Zhang et al., 2016)

(continued on next page)

Table 1 (continued)

Therapeutic strategies	Molecular mechanism	References
Mitophagy Targeting (PINK1, Parkin)	metabolism and copper homeostasis, leading to cuproptosis. ATO inhibits BC growth, promoting apoptosis and activating key apoptotic proteins. Targeting mitophagy-specific proteins, such as PINK1 and Parkin, addresses mitochondrial dysfunction in cancer, offering new therapeutic opportunities	(Dong and Zhang, 2024; Li et al., 2022; Narendra and Youle, 2024; Wang et al., 2023)

disseminated tumor cells in circulation and at secondary sites by reducing oxidative stress, allowing them to establish metastatic colonies. Targeting these processes therapeutically could reduce the metastatic potential of BC and improve long-term survival outcomes (Grandvallet et al., 2020; Wu and Sharma, 2023).

### 3.2. Lung cancer

Lung cancer (LC) is the leading cause of cancer-related mortality worldwide, with non-small cell lung cancer (NSCLC) and small cell lung cancer (SCLC) accounting for approximately 85 % and 15 % of cases, respectively (Lemjabbar-Alaoui et al., 2015; Molina et al., 2008). LC is

significantly influenced by various risk factors, with cigarette smoking being the predominant cause (Chen, 2023; Schwartz and Cote, 2016), although other contributors include aging, air pollution, radon, occupational exposures, and chronic lung diseases (de Groot et al., 2018). Tobacco smoke contains harmful chemicals, including polycyclic aromatic hydrocarbons (PAHs), nitrosamines, and heavy metals like cadmium (Hecht, 2011; Lemjabbar-Alaoui et al., 2015; Liu et al., 2009). These toxins generate excessive ROS, overwhelming the antioxidant defenses and leading to DNA damage, triggering mutations in key tumor suppressor genes like p53 and activating oncogenes like KRAS (Drosten and Barbacid, 2022; Luo et al., 2022; Suzawa et al., 2019). Moreover, tobacco smoke accelerates epigenetic changes, including DNA methylation and histone modification, which contribute to lung carcinogenesis (Seiler et al., 2020; Xie et al., 2021; Zong et al., 2019). On the other hand, air pollution, particularly fine particulate matter (PM2.5 and PM10), also plays a significant role in LC. These fine particles, often carrying toxic metals and chemicals, reach deep into the lungs, where they fuel chronic inflammation and oxidative stress. Studies have shown that prolonged exposure to PM2.5 increases LC risk by activating inflammatory pathways such as NF-κB and STAT3, which encourage tumor growth by stimulating the production of inflammatory molecules and new blood vessels (Deng et al., 2020; Grivennikov and Karin, 2010; Lim and Yoon, 2022; Liu et al., 2023; Zhao et al., 2021).

Chronic inflammation, triggered by infection or long-term exposure to pollutants, contributes to LC development (Budisan et al., 2021), creating a microenvironment rich in ROS and RNS, which further damage DNA and proteins. Pro-inflammatory cytokines such as tumor

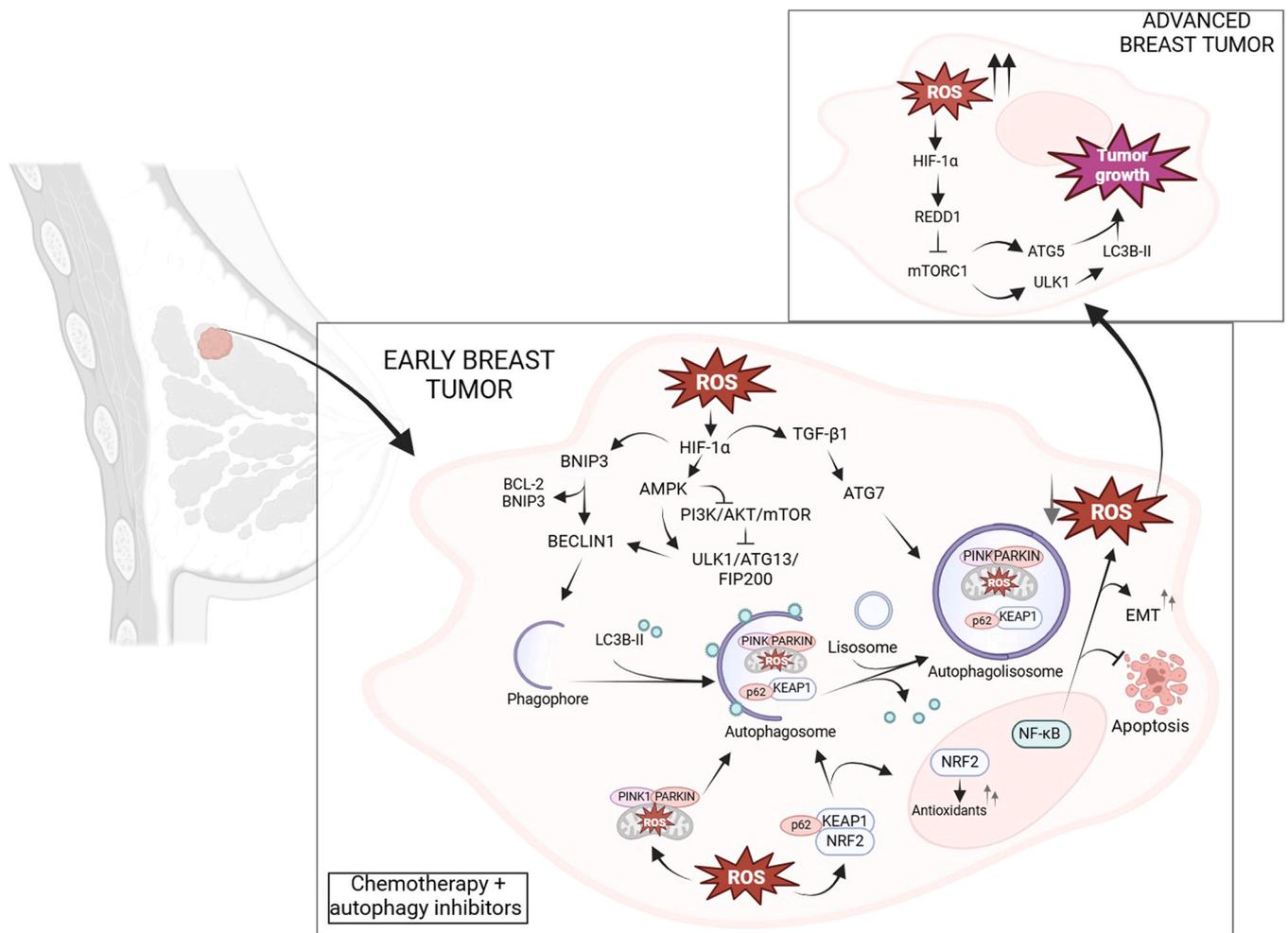


Fig. 2. Molecular pathways involved in determining early and advanced BC.

necrosis factor-alpha (TNF-α) and interleukin-6 (IL-6) are upregulated in such environments and promote cancer cell proliferation and survival through the PI3K/AKT/mTOR and JAK/STAT signaling pathways (Deng et al., 2020; Rasková, 2022; Shang et al., 2017; Wu and Zhou, 2010).

Exposure to carcinogens also induces changes in the tumor micro-environment characterized by immune evasion and angiogenesis. Air pollutants such as benzene and formaldehyde contribute to these changes by exacerbating oxidative stress and impairing immune surveillance (Shankar et al., 2019; Xiong et al., 2024).

In the past few decades, our understanding of the biology and progression mechanisms of the disease has significantly improved. It has become clear that autophagy and ROS can regulate each other to influence the initiation and progression of LC. These processes initially promote cancer through oxidative stress and DNA mutations, while at later stages, they can induce apoptosis (ArulJothi et al., 2022). Autophagy is differentially regulated in NSCLC and SCLC. While NSCLC often shows upregulation of autophagy-related genes, SCLC exhibits a mixed autophagy profile, with both up- and down-regulated genes compared to normal tissue, possibly reflecting different tumor microenvironments and metabolic needs (Shen et al., 2022; Wang et al., 2023; Zinn et al., 2013). These findings highlight the need for subtype-specific approaches when targeting autophagy in LC therapies (Filomeni et al., 2015).

In NSCLC, the AKR1C1 (Aldo-keto reductase family 1 member C1) is overexpressed and is associated with a poor prognosis (Zhu et al., 2018). AKR1C1, directly interacts with SQSTM1, producing its oligomerization and promoting autophagy, contributing to cancer cells' drug resistance. Under oxidative stress conditions, the AKR1C1-SQSTM1 complex stabilizes a signaling loop that involves AKR1C1, SQSTM1, and Nrf2, increasing antioxidant defense mechanisms of tumor cells (Chang et al., 2022). Autophagy also maintains mitochondrial homeostasis during

EMT, especially under TGF-β1 stimulation, supporting ATP production and energy balance (Hwang et al., 2022), while its inhibition reduces mesenchymal marker expression at the protein level, without affecting mRNA levels, suggesting a post-transcriptional control (Alizadeh et al., 2018). Furthermore, its inhibition decreases ATP production, affecting mitochondrial function, which activates AMPK and alters protein translation (Hwang et al., 2022). The PI3K/AKT/mTOR axis, a central regulator of autophagy and cell growth, has been identified as a key therapeutic target in NSCLC (Tan, 2020; Yip, 2015). Dual inhibitors of PI3K and mTOR not only suppress autophagy but also disrupt oncogenic signaling pathways, thereby enhancing the efficacy of chemotherapy and immunotherapy (Huang et al., 2020; Shimizu et al., 2023). Targeting upstream regulators such as AMPK could also selectively impair autophagy in cancer cells without compromising its homeostatic functions in normal tissues (Wang et al., 2020; Yuan et al., 2023) (Fig. 3).

In SCLC, the activation of the PI3K/AKT/mTOR signaling pathway is associated with radiotherapy resistance and poor prognosis in particular in those patients carrying RAS mutations (Chen et al., 2020). Inhibiting this pathway accelerates G6PD degradation via HSC70-mediated autophagy, disrupting NADPH production and glutathione regeneration, thus enhancing oxidative damage and apoptosis, particularly when combined with radiation (Deng et al., 2023). This reduction of G6PD levels typically helps tumor cells survive oxidative stress damage by generating NADPH and reduced GSH. The inhibition of G6PD reduces cancer cell viability, migration and colony formation and increases oxidative stress damage and induces apoptosis when combined with radiotherapy (Deng et al., 2023). In addition, in cisplatin-resistant LC cells, G6PD suppression reverses drug resistance by altering redox homeostasis (Hong et al., 2018) and triggers immunogenic cell death, enhancing the efficacy of immunotherapies in LC models (Nakamura

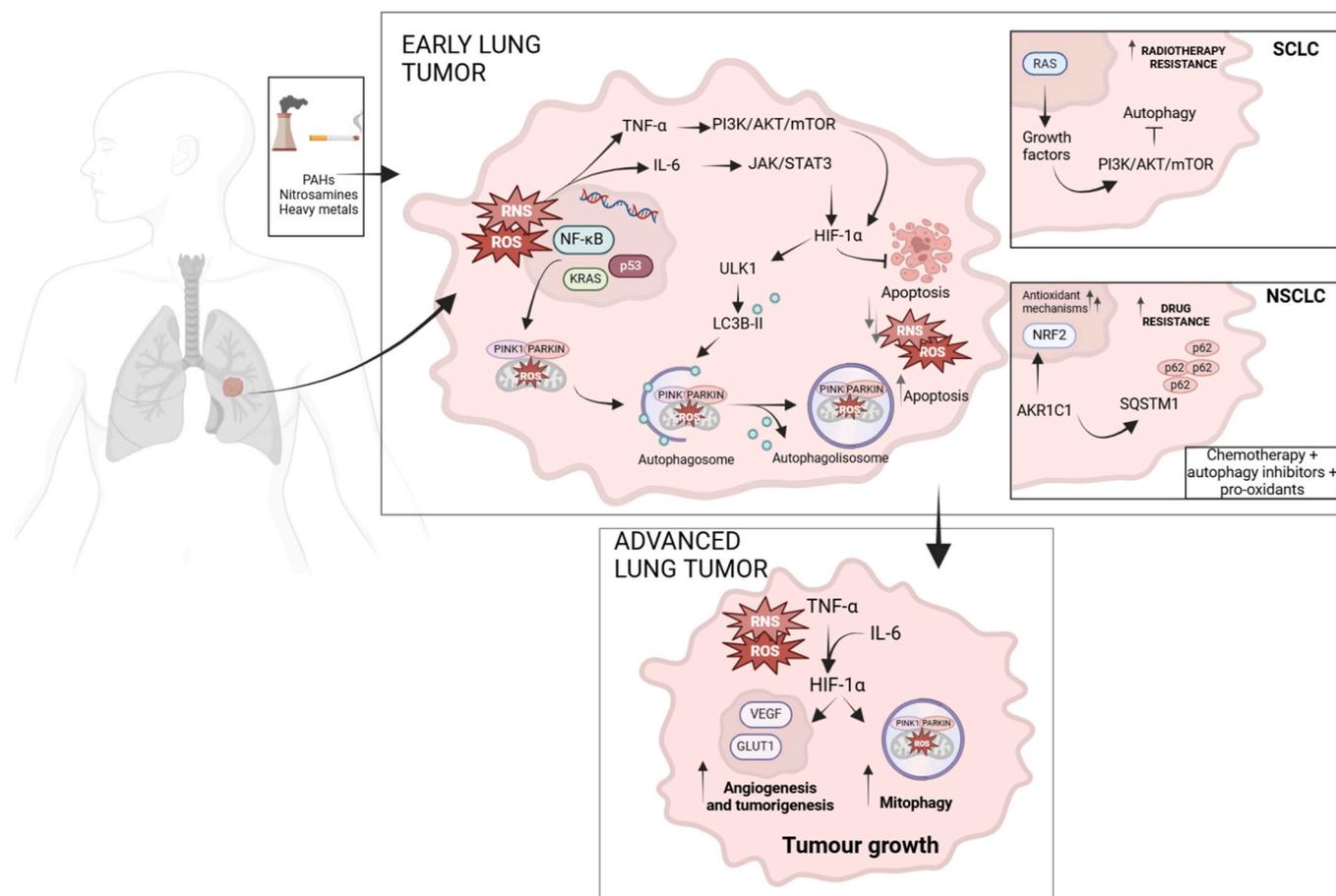


Fig. 3. Molecular pathways involved in determining early and advanced LC.

et al., 2024). Hu et al. (2023) also demonstrated that the activation of Copper Chaperone for Superoxide Dismutase (CCS) through H3K27 acetylation reduces oxidative stress in LC cells, promoting autophagy and inhibiting cellular apoptosis (Hu et al., 2023). Inhibition of CCS increases ROS levels, thereby enhancing oxidative stress and reducing cell survival. Furthermore, CCS inhibition suppresses autophagy and activates apoptosis by increasing the expression of the pro-apoptotic gene BAX and decreasing the expression of the anti-apoptotic gene Bcl-2. CCS inhibition also plays a role in tumoral microenvironment and immune response in LC (Zhou et al., 2024) (Fig. 3).

Tumor cells under oxidative stress can suffer mitochondrial dysfunction, which is associated with a reduction in the number of mitochondrial DNA (mtDNA) copies (Scheid et al., 2021). This stress is also induced by the transfer of mtDNA from stromal cells to tumor cells, resulting in the restoration of mitochondrial function and the promotion of tumor progression (Tan et al., 2015). ROS induce mitophagy in cancer-associated fibroblasts (CAFs), leading to the release of mtDNA and facilitating communication between CAFs and tumor cells through autophagy and oxidative stress. Additionally, tumor-derived exosomes play a crucial role in CAF activation and tumor progression by transferring molecules like miRNAs, which reprogram CAFs to a cancer-promoting phenotype (Zhou et al., 2024).

The role of autophagy in drug resistance is particularly well documented. Increased autophagic activity in LC cells has been shown to mitigate the cytotoxic effects of chemotherapeutic agents such as cisplatin and paclitaxel (Chen et al., 2018; Ornatowski et al., 2020). Similarly, during radiotherapy, which generates lethal levels of ROS, cancer cells upregulate autophagy-related genes such as LC3B and ATG7 to limit oxidative damage and maintain mitochondrial integrity (Chen et al., 2018; Shadab et al., 2020; Zinn et al., 2013). Autophagy inhibitors such as CQ and HCQ have also shown efficacy in preclinical models, particularly when combined with chemotherapy or immunotherapy (Zinn et al., 2013). These agents increase oxidative stress in cancer cells by preventing the removal of damaged mitochondria, thereby pushing them towards apoptosis (Chen et al., 2018; Takano et al., 2023). In addition to HCQ, novel inhibitors SBI0206965, targeting specific autophagy-related proteins such as ULK1, are being developed to improve therapeutic precision and minimize off-target effects (Ghazi et al., 2024; Tang et al., 2017) (Fig. 3).

### 3.3. Prostate cancer (PCa)

Prostate cancer (PCa) is the second most frequently diagnosed malignancy and a leading cause of cancer-related mortality in men worldwide (Bergengren et al., 2023). Its global incidence has risen over the past two decades, driven by population aging, advancements in diagnostic techniques, and increased screening efforts (Rawla, 2019). Although mortality rates have declined, PCa remains the sixth leading cause of cancer-related death globally. Established risk factors include age, ethnicity, and family history, while environmental influences also contribute significantly (Bergengren et al., 2023). Additional potential risk factors include smoking, diet, physical activity, certain medications, and occupational exposures (Bergengren et al., 2023). The widespread adoption of prostate-specific antigen (PSA) screening, along with advanced diagnostic tools such as MRI and novel biomarkers, has shaped PCa epidemiology and detection strategies (Pernar, 2018). PCa is a highly heterogeneous disease, ranging from indolent to aggressive forms (Adamaki and Zoumpourlis, 2021). Autophagy and ROS play a pivotal role in PCa initiation and progression (Wang et al., 2016), with their interplay influencing tumor behavior in a context-dependent manner (Kongara and Karantza, 2012).

On the pro-tumorigenic side, androgen receptor (AR) signaling induces autophagy, promoting PCa cell proliferation and survival (Ashrafizadeh et al., 2022). Autophagy can act as a pro-survival mechanism, especially in response to chemotherapy, with autophagy induction by NPRL2 leading to resistance against drugs like everolimus and

docetaxel (Luo et al., 2020). Similarly, recent evidence shows that modulation of autophagy and oxidative stress also mediates resistance to bortezomib in prostate cancer, further highlighting the role of these processes in therapeutic response (Zafeiropoulou et al., 2024). NPRL2 modulates mTOR signaling and the response to stress conditions. Its overexpression inhibits mTOR, favoring autophagy and promoting the survival of tumor cells in hostile environments. Cells with high NPRL2 expression are less sensitive to this treatment, while NPRL2 inhibition enhances the therapeutic response by promoting apoptosis. This suggests that NPRL2 may be modulating lysosomal acidification, a key process in the late phase of autophagy (Ashrafizadeh et al., 2022; Chen et al., 2019). Mesenchymal stem cells secrete TGF- $\beta$ 1 which regulate Nox4 and promotes PCa progression by generating ROS and activating fibroblasts which produces tumor microenvironment remodeling and facilitates cancer cell migration and invasion (Sampson et al., 2018). Furthermore, Nox4 influences autophagy, promoting cell survival under adverse conditions and modulating communication between stromal and epithelial cells. Its interaction with TGF $\beta$ 1 creates a feedback loop that reinforces stromal activation and contributes to tumor progression, consolidating its position as a central regulator of oxidative stress and tumor dynamics in PCa (Sampson et al., 2018). Oxidative stress regulates gene expression in CAFs. Hydrogen peroxide promotes changes in the expression of growth factors, adhesion molecules and metalloproteinases, facilitating the conversion of fibroblasts to active myofibroblasts, enhancing invasive cancer growth and metastasis (Cat et al., 2006). CAF becomes resistant to chemotherapy and radiotherapy, contributing to tumor relapses. In addition, ROS increases Matrix Metalloproteinase-3 (MMP-3) expression in PCa cells by miR-128, suggesting an interaction between oxidative stress and metalloproteinases regulation (Hsieh et al., 2017) (Fig. 4).

On the anti-tumorigenic side, ROS-induced oxidative stress can trigger autophagic cell death in PCa cells, with agents like inactivated Sendai virus (HVJ-E) has been shown to exerting anticancer effects via ROS-dependent autophagic cell death (Qian et al., 2018). For instance, HVJ-E induces autophagy in PCa cells through the activation of the PI3K/Beclin-1 pathway, favoring cell death. Modulators such as rapamycin increase apoptosis mediated by the virus, while CQ does not alter the anticancer effects. Moreover, this virus generates ROS in tumor cells, activating MAPK (JNK and p38) pathways, which are associated with cytotoxicity and apoptosis (Gao et al., 2014; Qian et al., 2018). Additionally, downregulation of DNML1/DRP1 induces autophagy, leading to inhibited proliferation. DRP1 is positively regulated by AR signaling and modulates the VDAC-MPC complex, which regulates mitochondrial pyruvate import, promoting the growth and survival of PCa cells. Moreover, microenvironmental stress, such as hypoxia and oxidative stress, activates adaptive stress responses, including autophagy, which are closely linked to mitochondrial functions. Under low oxygen conditions, mitochondrial fragmentation mediated by DRP1 is associated with higher ROS levels, enhancing cell survival. The inhibition of DRP1, combined with autophagy inhibition, increases cell death in PCa (Lee et al., 2020). At the molecular level, AR signaling regulates autophagy-related genes such as *TFEB*, *ATG4B*, *ATG4D*, *ULK1*, and *ULK2*, while *GABARAPL1*, modulated by androgen, plays a key role in autophagy and cell proliferation (Ashrafizadeh et al., 2022). Furthermore, during ER stress, upregulation of ERN1/IRE1 activates autophagy to restore homeostasis. ERN1/IRE1, along with other proteins such as ATF6 and CHOP, is overexpressed in tumor cells due to miR-200c-3p activation. miR-200c-3p plays a significant role in autophagy regulation and the ER stress response, inducing an increase in LC3-II expression and the accumulation of LC3 puncta, while its inhibition blocks autophagy induced by starvation. This suggests that miR-200c-3p can enhance survival mechanisms through autophagy activation and the IRE1 pathway in PCa (Sohn, 2018) (Fig. 4).

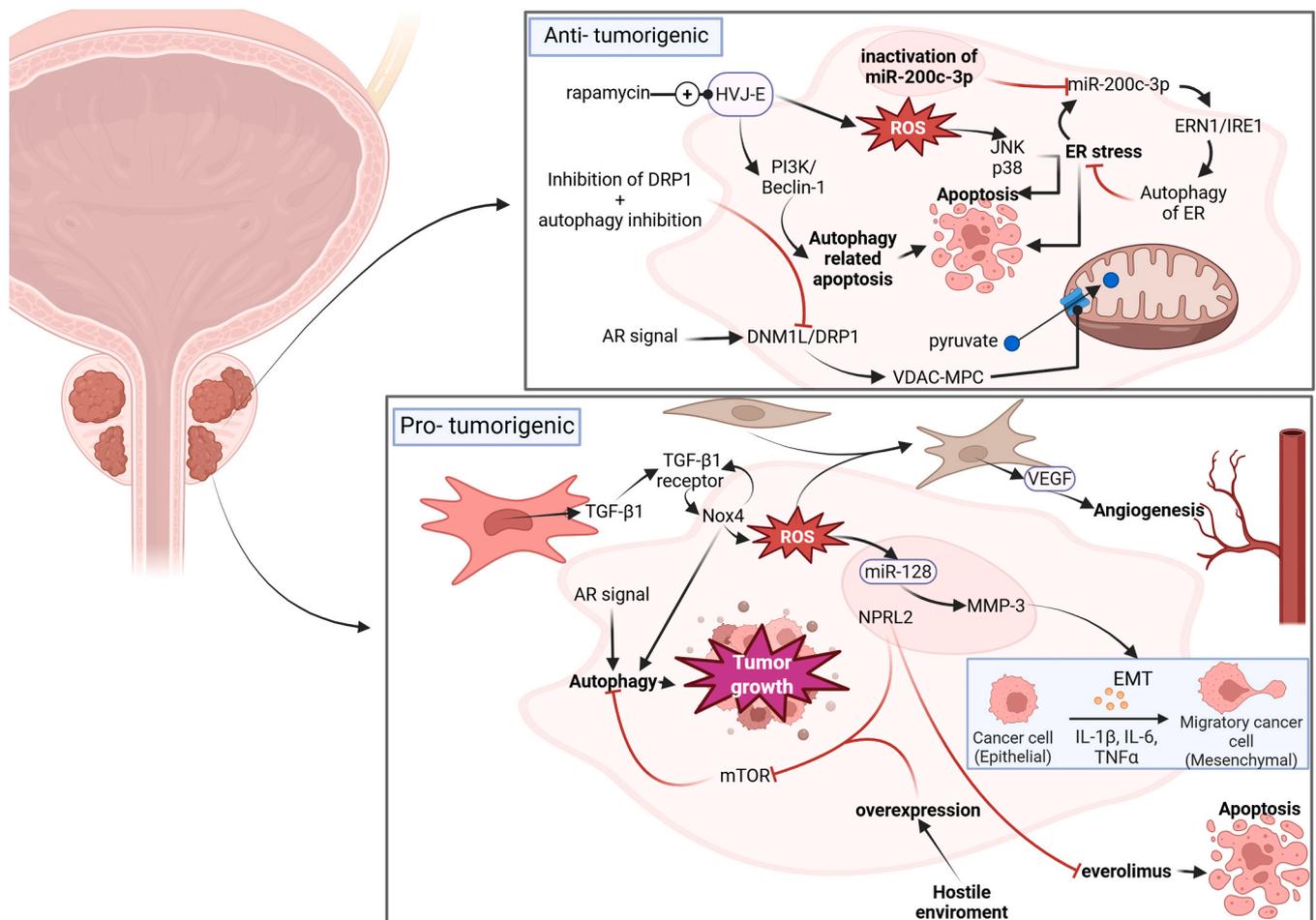


Fig. 4. Molecular pathways involved in determining early and advanced PCa.

### 3.4. Colorectal cancer

Colorectal cancer (CRC) is the third most prevalent cancer in developed countries and the second leading cause of cancer-related morbidity and mortality worldwide in both men and women (Lauzier et al., 2019; Siegel et al., 2013). According to GLOBOCAN 2022 data, there were approximately 1.926 million new CRC cases and 903,859 deaths globally (Bray et al., 2024). Despite significant advancements in diagnosis, surgery, and treatment, approximately 40 % of CRC patients succumb to the disease (Guo et al., 2024). While CRC incidence is rising globally, especially in developing countries adopting Western lifestyles, mortality rates have decreased in developed nations due to improved screening and treatment options. Risk factors for CRC include age, genetic predisposition, gastrointestinal disorders, and lifestyle factors such as obesity, physical inactivity, red meat consumption, alcohol intake, and smoking (Sawicki et al., 2021). While the molecular mechanisms underlying CRC progression are still being elucidated, growing evidence highlights a dynamic interplay between autophagy and oxidative stress in regulating tumor initiation, progression, treatment resistance, and metastasis.

On one hand, ULK1 is a key regulator in autophagy initiation and its expression increases in CRC progression. In CRC, ULK1-driven autophagy promotes clearance of cytotoxic components in hypoxic, nutrient-deprived environments (Zou et al., 2015). In addition, the AMPK/ULK1 axis is involved in the treatment response of oridonin, a natural compound belonging to the ent-kaurane diterpenoids, in CRC cells. In DLD-1 colon cancer cells, oridonin increases AMPK phosphorylation while decreasing mTOR and ULK1 phosphorylation in a concentration-dependent manner, promoting autophagy and apoptosis

(Bu et al., 2020). AMPK inhibition reversed cytotoxicity and reduced apoptosis, confirming its key role in this process. Furthermore, in animal models, oridonin significantly reduced tumor growth, with changes in AMPK, mTOR, and ULK1 levels like those observed in vitro, supporting the importance of the AMPK/mTOR/ULK1 pathway in regulating cell death in CRC (Bu et al., 2020). Liu et al. (2019) have demonstrated that autophagy plays a survival role when CRC cells are treated with the inhibitor of PI3K/mTOR denominated NVP-BEZ235 so blocking autophagy by AMPK/ULK1 inhibition enhances the treatment response (Liu et al., 2019). Ras mutations lead to the uncontrolled activation of several signaling pathways, such as the Raf-MAPK, MEK-ERK, and PI3K-Akt pathways. This alteration in Ras also increases superoxide production through the activation of NOX1 in the MAPK pathway. Oncogenic KRAS allows cells to evade apoptosis through the MAPK, PI3K, and Rac1 pathways. Furthermore, oncogenic Ras initiates an antioxidant program that helps the tumor grow, facilitating its adaptation to the environment, proliferation, and resistance to cell death (Catalano et al., 2025; Ternet and Kiel, 2021). ERK regulates autophagy and oxidative stress in CRC. When ERK is inhibited, both autophagy and apoptosis are induced in CRC cells. Additionally, inhibiting ERK leads to an increase in ROS levels, which activates autophagy, potentially protecting the cells from treatment effects (Mi et al., 2021).

ROS play a crucial role in the activation of ferroptosis. In CRC cells, the loss of p53 is associated with increased activity of DPP4 (dipeptidyl peptidase-4) on the plasma membrane, which facilitates lipid peroxidation mediated by NOX and promotes ferroptosis. In an antagonistic role, p53 protects against ferroptosis in CRC by promoting the translocation of DPP4 to the nucleus, where it forms a complex with p53. This complex blocks the formation of the DPP4-NOX link, preventing lipid



Wnt/ $\beta$ -catenin, and Notch pathways play a dynamic role in HCC, influencing tumorigenesis, therapy resistance (Yuanyuan Li et al., 2023), and metabolic reprogramming through stage-specific mechanisms. The Keap1/Nrf2/ARE pathway intersects with autophagy in HCC through p62-mediated mechanisms (Orrù et al., 2020), where phosphorylated p62 sequesters Keap1, stabilizing Nrf2 and driving antioxidant and detoxification responses. Autophagy deficiency exacerbates this effect by accumulating p62-Keap1 aggregates, leading to sustained Nrf2 activation, which supports tumor progression. In early HCC, Nrf2 or Keap1 mutations predominate, while in advanced stages, p62 accumulation replaces mutations as the primary driver of Nrf2 activation (Lau et al., 2010). During oxidative stress, modifications to Keap1 allow Nrf2 to translocate into the nucleus, where it binds to the antioxidant response element (ARE) and activates the expression of antioxidant genes (Yuanyuan Li et al., 2023). Furthermore, Nrf2 promotes immune evasion by upregulating immunosuppressive cytokines such as IL-10 and TGF- $\beta$ , improving the tumor microenvironment and reducing the effectiveness of immune checkpoint inhibitors (ICIs) (Gan et al., 2024; Raghunath et al., 2018). In this way, Nrf2 activation is linked to multidrug resistance (MDR) and poor prognosis (Yuanyuan Li et al., 2023). Another sign of poor prognosis is given by PI3K/AKT/mTOR which, altered by mutations in EGFR, promotes tumor progression. FOXO 3 acts as a tumor promoter in HCC, where its overexpression induces both oxidative stress and tumorigenesis. Although FOXO3 regulates both the production and the deletion of ROS, its activation in hepatocytes favors cell damage and malignant transformation. It is observed that FOXO3 activates the signalization pathway Akt/mTORC2 promoting a positive feedback loop which maintains its activation. Moreover, FOXO3 increases the PPP pathway and its relationship with NADPH/NADP<sup>+</sup> protecting cells from oxidative stress (Yang et al., 2021).

Likewise, the overproduction of endothelial and inducible nitric oxide synthases (eNOS and iNOS) raises NO levels, which disrupts normal autophagy. This happens because NO interferes with the binding of BECN1 and VPS34 while strengthening the interaction between Bcl-2 and BECN1, ultimately pushing cancer cells toward apoptosis (O'Connor et al., 2010). Silica nanoparticles, however, affect the NO/NOS system by blocking the PI3K/AKT/mTOR pathway. As a result, eNOS activity drops while iNOS activity increases significantly, leading to inflammation, autophagy activation, and eventually, endothelial dysfunction (Duan et al., 2014).

Hypoxia-induced autophagy is mediated by HIF-1 $\alpha$  and its downstream targets BNIP3 and BNIP3L, which regulate BCL-2 interactions (Nakamura et al., 2024). In addition, the Wnt/ $\beta$ -catenin signaling pathway plays a crucial role in regulating autophagy in HCC (Zhao, 2024). Under nutrient stress,  $\beta$ -catenin interacts with LC3 for autophagic degradation, suppressing autophagosome formation and inhibiting the expression of p62, reducing its activity and supporting cellular adaptation (Petherick et al., 2013). Autophagy can degrade  $\beta$ -catenin, promoting cellular adaptation to metabolic stress and creating a feedback loop where  $\beta$ -catenin inhibits autophagy. In HCC, Wnt/ $\beta$ -catenin-mediated suppression of autophagy may support tumor growth, while autophagy activation can enhance  $\beta$ -catenin degradation and potentially inhibit tumor progression (Inagawa et al., 2002). This pathway also contributes to drug resistance and metastasis in HCC. Targeting autophagy or modulating the Wnt/ $\beta$ -catenin signaling axis could offer promising therapeutic strategies to improve treatment outcomes in HCC. In addition, USP8, an ubiquitin ligase, is associated with ferroptosis resistance in HCC. USP8 stabilizes  $\beta$ -catenin, which upregulates GPX4 and its depletion of USP8 reduces GPX4 levels, promoting ferroptosis (Tang, 2023) (Fig. 5).

In primary hepatocellular carcinoma (HCC) tumors, the *RPRD1A* gene (Regulation of Polymerase II Transcription DNA Repair and Damage 1 A) is overexpressed and correlates with aggressive clinicopathological characteristics. Its function involves disrupting the interaction between TRIM21 and p62, thereby blocking ubiquitination and

enhancing Keap1 sequestration. This leads to the stabilization of NRF2, helping to reduce cellular damage caused by ROS and promoting HCC progression. Additionally, RPRD1A regulates the cell cycle and upregulates Wnt/ $\beta$ -catenin signaling. Therefore, the RPRD1A-TRIM21-p62 axis may represent a novel therapeutic target in HCC cells (Feng et al., 2021). Inflammation is a major cause of ROS production in chronic liver disease. Pro-inflammatory cytokines such as TNF- $\alpha$  and IL-6 exacerbate oxidative damage by inducing NADPH oxidase activity and disrupting mitochondrial function (Cichoż-Lach and Michalak, 2014; Liang et al., 2016; Singh et al., 2017; Tang et al., 2022; Vachliotis and Polyzos, 2023). Chronic liver diseases, including viral hepatitis B and C, NAFLD and alcohol-induced liver injury, are often associated with sustained ROS production due to persistent inflammation, mitochondrial dysfunction and lipid peroxidation (Che et al., 2023; Cichoż-Lach and Michalak, 2014; Sharma et al., 2023) (Fig. 6).

Mitochondrial dysfunction, characterized by ultrastructural lesions, altered dynamics, and impaired respiratory chain activity, plays a crucial role in the pathogenesis of these conditions (Mansouri et al., 2018; Zhang et al., 2022). Persistent ROS production, primarily from mitochondrial NADH and NADPH oxidases, leads to lipid peroxidation and depletion of antioxidant mechanisms. This oxidative stress disrupts redox balance, modulates inflammatory pathways, and contributes to disease progression, including fibrosis, cirrhosis, and HCC (Shimizu et al., 2012).

#### 4. Therapeutic potential

Autophagy and oxidative stress are crucial factors in the initiation, progression, and treatment of solid tumors. Their therapeutic value lies in modulating these processes and leveraging their interactions to improve cancer treatment strategies (Yang et al., 2011). In established tumors, autophagy often promotes cell survival under stress conditions, leading to treatment resistance. On the other hand, modulating oxidative stress offers several therapeutic approaches, such as pro-oxidant therapies, antioxidant enzyme inhibition or targeted therapies (Gorrini et al., 2013).

##### 4.1. Breast cancer therapeutic potential

BC can be classified in three principal types: tumors with positive hormonal receptors (BC HR<sup>+</sup>) characterized by a higher expression of estrogen and/or progesterone levels (ER<sup>+</sup> and/or PR<sup>+</sup>); BC; breast cancer positive HER2 positive that overexpress the oncogene ERBB2; and triple negative breast cancer (TNBC), which lacks both hormonal receptors and HER2 overexpression. These subtypes have differences in terms of aggressiveness, therapeutic response, progression risk and tendency to develop metastases in specific organs (Prat and Perou, 2011). The dysregulation of the PI3K/AKT/mTOR pathway is involved in endocrine resistance in patients with BC HR<sup>+</sup>. It is observed that the overactivation of the growth factor signaling pathway, including PI3K/AKT/mTOR, contributes to resistance to endocrine therapy (ET) and CDK4/6 inhibitors. The interaction of these pathways plays a crucial role in the treatment response of breast cancer. In preclinical studies the simultaneous inhibition of the estrogen receptor, CDK4/6 and PI3K has demonstrated synergistic effects, favoring the cell cycle arrest, apoptosis induction and tumoral regression in both, *in vitro* and *in vivo* models (Jhaveri et al., 2024). A phase I/Ib clinical trial showed that the treatment with inavolisib, a PI3K inhibitor administered daily, in combination with palbociclib, an inhibitor of CDK4/6 and endocrine therapy (letrozole or fulvestrant), resulted in a reduction in the expression of PI3K/AKT biomarker in tumoral tissue, confirming the efficacy of this treatment (Jhaveri et al., 2024), ClinicalTrials.gov identifier: NCT03006172). Another clinical trial in phase III, INAVO120, is evaluating the combination of inavolisib, palbociclib and fulvestrant in HR<sup>+</sup>, HER2- metastatic BC patients with PIK3CA mutation. The preliminary results of this study have demonstrated a significant

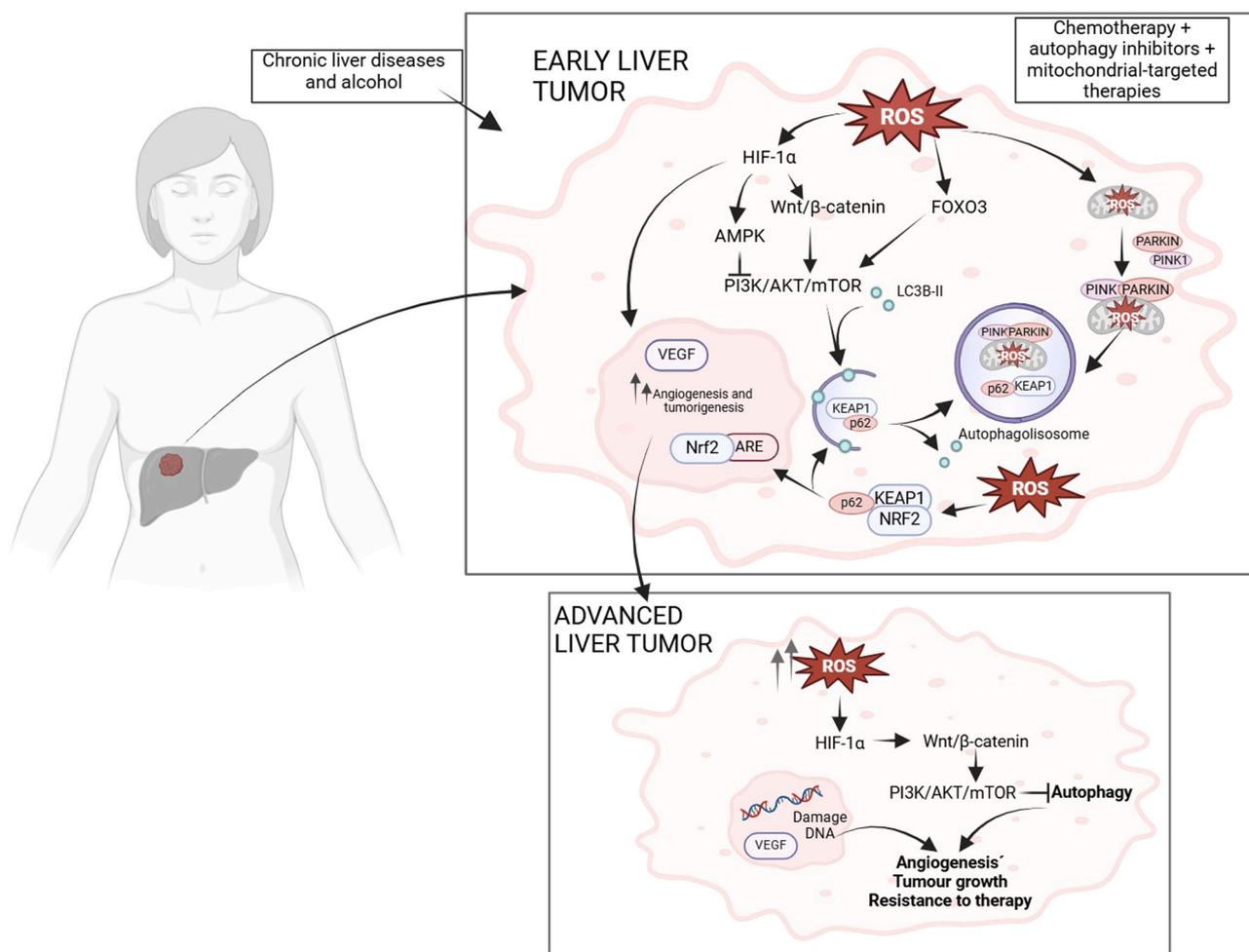


Fig. 6. Molecular pathways involved in determining early and advanced HCC.

improvement in progression-free survival (ClinicalTrials.gov identifier: NCT04191499) (Juric et al., 2024).

Additionally, autophagy contributes to drug resistance in HER2 + BC by HER2 + protected autophagy mechanism. HER2 is a transmembrane tyrosine kinase receptor belonging to the epidermal growth receptor family. Its overexpression triggers excessive activation of various signaling pathways, including MAPK, JAK/STAT, RAS/MEK/ERK and PI3K/AKT/mTOR, promoting proliferation and differentiation of tumor cells. Evidence suggests that the HER2 receptor inhibits autophagy interacting with Beclin-1, moreover, the activation of Beclin-1 signaling pathway induces mTORC1 activation, a key inhibitor in autophagy. It has been reported that the HER2-targeted treatments can induce autophagy as a resistance mechanism, and its inhibition may enhance the drug response (M. Wang et al., 2024).

Di Donato et al. (2022) investigated the role of the p110 $\delta$  PI3K and Vps34 isoforms (Di Donato et al., 2022). The p110 $\delta$  isoform is involved in the proliferation and cell migration modulation and its inhibition with the IC87114 drug decreases cell migration in spheroids of both the MCF-7 (BR HR+ cell line) and MDA-MB-231 (TNBC cell line). In contrast, Vps34, a class III PI3K kinase regulates autophagy and influences cancer development. Its inhibition with Vps34-IN1 increases sunitinib and erlotinib drug's efficacy, showing synergistic effects in the cell migration and proliferation. Vsp34 belongs to class III PI3K, and its pharmacological inhibition mediates vesicle transport and autophagy phenomena (Di Donato et al., 2022).

MitoQ, a derivative of Coenzyme Q10 that specifically targets mitochondria. It has antioxidant properties but, paradoxically, it also induces the production of ROS in cells. The increased ROS levels trigger

a protective response that involves autophagy. MitoQ generates the degradation of Keap 1 through autophagy, which activates the Keap1/Nrf2 pathway. The deletion of Keap 1 increases the transcriptional activity of Nrf2 leading to the expression of antioxidants genes such as p62, protecting cells from oxidant damage. The p62 protein is associated with Keap1 and activates Nrf2, producing a positive feedback loop that protects cells from oxidative damage. Autophagy inhibition produces an accumulation of p62 and the activation of Nrf2, producing to the tumor cells a defense against chemotherapeutic agents. Therefore, inhibiting Nrf2 could be a potential strategy to improve the function of chemotherapeutic drugs (Gonzalez et al., 2014).

Various studies have explored the role of REDD1 in growth regulation, differentiation, stress response, survival and cellular metabolism. Its main function is to inhibit the mTORC1 complex, which consists of the serine/threonine mTOR kinase and the Raptor protein. mTORC1 is activated by mitogens and nutrients, coordinating key cellular processes, including autophagy (Zhidkova et al., 2022). Hypoxia, energy stress, oxidative stress, glucocorticoids, or specific pharmacological activators have been shown to upregulate REDD1, leading to mTORC1 inhibition and reduced cancer cell proliferation. Furthermore, the increased REDD1 expression following chemotherapy has been linked to lower BC cell survival. In HER2-positive and triple-negative BC, tumor cell proliferation and survival in hypoxic environments are associated with REDD1 downregulation and HIF-1 $\alpha$  stabilization (Lan et al., 2013; Tirado-Hurtado et al., 2018; Zhidkova et al., 2020).

Pro-oxidants such as elesclomol and arsenic trioxide exploit the redox vulnerabilities of BC cells by selectively increasing ROS (Alli and Ford, 2012; Qu et al., 2010), overwhelming antioxidant defenses, and

inducing apoptosis (Kirshner et al., 2023). Their efficacy is further enhanced when combined with autophagy inhibitors, revealing a promising dual-targeting strategy (Yang Li et al., 2023). Elesclomol induces oxidative stress by disrupting mitochondrial metabolism and copper homeostasis, leading to ROS accumulation and cuproptosis, a copper-dependent cell death pathway (Nagai et al., 2012). It also targets cancer stem-like and drug-resistant cells, including cisplatin-resistant populations, by elevating ROS levels beyond their antioxidant capacity (Buccarelli et al., 2021). Similarly, arsenic trioxide (ATO) inhibits BC growth by downregulating Notch-1 signaling, reducing Bcl-2 and NF- $\kappa$ B levels, and promoting apoptosis (Xia et al., 2014). It exhibits dose-dependent cytotoxicity in triple-negative (MDA-MB-231), ER+ (MCF-7), and HER2 + (SKBR-3) BC cells (Alzuwaidi and Khalil, 2019; Chow et al., 2004; Rajaram et al., 2024; Wang et al., 2011). ATO also upregulates p53 expression, inhibits cell migration and invasion while down regulating Bcl-2 (Zhang et al., 2016), and activates key apoptotic proteins, including caspases and cytochrome C (Alzuwaidi and Khalil, 2019), which further contribute to apoptosis induction (Chow et al., 2004). Combining elesclomol or ATO with autophagy inhibitors like bafilomycin A1 or CQ prevents ROS clearance (Wang et al., 2023), leading to excessive oxidative damage and increased apoptosis. Additionally, ATO synergizes with tetrandrine or miR-27a modulation, further suppressing antioxidant defenses and metastatic potential (Zhang et al., 2016). Furthermore, the combination of ATO and BIBR1532, an hTERT inhibitor, synergistically inhibits BC cell proliferation by targeting the hTERT and NF- $\kappa$ B pathways (Nasrollahzadeh, 2020). Likewise, it has been shown that the combination of ATO, carboplatin, and cyclophosphamide induces synergistic effects in promoting apoptosis and autophagy in triple-negative BC cell lines (Moomivand et al., 2024). Similarly, dichloroacetate (DCA) potentiates ATO's cytotoxicity by targeting cancer cell metabolism and downregulating c-Myc and HIF-1 $\alpha$  (Sun et al., 2011). Melatonin also enhances ATO-induced apoptosis in MCF-7 cells by increasing p53 activity and the Bax/Bcl-2 ratio while suppressing survivin, c-Myc, and hTERT expression (Nooshinfar et al., 2016).

Preclinical studies have offered promising strategies for improving the therapeutic potential in breast cancer treatment, demonstrating that autophagy inhibitors enhance the cytotoxic effects of pro-oxidants in BC models (Wang et al., 2023). Another resistance-overcoming strategy involves CAT suppression, as elesclomol sensitizes resistant BC cells by reducing CAT expression, a key antioxidant enzyme responsible for neutralizing hydrogen peroxide (Glorieux and Calderon, 2018). This reduction lowers the threshold for ROS-induced apoptosis, particularly in cells that have adapted to oxidative stress. Clinically, a potential strategy to overcome treatment resistance is the combination of ROS-inducing agents with chemotherapy (e.g., elesclomol + paclitaxel) (Kirshner et al., 2008), metabolic modulators (e.g., ATO + dichloroacetate, 2-deoxy-D-glucose or 3-bromopyruvate) (Modica-Napolitano et al., 2024), autophagy inhibitors (e.g., ATO + CQ) (Wang et al., 2023), or antioxidant pathway inhibitors (e.g., elesclomol + N-acetylcysteine (NAC)) presents a potential strategy to overcome treatment resistance.

On the other hand, dysregulated mitophagy in BC cells leads to excessive ROS accumulation, driving both tumorigenesis and metastasis (Dong and Zhang, 2024; Wang et al., 2023). Targeting mitophagy-specific proteins, such as PINK1 and Parkin, is an emerging area of research and offers new therapeutic opportunities to address mitochondrial dysfunction in cancer (Li et al., 2022; Narendra and Youle, 2024). These findings underscore the therapeutic potential of targeting both ROS and autophagy pathways to improve BC treatment outcomes (Table 1).

#### 4.2. Lung cancer therapeutic potential

The therapeutic potential of targeting autophagy and oxidative stress in LC is an area of significant interest, as these processes play critical roles in cancer cell survival, progression, and treatment resistance. Jeon,

S. J. et al. (2019) have shown that TOR Signaling Pathway Regulator-Like Protein (TIPRL) binds to eIF2 $\alpha$  and promotes its phosphorylation, thereby activating the eIF2 $\alpha$ -ATF4 pathway (Jeon, 2019). As a result, tumor cells increase their ability to resist metabolic stress, promoting survival and contributing to tumor progression through autophagy activation. In contrast, TIPRL deletion decreases the autophagy flux, reducing the tumor cell survival and increasing the cell death. The combination of metabolic and ER stress induction with autophagy inhibition through TIPRL deletion could present a potential therapeutic target to treat LC (Jeon, 2019). Likewise, autophagy inhibition by CQ or specific autophagy inhibition potentiate T CD8 + infiltration and increase MHC-I expression molecules on tumor cell surface, thereby enhancing immunotherapy efficacy. Consequently, tumor growth is reduced. This process boosts the immune response through increasing tumor antigen exposure to T cells and activating the adaptive immune response (Guo et al., 2022) (Table 2).

Wiel C. et al. (2019) have demonstrated long-term supplementation with antioxidants including N-acetylcysteine and vitamin E, can enhance KRAS-related LC metastasis (Wiel et al., 2019). These antioxidants produce the stabilization of BACH1, activating glycolysis genes like hexokinase 2 and GAPDH, which promote metastasis. Inhibiting BACH1 reduces glycolysis and prevents antioxidant-induced metastasis. BACH1 plays a key role in metastasis under oxidative stress conditions and may serve as a potential therapeutic target for LC (Wiel et al., 2019).

Natural products are also under investigation; Red ginseng, specifically the Rg3-extract, has been demonstrated to promote apoptosis and mitophagy in lung tumor cells, through the activation of ROS and PINK1-Parkin signaling pathway (Nakamura et al., 2024). A ginsenoside metabolite, Compound K, induces apoptosis through autophagy in A549 and H1975 cells through AMPK/mTOR pathways and JNK. Therefore, in cisplatin treatment, Rh2 ginsenoside enhances apoptosis by autophagy inhibition, which reduces drug resistance in LC (Qin et al., 2024). Luo D. et al. (2023) discovered a novel alkaloid derivative from *Sophora flavescens* roots denominated Sophflarine A (SFA) (Luo et al., 2023). SFA induces ROS accumulation leading to autophagy activation by inhibiting PI3K/Akt/mTOR pathway. Additionally, ROS promotes NLRP3 inflammasome activation, triggering pyroptosis and cancer cell death. Furthermore, SFA reduces metastasis blocking epithelial-mesenchymal transition and suppressing angiogenesis. These findings suggest that SFA can be a drug candidate for the treatment of LC (Luo et al., 2023). Rhabdothermin E is an ent-kaurene diterpenoid. The mechanism action of rhabdothermin E involves ROS accumulation which activates p38/MAPK/JNK pathway, promoting both apoptosis and ferroptosis. Moreover, it reduces GSH and GPX4 levels while increasing lipid peroxidation, confirming its ability to induce ferroptosis through SLC7A11/GPX4 pathway. In murine models, rhabdothermin E decreases tumor growth more effectively than treatments like cisplatin. In addition, it reduces the expression of tumor proliferative index Ki67 (tumor proliferation indicator) expression and increases Bax/Bcl-2 ratio, favoring apoptosis. These findings highlight the strong antitumor potential of rhabdothermin E, making it a promising therapeutic candidate for LC (Jin et al., 2024). Another study has shown that the synergistic effect of cisplatin with ginkgetin, a metabolite derived from *Ginkgo biloba*, enhances the cytotoxicity in LC cells, specifically NSCLC with wild EGFR both in vitro and in vivo. Ginkgetin, a flavonoid from *Ginkgo biloba*, enhances cisplatin cytotoxicity in NSCLC by promoting ROS accumulation and ferroptosis. It inhibits SLC7A11 and GPX4, disrupting redox homeostasis and sensitizing cells to lipid peroxidation, without affecting healthy lung tissue (Lou et al., 2021). Since resistance to apoptosis and restoration of redox balance are factors limiting the effectiveness of DDP, induction of ferroptosis and disruption of redox homeostasis by ginkgetin offer a promising strategy to improve the efficacy of this drug in the treatment of NSCLC (Lou et al., 2021). Another natural compound that may offer a new therapeutic option for LC is celastrol (a natural compound belonging to the triterpenoids family), either alone or in combination with TRAIL. Celastrol inhibits autophagic flux, enhancing

**Table 2**  
Therapeutic strategies in LC.

Therapeutic strategies	Molecular mechanism	References
<b>Preclinical study:</b> TIPRL deletion + metabolic and ER stress induction could present a potential therapeutic target	Decreases the autophagy flux, reducing the tumor cell survival and increasing the cell death	(Jeon, 2019)
<b>Preclinical study:</b> Autophagy inhibition by CQ or specific autophagy inhibition enhance immunotherapy efficacy	Potentiates T CD8 + infiltration and increase MHC-I expression molecules on tumor cell surface,	(Guo et al., 2022)
<b>Preclinical study:</b> BACH1 inhibitors	Reduce glycolysis and prevents antioxidant-induced metastasis, since long-term supplementation with antioxidants including N-acetylcysteine and vitamin E, produce the stabilization of BACH1, activating glycolysis genes like <i>HK2</i> and <i>GAPDH</i> , which promote metastasis	(Wiel et al., 2019)
<b>Preclinical study:</b> Red ginseng, Rg3-extract and ginsenoside metabolite, Compound K	Promotes apoptosis and mitophagy through the activation of ROS, PINK1-PARKIN, AMPK/mTOR pathways and JNK	(Nakamura et al., 2024; Qin et al., 2024)
<b>Preclinical study:</b> Novel alkaloid derivative from <i>Sophora flavescens</i> roots denominated Sophflarin A (SFA)	Induces ROS accumulation that inhibits PI3K/Akt/mTOR pathway leading to autophagy activation, and activates NLRP3 inflammasome, triggering pyroptosis and cancer cell death, SFA also blocks epithelial-mesenchymal transition and suppresses angiogenesis	(Luo et al., 2023)
<b>Preclinical study:</b> Rhabdothermin E, an entkauran diterpenoid, with no previously reported antitumor potential	Induces ROS accumulation that activates p38/MAPK/JNK pathway, reduces Ki-67 expression, increases Bax/Bcl-2 ratio, promoting both apoptosis and ferroptosis through SLC7A11/GPX4 pathway. In murine models, tumor growth decreases more effectively than treatments like cisplatin	(Jin et al., 2024)
<b>Preclinical study:</b> Synergistic effect <i>in vitro</i> and <i>in vivo</i> of cisplatin + ginkgetin, a metabolite derived from <i>Ginkgo biloba</i>	Enhances the cytotoxicity in NSCLC cells with wild EGFR by inducing ROS accumulation which activates p38/MAPK/JNK pathway and triggers apoptosis and ferroptosis	(Lou et al., 2021)
<b>Preclinical study:</b> Celastrol (a natural compound belonging to the triterpenoids family)	Inhibits autophagic flux, increases ROS levels and decreases mitochondrial membrane potential, contributing to cell death, enhancing apoptosis. The combined treatment of TRAIL and celastrol increases cell death compared to the individual treatments, moreover, celastrol elevates LC3-II and p62 levels and potentiates caspase 3 and 8 activation, which are correlated with apoptosis	(Nazim et al., 2019)

apoptosis mediated by TRAIL pathway. The combined treatment of TRAIL and celastrol increases cell death compared to the individual treatments, moreover, celastrol elevates LC3-II and p62 levels and potentiates caspase 3 and 8 activation, which are correlated with apoptosis

induction. Additionally, autophagy inhibition by celastrol increases ROS levels and decreases  $\Delta\Psi_m$  (mitochondrial membrane potential), contributing to cell death (Nazim et al., 2019).

Clinical trials are currently investigating the combination of autophagy inhibitors with immune checkpoint inhibitors, such as anti-PD-1/PD-L1 antibodies, in NSCLC. Preliminary results indicate that autophagy inhibition enhances immunotherapy efficacy by promoting tumor cell death and reversing immune suppression in the tumor microenvironment (Gao and Chen, 2021; Guo et al., 2022; Li et al., 2024). Furthermore, the integration of pro-oxidants with autophagy inhibitors is another promising avenue. By simultaneously increasing ROS levels and blocking the protective effects of autophagy, this dual-targeting approach could induce synergistic cytotoxicity in LC cells while sparing normal tissues (Shimizu et al., 2023; W. Wang et al., 2022).

Pro-oxidants such as elesclomol and arsenic trioxide selectively exploit the elevated ROS vulnerability of cancer cells, inducing apoptosis with minimal toxicity to normal cells (You and Park, 2012). Preclinical studies have shown that combining pro-oxidants with autophagy inhibitors further sensitizes LC cells to oxidative damage, highlighting the potential of dual-targeting strategies (Kaminsky et al., 2012; Wang et al., 2024). Conversely, antioxidants like NAC may protect normal tissues from therapy-induced oxidative damage, improving tolerance during radiotherapy (Shin et al., 2020). For example, N-acetylcysteine (NAC) has been shown to protect normal lung tissue during radiotherapy while maintaining the efficacy of cancer treatment (Moldeus et al., 1986; Sala, 1993). However, the use of antioxidants in cancer therapy remains controversial, as they may also protect tumor cells from ROS-mediated cytotoxicity.

#### 4.3. PCa therapeutic potential

The interplay between autophagy and ROS presents significant therapeutic potential in PCa (Dong et al., 2023). The activation of autophagy by chemotherapeutic agents, such as docetaxel, plays a crucial role in promoting drug resistance in PCa. Increased expression of NPRL2 and BECN1 has been linked to autophagy-induced resistance, whereas mTOR pathway activation counteracts this effect by suppressing autophagy and improving docetaxel efficacy. Additionally, TGF- $\beta$ 1 stimulates autophagy and reduces the effectiveness of docetaxel. Inhibiting TGF- $\beta$ 1 secretion can prevent autophagy activation, thereby enhancing the drug's impact on PCa cells. KLF5, a transcription factor downregulated by docetaxel via the AMPK-mTOR-p70S6K axis, suppresses autophagy and sensitizes cells to therapy. Similarly, AMPK activation under androgen deprivation induces protective autophagy, promoting cell survival (Jia et al., 2019; Wang et al., 2018; Yu, 2021) (Table 3).

Ramos-Torres et al. (2016) investigated the effect of capsaicin in PCa cells, showing that this substance has an antiproliferative and cytotoxic effect, specifically in PC-3 cells, insensitive to androgens. Capsaicin also inhibits PI3K/Akt/mTOR pathway, which is overexpressed in PCa and increases LC3-II levels. However, it also produces an accumulation of p62, suggesting that capsaicin inhibits the autophagy pathway. Capsaicin also generates ROS, contributing to apoptosis when autophagy is inhibited, supporting its use as a potential PCa therapy (Ramos-Torres et al., 2016). Another study has shown the effects of Hydroxyl-Nor-Kavain (HNK), a natural anticancer agent, on autophagy in PCa cells. Autophagy induced by HNK plays a protective role in the cells, due to autophagy inhibition increases apoptosis in cells treated with HNK and it inhibits mTOR and AKT phosphorylation. In addition, ROS also plays a significant role in autophagy induction by HNK. However, autophagy inhibition with antioxidant substances does not affect apoptosis induced by HNK. Treatment with HNK induces protective autophagy in PCa cells, and inhibiting autophagy could enhance HNK's anticancer effect, highlighting the therapeutic potential of this natural agent (Hahm et al., 2014). Mukha et al. (2021) demonstrated that autophagy and oxidative stress play a key role in PCa

**Table 3**  
Therapeutic strategies in PCa.

Therapeutic strategies	Molecular mechanism	References
<i>Preclinical study:</i> Capsaicin	Inhibits PI3K/Akt/mTOR, increases LC3-II, but also accumulates p62, suggesting autophagy inhibition. Generates ROS. Induces cytotoxicity and apoptosis in androgen-insensitive PC-3 cells, especially in combination with autophagy inhibitors	(Ramos-Torres et al., 2016)
<i>Preclinical study:</i> Hydroxyl-Nor-Kavain (HNK)	Induces protective autophagy, inhibits mTOR/AKT phosphorylation, increases ROS. Enhances apoptosis when autophagy is inhibited, improving therapeutic potential	(Hahm et al., 2014)
<i>Preclinical study:</i> CB-839 (Glutamine Metabolism Inhibitor)	Inhibits glutamine metabolism, increasing ROS and DNA damage. Induces autophagy as a survival mechanism. Sensitizes PCa cells to radiotherapy; combination with autophagy inhibitors (e.g., CQ) enhances oxidative stress and promotes apoptosis	(Mukha et al., 2021)
<i>Preclinical study:</i> Metformin	Downregulates MYC and GLS1, inhibits autophagy, and increases oxidative stress. Acts as a radiosensitizer, enhancing the effects of radiotherapy	(Mukha et al., 2021)
<i>Preclinical study:</i> Monoamine oxidase A (MAOA) inhibitors	Generates ROS, inhibits apoptosis (reduces p53 activation), and induces mitophagy. Inhibiting MAOA suppresses autophagy and promotes apoptosis, making it a potential therapy for castration-resistant PCa	(Lin et al., 2017)
<i>Preclinical study:</i> Sertraline (STL)	Induces ROS, mitochondrial dysfunction, autophagy and apoptosis. Inhibits PCa stem cell (PCSC) markers and EMT. Reduces PCa cell proliferation, migration, and stemness	(Chinnapaka et al., 2020)
<i>Preclinical study:</i> Calotropis procera extract (CPE)	Modulates autophagy, reduces ROS, increases p27 (cell cycle inhibitor), and inhibits NF-κB. Reduces PCa cell viability and migration; autophagy effects are cell type dependent. Preclinical study (clinical trials ongoing): PT-112. Generates mitochondrial stress via ROS, alters mitochondrial respiration, induces autophagy and immunogenic cell death (ICD). Reduces PCa cell viability and migration; autophagy effects depend on cell type. Selectively induces tumor cell death, enhances mitochondrial damage, and stimulates immune response	(Singh et al., 2024)
<i>Preclinical study:</i> PT-112 in PCa cells	PT-112 induces mitochondrial stress and ROS production, activates autophagy, and promotes immunogenic cell death (ICD) via DAMPS release,	(Soler-Agesta et al., 2024)

**Table 3 (continued)**

Therapeutic strategies	Molecular mechanism	References
<i>Preclinical study:</i> miRNA-based therapies (tumor-suppressing/promoting miRNAs, siRNAs)	enhancing tumor cell targeting miRNAs regulate autophagy in cancer cells. miRNA replacement or inhibition strategies may suppress tumor progression by targeting autophagy-related pathways	(Ashrafizadeh et al., 2022; John Clotaire et al., 2016)
<i>Preclinical study:</i> Combating resistance to androgen deprivation therapy (ADT)	In prostate cancer, increased autophagic flux enables cancer cells to degrade AR inhibitors and sustain AR signaling, contributing to resistance against ADT	(Elshazly and Gewirtz, 2023; Nguyen et al., 2014; Quan, 2020)

radioresistance through glutamine (Gln) metabolism (Mukha et al., 2021). Tumor cells rely on Gln to maintain redox balance, by glutathione production (GSH), which reduces ROS levels and protects against DNA damage. Gln metabolism inhibition with drugs such as CB-839 increases ROS levels, accumulation of DNA damage and enhances cells sensibility to radiotherapy. However, PCa cells activate autophagy like a pro-survival mechanism in response to Gln inhibition, leading to cellular adaptation and therapy resistance. Therefore, autophagy inhibition by ATG 5 inhibition or with CQ, enhances the radiosensitizer effect of Gln inhibition, increasing oxidative damage and promoting cell death. Furthermore, metformin, an antidiabetic drug, shows a radiosensitizer effect through downregulation of MYC and GLS1 expression, autophagy inhibition and increasing oxidative stress in tumor cells. The combination of Gln inhibitors and autophagy inhibitors can enhance the radiotherapy efficacy in PCa by promoting oxidative damage and reducing the tumor cell resistance to treatments (Mukha et al., 2021). Monoamine oxidase A (MAOA), a target gene of repressor element 1-silencing transcription factor (REST), plays a crucial role in autophagy activation and apoptosis inhibition, specifically in androgen deprivation-induced neuroendocrine differentiated PCa cells. MAOA generates ROS that inhibit apoptosis by reducing p53 activation, while simultaneously activating autophagy through mitophagy induction. These mechanisms are essential for PCa progression due to autophagy activation and apoptosis inhibition contribute to cell survival and treatment resistance. In addition, the study highlights that MAOA regulation by REST and its relationship with oxidative stress are key factors in autophagy activation and neuroendocrine differentiation (NED) in PCa cells. MAOA inhibition could be a therapeutic strategy for advanced PCa patients, including those resistant to castration and chemotherapy (Lin et al., 2017).

Other authors have investigated the impact of the antidepressant sertraline (STL) on PCa stem cells (PCSC), highlighting its potential as a treatment in PCa. STL inhibits growth and metastasis of PCSC, decreasing cell viability in various PCa cell lines. Moreover, STL reduces cell proliferation and migration of PCSC, through inhibiting EMT markers, such as TCF8 and LEF1. STL's cytotoxicity is associated with apoptosis induction and ROS generation, besides, STL induces cell cycle arrest in the G0 phase and activates mitochondrial dysfunction, contributing to cell death. Oxidative stress plays a significant role in this process, as ROS generation and the use of STL negatively regulates stem cell marker expressions including ALDH1 and anti-apoptotic proteins such as TCTP. Furthermore, STL induces autophagy in PCSC, and the combination of STL with antioxidants suggests that apoptosis induction is mediated by ROS. STL acts on PCSC through oxidative stress, apoptosis activation and autophagy, affecting PCSC mark expression, and could be a potential therapy to advanced PCa (Chinnapaka et al., 2020).

Natural compounds as leaf extract of *Calotropis procera* (CPE), reduce cell viability, division, and migration of PCa cells, both androgen-

sensitive and androgen-independent. CPE treatment in cells increases p27 protein expression, producing inflammatory inhibition and cell cycle regulation, additionally, it also reduces NF- $\kappa$ B activation. Furthermore, CPE decreases ROS levels, which in turn reduces cell viability and migration in PCA cells. The effect of CPE on autophagy requires further study; in PC-3 cell type, autophagy activation is associated with increased levels of p62, LC3B and Beclin-1, in contrast, in another 22Rv1 cell type, treatment with CPE decreases Beclin-1 and LC3-II expression, suggesting autophagy inhibition. Thus, the effects of autophagy depend on the cell type. Therefore, CPE inhibits proliferation and migration in PCa cells by autophagy modulation and reducing ROS levels. It could be a new therapy to PCa but more studies are needed to fully understand the role of autophagy (Singh et al., 2024).

Soler-Agesta et al. (2024), have shown the anticancer effects of PT-112 in PCa cells. PT-112 generates mitochondrial stress by ROS production, modifying mitochondrial membrane potential, mitochondrial respiration and mitochondrial morphology. In addition, PT-112 activates autophagy and generates immunogenic cell death by DAMPS (ICD) in tumor cells and does not affect to not tumor cells. Mitochondrial stress and DAMPs release are correlated due to the ATP release, with PT-112 promoting ICD that could enhance mitochondrial damage, cell death and apoptosis. Autophagy, influenced by mitochondrial stress is also involved in the ICD induction, and the ribosome biogenesis altered by PT-112 could also be involved in autophagy regulation contributing to the selective targeting of tumor cells by PT-112 (Soler-Agesta et al., 2024).

miRNA-based approaches, including tumor-suppressing and tumor-promoting miRNAs, regulating autophagy in PCa, and miRNA replacement therapy and siRNAs could be employed to modulate autophagy and suppress PCa progression (Ashrafizadeh et al., 2022; John Clotaire et al., 2016). Resistance to androgen deprivation therapy (ADT), the cornerstone of PCa treatment, is often associated with increased autophagic flux. Cancer cells use autophagy to degrade androgen receptor (AR) inhibitors and maintain AR signaling, allowing them to escape therapy-induced cell death (Elshazly and Gewirtz, 2023; Nguyen et al., 2014; Quan, 2020).

#### 4.4. Colorectal cancer therapeutic potential

Autophagy and ROS play crucial roles in the development and progression of CRC. Wang et al. (2023) showed that ANXA10, overexpressed in BRAF-mutant CRC, suppresses ferroptosis by reducing Fe<sup>2+</sup> accumulation and enhancing transferrin Receptor 1 degradation (TFRC). ANXA10 inhibition increased ferroptosis and autophagic flux, accompanied by SQSTM1 accumulation, indicating a regulatory role in both processes (Wang et al., 2023). Other authors have explored the role of glutaminase 1 (GLS1), a key enzyme in Gln metabolism, in the growth and migration of CRC cells. GLS1 is upregulated in CRC and promotes proliferation and migration by converting Gln to glutamate, lowering ROS levels, and upregulating NRF2. GLS1 inhibition increased ROS and decreased NRF2, impaired autophagy, and suppressed tumor growth, suggesting it as a dual regulator of redox and autophagic balance (Liu et al., 2021) (Table 4).

Zhang et al. (2024) demonstrated that ATG7 suppression can enhance T CD8 + cells activation, offering a novel mechanism to immunotherapy in CCR (W. Zhang et al., 2024). This effect occurs by ROS/NF- $\kappa$ B pathway activation, which increases the expression of major histocompatibility complex class I (MHC-I) on the cell membrane. Moreover, the authors showed that the combination of an ATG7 inhibitor with atorvastatin, a drug belonging to the statin class, used primarily to reduce cholesterol and triglyceride levels in the blood, which inhibits HMG-CoA (3-hydroxy-3-methylglutaryl-CoA) reductase (HMGCR) could be a potential therapeutic target. Consequently, the simultaneous inhibition of ATG7 and cholesterol regulation can enhance the immunotherapy response in patients with colorectal cancer and microsatellite instability-high (MSI-H) (Zhang et al., 2024). Gingerol

**Table 4**  
Representative Therapeutic Strategies Investigated in CRC.

Therapeutic strategies	Molecular mechanism	Ref.
<b>Preclinical study: ANXA10 inhibition</b>	Induces ferroptosis by increasing Fe <sup>2+</sup> levels and reducing TFRC degradation; enhances autophagic flux and SQSTM1 accumulation	(Wang et al., 2023)
<b>Preclinical study: GLS1 inhibition</b>	Disrupts glutamine metabolism, increases ROS, reduces Nrf2 expression, and alters autophagy, suppressing CRC cell growth and migration	(Liu et al., 2021)
<b>Preclinical study: ATG7 inhibitor + atorvastatin</b>	Activates ROS/NF- $\kappa$ B signaling, increasing MHC-I expression and enhancing CD8 + T-cell activation. Atorvastatin, an inhibitor of HMGCR, improves immunotherapy response	(Zhang et al., 2024)
<b>Preclinical study: Gingerol acid (GA)</b>	Induces ferroptosis by increasing lipid peroxidation, GSH depletion, and ROS/Fe2 + accumulation. Ferroptosis inhibitors prevent GA-induced cell death.	(Xiao et al., 2024)
<b>Preclinical study: Cysteine depletion + mTOR/autophagy inhibition</b>	Cysteine is crucial for tumor growth and redox balance. Its depletion activates autophagy via the mTOR/ULK pathway. The combination enhances treatment response	(Lin et al., 2024)
<b>Preclinical study: KRAS mutation and mitophagy</b>	KRAS G12/13 mutations increase autophagic flux. Activating mutant KRAS induces mitophagy and mitochondrial fission via AKT phosphorylation (S473) and ROS generation, leading to cancer cell death	(Iskandar et al., 2024)
<b>Preclinical study: Heat shock protein 90 (HSP90) JD-02 inhibition</b>	Disrupts protein homeostasis, induces G0/G1 arrest, apoptosis, and autophagy via ROS/SRC signaling. Preclinical/clinical studies: Immune checkpoint inhibitors (ICIs, anti-PD-1/CTLA-4 antibodies) + autophagy inhibition (CQ/HCQ). Restore immune surveillance. Autophagy inhibition enhances antigen presentation and T-cell infiltration, boosting ICI response	(Liu and Sun, 2021; Wang et al., 2022)
<b>Preclinical study: Conventional chemotherapy and radiotherapy</b>	Induce cytotoxicity through ROS generation. However, cancer cells, especially cancer stem cells—activate antioxidant mechanisms that reduce treatment efficacy	(Lyons et al., 2023)
<b>Preclinical study: Emerging therapies (photodynamic, sonodynamic, radiodynamic)</b>	Utilize targeted ROS generation to induce cell death. Nevertheless, adaptive mechanisms in cancer cells reduce the effectiveness of ROS-based drugs like 5-FU and oxaliplatin	(Levy and Thorburn, 2011)
<b>Preclinical study: <math>\alpha</math>-hederin</b>	Increases ROS production through the activation of the AMPK pathway, inhibition of mTOR and leads to autophagic cell death in cancer cells	(Sun et al., 2019)

(continued on next page)

Table 4 (continued)

Therapeutic strategies	Molecular mechanism	Ref.
<b>Preclinical study:</b> Cucurbitacin CC 90003	Promote ROS generation and induces autophagy	(Zhang et al., 2012)
<b>Preclinical study:</b> Hydroperoxy fatty acid 13 (S)-HpODE	Generate ROS, which leads to BAX activation and BCL-2 inhibition	(Biswas et al., 2023)
<b>Preclinical study:</b> Dimethyl fumarate	Induces glutathione depletion, ROS generation, and MAPK activation, which leads to necroptosis induction	(Han et al., 2018)
<b>Preclinical study:</b> Oleanolic acid	Promote ROS generation leading to G1/S cell cycle arrest and inhibition of cell proliferation	(Guo et al., 2017)
<b>Preclinical study:</b> Cucurbitacin E	Promote ROS generation and impede tumor growth by arresting the cell cycle at the G2/M phase	(Zhang et al., 2012)
<b>Preclinical study:</b> CNN16, Psoralidin, Alantolactone, TPEN, Graphene oxide, CopA3, Extracts from marine invertebrates	Induce the generation of ROS or ROS accumulation causing DNA damage	(Arslan et al., 1985; da Silva et al., 2022; Dey and Kang, 2021; Ding et al., 2016; Krasteva et al., 2019; Ruiz-Torres et al., 2019; Sun et al., 2022)
Clinical trial: NCT02106806	Analyzes the oxidative stress (SOD, GPx, MDA, Isoprostane, Vitamin C, Vitamin E) during cycles of chemotherapy in patients after surgery for CRC, with or without oral zinc supplementation	(Zhang et al., 2024)

acid (GA), an active compound of ginger, inhibits CRC progression by targeting the SLC7A11 protein and promoting its ubiquitin-mediated degradation, thereby inducing ferroptosis. GA treatment decreases significantly programmed CRC cellular proliferation both in vitro and in vivo, without showing toxic effect in vital organs. The mechanism consists of GA trigger ferroptosis in CRC cells by lipid peroxidation, GSH exhaustion and ROS and Fe<sup>2+</sup> accumulation. Additionally, treatment with apoptosis inhibitors and autophagy don't protect significantly against induced cell death by GA, while ferroptosis inhibitors did, suggesting that ferroptosis is key in the antitumor action of GA. In summary, this study shows that GA promotes ferroptosis in CRC and suggests that it may be a promising candidate for targeted treatment of colorectal cancer (Xiao et al., 2024). On the other hand, cysteine is a key metabolite in CRC and is overexpressed in tumor cells. It has been demonstrated that cysteine is essential for tumor growth, and the inhibition of cysteine transporters reduces cysteine levels and can induce ferroptosis. ATF4, a transcription factor, regulates the expression of cysteine transporters in response to microenvironment tumor stress, such as hypoxia and oxidative stress. Cysteine is utilized for the GSH synthesis, allowing CRC cells to maintain ROS levels in a favorable balance to the tumor growth. Combining cysteine depletion with mTOR or autophagy inhibitors enhances tumor cell sensitivity and may overcome redox adaptation (Lin et al., 2024).

Furthermore, it has been demonstrated that G12/13 mutations in the KRAS gene in CRC increase autophagic flux in response to cellular stress. Pharmacological activation of mutant KRAS stimulates both mitophagy and mitochondrial fission, a process mediated by AKT phosphorylation at S473 and ROS generation, ultimately leading to the death of mutant KRAS-driven cancer cells. However, the relationship between AKT signaling, mTORC1, and mitophagy remains debated. AKT phosphorylation at S473 by mTORC2 is essential for mitophagy and mitochondrial fission to occur following drug-induced activation of mutant KRAS (Iskandar et al., 2024). Lan et al. (2024) identified JD-02 as a novel heat

shock protein 90 (HSP90) inhibitor that binds to the ATP-binding site in the N-terminal domain, leading to protein destabilization and degradation. In CRC cells, JD-02 induces G0/G1 cell cycle arrest, apoptosis and autophagy primarily through the generation of ROS and modulation of Hsp90/SRC signalling axis. The combination of JD-02 with CQ, significantly enhances apoptotic cell death, suggesting that autophagy acts as a cytoprotective mechanism in this context (Lan et al., 2024). In line with this N-acetylcysteine (NAC) has shown to inhibit autophagy and apoptosis by modulating the ROS/JNK/Bcl-2 axis in liver injury models (Wang et al., 2014) and the AMPK/mTOR pathway in macrophages (Lin et al., 2020; Su et al., 2024). Given that both pathways are involved in CRC progression and chemoresistance, these mechanisms are likely conserved. Moreover, NAC has been reported to reduce inflammation-associated tumorigenesis in murine CRC models by suppressing NF-κB activity and oxidative DNA damage, thereby reducing tumor burden and dysplasia severity. However, its effects are dose- and context-dependent. For example, under conditions of high ER stress, NAC may paradoxically inhibit autophagy, impair cellular adaptation and promote apoptosis (Xiang et al., 2016). Therefore, while NAC shows promise as an adjuvant to modulate redox balance and autophagy in CRC, further studies are needed to delineate its precise therapeutic window and mechanism of action in this cancer type. Taken together, these results highlight that autophagy inhibitors may enhance the efficacy of JD-02 treatment, especially when combined with autophagy inhibitors such as CQ or NAC (Lan et al., 2024).

On the other hand, combination therapies integrating ICIs with autophagy inhibitors are advancing in CRC research. ICIs, such as anti-PD-1 and anti-CTLA-4 antibodies, restore immune surveillance but are often limited by the immunosuppressive CRC microenvironment. Combining ICIs with autophagy inhibitors, such as CQ or HCQ, disrupts tumor-induced immune evasion and enhances antigen presentation and cytotoxic T-cell infiltration, thereby increasing the therapeutic potential of ICIs (Liu and Sun, 2021; Wang et al., 2022). This dual-targeting strategy overcomes immune resistance and exploits autophagy vulnerabilities.

Conventional CRC therapies, including chemotherapy and radiotherapy, derive their cytotoxicity from ROS generation (Lyons et al., 2023). However, elevated detoxification mechanisms in cancer cells, particularly cancer stem cells, can lead to therapy resistance. Emerging approaches such as photodynamic, sonodynamic, and radiodynamic therapies aim to harness ROS for therapeutic benefit (Lyons et al., 2023). While this adaptive mechanism allows cancer cells to mitigate oxidative damage, it also contributes to therapy resistance by reducing the efficacy of ROS-inducing chemotherapies such as 5-fluorouracil (5-FU) and oxaliplatin (Levy and Thorburn, 2011).

#### 4.5. Hepatocellular carcinoma therapeutic potential

Autophagy and ROS play a crucial role in HCC progression, influencing both tumor development and therapeutic responses. Sorafenib, a chemotherapeutic drug for HCC, exerts its effects through the production of O<sub>2</sub><sup>-</sup> (superoxide anion). Sorafenib induces ROS generation, specifically O<sub>2</sub><sup>-</sup>, in tumor and endothelial cells which produce irreversible cellular damage such as cell death or inhibition of proliferation. O<sub>2</sub><sup>-</sup> is produced by NADPH oxidase, which is activated by sorafenib and mediated by VEGF, contributing to ROS production and tumor hypoxia. In patients, the concentration of advanced oxidative protein products (AOPP) in serum, a marker of oxidative stress, is correlated with sorafenib efficacy. A serum AOPP level of ≥ 0.2 μmol/L is a good predictor of patient survival and treatment response (Coriat et al., 2012). Additionally, it has been observed that the combination of Agrimol B with sorafenib, inhibits the growth and proliferation of HCC cells, inducing both apoptosis and autophagic cell death. Specifically, Agrimol B, a compound derived from *Agrimonia pilosa Ledeb.*, induces autophagic cytotoxic in HCC by negatively regulating NADH: ubiquinone oxidoreductase subunit 1 (NDUFS1) which produces an accumulation of

mitochondrial ROS (Dong et al., 2024).

Oxaliplatin treatment induces autophagy in HCC cells, by autophagosomes generation and the accumulation of LC3-II. However, survival oxaliplatin-treated cells have shown higher metastatic potential. Inhibiting autophagy with CQ and 3-methylamphetamine enhances cell death induced by oxaliplatin both *in vivo* and *in vitro* suggesting that autophagy contributes to tumor cell survival treated with oxaliplatin. Drug resistance to platinum compounds can be correlated with reduced ROS, and autophagy helps to delete damaged proteins and organelles generated by ROS. Inhibiting autophagy combined with oxaliplatin increases ROS generation and potentiates treatment cytotoxicity (Ciccarone et al., 2019).

Xu et al. (2021) investigated the role of URI1, a protein overexpressed in HCC cells exposed to irradiation, which appears to inhibit radiotherapy-induced apoptosis. URI1 reduces ROS levels induced by irradiation, suggesting that URI1 can protect tumor cells against oxidative stress and contributes to HCC radioresistance. Furthermore, URI1 activates autophagy to eliminate excess ROS and maintain redox balance, thereby protecting cells from oxidative damage. This process is regulated by AMPK/FOXO3 signaling pathway, where URI1 promotes AMPK and FOXO3 activation, favoring autophagy. These findings suggest that modulating the URI1 pathway could enhance radiotherapy treatments in HCC (Xu et al., 2021).

Dihydromyricetin (DHM), a natural flavonoid, induces apoptosis in a concentration-dependent manner, reducing cellular proliferation without affecting normal hepatic cells. This effect is correlated with a decrease in ROS production in HCC cells, disrupting redox balance, blocking ROS-mediated transduction signals and promoting apoptosis. Additionally, DHM decreases GSH levels and ATP in tumor cells, while inducing heme oxygenase-1 expression (HO-1) which further favors apoptosis. ROS imbalance plays a key role in the apoptosis process induced by DHM, as even a small amount of ROS production is essential for cancer cell growth and that an imbalance in the cellular environment can trigger cell death in HCC cells (Liu et al., 2014).

Another molecule with potential interest for HCC therapy is FDX1, which plays a crucial role in cellular metabolism and HCC progression. FDX1 converts  $\text{Cu}^{2+}$  to  $\text{Cu}^{1+}$  and it is involved in diverse tumoral process, including proliferation, migration and tumor cell invasion. Its deficiency provokes metabolic alterations in mitochondria generating oxidative stress and activating mitophagy by PINK1/Parkin pathway. Additionally, FDX1 interacts with the PI3K/AKT pathway, which is associated with autophagy in malignant tumors, promoting HCC progression when its expression is low. From a therapeutic perspective, FDX1 is a tumor suppressor, and its activation can represent a potential strategy against HCC. Currently, diverse options to enhance its function are being explored which involve: 1) the use of copper ionophores as elesclomol, drugs that induce its expression (CX-5461, temozolomide or erlotinib among others), and 2) strategies for redox balance regulation through ROS scavengers. Moreover, FDX1 role in immunomodulation suggests that its combination with immunotherapy could improve treatment response and patients' survival (Sun et al., 2024). On the other hand, patients treated with incomplete radiofrequency ablation (iRFA), a therapy used for HCC, can achieve outcomes comparable to surgical resection when the tumor diameter is  $< 1$  cm. However, HCC recurrence after this treatment remains a major challenge and is associated with poor prognosis, however HCC recurrence after this treatment remains a major challenge and is associated with poor prognosis. In the short term, after treatment, ROS levels increase and have a pro-survival effect. NADPH oxidase (NOX4), located in mitochondria, plays an important role in mitochondrial ROS production induced by thermal stress. This ROS increase activates mitophagy dependent on PINK1, promoting the deletion of damaged mitochondria and avoiding apoptosis. NOX4 inhibitors reduce mitochondrial ROS, highlighting its key role in response to thermal stress. Additionally, the transcription factor Nrf2 is activated in response to oxidative and thermal stress, promoting metabolic reprogramming and increased glycolysis and the regulation of key genes

like PINK1. Nrf2 could also be involved in ferroptosis resistance after iRFA, although this aspect requires further investigation (Peng et al., 2023).

Caryophyllene oxide, a chemical compound found in essential oils, increased ROS formation and reduced intracellular antioxidants such as NRF2, HO-1, GPX4, and NQO1, which protect cells from ferroptosis. Additionally, caryophyllene oxide increased  $\text{Fe}^{2+}$  levels, promoting lipid peroxidation and cellular damage. It also regulated ferritinophagy by enhancing NCOA4 and LC3 II expression, facilitating ferritin autophagy and reducing FTH1 levels, thus contributing to ferroptosis (Xiu et al., 2022). (Table 5)

**Table 5**  
Therapeutic strategies in HCC.

Therapeutic strategies	Molecular mechanism	References
<b>Clinical study: Sorafenib</b>	Induces ROS ( $\text{O}_2^-$ ) via NADPH oxidase activation, leading to tumor hypoxia and cellular damage. Serum advanced oxidative protein products (AOPP) levels correlate with treatment efficacy	(Coriat et al., 2012)
<b>Preclinical study: Agrimol B</b>	Inhibits NDUFS1, leading to mitochondrial ROS accumulation and autophagic cytotoxicity. Enhances apoptosis and autophagy when combined with sorafenib	(Dong et al., 2024)
<b>Preclinical study: Oxaliplatin + autophagy inhibitors (CQ, 3-MA)</b>	Induces autophagy (LC3-II accumulation) and autophagy inhibition increases ROS, enhancing oxaliplatin cytotoxicity	(Ciccarone et al., 2019)
<b>Preclinical study: URI1 inhibition</b>	URI1 overexpression reduces ROS and promotes autophagy via AMPK/FOXO3, contributing to radiotherapy resistance. Inhibiting URI1 may improve radiotherapy efficacy.	(Xu et al., 2021)
<b>Preclinical study: Dihydromyricetin (DHM)</b>	Induces apoptosis by disrupting redox balance, reducing ROS, ATP, and GSH, and increasing HO-1 expression	(Liu et al., 2014)
<b>Preclinical study: FDX1 activation</b>	Regulates $\text{Cu}^{2+}$ to $\text{Cu}^{1+}$ conversion, affecting metabolism, oxidative stress, and mitophagy (PINK1/Parkin pathway). Low FDX1 promotes HCC progression. Therapeutic approaches include copper ionophores and redox balance regulators	(Sun et al., 2024)
<b>Preclinical/clinical study: Incomplete radiofrequency ablation (iRFA) + NOX4/Nrf2 inhibition</b>	iRFA-induced ROS activates NOX4, leading to mitophagy (PINK1-dependent) and ferroptosis resistance via Nrf2. NOX4 inhibition reduces mitochondrial ROS and may improve outcomes	(Peng et al., 2023)
<b>Preclinical study: Caryophyllene oxide</b>	Caryophyllene oxide promotes ferroptosis by increasing ROS and $\text{Fe}^{2+}$ levels, reducing key antioxidants, and enhancing ferritinophagy through NCOA4 and LC3 II upregulation, leading to ferritin degradation	(Xiu et al., 2022)

## 5. Challenges and future directions

### 5.1. Therapeutic targeting and challenges in autophagy modulation

The modulation of autophagy is a promising approach in cancer therapy, targeting its dual role in tumor biology. However, this approach often causes unintended toxicity in normal tissues, although inhibiting autophagy can suppress its pro-survival functions in cancer cells. The optimal balance between risks and benefits requires further study to minimize side effects while maximizing therapeutic efficacy (Taucher et al., 2022). Developing selective inhibitors that target autophagy specifically in malignant cells without affecting normal cells remains a major challenge (Lim and Murthy, 2020). Identifying robust biomarkers to predict which patients will benefit most is critical for establishing this selectivity (Taucher et al., 2022). Notably, candidate biomarkers such as LC3, p62, and Nrf2-regulated genes offer promising avenues for patient stratification and monitoring autophagic and oxidative stress activity in tumors.

### 5.2. Complex interactions between autophagy, oxidative stress, and tumor biology

Autophagy's role in cancer is context-dependent, acting as either tumor suppressor or promoter based on tumor type, stage, and genetics (Debnath et al., 2023). This duality complicates therapy development and underscores the need for detailed understanding of autophagy's function in specific cancer contexts. The interplay between autophagy and oxidative stress is complex and dynamic, involving feedback loops and signaling pathways. Oxidative stress can stimulate or be attenuated by autophagy, forming a balance that influences cancer progression and treatment response (Debnath et al., 2023; Taucher et al., 2022). Targeted research is essential to dissect these interactions and refine therapies.

### 5.3. Exploration of combination therapies

Combining autophagy modulators with chemotherapy, radiotherapy, or novel targeted therapies shows synergistic potential to improve efficacy (Lim and Murthy, 2020). For instance, co-targeting autophagy with oxidative stress may enhance therapeutic outcomes by reducing cellular ROS levels and increasing cancer cell death. Furthermore, since autophagy shapes the tumor immune microenvironment, integrating autophagy modulation with immunotherapy is promising. Understanding these interactions could open the way to novel combination regimens optimizing immune response and suppressing tumor growth.

### 5.4. Advancing research tools and methodologies

Progress in this area depends on better experimental tools. Advanced genetic models are needed to clarify the role of specific ATGs in cancer progression and resistance to therapy (Lim and Murthy, 2020). Improved imaging techniques that allow real-time visualization and quantification of autophagy in living tissues will support characterization of its dynamic response to therapy (Taucher et al., 2022).

### 5.5. Considerations for antioxidant therapies in cancer treatment

The use of antioxidants in cancer therapy adds another layer of complexity. Antioxidants may interfere with chemotherapeutic agents that rely on oxidative mechanisms to induce cancer cell death (Jiang et al., 2023; Marioli-Sapsakou and Kourti, 2021). Therefore, antioxidant supplementation must be carefully balanced to avoid counterproductive effects. Antioxidants can protect normal cells but potentially promote cancer cell survival and recurrence (Jiang et al., 2023; Singh et al., 2018). However, antioxidant doses affect cells differently. Low doses

may protect healthy and tumor cells, while high doses may stop tumor growth. Finding the best dose for treatment while avoiding cancer cell survival remains a challenge (Singh et al., 2018). Effects vary across cancer types and treatments (Marioli-Sapsakou and Kourti, 2021; Singh et al., 2018).

### 5.6. Future directions

To fully harness autophagy modulation in cancer therapy and improve patient outcomes while reducing risks, future research should focus on overcoming key challenges. Firstly, it is crucial to develop personalized strategies that tailor autophagy and oxidative stress modulation based on individual tumor characteristics and genetic profiles. Additionally, exploring the synergistic effects of combining autophagy modulators with traditional chemotherapies or targeted therapies could enhance treatment efficacy. Reliable biomarkers need to be identified to predict and monitor the efficacy of therapies based on autophagy and oxidative stress modulation. Moreover, the discovery of new compounds that selectively modulate autophagy or oxidative stress pathways in cancer cells is key to developing more effective treatments. A deeper understanding of the molecular mechanisms underlying the interplay between autophagy, oxidative stress, and cancer progression is also required. This will enable the design of more specific and effective therapies. Furthermore, determining the optimal timing for autophagy modulation in different cancer types and stages is crucial to maximizing therapeutic benefit. Finally, investigating the tumor microenvironment's role in these processes is crucial to improve treatment response. Together, these research efforts are vital to advance autophagy modulation as a cancer therapeutic strategy.

## 6. Conclusions

Autophagy and oxidative stress play crucial and often contradictory roles in the development and progression of solid tumors. While autophagy can act as a protector mechanism by eliminating damaged organelles and reducing cellular damage, it can also promote tumor cell survival under stress conditions promoting drug resistance. On the other hand, oxidative stress, characterized by the production of ROS, can induce cellular damage and apoptosis. However, it is a double-edged sword, and it can also stimulate tumor cell proliferation and adaptation.

In solid tumors such as BC, LC, CRC, PCa or HCC, the interaction between autophagy and oxidative stress influences the progression and resistance to chemotherapy and immune evasion. From a therapeutic perspective, regulation of autophagy and oxidative stress is a key strategy to enhance the efficacy of cancer treatments. Autophagy inhibition has been explored to increase tumor susceptibility to chemotherapy and radiotherapy, at the same time, oxidative stress modulation by antioxidants or prooxidants substances can be used to induce tumor programmed cell death. However, due to the complexity of these processes and their dual roles, a personalized approach is necessary to determine whether their activation or inhibition would be more beneficial for each specific cancer type.

## Funding

This work was partially supported by Instituto de Salud Carlos III – ISCIII (Spanish Government) co-funded by FEDER funds/European Regional Development Fund (ERDF) - a way to build Europe (PI20/01845). Additional support came from the Consejería de Transformación Económica, Industria, Conocimiento y Universidades and FEDER (PY20/01282). Funding for open access charge: Universidad de Granada/CBUA.

## CRediT authorship contribution statement

Conceptualization, A.J.C.-S., J.M.S.-M., F.J.R.Z. and J.S.;

methodology, A.J.C.-S., J.M.S.-M, M.C, C.G.O. and J.S.; writing—original draft preparation, A.J.C.-S., J.M.S.-M., C.G.O., M.C., L.D.-B., F.J.R.Z. and J.S.; writing—review and editing, all authors; Illustration, F.J.R.Z.; supervision, F.J.R.Z. and J.S.; project administration, J.S.; funding acquisition, J.S. All authors have read and agreed to the published version of the manuscript.

## Declaration of Competing Interest

The authors declare no conflict of interest. The funders had no role in the writing of the manuscript, or in the decision to publish the reviewed material.

## References

- Adamaki, M., Zoumpourlis, V., 2021. Prostate cancer biomarkers: from diagnosis to prognosis and precision-guided therapeutics. *Pharmacol. Ther.* 228, 107932.
- Aita, V.M., Liang, X.H., Murty, V.V.S., Pincus, D.L., Yu, W., Cayanis, E., Kalachikov, S., Gilliam, T.C., Levine, B., 1999. Cloning and genomic organization of beclin 1, a candidate tumor suppressor gene on chromosome 17q21. *Genomics* 59, 59–65.
- Alizadeh, J., Glogowska, A., Thliveris, J., Kalantari, F., Shojaei, S., Hombach-Klonisch, S., Klonisch, T., Ghavami, S., 2018. Autophagy modulates transforming growth factor beta 1 induced epithelial to mesenchymal transition in non-small cell lung cancer cells. *Biochim. Biophys. Acta Mol. Cell Res.* 1865, 749–768.
- Alli, E., Ford, J.M., 2012. Breast cancers with compromised DNA repair exhibit selective sensitivity to elesclomol. *DNA Repair (Amst.)* 11, 522–524.
- Alzuwaidi, S., Khalil, R., 2019. Mechanism of arsenic trioxide inhibition of cell growth and induction of apoptosis in triple negative MDA-MB-231 breast cancer cell line. *FASEB J.* 33. [https://doi.org/10.1096/fasebj.2019.33.1\\_supplement.lb94](https://doi.org/10.1096/fasebj.2019.33.1_supplement.lb94).
- An, X., Yu, W., Liu, J., Tang, D., Yang, L., Chen, X., 2024. Oxidative cell death in cancer: mechanisms and therapeutic opportunities. *Cell Death Dis.* 15, 556.
- Aqbi, H.F., Tyutyunyk-Massey, L., Keim, R.C., Butler, S.E., Thekkudan, T., Joshi, S., Smith, T.M., Bandyopadhyay, D., Idowu, M.O., Bear, H.D., Payne, K.K., Gewirtz, D.A., Manjili, M.H., 2018. Autophagy-deficient breast cancer shows early tumor recurrence and escape from dormancy. *Oncotarget* 9, 22113–22122.
- Arslan, P., Di Virgilio, F., Beltrame, M., Tsien, R.Y., Pozzan, T., 1985. Cytosolic Ca<sup>2+</sup> homeostasis in Ehrlich and Yoshida carcinomas. A new, membrane-permeant chelator of heavy metals reveals that these ascites tumor cell lines have normal cytosolic free Ca<sup>2+</sup>. *J. Biol. Chem.* 260, 2719–2727.
- Aruljothi, K.N., Kumaran, K., Senthil, S., Nidhu, A.B., Munaff, N., Janitri, V.B., Kirubakaran, R., Singh, S.K., Gupt, G., Dua, K., Krishnan, A., 2022. Implications of reactive oxygen species in lung cancer and exploiting it for therapeutic interventions. *Med. Oncol.* 40, 43.
- Ashrafizadeh, M., Paskah, M.D.A., Mirzaei, S., Gholami, M.H., Zarrabi, A., Hashemi, F., Hushmandi, K., Hashemi, M., Nabavi, N., Crea, F., Ren, J., Klionsky, D.J., Kumar, A.P., Wang, Y., 2022. Targeting autophagy in prostate cancer: preclinical and clinical evidence for therapeutic response. *J. Exp. Clin. Cancer Res.* 41, 105.
- Avivar-Valderas, A., Salas, E., Bobrovnikova-Marjon, E., Diehl, J.A., Nagi, C., Debnath, J., Aguirre-Ghiso, J.A., 2011. PERK integrates autophagy and oxidative stress responses to promote survival during extracellular matrix detachment. *Mol. Cell. Biol.* 31, 3616–3629.
- Behrends, C., Sowa, M.E., Gygi, S.P., Harper, J.W., 2010. Network organization of the human autophagy system. *Nature* 466, 68–76.
- Bergengren, O., Pekala, K.R., Matsoukas, K., Fainberg, J., Mungovan, S.F., Bratt, O., Bray, F., Brawley, O., Luckenbaugh, A.N., Mucci, L., Morgan, T.M., Carlsson, S.V., 2023. 2022 update on prostate cancer epidemiology and risk factors—a systematic review. *Eur. Urol.* 84, 191–206.
- Biswas, P., Swaroop, S., Dutta, N., Arya, A., Ghosh, S., Dhabal, S., Das, P., Majumder, C., Pal, M., Bhattacharjee, A., 2023. IL-13 and the hydroperoxy fatty acid 13(S)HpODE play crucial role in inducing an apoptotic pathway in cancer cells involving MAO-A/ROS/p53/p21 signaling axis. *Free Radic. Biol. Med.* 195, 309–328.
- Bray, F., Laversanne, M., Sung, H., Ferlay, J., Siegel, R.L., Soerjomataram, I., Jemal, A., 2024. Global cancer statistics 2022: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. *CA Cancer J. Clin.* 74, 229–263.
- Buccarelli, M., D'Alessandris, Q.G., Matarrese, P., Mollinari, C., Signore, M., Cappannini, A., Martini, M., D'Aliberti, P., De Luca, G., Pedini, F., Boe, A., Biffoni, M., Pallini, R., Ricci-Vitiani, L., 2021. Elesclomol-induced increase of mitochondrial reactive oxygen species impairs glioblastoma stem-like cell survival and tumor growth. *J. Exp. Clin. Cancer Res.* 40, 228.
- Budanov, A.V., 2011. Stress-responsive sestrins link p53 with redox regulation and mammalian target of rapamycin signaling. *Antioxid. Redox Signal* 15, 1679–1690.
- Budanov, A.V., Karin, M., 2008. p53 target genes sestrin1 and sestrin2 connect genotoxic stress and mTOR signaling. *Cell* 134, 451–460.
- Budanov, A.V., Lee, J.H., Karin, M., 2010. Stressin' sestrins take an aging fight. *EMBO Mol. Med.* 2, 388–400.
- Budisan, L., Zanoaga, O., Braicu, C., Pirllog, R., Covaliu, B., Esanu, V., Korban, S.S., Berindan-Neagoe, I., 2021. Links between infections, lung cancer, and the immune system. *Int. J. Mol. Sci.* 22, 9394.
- Bu, H., Liu, D., Zhang, G., Chen, L., Song, Z., 2020. AMPK/mTOR/ULK1 axis-mediated pathway participates in apoptosis and autophagy induction by oridonin in colon cancer DLD-1 cells. *Onco. Targets Ther.* 13, 8533–8545.
- Cabrera-Serrano, A.J., Sánchez-Maldonado, J.M., González-Olmedo, C., Carretero-Fernández, M., Díaz-Beltrán, L., Gutiérrez-Bautista, J.F., García-Verdejo, F.J., Gálvez-Montosa, F., López-López, J.A., García-Martín, P., Pérez, E.M., Sánchez-Rovira, P., Reyes-Zurita, F.J., Sainz, J., 2025. Crosstalk between autophagy and oxidative stress in hematological malignancies: mechanisms, implications, and therapeutic potential. *Antioxid. (Basel)* 14. <https://doi.org/10.3390/antiox14030264>.
- Catalano, T., Selvaggi, F., Cotelleso, R., Aceto, G.M., 2025. The role of reactive oxygen species in colorectal cancer initiation and progression: perspectives on therapeutic approaches. *Cancers (Basel)* 17. <https://doi.org/10.3390/cancers17050752>.
- Cat, B., Stuhlmann, D., Steinbrenner, H., Alili, L., Holtkötter, O., Sies, H., Brenneisen, P., 2006. Enhancement of tumor invasion depends on transdifferentiation of skin fibroblasts mediated by reactive oxygen species. *J. Cell Sci.* 119, 2727–2738.
- Chang, L.-L., Li, Y.-K., Zhao, C.-X., Zeng, C.-M., Ge, F.-J., Du, J.-M., Zhang, W.-Z., Lu, P.-H., He, Q.-J., Zhu, H., Yang, B., 2022. AKR1C1 connects autophagy and oxidative stress by interacting with SQSTM1 in a catalytic-independent manner. *Acta Pharmacol. Sin.* 43, 703–711.
- Chavez-Dominguez, R., Perez-Medina, M., Lopez-Gonzalez, J.S., Galicia-Velasco, M., Aguilar-Cazares, D., 2020. The double-edge sword of autophagy in cancer: from tumor suppression to pro-tumor activity. *Front. Oncol.* 10, 578418.
- Chen, H., 2023. A review of the recent research progress on risk factors of lung cancer. *Theor. Nat. Sci.* 21, 291–295.
- Chen, J., Zhang, L., Zhou, H., Wang, W., Luo, Y., Yang, H., Yi, H., 2018. Inhibition of autophagy promotes cisplatin-induced apoptotic cell death through Atg5 and Beclin 1 in A549 human lung cancer cells. *Mol. Med. Rep.* <https://doi.org/10.3892/mmr.2018.8686>.
- Chen, K., Shang, Z., Dai, A.-L., Dai, P.-L., 2020. Novel PI3K/Akt/mTOR pathway inhibitors plus radiotherapy: strategy for non-small cell lung cancer with mutant RAS gene. *Life Sci.* 255, 117816.
- Chen, Y., McMillan-Ward, E., Kong, J., Israels, S.J., Gibson, S.B., 2008. Oxidative stress induces autophagic cell death independent of apoptosis in transformed and cancer cells. *Cell Death Differ.* 15, 171–182.
- Chen, Z., Jiang, Q., Zhu, P., Chen, Y., Xie, X., Du, Z., Jiang, L., Tang, W., 2019. NPRL2 enhances autophagy and the resistance to Everolimus in castration-resistant prostate cancer. *Prostate* 79, 44–53.
- Che, Z., Zhou, Z., Li, S.-Q., Gao, L., Xiao, J., Wong, N.-K., 2023. ROS/RNS as molecular signatures of chronic liver diseases. *Trends Mol. Med.* 29, 951–967.
- Chinnappa, S., Bakthavachalam, V., Muniarathnam, G., 2020. Repurposing antidepressant sertraline as a pharmacological drug to target prostate cancer stem cells: dual activation of apoptosis and autophagy signaling by deregulating redox balance. *Am. J. Cancer Res.* 10, 2043–2065.
- Choi, K.S., 2012. Autophagy and cancer. *Exp. Mol. Med.* 44, 109–120.
- Choi, K.S., Yan, C., Li, T.S., 2012. Autophagy and autophagy-related proteins in cancer. *Anticancer Res.* 44.
- Chow, S.K.Y., Chan, J.Y.W., Fung, K.P., 2004. Inhibition of cell proliferation and the action mechanisms of arsenic trioxide (As<sub>2</sub>O<sub>3</sub>) on human breast cancer cells. *J. Cell. Biochem.* 93, 173–187.
- Ciccarone, F., Castelli, S., Ciriolo, M.R., 2019. Oxidative stress-driven autophagy across onset and therapeutic outcome in hepatocellular carcinoma. *Oxid. Med. Cell. Longev.* 2019, 6050123.
- Cichoż-Lach, H., Michalak, A., 2014. Oxidative stress as a crucial factor in liver diseases. *World J. Gastroenterol.* 20, 8082–8091.
- Corazzari, M., Rapino, F., Ciccosanti, F., Giglio, P., Antonioli, M., Conti, B., Fimia, G.M., Lovat, P.E., Piacentini, M., 2015. Oncogenic BRAF induces chronic ER stress condition resulting in increased basal autophagy and apoptotic resistance of cutaneous melanoma. *Cell Death Differ.* 22, 946–958.
- Coriat, R., Nicco, C., Chéreau, C., Mir, O., Alexandre, J., Ropert, S., Weill, B., Chaussade, S., Goldwasser, F., Batteux, F., 2012. Sorafenib-induced hepatocellular carcinoma cell death depends on reactive oxygen species production in vitro and in vivo. *Mol. Cancer Ther.* 11, 2284–2293.
- da Silva, E.L., Mesquita, F.P., Ramos, I.N., de, F., Gomes, C.B., de, S.M.R., Moreira, C., dos, S., Ferreira, V.F., da Rocha, D.R., Bahia, M., de, O., Moreira-Nunes, C.A., de Souza, C.R.T., Burbano, R.M.R., Montenegro, R.C., 2022. Antitumoral effect of novel synthetic 8-hydroxy-2-((4-nitrophenyl)thio)naphthalene-1,4-dione (CNN16) via ROS-mediated DNA damage, apoptosis and anti-migratory effect in colon cancer cell line. *Toxicol. Appl. Pharm.* 456, 116256.
- Daskalaki, I., Gkikas, I., Tavernarakis, N., 2018. Hypoxia and selective autophagy in cancer development and therapy. *Front. Cell Dev. Biol.* 6, 104.
- Debnath, J., Gammoh, N., Ryan, K.M., 2023. Autophagy and autophagy-related pathways in cancer. *Nat. Rev. Mol. Cell Biol.* 24, 560–575.
- de Groot, P.M., Wu, C.C., Carter, B.W., Munden, R.F., 2018. The epidemiology of lung cancer. *Transl. Lung Cancer Res.* 7, 220–233.
- Deng, H., Chen, Y., Wang, L., Zhang, Y., Hang, Q., Li, P., Zhang, P., Ji, J., Song, H., Chen, M., Jin, Y., 2023. PI3K/mTOR inhibitors promote G6PD autophagic degradation and exacerbate oxidative stress damage to radiosensitive small cell lung cancer. *Cell Death Dis.* 14, 652.
- Deng, S., Ramos-Castaneda, M., Velasco, W.V., Clowers, M.J., Gutierrez, B.A., Noble, O., Dong, Y., Zarghooni, M., Alvarado, L., Caetano, M.S., Yang, S., Ostrin, E.J., Behrens, C., Wistuba, I.I., Stabile, L.P., Kadara, H., Watowich, S.S., Moghaddam, S.J., 2020. Interplay between estrogen and Stat3/NF-κB-driven immunomodulation in lung cancer. *Carcinogenesis* 41, 1529–1542.
- Dey, D.K., Kang, S.C., 2021. CopA3 peptide induces permanent cell-cycle arrest in colorectal cancer cells. *Mech. Ageing Dev.* 196, 111497.
- Ding, Y., Wang, H., Niu, J., Luo, M., Gou, Y., Miao, L., Zou, Z., Cheng, Y., 2016. Induction of ROS overload by alantolactone prompts oxidative DNA damage and apoptosis in colorectal cancer cells. *Int. J. Mol. Sci.* 17, 558.

- Divella, R., Mazzocca, A., Daniele, A., Sabbà, C., Paradiso, A., 2019. Obesity, nonalcoholic fatty liver disease and adipocytokines network in promotion of cancer. *Int. J. Biol. Sci.* 15, 610–616.
- Dodson, M., Redmann, M., Rajasekaran, N.S., Darley-Usmar, V., Zhang, J., 2015. KEAP1-NRF2 signalling and autophagy in protection against oxidative and reductive proteotoxicity. *Biochem. J.* 469, 347–355.
- Donato, D., Giovannelli, P., Migliaccio, A., Bilancio, A., 2022. Inhibition of Vps34 and p110delta PI3K impairs migration, invasion and three-dimensional spheroid growth in breast cancer cells. *Int. J. Mol. Sci.* <https://doi.org/10.3390/ijms23169008>.
- Dong, L., He, J., Luo, L., Wang, K., 2023. Targeting the interplay of autophagy and ROS for cancer therapy: an updated overview on phytochemicals. *Pharm. (Basel)* 16, 92.
- Dong, L., Luo, L., Wang, Z., Lian, S., Wang, M., Wu, X., Fan, J., Zeng, Y., Li, S., Lv, S., Yang, Y., Chen, R., Shen, E., Yang, W., Li, C., Wang, K., 2024. Targeted degradation of NDUFS1 by agrimol B promotes mitochondrial ROS accumulation and cytotoxic autophagy arrest in hepatocellular carcinoma. *Free Radic. Biol. Med.* 220, 111–124.
- Dong, Y., Zhang, X., 2024. Targeting cellular mitophagy as a strategy for human cancers. *Front. Cell Dev. Biol.* 12, 1431968.
- Dorval, J., Hontela, A., 2003. Role of glutathione redox cycle and catalase in defense against oxidative stress induced by endosulfan in adrenocortical cells of rainbow trout (*Oncorhynchus mykiss*). *Toxicol. Appl. Pharm.* 192, 191–200.
- Drosten, M., Barbacid, M., 2022. Targeting KRAS mutant lung cancer: light at the end of the tunnel. *Mol. Oncol.* 16, 1057–1071.
- Duan, J., Yu, Y., Yu, Y., Li, Y., Wang, J., Geng, W., Jiang, L., Li, Q., Zhou, X., Sun, Z., 2014. Silica nanoparticles induce autophagy and endothelial dysfunction via the PI3K/Akt/mTOR signaling pathway. *Int. J. Nanomed.* 9, 5131–5141.
- Duan, W.-J., Li, Q.-S., Xia, M.-Y., Tashiro, S.-I., Onodera, S., Ikejima, T., 2011. Silibinin activated p53 and induced autophagic death in human fibrosarcoma HT1080 cells via reactive oxygen species-p38 and c-Jun N-terminal kinase pathways. *Biol. Pharm. Bull.* 34, 47–53.
- Đuračková, Z., 2010. Some current insights into oxidative stress. *Physiol. Res.* 59, 459–469.
- Dwyer, S., Ruth, J., Seidel, H.E., Raz, A.A., Chodosh, L.A., 2024. Autophagy is required for mammary tumor recurrence by promoting dormant tumor cell survival following therapy. *Breast Cancer Res* 26, 143.
- El Hout, M., Cosialls, E., Mehrpour, M., Hamai, A., 2020. Crosstalk between autophagy and metabolic regulation of cancer stem cells. *Mol. Cancer* 19, 27.
- Elshazly, A.M., Gewirtz, D.A., 2023. Making the case for autophagy inhibition as a therapeutic strategy in combination with androgen-targeted therapies in prostate cancer. *Cancers (Basel)* 15. <https://doi.org/10.3390/cancers15205029>.
- Feng, X., Jiang, T., Yang, C., Pang, S., Ding, Z., Hu, H., Wang, H., Dong, L., Yang, N., 2021. RPRD1A stabilizes NRF2 and aggravates HCC progression through competing with p62 for TRIM21 binding. *Cell Death Dis.* 13, 6.
- Filomeni, G., De Zio, D., Ceconi, F., 2015. Oxidative stress and autophagy: the clash between damage and metabolic needs. *Cell Death Differ.* 22, 377–388.
- Galluzzi, L., Pietrocola, F., Bravo-San Pedro, J.M., Amaravadi, R.K., Baehrecke, E.H., Ceconi, F., Codogno, P., Debnath, J., Gewirtz, D.A., Karantza, V., Kimmelman, A., Kumar, S., Levine, B., Maiuri, M.C., Martin, S.J., Penninger, J., Piacentini, M., Rubinsztein, D.C., Simon, H.-U., Simonsen, A., Thorburn, A.M., Velasco, G., Ryan, K.M., Kroemer, G., 2015. Autophagy in malignant transformation and cancer progression. *EMBO J.* 34, 856–880.
- Gan, L., Wang, W., Jiang, J., Tian, K., Liu, W., Cao, Z., 2024. Dual role of Nrf2 signaling in hepatocellular carcinoma: promoting development, immune evasion, and therapeutic challenges. *Front. Immunol.* 15, 1429836.
- Gao, H., Gong, X.C., Chen, Z.D., Xu, X.S., Zhang, Q., Xu, X.M., 2014. Induction of apoptosis in hormone-resistant human prostate cancer PC3 cells by inactivated Sendai virus. *Biomed. Environ. Sci.* 27, 506–514.
- Gao, L., Chen, Y., 2021. Autophagy controls programmed death-ligand 1 expression on cancer cells (Review). *Biomed. Rep.* 15, 84.
- Ghazi, P.C., O'Toole, K.T., Srinivas Boggaram, S., Scherzer, M.T., Silvis, M.R., Zhang, Y., Bogdan, M., Smith, B.D., Lozano, G., Flynn, D.L., Snyder, E.L., Kinsey, C.G., McMahon, M., 2024. Inhibition of ULK1/2 and KRASG12C controls tumor growth in preclinical models of lung cancer. *Elife* 13. <https://doi.org/10.7554/elifesciences.96992.3>.
- Glorieux, C., Calderon, P.B., 2018. Catalase down-regulation in cancer cells exposed to arsenic trioxide is involved in their increased sensitivity to a pro-oxidant treatment. *Cancer Cell Int* 18. <https://doi.org/10.1186/s12935-018-0524-0>.
- Gonzalez, Y., Aryal, B., Chehab, L., Rao, V.A., 2014. Atg7- and Keap1-dependent autophagy protects breast cancer cell lines against mitochon-drial oxidative stress. *Oncotarget* 5, 1526–1537.
- Gorrini, C., Harris, I.S., Mak, T.W., 2013. Modulation of oxidative stress as an anticancer strategy. *Nat. Rev. Drug Discov.* 12, 931–947.
- Grandvalet, C., Feugeas, J.P., Monnier, F., Despouy, G., Valérie, P., Michaël, G., Hervouet, E., Peixoto, P., 2020. Autophagy is associated with a robust specific transcriptional signature in breast cancer subtypes. *Genes Cancer* 11, 154–168.
- Grivninkos, S.I., Karin, M., 2010. Dangerous liaisons: STAT3 and NF-κB collaboration and crosstalk in cancer. *Cytokine Growth Factor Rev.* 21, 11–19.
- Guo, L.W., Zhang, X.L., Cai, L., Zhu, C.X., Fang, Y., Yang, H.Y., Chen, H.D., 2024. Current status of global colorectal cancer prevalence, prevention and control. *Zhonghua Zhong Liu Za Zhi* 46, 57–65.
- Guo, W., Du, K., Luo, S., Hu, D., 2022. Recent advances of autophagy in non-small cell lung cancer: from basic mechanisms to clinical application. *Front. Oncol.* 12, 861959.
- Guo, Y., Han, B., Luo, K., Ren, Z., Cai, L., Sun, L., 2017. NOX2-ROS-HIF-1α signaling is critical for the inhibitory effect of oleoic acid on rectal cancer cell proliferation. *Biomed. Pharm.* 85, 733–739.
- Hahn, E.-R., Sakao, K., Singh, S.V., 2014. Honokiol activates reactive oxygen species-mediated cytoprotective autophagy in human prostate cancer cells. *Prostate* 74, 1209–1221.
- Han, Q., Deng, Y., Chen, S., Chen, R., Yang, M., Zhang, Z., Sun, X., Wang, W., He, Y., Wang, F., Pan, X., Li, P., Lai, W., Luo, H., Huang, P., Guan, X., Deng, Y., Yan, J., Xu, X., Wen, Y., Chen, A., Hu, C., Li, X., Li, S., 2017. Downregulation of ATG5-dependent macroautophagy by chaperone-mediated autophagy promotes breast cancer cell metastasis. *Sci. Rep.* 7, 4759.
- Han, Q., Ma, Y., Wang, H., Dai, Y., Chen, C., Liu, Y., Jing, L., Sun, X., 2018. Resibufogenin suppresses colorectal cancer growth and metastasis through RIP3-mediated necroptosis. *J. Transl. Med.* 16, 201.
- Hasan, A., Rizvi, S.F., Parveen, S., Pathak, N., Nazir, A., Mir, S.S., 2022. Crosstalk between ROS and autophagy in tumorigenesis: understanding the multifaceted paradox. *Front. Oncol.* 12, 852424.
- Hayes, J.D., Dinkova-Kostova, A.T., Tew, K.D., 2020. Oxidative stress in cancer. *Cancer Cell* 38, 167–197.
- Hecht, S.S., 2011. Tobacco smoke carcinogens and lung cancer. in: *Chemical Carcinogenesis*. Humana Press, Totowa, NJ, pp. 53–74.
- He, Y., She, H., Zhang, T., Xu, H., Cheng, L., Yepes, M., Zhao, Y., Mao, Z., 2018. p38 MAPK inhibits autophagy and promotes microglial inflammatory responses by phosphorylating ULK1. *J. Cell Biol.* 217, 315–328.
- Hong, W., Cai, P., Xu, C., Cao, D., Yu, W., Zhao, Z., Huang, M., Jin, J., 2018. Inhibition of glucose-6-phosphate dehydrogenase reverses cisplatin resistance in lung cancer cells via the redox system. *Front. Pharm.* 9, 43.
- Hsieh, C.-L., Liu, C.-M., Chen, H.-A., Yang, S.-T., Shigemura, K., Kitagawa, K., Yamamichi, F., Fujisawa, M., Liu, Y.-R., Lee, W.-H., Chen, K.-C., Shen, C.-N., Lin, C.-C., Chung, L.W.K., Sung, S.-Y., 2017. Reactive oxygen species-mediated switching expression of MMP-3 in stromal fibroblasts and cancer cells during prostate cancer progression. *Sci. Rep.* 7, 9065.
- Huang, X., Wan, J., Leng, D., Zhang, Y., Yang, S., 2020. Dual-targeting nanomicelles with CD133 and CD44 aptamers for enhanced delivery of gefitinib to two populations of lung cancer-initiating cells. *Exp. Ther. Med.* 19, 192–204.
- Hu, M., Yuan, L., Zhu, J., 2024. The dual role of NRF2 in colorectal cancer: targeting NRF2 as a potential therapeutic approach. *J. Inflamm. Res.* 17, 5985–6004.
- Hu, W., Zhang, C., Wu, R., Sun, Y., Levine, A., Feng, Z., 2010. Glutaminase 2, a novel p53 target gene regulating energy metabolism and antioxidant function. *Proc. Natl. Acad. Sci. U. S. A.* 107, 7455–7460.
- Hu, Y.-L., DeLay, M., Jahangiri, A., Molinaro, A.M., Rose, S.D., Carbonell, W.S., Aghi, M. K., 2023. Data from hypoxia-induced autophagy promotes tumor cell survival and adaptation to antiangiogenic treatment in glioblastoma. <https://doi.org/10.1158/0008-5472.ccr.23.0121>.
- Hu, Y., Mu, H., Deng, Z., 2023. H3K27 acetylation activated-CCS regulates autophagy and apoptosis of lung cancer by alleviating oxidative stress. *Tissue Cell* 80, 101964. <https://doi.org/10.1016/j.tice.2022.101964>.
- Hwang, J.S., 2022. Regulation of TGF-beta1-induced EMT by Autophagy-Dependent Energy Metabolism in Cancer Cells. *Cancers* 2036. <https://doi.org/10.3390/cancers14194845>.
- Inagawa, S., Itabashi, M., Adachi, S., Kawamoto, T., Hori, M., Shimazaki, J., Yoshimi, F., Fukao, K., 2002. Expression and prognostic roles of beta-catenin in hepatocellular carcinoma: correlation with tumor progression and postoperative survival. *Clin. Cancer Res.* 8, 450–456.
- Iqbal, M.J., 2024. Interplay of oxidative stress, cellular communication and signaling 1281 pathways in cancer. *Cell Commun. Signal* 22.
- Iskandar, K., Foo, J., Liew, A.Q.X., Zhu, H., Raman, D., Hirpara, J.L., Leong, Y.Y., Babak, M.V., Kirsanova, A.A., Armand, A.-S., Oury, F., Bellot, G., Pervaiz, S., 2024. A novel MTORC2-AKT-ROS axis triggers mitofission and mitophagy-associated execution of colorectal cancer cells upon drug-induced activation of mutant KRAS. *Autophagy* 20, 1418–1441.
- Jalali, P., Shahmoradi, A., Samii, A., Mazloomnejad, R., Hatamnejad, M.R., Saeed, A., Namdar, A., Salehi, Z., 2025. The role of autophagy in cancer: from molecular mechanism to therapeutic window. *Front. Immunol.* 16, 1528230.
- Jeon, S.J., 2019. TIPRL potentiates survival of lung cancer by inducing autophagy through the eIF2alpha-ATF4 pathway. *Cell Death Dis.* 10.
- Jhaveri, K.L., Accordino, M.K., Bedard, P.L., Cervantes, A., Gambardella, V., Hamilton, E., Italiano, A., Kalinsky, K., Krop, I.E., Oliveira, M., Schmid, P., Saura, C., Turner, N.C., Varga, A., Cheeti, S., Hilz, S., Hutchinson, K.E., Jin, Y., Royer-Joo, S., Peters, U., Shankar, N., Schutzman, J.L., Juric, D., 2024. Phase I/II trial of inavolisib plus palbociclib and endocrine therapy for PIK3CA-mutated, hormone receptor-positive, human epidermal growth factor receptor 2-negative advanced or metastatic breast cancer. *J. Clin. Oncol.* 42, 3947–3956.
- Jia, J., Zhang, H.-B., Shi, Q., Yang, C., Ma, J.-B., Jin, B., Wang, X., He, D., Guo, P., 2019. KLF5 downregulation desensitizes castration-resistant prostate cancer cells to docetaxel by increasing BECN1 expression and inducing cell autophagy. *Theranostics* 9, 5464–5477.
- Jiang, H., Zuo, J., Li, B., Chen, R., Luo, K., Xiang, X., Lu, S., Huang, C., Liu, L., Tang, J., Gao, F., 2023. Drug-induced oxidative stress in cancer treatments: angel or devil? *Redox Biol.* 63, 102754.
- Jiang, T., Harder, B., Rojo de la Vega, M., Wong, P.K., Chapman, E., Zhang, D.D., 2015. p62 links autophagy and Nrf2 signaling. *Free Radic. Biol. Med.* 88, 199–204.
- Jin, J., Nan, J., Si, Y., Chen, X., Wang, H., Wang, X., Huang, J., Guo, T., 2024. Exploring the therapeutic potential of rabadotermine E in lung cancer treatment: targeting the ROS/p38 MAPK/JNK signaling pathway. *Mol. Med. Rep.* 30. <https://doi.org/10.3892/mmr.2024.13330>.
- John Clotaire, D.Z., Zhang, B., Wei, N., Gao, R., Zhao, F., Wang, Y., Lei, M., Huang, W., 2016. MiR-26b inhibits autophagy by targeting ULK2 in prostate cancer cells. *Biochem. Biophys. Res. Commun.* 472, 194–200.

- Juric, D., Kalinsky, K., Turner, N.C., Jhaveri, K.L., Schmid, P., Loi, S., Saura, C., Im, S.-A., Sunpaweravong, P., Li, H., Musolino, A., Zhang, Q., Nowecki, Z., Leung, R.C.-Y., Thanopoulou, E., Shankar, N., Lei, G., Devine, J., Stout, T.J., Loibl, S., 2024. First-line inavolisib/placebo + palbociclib + fulvestrant (Inavo/Pbo+Palbo+Fulv) in patients (pts) with *PIK3CA*-mutated, hormone receptor-positive, HER2-negative locally advanced/metastatic breast cancer who relapsed during/within 12 months (mo) of adjuvant endocrine therapy completion: INAVO120 Phase III randomized trial additional analyses. *J. Clin. Oncol.* 42, 1003–1003.
- Kaminsky, V.O., Piskunova, T., Zborovskaya, I.B., Tchekina, E.M., Zhivotovsky, B., 2012. Suppression of basal autophagy reduces lung cancer cell proliferation and enhances caspase-dependent and -independent apoptosis by stimulating ROS formation. *Autophagy* 8, 1032–1044.
- Katsuragi, Y., Ichimura, Y., Komatsu, M., 2015. p62/SQSTM1 functions as a signaling hub and an autophagy adaptor. *FEBS J.* 282, 4672–4678.
- Kelsey, J.L., Horn-Ross, P.L., 1993. Breast cancer: magnitude of the problem and descriptive epidemiology. *Epidemiol. Rev.* 15, 7–16.
- Kern, J.C., Kehrer, J.P., 2005. Free radicals and apoptosis: relationships with glutathione, thioredoxin, and the BCL family of proteins. *Front. Biosci.* 10, 1727–1738.
- Kim, J., Kundu, M., Viollet, B., Guan, K.-L., 2011. AMPK and mTOR regulate autophagy through direct phosphorylation of Ulk1. *Nat. Cell Biol.* 13, 132–141.
- Kirshner, J.R., He, S., Balasubramanyam, V., Kepros, J., Yang, C.-Y., Zhang, M., Du, Z., Barsoum, J., Bertin, J., 2023. Data from Elesclomol induces cancer cell apoptosis through oxidative stress. <https://doi.org/10.1158/1535-7163.ccr.6532011.v1>.
- Kirshner, J.R., He, S., Balasubramanyam, V., Kepros, J., Yang, C.-Y., Zhang, M., Du, Z., Barsoum, J., Bertin, J., 2008. Elesclomol induces cancer cell apoptosis through oxidative stress. *Mol. Cancer Ther.* 7, 2319–2327.
- Kma, L., Baruah, T.J., 2022. The interplay of ROS and the PI3K/Akt pathway in autophagy regulation. *Biotechnol. Appl. Biochem.* 69, 248–264.
- Kongara, S., Karantza, V., 2012. The interplay between autophagy and ROS in tumorigenesis. *Front. Oncol.* 2, 171.
- Krasteva, N., Keremidarska-Markova, M., Hristova-Panusheva, K., Andreeva, T., Speranza, G., Wang, D., 2019. Aminated graphene oxide as a potential new therapy for colorectal cancer. *Oxidative Med. Oxid. Med. Cell. Longev.* 2019.
- Lan, N., Su, Y., Zeng, Q., Zhou, P., Hu, Y., Zhang, Z., Wang, Y., Liu, K., 2024. JD-02, a novel Hsp90 inhibitor, induces ROS/SRC axis-dependent cytoprotective autophagy in colorectal cancer cells. *Mol. Carcinog.* 63, 1038–1050.
- Lan, Y.-C., Chang, C.-L., Sung, M.-T., Yin, P.-H., Hsu, C.-C., Wang, K.-C., Lee, H.-C., Tseng, L.-M., Chi, C.-W., 2013. Zoledronic acid-induced cytotoxicity through endoplasmic reticulum stress triggered REDD1-mTOR pathway in breast cancer cells. *Anticancer Res* 33, 3807–3814.
- Lau, A., Wang, X.-J., Zhao, F., Villeneuve, N.F., Wu, T., Jiang, T., Sun, Z., White, E., Zhang, D.D., 2010. A noncanonical mechanism of Nrf2 activation by autophagy deficiency: direct interaction between Keap1 and p62. *Mol. Cell. Biol.* 30, 3275–3285.
- Lauzier, A., Normandeau-Guimond, J., Vaillancourt-Lavigne, V., Boivin, V., Charbonneau, M., Rivard, N., Scott, M.S., Dubois, C.M., Jean, S., 2019. Colorectal cancer cells respond differentially to autophagy inhibition in vivo. *Sci. Rep.* 9, 11316.
- Lee, Y.G., Nam, Y., Shin, K.J., Yoon, S., Park, W.S., Joung, J.Y., Seo, J.K., Jang, J., Lee, S., Nam, D., Caino, M.C., Suh, P.-G., Chan Chae, Y., 2020. Androgen-induced expression of DRP1 regulates mitochondrial metabolic reprogramming in prostate cancer. *Cancer Lett.* 471, 72–87.
- Lemjabbar-Alaoui, H., Hassan, O.U., Yang, Y.-W., Buchanan, P., 2015. Lung cancer: Biology and treatment options. *Biochim. Biophys. Acta* 1856, 189–210.
- Leslie, N.R., 2006. The redox regulation of PI 3-kinase-dependent signaling. *Antioxid. Redox Signal* 8, 1765–1774.
- Levine, B., Kroemer, G., 2019. Biological functions of autophagy genes: a disease perspective. *Cell* 176, 11–42.
- Levy, J.M.M., Thorburn, A., 2011. Targeting autophagy during cancer therapy to improve clinical outcomes. *Pharmacol. Ther.* 131, 130–141.
- Liang, C., Feng, P., Ku, B., Dotan, I., Canaani, D., Oh, B.-H., Jung, J.U., 2006. Autophagic and tumour suppressor activity of a novel Beclin1-binding protein UVRAG. *Nat. Cell Biol.* 8, 688–699.
- Liang, S., Kisseleva, T., Brenner, D.A., 2016. The role of NADPH oxidases (NOXs) in liver fibrosis and the activation of myofibroblasts. *Front. Physiol.* 7, 17.
- Li, C., Wei, C., Zhao, G., Yang, X., 2023. Cancer cells remodeling and quality control are inextricably linked to autophagy. *AIMS Mol. Sci.* 10, 109–126.
- Li, J., Quan, C., He, Y.-L., Cao, Y., Chen, Y., Wang, Y.-F., Wu, L.-Y., 2022. Autophagy regulated by the HIF/REDD1/mTORC1 signaling is progressively increased during erythroid differentiation under hypoxia. *Front. Cell Dev. Biol.* 10, 896893.
- Lim, J., Murthy, A., 2020. Targeting autophagy to treat cancer: challenges and opportunities. *Front. Pharm.* 11, 590344.
- Lim, J.U., Yoon, H.K., 2022. Narrative review: association between lung cancer development and ambient particulate matter in never-smokers. *J. Thorac. Dis.* 14, 553–563.
- Li, M., Zhao, L., Liu, J., Liu, A., Jia, C., Ma, D., Jiang, Y., Bai, X., 2010. Multi-mechanisms are involved in reactive oxygen species regulation of mTORC1 signaling. *Cell. Signal* 22, 1469–1476.
- Lin, X., Wei, M., Song, F., Xue, D.I., Wang, Y., 2020. N-acetylcysteine (NAC) attenuating apoptosis and autophagy in RAW264.7 cells in response to incubation with mycolic acid from bovine mycobacterium tuberculosis complex. *Pol. J. Microbiol.* 69, 223–229.
- Lin, Y.-C., Chang, Y.-T., Campbell, M., Lin, T.-P., Pan, C.-C., Lee, H.-C., Shih, J.C., Chang, P.-C., 2017. MAOA-A novel decision maker of apoptosis and autophagy in hormone refractory neuroendocrine prostate cancer cells. *Sci. Rep.* 7, 46338.
- Li, N., Zuo, R., He, Y., Gong, W., Wang, Y., Chen, L., Luo, Y., Zhang, C., Liu, Z., Chen, P., Guo, H., 2024. PD-L1 induces autophagy and primary resistance to EGFR-TKIs in EGFR-mutant lung adenocarcinoma via the MAPK signaling pathway. *Cell Death Dis.* 15, 555.
- Lin, Z., Yang, S., Qiu, Q., Cui, G., Zhang, Y., Yao, M., Li, X., Chen, C., Gu, J., Wang, T., Yin, P., Sun, L., Hao, Y., 2024. Hypoxia-induced cysteine metabolism reprogramming is crucial for the tumorigenesis of colorectal cancer. *Redox Biol.* 75, 103286.
- Li, Q., Chu, Y., Li, S., Yu, L., Deng, H., Liao, C., Liao, X., Yang, C., Qi, M., Cheng, J., Chen, G., Huang, L., 2022. The oncoprotein MUC1 facilitates breast cancer progression by promoting Pink1-dependent mitophagy via ATAD3A destabilization. *Cell Death Dis.* 13, 899.
- Li, R., Jia, Z., Trush, M.A., 2016. *React. Oxyg. 1279 Species (Apex)* 1, 9–21.
- Lisanti, M.P., Martinez-Outschoorn, U.E., Chiavarina, B., Pavlides, S., Whitaker-Menezes, D., Tsirigos, A., Witkiewicz, A., Lin, Z., Balliet, R., Howell, A., Sotgia, F., 2010. Understanding the “lethal” drivers of tumor-stroma co-evolution: emerging role(s) for hypoxia, oxidative stress and autophagy/mitophagy in the tumor micro-environment. *Cancer Biol. Ther.* 10, 537–542.
- Liu, B., Tan, X., Liang, J., Wu, S., Liu, J., Zhang, Q., Zhu, R., 2014. A reduction in reactive oxygen species contributes to dihydromyricetin-induced apoptosis in human hepatocellular carcinoma cells. *Sci. Rep.* 4, 7041.
- Liu, C., Yang, D., Liu, Y., Piao, H., Zhang, T., Li, X., Zhao, E., Zhang, D., Zheng, Y., Tang, X., 2023. The effect of ambient PM2.5 exposure on survival of lung cancer patients after lobectomy. *Environ. Health* 22, 23.
- Liu, F., Lee, J.Y., Wei, H., Tanabe, O., Engel, J.D., Morrison, S.J., Guan, J.-L., 2010. FIP200 is required for the cell-autonomous maintenance of fetal hematopoietic stem cells. *Blood* 116, 4806–4814.
- Liu, H.-Y., Zhang, H.-S., Liu, M.-Y., Li, H.-M., Wang, X.-Y., Wang, M., 2021. GLS1 depletion inhibited colorectal cancer proliferation and migration via redox/Nrf2/autophagy-dependent pathway. *Arch. Biochem. Biophys.* 708, 108964.
- Liu, J., 2023. Oxidative stress and autophagy-mediated immune patterns and tumor microenvironment infiltration characterization in gastric cancer. *Aging (Albany NY)*. *Aging (Albany NY)* 15, 12513–12536.
- Liu, J., Long, S., Wang, H., Liu, N., Zhang, C., Zhang, L., Zhang, Y., 2019. Blocking AMPK/ULK1-dependent autophagy promoted apoptosis and suppressed colon cancer growth. *Cancer Cell Int* 19, 336.
- Liu, J., Qu, W., Kadiiska, M.B., 2009. Role of oxidative stress in cadmium toxicity and carcinogenesis. *Toxicol. Appl. Pharm.* 238, 209–214.
- Liu, Y.-T., Sun, Z.-J., 2021. Turning cold tumors into hot tumors by improving T-cell infiltration. *Theranostics* 11, 5365–5386.
- Liu, Z., Ma, C., Wang, Q., Yang, H., Lu, Z., Bi, T., Xu, Z., Li, T., Zhang, L., Zhang, Y., Liu, J., Wei, X., Li, J., 2022. Targeting FAM134B-mediated reticulophagy activates sorafenib-induced ferroptosis in hepatocellular carcinoma. *Biochem. Biophys. Res. Commun.* 589, 247–253.
- Li, X., He, S., Ma, B., 2020. Autophagy and autophagy-related proteins in cancer. *Mol. Cancer* 19, 12.
- Li, X., Zhao, H., 2025. Targeting secretory autophagy in solid cancers: mechanisms, immune regulation and clinical insights. *Exp. Hematol. Oncol.* 14, 12.
- Li, Y., Yu, Y., Yang, L., Wang, R., 2023. Insights into the role of oxidative stress in hepatocellular carcinoma development. *Front. Biosci. (Landmark Ed.)* 28, 286.
- Li, Y., Zhang, X., Wang, Z., Li, B., Zhu, H., 2023. Modulation of redox homeostasis: a strategy to overcome cancer drug resistance. *Front. Pharm.* 14, 1156538.
- Llovet, J.M., Kelley, R.K., Villanueva, A., Singal, A.G., Pikarsky, E., Roayaie, S., Lencioni, R., Koike, K., Zucman-Rossi, J., Finn, R.S., 2021. Hepatocellular carcinoma. *Nat. Rev. Dis. Prim.* 7, 6.
- Lock, R., Kenific, C.M., Leidal, A.M., Salas, E., Debnath, J., 2014. Autophagy-dependent production of secreted factors facilitates oncogenic RAS-driven invasion. *Cancer Discov.* 4, 466–479.
- Lou, J.-S., Zhao, L.-P., Huang, Z.-H., Chen, X.-Y., Xu, J.-T., Tai, W.C.-S., Tsim, K.W.K., Chen, Y.-T., Xie, T., 2021. Ginkgetin derived from Ginkgo biloba leaves enhances the therapeutic effect of cisplatin via ferroptosis-mediated disruption of the Nrf2/HO-1 axis in EGFR wild-type non-small-cell lung cancer. *Phytomedicine* 80, 153370.
- Luo, D., Dai, X., Tian, H., Fan, C., Xie, H., Chen, N., Wang, J., Huang, L., Wang, H., Wang, G., Zhang, Y., 2023. Sophoflaurin A, a novel matrine-derived alkaloid from *Sophora flavescens* with therapeutic potential for non-small cell lung cancer through ROS-mediated pyroptosis and autophagy. *Phytomedicine* 116, 154909.
- Luo, J., Ostrem, J., Pellini, B., Imbody, D., Stern, Y., Solanki, H.S., Haura, E.B., Villaruz, L.C., 2022. Overcoming KRAS-mutant lung cancer. *Am. Soc. Clin. Oncol. Educ. Book* (42), 1–11.
- Luo, S., Shao, L., Chen, Z., Hu, D., Jiang, L., Tang, W., 2020. NPRL2 promotes docetaxel chemoresistance in castration resistant prostate cancer cells by regulating autophagy through the mTOR pathway. *Exp. Cell Res.* 390, 111981.
- Lyons, N.J., Giri, R., Begun, J., Clark, D., Proud, D., He, Y., Hooper, J.D., Kryza, T., 2023. Reactive oxygen species as mediators of disease progression and therapeutic response in colorectal cancer. *Antioxid. Redox Signal* 39, 186–205.
- Maiuri, M.C., Galluzzi, L., Morselli, E., Kepp, O., Malik, S.A., Kroemer, G., 2010. Autophagy regulation by p53. *Curr. Opin. Cell Biol.* 22, 181–185.
- Mansouri, A., Gattoliat, C.-H., Asselah, T., 2018. Mitochondrial dysfunction and signaling in chronic liver diseases. *Gastroenterology* 155, 629–647.
- Marioli-Sapsakou, G.-K., Kourti, M., 2021. Targeting production of reactive oxygen species as an anticancer strategy. *Anticancer Res* 41, 5881–5902.
- Martinez-Outschoorn, U.E., 2010. Autophagy in cancer associated fibroblasts promotes tumor cell survival: role of hypoxia, HIF1 induction and NFκB activation in the tumor stromal microenvironment. *Cell Cycle* 9, 3515–3533.
- Mathew, R., Karp, C.M., Beaudoin, B., Vuong, N., Chen, G., Chen, H.-Y., Bray, K., Reddy, A., Bhanot, G., Gelinas, C., DiPaola, R.S., Karantza-Wadsworth, V., White, E.,

2009. Autophagy suppresses tumorigenesis through elimination of p62. *Cell* 137, 1062–1075.
- Mi, W., Wang, C., Luo, G., Li, J., Zhang, Y., Jiang, M., Zhang, C., Liu, N., Jiang, X., Yang, G., Zhang, L., Zhang, G., Zhang, Y., Fu, Y., 2021. Targeting ERK induced cell death and p53/ROS-dependent protective autophagy in colorectal cancer. *Cell Death Discov.* 7, 375.
- Modica-Napolitano, J.S., Murray, M., Thibault, J., Haley-Read, J.-P., Nixdorf, L., Shanahan, B., Iacovella, N., Reyes, C., 2024. The in vitro cytotoxic effect of elesclomol on breast adenocarcinoma cells is enhanced by concurrent treatment with glycolytic inhibitors. *Cancers (Basel)* 16, 4054.
- Moldeus, P., Cotgreave, I.A., Berggren, M., 1986. Lung Protection by a Thiol-Containing Antioxidant - N-Acetylcysteine. *Respiration* 50, 31–42.
- Molina, J.R., Yang, P., Cassivi, S.D., Schild, S.E., Adjei, A.A., 2008. Non-small cell lung cancer: epidemiology, risk factors, treatment, and survivorship. *Mayo Clin. Proc.* 83, 584–594.
- Moomivand, S., Nikbakht, M., Majd, A., Bikhof Torbati, M., Mousavi, S.A., 2024. Combining chemotherapy agents and autophagy modulators for enhanced breast cancer cell death. *Adv. Pharm. Bull.* 14, 908–917.
- Mukha, A., Kahya, U., Linge, A., Chen, O., Löck, S., Lukiyanchuk, V., Richter, S., Alves, T. C., Peitzsch, M., Telychko, V., Skvortsov, S., Negro, G., Aschenbrenner, B., Skvortsova, I.-I., Mirtschink, P., Lohaus, F., Hölscher, T., Neubauer, H., Rivandi, M., Labitzky, V., Lange, T., Franken, A., Behrens, B., Stoeklein, N.H., Toma, M., Sommer, U., Zschaek, S., Rehm, M., Eisenhofer, G., Schwager, C., Abdollahi, A., Groeben, C., Kunz-Schughart, L.A., Baretton, G.B., Baumann, M., Krause, M., Peitzsch, C., Dubrovskaya, A., 2021. GLS-driven glutamine catabolism contributes to prostate cancer radiosensitivity by regulating the redox state, stemness and ATG5-mediated autophagy. *Theranostics* 11, 7844–7868.
- Nagai, M., Vo, N.H., Shin Ogawa, L., Chimmamanada, D., Inoue, T., Chu, J., Beaudette-Zlatanova, B.C., Lu, R., Blackman, R.K., Barsoum, J., Koya, K., Wada, Y., 2012. The oncology drug elesclomol selectively transports copper to the mitochondria to induce oxidative stress in cancer cells. *Free Radic. Biol. Med.* 52, 2142–2150.
- Nakamura, M., Magara, T., Yoshimitsu, M., Kano, S., Kato, H., Yokota, K., Okuda, K., Morita, A., 2024. Blockade of glucose-6-phosphate dehydrogenase induces immunogenic cell death and accelerates immunotherapy. *J. Immunother. Cancer* 12, e008441.
- Narendra, D.P., Youle, R.J., 2024. The role of PINK1-Parkin in mitochondrial quality control. *Nat. Cell Biol.* 26, 1639–1651.
- Nasrollahzadeh, A., 2020. Arsenic trioxide and BIBR1532 synergistically inhibit breast cancer cell proliferation through attenuation of NF-kappaB signaling pathway. *Life Sci.* 257.
- Nazim, U.M., Yin, H., Park, S.-Y., 2019. Autophagy flux inhibition mediated by celastrol sensitized lung cancer cells to TRAIL-induced apoptosis via regulation of mitochondrial transmembrane potential and reactive oxygen species. *Mol. Med. Rep.* 19, 984–993.
- Nguyen, H.G., Yang, J.C., Kung, H.-J., Shi, X.-B., Tilki, D., Lara, P.N., Jr, DeVere White, R.W., Gao, A.C., Evans, C.P., 2014. Targeting autophagy overcomes Enzalutamide resistance in castration-resistant prostate cancer cells and improves therapeutic response in a xenograft model. *Oncogene* 33, 4521–4530.
- Nooshinfar, E., Bashash, D., Safaroghli-Azar, A., Bayati, S., Rezaei-Tavirani, M., Ghaffari, S.H., Akbari, M.E., 2016. Melatonin promotes ATO-induced apoptosis in MCF-7 cells: Proposing novel therapeutic potential for breast cancer. *Biomed. Pharm.* 83, 456–465.
- Nurdinon, N., 2020. LC3 and beclin-1 as markers of autophagic activity in breast cancer. *Erciyes Med. J.* <https://doi.org/10.14744/etd.2020.99997>.
- O'Connor, P.M., Lapointe, T.K., Beck, P.L., Buret, A.G., 2010. Mechanisms by which inflammation may increase intestinal cancer risk in inflammatory bowel disease. *Inflamm. Bowel Dis.* 16, 1411–1420.
- Onorati, A.V., Dyczynski, M., Ojha, R., Amaravadi, R.K., 2018. Targeting autophagy in cancer. *Cancer* 124, 3307–3318.
- Ornatowski, W., Lu, Q., Yegambaram, M., Garcia, A.E., Zemskov, E.A., Maltepe, E., Fineman, J.R., Wang, T., Black, S.M., 2020. Complex interplay between autophagy and oxidative stress in the development of pulmonary disease. *Redox Biol.* 36, 101679.
- Orrù, C., Perra, A., Kowalik, M.A., Rizzolio, S., Puliga, E., Cabras, L., Giordano, S., Columbano, A., 2020. Distinct mechanisms are responsible for Nrf2-Keap1 pathway activation at different stages of rat hepatocarcinogenesis. *Cancers (Basel)* 12, 2305.
- Pandey, P., Garg, A., Singh, V., Rai, G., Mishra, N., 2024. Clinical trials and future prospects of autophagy and ROS in cancer. *Cancer Drug Discovery and Development, Cancer Drug Discovery and Development. Springer Nature Switzerland, Cham*, pp. 337–369.
- Pani, G., Galeotti, T., 2011. Role of MnSOD and p66shc in mitochondrial response to p53. *Antioxid. Redox Signal* 15, 1715–1727.
- Peng, C., Li, X., Ao, F., Li, T., Guo, J., Liu, J., Zhang, X., Gu, J., Mao, J., Zhou, B., 2023. Mitochondrial ROS driven by NOX4 upregulation promotes hepatocellular carcinoma cell survival after incomplete radiofrequency ablation by inducing of mitophagy via Nrf2/PINK1. *J. Transl. Med.* 21, 218.
- Pernar, C.H., 2018. The Epidemiology of Prostate Cancer. *Cold Spring Harb. Perspect. Med.*
- Petherick, K.J., Williams, A.C., Lane, J.D., Ordóñez-Morán, P., Huelsken, J., Collard, T.J., Smart, H.J.M., Batson, J., Malik, K., Paraskeva, C., Greenhough, A., 2013. Autolysosomal  $\beta$ -catenin degradation regulates Wnt-autophagy-p62 crosstalk. *EMBO J.* 32, 1903–1916.
- Poillet-Perez, L., Despouy, G., Delage-Mourroux, R., Boyer-Guittaut, M., 2015. Interplay between ROS and autophagy in cancer cells, from tumor initiation to cancer therapy. *Redox Biol.* 4, 184–192.
- Prat, A., Perou, C.M., 2011. Deconstructing the molecular portraits of breast cancer. *Mol. Oncol.* 5, 5–23.
- Qian, M., Tan, H.M., Yu, N., Wang, T., Zhang, Q., 2018. Inactivated Sendai virus induces ROS-dependent apoptosis and autophagy in human prostate cancer cells. *Biomed. Environ. Sci.* 31, 280–289.
- Qin, P., Li, Q., Zu, Q., Dong, R., Qi, Y., 2024. Natural products targeting autophagy and apoptosis in NSCLC: a novel therapeutic strategy. *Front. Oncol.* 14, 1379698.
- Quan, Z., 2020. PLCvarepsilon maintains the functionality of AR signaling in prostate cancer via an autophagy-dependent mechanism. *Cell Death Dis.* 11.
- Qu, Y., Wang, J., Sim, M.-S., Liu, B., Giuliano, A., Barsoum, J., Cui, X., 2010. Elesclomol, counteracted by Akt survival signaling, enhances the apoptotic effect of chemotherapy drugs in breast cancer cells. *Breast Cancer Res. Treat.* 121, 311–321.
- Raghunath, A., Sundarraj, K., Arfuso, F., Sethi, G., Perumal, E., 2018. Dysregulation of Nrf2 in hepatocellular carcinoma: role in cancer progression and chemoresistance. *Cancers (Basel)* 10, 481.
- Rahman, M.A., Park, M.N., Rahman, M.H., Rashid, M.M., Islam, R., Uddin, M.J., Hannan, M.A., Kim, B., 2022. P53 modulation of autophagy signaling in cancer therapies: perspectives mechanism and therapeutic targets. *Front. Cell Dev. Biol.* 10, 761080.
- Rajaram, S., Synnott, N.C., Crown, J., Madden, S.F., Duffy, M.J., 2024. Targeting mutant p53 with arsenic trioxide: a preclinical study focusing on triple negative breast cancer. *Transl. Oncol.* 46, 102025.
- Ramos-Torres, A., Bort, A., Morell, C., Rodríguez-Henche, N., Díaz-Laviada, I., 2016. The pepper's natural ingredient capsaicin induces autophagy blockage in prostate cancer cells. *Oncotarget* 7, 1569–1583.
- Rasková, M., 2022. The Role of IL-6 in cancer cell invasiveness and metastasis-overview and therapeutic opportunities. *Cells.*
- Rawla, P., 2019. Epidemiology of prostate cancer. *World J. Oncol.* 10, 63–89.
- Redza-Dutordoir, M., Averill-Bates, D.A., 2021. Interactions between reactive oxygen species and autophagy. Special issue Death mechanisms cellular homeostasis. *Biochim. Biophys. Acta Mol. Cell Res.* 1868, 119041.
- Reuter, S., Gupta, S.C., Chaturvedi, M.M., Aggarwal, B.B., 2010. Oxidative stress, inflammation, and cancer: how are they linked? *Free Radic. Biol. Med.* 49, 1603–1616.
- Roessner, A., Kuester, D., Malfertheiner, P., Schneider-Stock, R., 2008. Oxidative stress in ulcerative colitis-associated carcinogenesis. *Pathol. Res. Pr.* 204, 511–524.
- Ruiz-Torres, V., Rodríguez-Pérez, C., Herranz-López, M., Martín-García, B., Gómez-Caravaca, A.-M., Arráez-Román, D., Segura-Carretero, A., Barrajón-Catalán, E., Micol, V., 2019. Marine invertebrate extracts induce colon cancer cell death via ROS-mediated DNA oxidative damage and mitochondrial impairment. *Biomolecules* 9, 771.
- Sala, R., 1993. Protection by N-acetylcysteine against pulmonary endothelial-cell damage induced by oxidant injury. *Eur. Respir. J.* 6, 440–446.
- Samant, H., Amiri, H.S., Zibari, G.B., 2021. Addressing the worldwide hepatocellular carcinoma: epidemiology, prevention and management. *J. Gastrointest. Oncol.* 12, S361–S373.
- Sampson, N., Brunner, E., Weber, A., Pühr, M., Schäfer, G., Szyndralewicz, C., Klocker, H., 2018. Inhibition of Nox4-dependent ROS signaling attenuates prostate fibroblast activation and abrogates stromal-mediated protumorigenic interactions. *Int. J. Cancer* 143, 383–395.
- Sarkar, S., Korolchuk, V.I., Renna, M., Imarishi, S., Fleming, A., Williams, A., Garcia-Arencibia, M., Rose, C., Luo, S., Underwood, B.R., Kroemer, G., O'Kane, C.J., Rubinshtein, D.C., 2011. Complex inhibitory effects of nitric oxide on autophagy. *Mol. Cell* 43, 19–32.
- Sawicki, T., Ruskowska, M., Danielewicz, A., Niedźwiedzka, E., Artukowicz, T., Przybyłowicz, K.E., 2021. A review of colorectal cancer in terms of epidemiology, risk factors, development, symptoms and diagnosis. *Cancers (Basel)* 13, 2025.
- Scheid, A.D., Beadnell, T.C., Welch, D.R., 2021. Roles of mitochondria in the hallmarks of metastasis. *Br. J. Cancer* 124, 124–135.
- Scholten, A., 2024. Non-alcoholic fatty liver disease and the risk of future hepatocarcinoma. *Sci. Insights* 44, 1357–1364.
- Schwartz, A.G., Cote, M.L., 2016. Epidemiology of lung cancer, in: *Advances in Experimental Medicine and Biology. Advances in Experimental Medicine and Biology. Springer International Publishing, Cham*, pp. 21–41.
- Seiler, C.L., Song, J.U.M., Kotandeniya, D., Chen, J., Kono, T.J.Y., Han, Q., Colwell, M., Auch, B., Sarver, A.L., Upadhyaya, P., Ren, Y., Faulk, C., De Flora, S., La Maestra, S., Chen, Y., Kassie, F., Tretyakova, N.Y., 2020. Inhalation exposure to cigarette smoke and inflammatory agents induces epigenetic changes in the lung. *Sci. Rep.* 10, 11290.
- Shadab, M., Millar, M.W., Slavin, S.A., Leonard, A., Fazzal, F., Rahman, A., 2020. Autophagy protein ATG7 is a critical regulator of endothelial cell inflammation and permeability. *Sci. Rep.* 10, 13708.
- Shahgoli, V.K., Noorolyai, S., Ahmadvour Youshanlui, M., Saedi, H., Nasiri, H., Mansoori, B., Holmskov, U., Baradaran, B., 2024. Inflammatory bowel disease, colitis, and cancer: unmasking the chronic inflammation link. *Int. J. Colorectal Dis.* 39, 173.
- Shan, C., Wang, Y., Wang, Y., 2024. The crosstalk between autophagy and Nrf2 signaling in cancer: From biology to clinical applications. *Int. J. Biol. Sci.* 20, 6181–6206.
- Shang, G.-S., Liu, L., Qin, Y.-W., 2017. IL-6 and TNF- $\alpha$  promote metastasis of lung cancer by inducing epithelial-mesenchymal transition. *Oncol. Lett.* 13, 4657–4660.
- Shankar, A., Dubey, A., Saini, D., Singh, M., Prasad, C.P., Roy, S., Bharati, S.J., Rinki, M., Singh, N., Seth, T., Khanna, M., Sethi, N., Kumar, S., Sirohi, B., Mohan, A., Guleria, R., Rath, G.K., 2019. Environmental and occupational determinants of lung cancer. *Transl. Lung Cancer Res.* 8, S31–S49.
- Sharifi-Rad, M., Anil Kumar, N.V., Zucca, P., Varoni, E.M., Dini, L., Panzarini, E., Rajkovic, J., Tsouh Fokou, P.V., Azzini, E., Peluso, I., Prakash Mishra, A., Nigam, M.,

- El Rayess, Y., Beyrouthy, M.E., Polito, L., Iriti, M., Martins, N., Martorell, M., Docea, A.O., Setzer, W.N., Calina, D., Cho, W.C., Sharifi-Rad, J., 2020. Lifestyle, oxidative stress, and antioxidants: Back and forth in the pathophysiology of chronic diseases. *Front. Physiol.* 11, 694.
- Sharma, P., Nandave, M., Nandave, D., Yadav, S., Vargas-De-La-Cruz, C., Singh, S., Tandon, R., Ramniwas, S., Behl, T., 2023. Reactive oxygen species (ROS)-mediated oxidative stress in chronic liver diseases and its mitigation by medicinal plants. *Am. J. Transl. Res.* 15, 6321–6341.
- Shen, W., Luo, P., Sun, Y., Zhang, W., Zhou, N., Zhan, H., Zhang, Q., Shen, J., Lin, A., Cheng, Q., Wang, Q., Zhang, J., Wang, H.-H., Wei, T., 2022. NRBF2 regulates the chemoresistance of small cell lung cancer by interacting with the P62 protein in the autophagy process. *iScience* 25, 104471.
- Shimizu, I., Shimamoto, N., Saiki, K., Furujio, M., Osawa, K., 2012. Lipid peroxidation in hepatic fibrosis. In: *Lipid Peroxidation*. InTech.
- Shimizu, T., Tolcher, A.W., Papadopoulos, K.P., Beeram, M., Rasco, D.W., Smith, L.S., Gunn, S., Smetzer, L., Mays, T.A., Kaiser, B., Wick, M.J., Alvarez, C., Cavazos, A., Mangold, G.L., Patnaik, A., 2023. Data from the clinical effect of the dual-targeting strategy involving PI3K/AKT/mTOR and RAS/MEK/ERK pathways in patients with advanced cancer. <https://doi.org/10.1158/1078-0432.ccr.20148.v1>.
- Shin, J., Song, M.-H., Oh, J.-W., Keum, Y.-S., Saini, R.K., 2020. Pro-oxidant actions of carotenoids in triggering apoptosis of cancer cells: a review of emerging evidence. *Antioxid. (Basel)* 9, 532.
- Siegel, R., Naishadham, D., Jemal, A., 2013. Cancer statistics, 2013. *CA Cancer J. Clin.* 63, 11–30.
- Singh, A., Koduru, B., Carlisle, C., Akhter, H., Liu, R.-M., Schroder, K., Brandes, R.P., Ojcius, D.M., 2017. NADPH oxidase 4 modulates hepatic responses to lipopolysaccharide mediated by Toll-like receptor-4. *Sci. Rep.* 7, 14346.
- Singh, K., Bhoori, M., Kasu, Y.A., Bhat, G., Marar, T., 2018. Antioxidants as precision weapons in war against cancer chemotherapy induced toxicity – Exploring the armoury of obscurity. *Saudi Pharm. J.* 26, 177–190.
- Singh, P., Dhole, B., Choudhury, J., Tuli, A., Pandey, D., Velpandian, T., Gupta, S., Chaturvedi, P.K., 2024. Calotropis procera extract inhibits prostate cancer through regulation of autophagy. *J. Cell. Mol. Med.* 28, e18050.
- Sohn, E.J., 2018. MicroRNA 200c-3p regulates autophagy via upregulation of endoplasmic reticulum stress in PC-3 cells. *Cancer Cell Int* 18.
- Soler-Agosta, R., Moreno-Loshuertos, R., Yim, C.Y., Congenie, M.T., Ames, T.D., Johnson, H.L., Stossi, F., Mancini, M.G., Mancini, M.A., Ripollés-Yuba, C., Marco-Brualla, J., Junquera, C., Martínez-De-Mena, R., Enríquez, J.A., Price, M.R., Jimeno, J., Anel, A., 2024. Cancer cell-selective induction of mitochondrial stress and immunogenic cell death by PT-112 in human prostate cell lines. *J. Transl. Med.* 22, 927.
- Strohecker, A.M., Guo, J.Y., Karsli-Uzunbas, G., Price, S.M., Chen, G.J., Mathew, R., McMahon, M., White, E., 2013. Autophagy sustains mitochondrial glutamine metabolism and growth of BrafV600E-driven lung tumors. *Cancer Discov.* 3, 1272–1285.
- Stumptner, C., Heid, H., Fuchsichler, A., Hauser, H., Mischinger, H.J., Zatloukal, K., Denk, H., 1999. Analysis of intracytoplasmic hyaline bodies in a hepatocellular carcinoma. *Demonstr. p62 Major Const. Am. J. Pathol.* 154, 1701–1710.
- Sun, B., Ding, P., Song, Y., Zhou, J., Chen, X., Peng, C., Liu, S., 2024. FDX1 downregulation activates mitophagy and the PI3K/AKT signaling pathway to promote hepatocellular carcinoma progression by inducing ROS production. *Redox Biol.* 75, 103302.
- Sun, C., Zhao, L., Wang, X., Hou, Y., Guo, X., Lu, J.-J., Chen, X., 2022. Psoralidin, a natural compound from *Psoralea corylifolia*, induces oxidative damage mediated apoptosis in colon cancer cells. *J. Biochem. Mol. Toxicol.* 36. <https://doi.org/10.1002/jbt.23051>.
- Sun, J., Feng, Y., Wang, Y., Ji, Q., Cai, G., Shi, L., Wang, Y., Huang, Y., Zhang, J., Li, Q., 2019.  $\alpha$ -hederin induces autophagic cell death in colorectal cancer cells through reactive oxygen species dependent AMPK/mTOR signaling pathway activation. *Int. J. Oncol.* 54, 1601–1612.
- Sun, R.C., Board, P.G., Blackburn, A.C., 2011. Targeting metabolism with arsenic trioxide and dichloroacetate in breast cancer cells. *Mol. Cancer* 10, 142.
- Su, R., Qiao, M., Gao, T., Gao, J., Nie, L., Li, S., Wang, Y., Pang, Y., Li, Q., 2014. Effect of N-acetylcysteine on apoptosis and autophagy of macrophages infected with *Mycobacterium tuberculosis*. *J. Infect. Dev. Ctries* 18, 1566–1575.
- Suzawa, K., Offin, M., Lu, D., Kurzatkowski, C., Vojnic, M., Smith, R.S., Sabari, J.K., Tai, H., Mattar, M., Khodos, I., de Stanchina, E., Rudin, C.M., Kris, M.G., Arcila, M.E., Lockwood, W.W., Drilon, A., Ladanyi, M., Somwar, R., 2019. Activation of KRAS mediates resistance to targeted therapy in MET Exon 14-mutant non-small cell lung cancer. *Clin. Cancer Res.* 25, 1248–1260.
- Suzuki, S., Tanaka, T., Poyurovsky, M.V., Nagano, H., Mayama, T., Ohkubo, S., Lokshin, M., Hosokawa, H., Nakayama, T., Suzuki, Y., Sugano, S., Sato, E., Nagao, T., Yokote, K., Tatsuno, I., Prives, C., 2010. Phosphate-activated glutaminase (GLS2), a p53-inducible regulator of glutamine metabolism and reactive oxygen species. *Proc. Natl. Acad. Sci. U. S. A.* 107, 7461–7466.
- Taha-Mehlitz, S., Bianco, G., Coto-Llerena, M., Kancherla, V., Bantug, G.R., Gallon, J., Ercan, C., Panebianco, F., Eppenberger-Castori, S., von Strauss, M., Staubli, S., Bolli, M., Peterli, R., Matter, M.S., Terracciano, L.M., von Flüe, M., Ng, C.K.Y., Soysal, S.D., Kollmar, O., Piscuoglio, S., 2021. Adenylosuccinate lyase is oncogenic in colorectal cancer by causing mitochondrial dysfunction and independent activation of NRF2 and mTOR-MYC-axis. *Theranostics* 11, 4011–4029.
- Takamura, A., Komatsu, M., Hara, T., Sakamoto, A., Kishi, C., Waguri, S., Eishi, Y., Hino, O., Tanaka, K., Mizushima, N., 2011. Autophagy-deficient mice develop multiple liver tumors. *Genes Dev.* 25, 795–800.
- Takano, N., Hiramoto, M., Yamada, Y., Kokuba, H., Tokuhisa, M., Hino, H., Miyazawa, K., 2023. Azithromycin, a potent autophagy inhibitor for cancer therapy, perturbs cytoskeletal protein dynamics. *Br. J. Cancer* 128, 1838–1849.
- Tan, A.C., 2020. Targeting the PI3K/Akt/mTOR pathway in non-small cell lung cancer (NSCLC). *Thorac. Cancer* 11, 511–518.
- Tan, A.S., Baty, J.W., Dong, L.-F., Bezawork-Geleta, A., Endaya, B., Goodwin, J., Bajzikova, M., Kovarova, J., Peterka, M., Yan, B., Pesar, E.A., Sobol, M., Filimonenko, A., Stuart, S., Vondrusova, M., Kluckova, K., Sachaphibulkij, K., Rohlena, J., Hozak, P., Truksa, J., Eccles, D., Haupt, L.M., Griffiths, L.R., Neuzil, J., Berridge, M.V., 2015. Mitochondrial genome acquisition restores respiratory function and tumorigenic potential of cancer cells without mitochondrial DNA. *Cell Metab.* 21, 81–94.
- Tang, F., Hu, P., Yang, Z., Xue, C., Gong, J., Sun, S., Shi, L., Zhang, S., Li, Z., Yang, C., Zhang, J., Xie, C., 2017. SBI0206965, a novel inhibitor of Ulk1, suppresses non-small cell lung cancer cell growth by modulating both autophagy and apoptosis pathways. *Oncol. Rep.* 37, 3449–3458.
- Tang, J., 2023. USP8 positively regulates hepatocellular carcinoma tumorigenesis and confers ferroptosis resistance through beta-catenin stabilization. *Cell Death Dis.* 14.
- Tang, J., Di, J., Cao, H., Bai, J., Zheng, J., 2015. p53-mediated autophagic regulation: a prospective strategy for cancer therapy. *Cancer Lett.* 363, 101–107.
- Tang, S.-P., Mao, X.-L., Chen, Y.-H., Yan, L.-L., Ye, L.-P., Li, S.-W., 2022. Reactive oxygen species induce fatty liver and ischemia-reperfusion injury by promoting inflammation and cell death. *Front. Immunol.* 13, 870239.
- Taucher, E., Mykoliuk, I., Fediuk, M., Smolle-Juettner, F.-M., 2022. Autophagy, oxidative stress and cancer development. *Cancers (Basel)* 14, 1637.
- Ternet, C., Kiel, C., 2021. Signaling pathways in intestinal homeostasis and colorectal cancer: KRAS at centre stage. *Cell Commun. Signal* 19, 31.
- Tirado-Hurtado, I., Fajardo, W., Pinto, J.A., 2018. DNA damage inducible transcript 4 gene: the switch of the metabolism as potential target in cancer. *Front. Oncol.* 8, 106.
- Tripathi, D.N., Chowdhury, R., Trudel, L.J., Tee, A.R., Slack, R.S., Walker, C.L., Wogan, G.N., 2013. Reactive nitrogen species regulate autophagy through ATM-AMPK-TSC2-mediated suppression of mTORC1. *Proc. Natl. Acad. Sci. U. S. A.* 110, E2950–E2957.
- Vachlitos, I.D., Polyzos, S.A., 2023. The role of tumor necrosis factor-alpha in the pathogenesis and treatment of nonalcoholic fatty liver disease. *Curr. Opin. Rep.* 12, 191–206.
- Wang, C., Chen, K., Xia, Y., Dai, W., Wang, F., Shen, M., Cheng, P., Wang, J., Lu, J., Zhang, Y., Yang, J., Zhu, R., Zhang, H., Li, J., Zheng, Y., Zhou, Y., Guo, C., 2014. N-acetylcysteine attenuates ischemia-reperfusion-induced apoptosis and autophagy in mouse liver via regulation of the ROS/JNK/Bcl-2 pathway. *PLoS One* 9, e108855.
- Wang, H., Lin, M., Chen, G., Xiao, Z., Shuai, X., 2023. Nanodrug regulates ROS homeostasis via enhancing fatty acid oxidation and inhibiting autophagy to overcome tumor drug resistance. *Biomater. Sci.* 11, 7179–7187.
- Wang, J., Tan, X., Yang, Q., Zeng, X., Zhou, Y., Luo, W., Lin, X., Song, L., Cai, J., Wang, T., Wu, X., 2016. Inhibition of autophagy promotes apoptosis and enhances anticancer efficacy of adriamycin via augmented ROS generation in prostate cancer cells. *Int. J. Biochem. Cell Biol.* 77, 80–90.
- Wang, K.-W., Wang, M.-D., Li, Z.-X., Hu, B.-S., Wu, J.-J., Yuan, Z.-D., Wu, X.-L., Yuan, Q.-F., Yuan, F.-L., 2022. An antigen processing and presentation signature for prognostic evaluation and immunotherapy selection in advanced gastric cancer. *Front. Immunol.* 13, 992060.
- Wang, M., Liu, M., Yang, C., Hu, Y., Liao, X., Liu, Q., 2024. Autophagy modulation in therapeutic strategy of breast cancer drug resistance. *J. Cancer* 15, 5462–5476.
- Wang, M., Yu, H., Wu, R., Chen, Z.-Y., Hu, Q., Zhang, Y.-F., Gao, S.-H., Zhou, G.-B., 2020. Autophagy inhibition enhances the inhibitory effects of ursolic acid on lung cancer cells. *Int. J. Mol. Med.* 46, 1816–1826.
- Wang, Q., He, W.-Y., Zeng, Y.-Z., Hossain, A., Gou, X., 2018. Inhibiting autophagy overcomes docetaxel resistance in castration-resistant prostate cancer cells. *Int. Urol. Nephrol.* 50, 675–686.
- Wang, R.-A., Zhang, M.-Y., Jiang, Y.-X., Wang, X.-D., Qu, J.-J., Yue, Y.-L., Qu, Y.-Q., 2023. Autophagy-related tumor subtypes associated with significant gene expression profiles and immune cell infiltration signatures to reveal the prognosis of non-small cell lung cancer. *J. Cancer* 14, 1427–1442.
- Wang, S., Long, H., Hou, L., Feng, B., Ma, Z., Wu, Y., Zeng, Y., Cai, J., Zhang, D.-W., Zhao, G., 2023. The mitophagy pathway and its implications in human diseases. *Signal Transduct. Target. Ther.* 8, 304.
- Wang, W., Sun, Y., Liu, X., Kumar, S.K., Jin, F., Dai, Y., 2022. Dual-targeted therapy circumvents non-genetic drug resistance to targeted therapy. *Front. Oncol.* 12, 859455.
- Wang, X., Li, P., Ji, H., Xu, Z., Xing, H., 2024. Single-cell transcriptomics reveals over-activated reactive oxygen species pathway in hepatocytes in the development of hepatocellular carcinoma. *Sci. Rep.* 14, 29809.
- Wang, X., Zhou, Y., Ning, L., Chen, J., Chen, H., Li, X., 2023. Knockdown of ANXA10 induces ferroptosis by inhibiting autophagy-mediated TFRC degradation in colorectal cancer. *Cell Death Dis.* 14, 588.
- Wang, Y., Zhang, Y., Yang, L., Cai, B., Li, J., Zhou, Y., Yin, L., Yang, L., Yang, B.F., Lu, Y. J., 2011. Arsenic trioxide induces the apoptosis of human breast cancer MCF-7 cells through activation of caspase-3 and inhibition of HERG channels. *Exp. Ther. Med.* 2, 481–486.
- Wei, H., Wei, S., Gan, B., Peng, X., Zou, W., Guan, J.-L., 2011. Suppression of autophagy by FIP200 deletion inhibits mammary tumorigenesis. *Genes Dev.* 25, 1510–1527.
- Wiel, C., Le Gal, K., Ibrahim, M.X., Jahangir, C.A., Kashif, M., Yao, H., Ziegler, D.V., Xu, X., Ghosh, T., Mondal, T., Kanduri, C., Lindahl, P., Sayin, V.I., Bergo, M.O., 2019. BACH1 stabilization by antioxidants stimulates lung cancer metastasis. *Cell* 178, 330–345.e22.

- Wu, Q.T., Sharma, D., 2023. Autophagy and breast cancer: connected in growth, progression, and therapy. *Cells*.
- Wu, Y., Zhou, B.P., 2010. TNF- $\alpha$ /NF- $\kappa$ B/Snail pathway in cancer cell migration and invasion. *Br. J. Cancer* 102, 639–644.
- Xia, J., Li, Y., Yang, Q., Mei, C., Chen, Z., Bao, B., Ahmad, A., Miele, L., Sarkar, F., Wang, Z., Xia, J., et al., 2014. Arsenic trioxide inhibits cell growth and induces apoptosis through inactivation of Notch signaling pathway in breast cancer. *Int. J. Mol. Sci.* 2012 13, 9627–9641. *Int. J. Mol. Sci.* 15, 14907–14908.
- Xiang, X.-Y., Yang, X.-C., Su, J., Kang, J.-S., Wu, Y., Xue, Y.-N., Dong, Y.-T., Sun, L.-K., 2016. Inhibition of autophagic flux by ROS promotes apoptosis during DTT-induced ER/oxidative stress in HeLa cells. *Oncol. Rep.* 35, 3471–3479.
- Xiao, H., Chen, C., Yuan, X., Yang, L., Zheng, Y., Yuan, J., Huang, S., Liang, J., Yuan, S., Li, M., Wang, J., 2024. Gingerone A induces ferroptosis in colorectal cancer via targeting suppression of SLC7A11 signaling pathway. *Biomed. Pharm.* 180, 117529.
- Xie, Z., Rahman, I., Goniewicz, M.L., Li, D., 2021. Perspectives on epigenetics alterations associated with smoking and vaping. *Funct. (Oxf.)* 2, zqab022.
- Xiong, Y., Du, K., Huang, Y., 2024. One-third of global population at cancer risk due to elevated volatile organic compounds levels. *Npj Clim. Atmos. Sci.* 7. <https://doi.org/10.1038/s41612-024-00598-1>.
- Xiu, Z., Zhu, Y., Han, J., Li, Y., Yang, X., Yang, G., Song, G., Li, S., Li, Y., Cheng, C., Li, Y., Fang, J., Li, X., Jin, N., 2022. Caryophyllene oxide induces ferritinophagy by regulating the NCOA4/FTH1/LC3 pathway in hepatocellular carcinoma. *Front. Pharm.* 13, 930958.
- Xu, Y., Ji, Y., Li, X., Ding, J., Chen, L., Huang, Y., Wei, W., 2021. URI1 suppresses irradiation-induced reactive oxygen species (ROS) by activating autophagy in hepatocellular carcinoma cells. *Int. J. Biol. Sci.* 17, 3091–3103.
- Yan, C., Li, T.-S., 2018. Dual role of mitophagy in cancer drug resistance. *Anticancer Res* 38, 617–621.
- Yang, S., Pang, L., Dai, W., Wu, S., Ren, T., Duan, Y., Zheng, Y., Bi, S., Zhang, X., Kong, J., 2021. Role of forkhead box O proteins in hepatocellular carcinoma biology and progression (review). *Front. Oncol.* 11, 667730.
- Yang, Z.J., Chee, C.E., Huang, S., Sinicrope, F.A., 2011. The role of autophagy in cancer: therapeutic implications. *Mol. Cancer Ther.* 10, 1533–1541.
- Yip, P.Y., 2015. Phosphatidylinositol 3-kinase-AKT-mammalian target of rapamycin (PI3K-Akt-mTOR) signaling pathway in non-small cell lung cancer. *Transl. Lung Cancer Res.* 4, 165–176.
- You, B.R., Park, W.H., 2012. Arsenic trioxide induces human pulmonary fibroblast cell death via increasing ROS levels and GSH depletion. *Oncol. Rep.* 28, 749–757.
- Youle, R.J., Narendra, D.P., 2011. Mechanisms of mitophagy. *Nat. Rev. Mol. Cell Biol.* 12, 9–14.
- Yuan, W., Fang, W., Zhang, R., Lyu, H., Xiao, S., Guo, D., Ali, D.W., Michalak, M., Chen, X.-Z., Zhou, C., Tang, J., 2023. Therapeutic strategies targeting AMPK-dependent autophagy in cancer cells. *Biochim. Biophys. Acta Mol. Cell Res.* 1870, 119537.
- Yu, Y., 2021. Mesenchymal stem cells desensitize castration-resistant prostate cancer to docetaxel chemotherapy via inducing TGF- $\beta$ 1-mediated cell autophagy. *Cell Biosci.* 11.
- Zafeiropoulou, K., Kalampounias, G., Alexis, S., Anastasopoulos, D., Symeonidis, A., Katsoris, P., 2024. Autophagy and oxidative stress modulation mediate Bortezomib resistance in prostate cancer. *PLoS One* 19, e0289904.
- Zatloukal, K., Stumptner, C., Fuchsbiçhler, A., Heid, H., Schnoelzer, M., Kenner, L., Kleinert, R., Prinz, M., Aguzzi, A., Denk, H., 2002. p62 is a common component of cytoplasmic inclusions in protein aggregation diseases. *Am. J. Pathol.* 160, 255–263.
- Zhang, C., Zhao, Y., Yu, M., Qin, J., Ye, B., Wang, Q., 2022. Mitochondrial dysfunction and chronic liver disease. *Curr. Issues Mol. Biol.* 44, 3156–3165.
- Zhang, Q., Wang, X., Cao, S., Sun, Y., He, X., Jiang, B., Yu, Y., Duan, J., Qiu, F., Kang, N., 2020. Berberine represses human gastric cancer cell growth in vitro and in vivo by inducing cytoskeletal autophagy via inhibition of MAPK/mTOR/p70S6K and Akt signaling pathways. *Biomed. Pharm.* 128, 110245.
- Zhang, S., Ma, C., Pang, H., Zeng, F., Cheng, L., Fang, B., Ma, J., Shi, Y., Hong, H., Chen, J., Wang, Z., Xia, J., 2016. Arsenic trioxide suppresses cell growth and migration via inhibition of miR-27a in breast cancer cells. *Biochem. Biophys. Res. Commun.* 469, 55–61.
- Zhang, T., Li, Y., Park, K.A., Byun, H.S., Won, M., Jeon, J., Lee, Y., Seok, J.H., Choi, S.-W., Lee, S.-H., Man Kim, J., Lee, J.H., Son, C.G., Lee, Z.-W., Shen, H.-M., Hur, G.M., 2012. Cucurbitacin induces autophagy through mitochondrial ROS production which counteracts to limit caspase-dependent apoptosis. *Autophagy* 8, 559–576.
- Zhang, W., Chen, L., Liu, J., Chen, B., Shi, H., Chen, H., Qi, H., Wu, Z., Mao, X., Wang, X., Huang, Y., Li, J., Yu, Z., Zhong, M., Wang, T., Li, Q., 2024. Inhibition of autophagy-related protein 7 enhances anti-tumor immune response and improves efficacy of immune checkpoint blockade in microsatellite instability colorectal cancer. *J. Exp. Clin. Cancer Res.* 43, 114.
- Zhang, W., Gai, C., Ding, D., Wang, F., Li, W., 2018. Targeted p53 on small-molecules-induced ferroptosis in cancers. *Front. Oncol.* 8, 507.
- Zhang, Y., Hao, M., Yang, X., Zhang, S., Han, J., Wang, Z., Chen, H.-N., 2024. Reactive oxygen species in colorectal cancer adjuvant therapies. *Biochim. Biophys. Acta Mol. Basis Dis.* 1870, 166922.
- Zhao, J., Wang, X., Mi, Z., Jiang, X., Sun, L., Zheng, B., Wang, J., Meng, M., Zhang, L., Wang, Z., Song, J., Yuan, Z., Wu, Z., 2021. STAT3/miR-135b/NF- $\kappa$ B axis confers aggressiveness and unfavorable prognosis in non-small-cell lung cancer. *Cell Death Dis.* 12, 493.
- Zhao, Y., Hu, X., Liu, Y., Dong, S., Wen, Z., He, W., Zhang, S., Huang, Q., Shi, M., 2017. ROS signaling under metabolic stress: cross-talk between AMPK and AKT pathway. *Mol. Cancer* 16, 79.
- Zhao, Y., Ye, X., Xiong, Z., Ihsan, A., Ares, I., Martínez, M., Lopez-Torres, B., Martínez-Larrañaga, M.-R., Anadón, A., Wang, X., Martínez, M.-A., 2023. Cancer metabolism: The role of ROS in DNA damage and induction of apoptosis in cancer cells. *Metabolites* 13. <https://doi.org/10.3390/metabo13070796>.
- Zhao, Z., 2024. Wnt/ $\beta$ -Catenin signaling pathway in hepatocellular carcinoma: pathogenic role and therapeutic target. *Front. Oncol.* 14.
- Zhidkova, E.M., Lylova, E.S., Grigoreva, D.D., Kirsanov, K.I., Osipova, A.V., Kulikov, E.P., Mertsalov, S.A., Belitsky, G.A., Budunova, L., Yakubovskaya, M.G., Lesovaya, E.A., 2022. Nutritional sensor REDD1 in cancer and inflammation: Friend or foe? *Int. J. Mol. Sci.* 23, 9686.
- Zhidkova, E.M., Lylova, E.S., Savinkova, A.V., Mertsalov, S.A., Kirsanov, K.I., Belitsky, G.A., Yakubovskaya, M.G., Lesovaya, E.A., 2020. A brief overview of the paradoxical role of glucocorticoids in breast cancer. *Breast Cancer (Auckl.)* 14, 1178223420974667.
- Zhong, B., Cheng, B., Huang, X., Xiao, Q., Niu, Z., Chen, Y.-F., Yu, Q., Wang, W., Wu, X.-J., 2021. Colorectal cancer-associated fibroblasts promote metastasis by up-regulating LRG1 through stromal IL-6/STAT3 signaling. *Cell Death Dis.* 13, 16.
- Zhou, J., Li, C., Yao, W., Alsidig, M.C., Huo, L., Liu, H., Miao, Y.-L., 2018. Hypoxia-inducible factor-1 $\alpha$ -dependent autophagy plays a role in glycolysis switch in mouse granulosa cells. *Biol. Reprod.* 99, 308–318.
- Zhou, W., Xu, G., Wang, Y., Xu, Z., Liu, X., Xu, X., Ren, G., Tian, K., 2017. Oxidative stress induced autophagy in cancer associated fibroblast enhances proliferation and metabolism of colorectal cancer cells. *Cell Cycle* 16, 73–81.
- Zhou, Y.-Y., Li, Y., Jiang, W.-Q., Zhou, L.-F., 2015. MAPK/JNK signalling: a potential autophagy regulation pathway. *Biosci. Rep.* 35. <https://doi.org/10.1042/BSR20140141>.
- Zhou, Z., Qu, C., Zhou, P., Zhou, Q., Li, D., Wu, X., Yang, L., 2024. Extracellular vesicles activated cancer-associated fibroblasts promote lung cancer metastasis through mitophagy and mtDNA transfer. *J. Exp. Clin. Cancer Res.* 43, 158.
- Zhu, H., Chang, L.-L., Yan, F.-J., Hu, Y., Zeng, C.-M., Zhou, T.-Y., Yuan, T., Ying, M.-D., Cao, J., He, Q.-J., Yang, B., 2018. AKR1C1 activates STAT3 to promote the metastasis of non-small cell lung cancer. *Theranostics* 8, 676–692.
- Zinn, R.L., Gardner, E.E., Dobromilskaya, I., Murphy, S., Marchionni, L., Hann, C.L., Rudin, C.M., 2013. Combination treatment with ABT-737 and chloroquine in preclinical models of small cell lung cancer. *Mol. Cancer* 12, 16.
- Zong, D., Liu, X., Li, J., Ouyang, R., Chen, P., 2019. The role of cigarette smoke-induced epigenetic alterations in inflammation. *Epigenetics Chromatin* 12, 65.
- Zou, Y., Chen, Z., He, X., He, X., Wu, X., Chen, Y., Wu, X., Wang, J., Lan, P., 2015. High expression levels of unc-51-like kinase 1 as a predictor of poor prognosis in colorectal cancer. *Oncol. Lett.* 10, 1583–1588.

**María Carretero-Fernández:** Early-career researcher involved in translational cancer research. She participates in multiple ongoing projects on solid tumours, gaining experience in molecular oncology and functional characterization of pathways such as autophagy and oxidative stress. She has begun contributing to publications and presenting her work at scientific meetings.

**Antonio José Cabrera-Serrano:** PhD. Researcher with a solid publication record and leadership in cancer research projects. His work integrates multi-omic approaches to study tumour biology in both solid and haematological cancers, focusing on autophagy and oxidative stress. He is active in mentoring and regularly presents at national and international conferences.

**José Manuel Sánchez-Maldonado:** PhD. Researcher with expertise in molecular oncology. He has co-authored multiple studies and meta-analyses on autophagy-related biomarkers in cancer. His work focuses on the role of oxidative stress in tumour progression and therapeutic response, with ongoing contributions to collaborative cancer research projects.

**Lucía Ruiz-Durán:** Early-career researcher focusing on the molecular mechanisms that drive tumor growth and therapy resistance in solid cancers. Her work involves the analysis of signaling pathways and cellular stress responses, with an emphasis on autophagy and oxidative stress. Lucía actively participates in scientific meetings to strengthen her expertise in translational oncology and biomarker discovery.

**Francisco Jiménez-Romera:** Francisco is involved in cancer research, focusing on the molecular mechanisms driving tumour progression. He participates in translational projects aimed at identifying novel biomarkers and therapeutic targets in solid tumours. Francisco has gained experience presenting his work at national and international conferences, contributing to the scientific community as a promising early-career researcher.

**Francisco José García-Verdejo:** MD, PhD. Medical Oncology Specialist, with a primary focus on solid tumours. He leads and participates in phase II and III clinical trials, contributing to the development and evaluation of novel therapeutic strategies in oncology. He is also involved in functional studies aimed at understanding molecular mechanisms underlying tumour biology, including pathways relevant to autophagy and oxidative stress. His work bridges clinical practice and translational research to improve patient outcomes in solid cancers.

**Carmen González-Olmedo:** PhD. Researcher specializing in cancer metabolomics. She has extensive experience in managing research projects and her recent work includes studies on metabolomics biomarkers in breast and colorectal cancer, and the interplay between autophagy and oxidative stress in solid tumours, contributing to advances in personalized cancer therapy.

**Aina Cardús:** Clinical researcher with expertise in the integration of clinical research with patient care, advancing the development of innovative therapies in oncology.

**Leticia Díaz-Beltrán:** PhD. Researcher leading projects in liquid biopsy, NGS, and multi-omics in cancer. Her work focuses on identifying metabolomic and miRNA biomarkers for diagnosis, prognosis, and treatment response, with significant contributions to scientific literature and precision oncology.

**Juan Francisco Gutiérrez-Bautista:** PhD. Researcher with extensive experience in translational immunology and oncology. His work focuses on tumour biology, especially mechanisms involving autophagy and oxidative stress, as well as immune responses relevant to cancer progression. He has contributed to multiple research projects and publications bridging molecular pathways with clinical implications in solid tumours.

**Yolanda Benavente:** PhD. Senior researcher with an extensive publication record in cancer epidemiology, with a particular focus on environmental and genetic risk factors for cancer.

**Fernando Gálvez-Montosa:** MD, PhD. Medical oncologist with expertise in tumour autophagy. His research elucidates how alterations in autophagy influence cancer progression and therapy resistance. He has contributed to defining autophagy as a therapeutic target.

**José Antonio López-López:** MD. Medical Oncology Specialist primarily focusing on digestive, lung, and head and neck cancers. He holds expert qualifications in Immunology and Medical Oncology, reflecting a strong clinical foundation in cancer treatment. He is actively engaged in multidisciplinary oncology care and research, contributing to advancing therapeutic approaches in solid tumours.

**Paloma García-Martín:** MD. Clinician and researcher with a strong focus on cancer biology and translational oncology. Her work centres on understanding mechanisms underlying tumour progression and treatment response, contributing to studies that aim to improve patient outcomes.

**Eva María Pérez:** MD. Clinical researcher with expertise in translational research. Her work focuses on the integration of clinical practice with advanced molecular tools, such as liquid biopsy, next-generation sequencing, and other omics technologies, to identify diagnostic and prognostic biomarkers for personalized treatment in cancer.

**Juan José Rodríguez-Sevilla:** MD, PhD. Clinical researcher leading projects on immune evasion in cancer, particularly NK and T-cell dysfunction. He has published in high-impact journals and his work contributes to understanding immune mechanisms in tumour progression.

**Delphine Casabonne:** PhD. Epidemiologist with expertise in environmental and genetic cancer risk factors. Her research includes the influence of viral infections, lifestyle, and anthropometrics on cancer development, contributing to large-scale studies and international consortia.

**Pedro Sánchez-Rovira:** MD, PhD. Renowned oncologist and expert in breast cancer. He has led over 50 clinical trials and several translational projects in tumour genomics and circulating tumour cells, contributing to major oncology journals and shaping clinical research in solid tumours.

**Fernando Jesús Reyes-Zurita:** PhD. Researcher in drug development targeting cancer pathways, with a focus on oxidative stress and autophagy. He has authored numerous publications and is involved in preclinical studies exploring new therapeutic strategies.

**Juan Sainz:** PhD. His research focuses on the genetic and molecular mechanisms underlying cancer development and progression. He has extensive experience in the study of cellular stress responses, such as autophagy and oxidative stress, and their roles in tumour initiation, immune evasion, and therapy resistance. Dr. Sainz actively participates in several international consortia on colorectal, pancreatic, and prostate cancer, where he integrates genomic, transcriptomic, and proteomic data to identify biomarkers and therapeutic targets. His work bridges basic and translational research, contributing to precision oncology through multi-omic and systems biology approaches.