RESEARCH Open Access



Nutritional impact of eicosapentaenoic acid supplementation (EPA) in patients with locally advanced head and neck cancer: a doubleblind placebo-controlled randomized trial

Lorena Arribas^{1,2*}, Laura Hurtós¹, Anna Esteve^{3,4}, Inmaculada Peiró^{1,2}, Ana Regina González-Tampán¹, Maryam Choulli^{1,2}, Maite Antonio⁵, Esther Vilajosana⁵, Alicia Lozano Borbalas⁶, Miren Taberna⁵ and Ricard Mesía³

Abstract

Background Evidence on the benefits of eicosapentaenoic acid (EPA) supplementation in cancer patients remains inconsistent, likely due to heterogeneity in tumor types, stages, treatment modalities, and nutritional strategies. This study aimed to assess the impact of EPA as a standalone supplement on body weight, body mass index (BMI), and muscle mass preservation in patients with locally advanced squamous cell carcinoma of the head and neck (SCCHN) undergoing curative-intent treatment.

Methods We conducted a double-blind, placebo-controlled randomized trial between December 2015 and September 2018, enrolling 54 patients with advanced SCCHN. Participants received either 2.7 g/day of oral EPA or placebo, alongside standard nutritional support according to European Society for Clinical Nutrition and Metabolism (ESPEN) guidelines. Assessments were performed at baseline (T0), post-induction chemotherapy (T1), and post-treatment (T2), including body composition, nutritional status, functionality, and inflammatory biomarkers. Plasma EPA (% of total fatty acids) was used as a biochemical marker of compliance.

Results Both groups were balanced at baseline (median age 58.9 years; 46.3% malnourished). Neither the intention-to-treat nor per-protocol analyses showed statistically significant differences between groups in weight loss, BMI, or body composition (p > 0.05). In the per-protocol analysis, estimated mean weight loss was -3.81 kg (95% CI: -5.70 to -1.91) in the EPA group, compared to -7.72 kg (95% CI: -12.8 to -6.40) in the placebo group. No differences were observed in nutritional status, energy or protein intake, physical function, or treatment-related toxicity. Complete response rates after oncological treatment were comparable (77.8% EPA vs. 66.7% placebo). Only half of the patients in the EPA group reached an increase in plasma EPA levels consistent with supplementation, and 31.5% discontinued the intervention due to intolerance.

Conclusions Our study did not demonstrate a clear benefit of EPA supplementation on weight, BMI, or muscle preservation in patients with locally advanced SCCHN receiving curative-intent treatment. While EPA was safe and

*Correspondence: Lorena Arribas larribas@iconcologia.net

Full list of author information is available at the end of the article



© The Author(s) 2025. **Open Access** This article is licensed under a Creative Commons Attribution-NonCommercial-NoDerivatives 4.0 International License, which permits any non-commercial use, sharing, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if you modified the licensed material. You do not have permission under this licence to share adapted material derived from this article or parts of it. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit http://creativecommons.org/licenses/by-nc-nd/4.0/.

Arribas et al. Nutrition Journal (2025) 24:140 Page 2 of 13

feasible, the findings do not provide sufficient evidence to support a clinically significant effect. Further research with larger sample sizes and more robust designs is warranted to better understand the potential role of EPA in this population.

Trial registration ClinicalTrials.gov identifier: NCT02715596 (registered on March 22, 2016 at ClinicalTrials.gov). **Keywords** Eicosapentaenoic acid, EPA, Head and neck cancer, Muscle mass, Weight loss, Nutritional intervention

Introduction

Reduced skeletal muscle mass is a known negative prognostic factor in cancer patients [1]. Weight loss has also been associated with decreased cancer survival [2]. In patients with squamous cell carcinoma of the head and neck (SCCHN), both weight loss and low muscle mass have been associated with chemotherapy dose-limiting toxicity [3, 4], late toxicity after radiotherapy [5] and reduced overall and progression-free survival [6]. Nutrition impact symptoms (i. e. symptoms that impair oral intake, such as odyno-dysphagia or anorexia) and severe treatment-related toxicity significantly affect nutritional status in this population [7, 8]. In patients with locally advanced SCCHN, induction chemotherapy may help alleviate initial nutritional impact symptoms and prevent further deterioration [8, 9]. However, during chemoradiation a significant decline of the nutritional status, particularly in muscle and adipose tissue, has been consistently reported, even with nutritional support [10-12]. Malnutrition affects approximately 42% of head and neck patients at diagnosis and increases to up to 77% during treatment [13, 14].

Several translational studies have demonstrated the role of inflammation in cancer-associated weight loss and muscle wasting. Omega-3 fatty acids have been suggested to reduce inflammation in cancer patients [15, 16] potentially contributing to improved body weight and muscle prevention. Eicosapentaenoic acid (EPA), a polyunsaturated fatty acid of the omega-3 class, can be rapidly incorporated into the cell membranes, contributing to metabolic regulation. Various small studies and randomized trials have assessed the effects of EPA or omega-3 supplementation across different cancer types and treatment settings. However, findings remain inconsistent regarding effects on weight loss, body composition and inflammatory biomarkers. A recent systematic review reported a positive effect in body weight in favour of omega-3 supplementation, but no effect on circulating inflammatory markers [17]. Another review concluded that EPA and/or docosahexaenoic acid (DHA) supplementation could help maintain or increase body weight, improve progression-free and overall survival, enhance quality of life, modulate immune parameters, and reduce serious adverse events [18]. Both reviews agreed that further studies are needed to clarify the role of EPA and omega-3 fatty acids during cancer treatment.

In HNC patients, most clinical trials have evaluated omega-3 fatty acids as components of commercial polymeric nutritional supplements [15, 16, 19–22]. A recent study [23] found that fish oils capsules were better tolerated than nutritional drinks with equivalent omega-3 content. To date, no study has evaluated the effect of EPA as a single-agent supplement, combined with an intensive nutritional intervention throughout the entire course of oncological treatment.

Thus, the aim of this study was to investigate whether EPA supplementation could have a positive effect on body weight, BMI and body composition in patients with locally advanced SCCHN undergoing a standardized curative-intent treatment protocol.

Methods

Study design and objectives

This randomized, double-blind placebo-controlled clinical trial (NCT02715596) assigned patients to receive either an EPA supplement or a placebo, using a block randomization method. All patients provided written informed consent prior to enrollment. The study was conducted in accordance with the principles of the Declaration of Helsinki and Good Clinical Practice, and was approved by the Ethics Committee for Clinical Research at Hospital Universitari de Bellvitge (PR261/14). The primary objective was to evaluate the potential beneficial effect of EPA supplementation on body weight, BMI and body composition. Secondary outcomes included EPA compliance, changes in nutritional status, functionality, treatment-related toxicity and inflammatory biomarkers including C-reactive protein (CRP), serum albumin, interleukin- 6 (IL-6) and neutrophil-to-lymphocyte ratio (NLR).

Patients and oncological treatment

Since cancer stage, treatment toxicity and response are closely related to progressive muscle and fat loss over time [24], we focused on a homogeneous treatment plan for patients with locally advanced SCCHN, who typically present with nutritional impact symptoms. To ensure cohort consistency, we excluded other treatment approaches as surgery and early-stage disease. Eligible patients were≥18 years old and diagnosed with stage III–IVa,b SCCHN (7th TNM edition). All were scheduled to receive curative-intent treatment consisting of induction chemotherapy followed by chemoradiotherapy (CRT) or radiotherapy (RT) plus

Arribas et al. Nutrition Journal (2025) 24:140 Page 3 of 13

cetuximab, according to local clinical guidelines [25]. Additional inclusion criteria required pathologically confirmed squamous cell carcinoma of the oral cavity, oropharynx, hypopharynx, larynx, or nasopharynx, with no prior history of recurrent disease.

The induction chemotherapy regimen (Fig. 1) was based on the TPF scheme: Taxanes, Platinum and 5-Fluorouracil [26]. For the subsequent radiotherapy phase, patients received either cisplatin (100 mg/m² every 21 days) [27] or cetuximab (250 mg/m² weekly, with a 400 mg/m² loading dose) [28], administered concurrently with intensity-modulated radiotherapy (IMRT) at a total dose of 69.96 Gy (2.12 Gy per session). Assignment to CRT or RT plus cetuximab was determined by the multidisciplinary head and neck tumor board, based on individual treatment response and/or toxicity during induction chemotherapy [29].

Study products and compliance

Patients were instructed to take one stick pack daily from the first day of chemotherapy and continuing throughout the entire treatment period (i.e. induction chemotherapy followed by CRT or RT plus cetuximab). Those in the intervention group received 2.7 g/day of EPA in liquid stick-pack format, while the placebo group received 2.7 g/day of mineral oil, matched in colour and odor to the EPA product. Given the poor compliance reported in previous studies [30, 31], a liquid formulation was chosen to enhance adherence, particularly in HNC patients who often experience swallowing difficulties. Both EPA and placebo supplements were manufactured, blinded, and supplied by Ferrer SA (Spain). The manufacturer had no access to the study results and did not influence the study's design or conclusions. The EPA formulation did not contain DHA.

Compliance was assessed at each chemotherapy cycle by tracking the number of stick packs distributed,

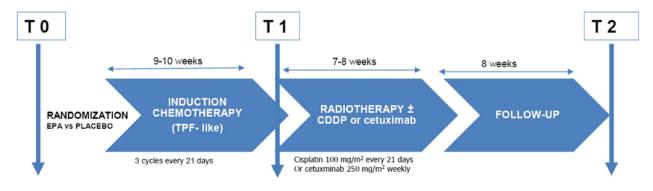
returned and the corresponding dates. In addition, plasma EPA levels were used as a biochemical marker of compliance. Total lipids were extracted from plasma samples, and fatty acid methyl esters (FAMEs) were analyzed by gas chromatography (GC). Results were expressed as the proportion of EPA relative to total identified plasma fatty acids (relative percent).

Assessments

Body weight, BMI and body composition were assessed before starting the oncological treatment (T0), after the induction chemotherapy (T1) and two months following completion of RT (T2). BMI was also calculated as [(weight (kg)/height (m²)] and categorized according to WHO standards.

All patients underwent an abdominal computed tomography (CT) scan prior to treatment and again approximately two months completing RT (i.e. an overall span of 7–8 months), as part of routine clinical practice. The third lumbar vertebra (L3) on axial cross-section images was used as the reference point for analysis, following established methodology [32-34]. Skeletal muscle and adipose tissue areas were determined using SliceOmatic© software (v5.0 Rev 8, Tomovision, Magog, Canada), applying standard Hounsfield Unit (HU) ranges: -29 to +150 HU for skeletal muscle, -150 to -50 HU for visceral adipose tissue (VAT), -190 to -30 HU for subcutaneous adipose tissue (SAT). Cross-sectional areas were normalized for height and reported as skeletal muscle index (SMI, cm²/m²) and total adipose tissue index (TATI, cm²/m², including SAT and VAT) [34–36]. Analyses were performed by a single observer blinded to clinical data, with precision of ±0.92 cm² for skeletal muscle area and ± 1.37 cm² for total adipose tissue area.

Nutritional data included baseline height, percentage of weight loss in the 3 months prior to induction chemotherapy, current weight, nutritional status



T0: Before starting the oncological treatment (baseline)

T1: After the induction chemotherapy

T2: 2-months after the end of the radiotherapy

Fig. 1 Oncological treatment plan

Arribas et al. Nutrition Journal (2025) 24:140 Page 4 of 13

(Patient-Generated Subjective Global Assessment, PG-SGA) [37, 38], and the type of nutritional support at each time point. Energy and protein intake were estimated via 24-h dietary recall using Dietsource© software (v3.0, InterCath, Spain), and expressed in kcal/day and g/day.

Inflammatory markers were assessed through routine laboratory testing. Albumin and CRP were used to calculate the Glasgow Prognostic Score (GPS) [39]; the NLR was derived from complete blood counts [40]. Interleukin-6 (IL-6) was measured using a human cytokine-specific ELISA kit (Biosource Europe, Belgium), following the manufacturer's instructions.

Functional status was assessed via handgrip strength, using a hydraulic dynamometer (SI Instruments Pty Ltd, Adelaide, Australia). The test was performed on the dominant hand, with the patient seated, shoulder in adduction and neutral rotation, and elbow flexed at 90°, always using the same hand [41].

Additional data were collected on nutrition impact symptoms and treatment-related toxicity, classified according to the National Cancer Institute – Common Terminology Criteria for Adverse Events (CTCAE), version 4.0 [42].

Sample size calculation

To determine the sample size, we considered the objective of the study to evaluate the effect of EPA supplementation, comparing two groups: one supplemented and one control. The expected mean relative weight loss from baseline (T0) in the control group was -10% and -3% in the supplemented group. Assuming a standard deviation (SD) of 9, with a significance level of 5% and a power of 80%, 54 patients were needed (27 per group) to find statistically significant differences between the two groups.

Statistical analyses

Descriptive statistics of baseline characteristics in the EPA and placebo participants were reported. The coprimary end points were the percentage change in weight and body composition from T0 and the absolute weight and body composition change from T0. Intention-to-treat (ITT) analyses as well as per-protocol (PP) analyses were conducted for outcomes of weight and body composition. Based on the ITT principle, all participants who were randomized were included in the analysis, regardless of protocol compliance. Per-protocol analysis compared intervention groups that included only those patients who completed the intervention.

The percentage of change from baseline in weight and body composition outcomes were assessed using an analysis of covariance model (ANCOVA) adjusted for age, sex, malnutrition. Each ANCOVA model further comprised the baseline variable of the respective outcome at T0. The dynamics estimations in weight

and body composition outcomes at each time point were obtained from linear mixed models (LMM) including intervention group and time as fixed factors, a group x time interaction term and a random intercept for subject. LMM were adjusted for age, sex and malnutrition, and each model further comprised the baseline variable of the respective outcome at T0. In the LMM models, p-values for the T2 vs T0 comparisons within groups were obtained using Tukey's method for multiple post-hoc comparisons between time and group. All results will be reported as mean and 95% confidence interval (CI), adjusted for the previously mentioned covariates.

In the ITT analysis, missing data due to noncompliance or dropout were imputed using the Reference-based multiple imputation method with the *jump to reference* (J2R) approach [43], with n = 100 imputations. The R package RefBasedMI [44], was used to build separate imputation models to each end point outcome and to generate imputed data considering multiple imputation under the assumption that, after dropout, participants in the EPA group no longer receive intervention and behave like those in the placebo group following their last observed time point. For the ANCOVA models, the transformation into percentage change in weight and body composition was performed later. The estimated marginal means (EMMs) were obtained using the R package emmeans [45].

The validity of PP effect estimates required correct adjustment for confounding due to incomplete adherence to the assigned interventions [46]. The Inverse Probability Weighting (IPW) enabled us to appropriately adjust for postrandomization biases due to protocol compliance. The probability of adherence in each group of treatment was estimated using logistic regression models considering age, sex, BMI and malnutrition as common regressors for the adherence status and the outcomes. Stabilized weights were computed from the estimated probabilities and further used in the ANCOVA and linear mixed models to reduce imbalance between groups of interventions in measured confounders [47].

Statistical analyses were performed using R version 4.4.2 with the RStudio 2024.04.0.

Results

Study population

Between December 2015 and September 2018, 54 patients with locally advanced SCCHN were enrolled and randomly assigned to receive EPA (n = 27) or placebo (n = 27). Figure 2 presents the participant flow diagram, including reasons for exclusion. Differences in sample size for the PP analysis were due to protocol deviations, as detailed in the Statistical Analyses section.

Arribas et al. Nutrition Journal (2025) 24:140 Page 5 of 13

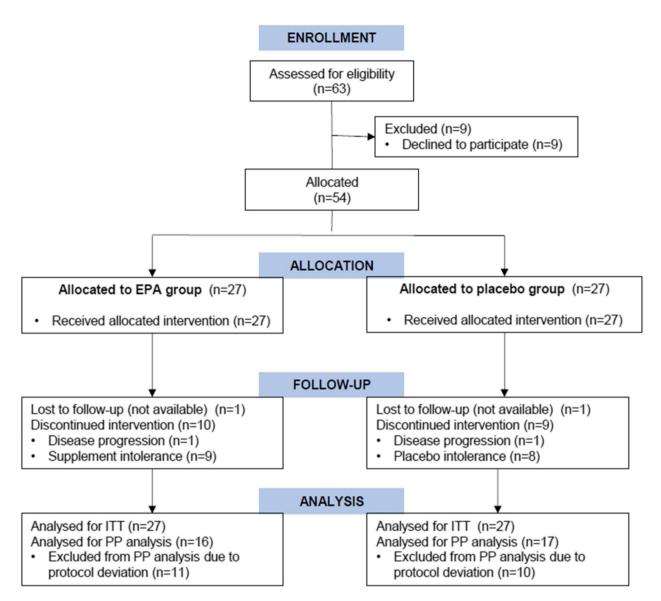


Fig. 2 Consort diagram

Baseline characteristics are shown in Table 1. The median age was 58.9 years (SD 6.8) and 78% of participants were male. The most common tumor site was larynx (37.0%). Following induction chemotherapy, 49.1% received RT and cetuximab. The median duration from diagnosis (T0) to completion of oncological treatment (T2) was 6.2 months (SD 2.1). Most patients had good baseline clinical status (ECOG performance status 0–1). At diagnosis, 54% were classified as well-nourished according to the PG-SGA, and 39% were overweight or obese. Sarcopenia, as defined by Martin et al. [2] was present in 22% of cases, with a mean skeletal muscle index (SMI) of 50.9 cm²/m² (SD 11.4).

After completion of treatment, complete response rates were similar between groups: 78% (18/27) in the EPA group vs. 67% (21/27) in the placebo group.

Coprimary end points: weight and body composition *ITT analysis*

In the ITT analysis, the mean percentage of weight loss from baseline (T0) to the end of treatment (T2) was -7.88% (95% CI: -13.4% to -2.40%) in the EPA group and -5.15% (95% CI: -11.4% to 1.11%) in the placebo group (Fig. 3A), with no statistically significant difference between groups (-2.73, 95% CI: -10.6; 5.17; p=0.474) (Table 2).

Modelling the dynamics of weight over time (T0, T1, T2), both groups showed significant weight loss. The estimated mean change was -4.25 kg (95% CI: -7.04 to -1.45; p = 0.0373) in the EPA group and -5.05 kg (95% CI: -8.20 to -1.90; p = 0.0256) in the placebo group. A similar pattern was observed for BMI (Fig. 4), with mean reductions of -1.55 kg/m² (95% CI: -2.54 to -0.56; p = 0.0286)

Arribas et al. Nutrition Journal (2025) 24:140 Page 6 of 13

Table 1 Characteristics of the participants at baseline (T0)

	Total (n = 54)	Placebo (<i>n</i> = 27)	EPA (n = 27)
Age, year	58.9 (6.8)	59.7 (6.0)	58.1 (7.6)
Sex, male, n (%)	42 (77.8%)	19 (70.4%)	23 (85.2%)
Primary tumour site, n (%)			
Oral cavity	9 (16.7%)	5 (18.5%)	4 (14.8%)
Oropharynx	10 (18.5%)	5 (18.5%)	5 (18.5%)
Larynx	20 (37.0%)	12 (44.4%)	8 (29.6%)
Hypopharynx	10 (18.5%)	3 (11.1%)	7 (25.9%)
Nasopharynx	5 (9.3%)	2 (7.4%)	3 (11.1%)
HPV ^a positive	3 (33.3%)	1 (25.0%)	2 (40.0%)
Pathological Stage (TNM), n (%)			
III	12 (22.2%)	5 (18.5%)	7 (25.9%)
IVa	31 (57.4%)	18 (66.7%)	13 (48.1%)
IVb	11 (20.4%)	4 (14.8%)	7 (25.9%)
Performance status, n (%)			
0	8 (14.8%)	3 (11.1%)	5 (18.5%)
1	43 (79.6%)	23 (85.2%)	20 (74.1%)
2	3 (5.6%)	1 (3.7%)	2 (7.4%)
Type of concomitant treatment, n (%)			
CDDP ^b	20 (37.7%)	9 (34.6%)	11 (40.7%)
Cetuximab	26 (49.1%)	14 (53.8%)	12 (44.4%)
Radiotherapy alone	4 (7.5%)	1 (3.8%)	3 (11.1%)
Others ^c	3 (5.7%)	2 (7.7%)	1 (3.7%)
Active smoker	17 (31.5%)	10 (37.0%)	7 (25.9%)
Active drinker	9 (16.7%)	4 (14.8%)	5 (18.5%)
PG-SGA ^d			
A—well nourished	29 (53.7%)	18 (66.7%)	11 (40.7%)
B—moderate malnutrition	16 (29.6%)	5 (18.5%)	11 (40.7%)
C—severe malnutrition	9 (16.7%)	4 (14.8%)	5 (18.5%)
Weight, kg	67.6 (15.0)	71.0 (16.2)	64.2 (13.0)
BMI ^{e,} kg/m ² , n (%)	24.4 (5.1)	25.7 (5.4)	23.1 (4.4)
Underweight (< 18.5 kg/m²)	4 (7.4%)	2 (7.4%)	2 (7.4%)
Normal weight (18.5–24.9 kg/m²)	29 (53.7%)	13 (48.1%)	16 (59.3%)
Overweight (25–29.9 kg/m²)	13 (24.1%)	6 (22.2%)	7 (25.9%)
Obesity (≥ 30 kg/m²)	8 (14.8%)	6 (22.2%)	2 (7.4%)
Albumin, g/L	44.8 (4.4)	45.5 (3.5)	44.1 (5.0)
CRP ^f , mg/L	22.0 (23.9)	21.4 (21.3)	22.7 (26.8)
IL-6 ^g , pg/mL	11.0 (10.9)	10.6 (10.8)	11.3 (11.2)
SMI ^{h,} cm ² /m ²	50.9 (11.4)	50.7 (12.5)	51.0 (10.4)
TATI ⁱ , cm²/m²	112.5 (50.8)	123.5 (54.2)	101.2 (45.4))
Energy intake, kcal/d	2120.1 (849.0)	2207.2 (975.0)	2233.0 (709.2)
Protein intake, g/d	80.2 (28.4)	78.9 (29.4)	81.6 (27.8)
Hand grip strength, kg	30.0 (10.5)	31.3 (12.4)	28.7 (8.4)

Values are presented as mean (standard deviation) for continuous variables and as number (percentage) for categorical variables. No formal statistical comparisons were conducted, as these represent baseline characteristics

^aHPV: Human papillomavirus

^bCDDP: cisplatin

 $^{^{}c}Others: include one patient loss of follow up before stating the concomitance and two patients with progression and change the oncological treatment\\$

 $^{{}^{\}rm d}{\rm PG\text{-}SGA:}\ {\rm Patient}\ {\rm Generated}\ {\rm Subjective}\ {\rm Global}\ {\rm Assessment}$

^eBMI: body mass index

^fCRP: C- Reactive Protein

gIL-6: Interleukin-6

^hSMI: Skeletal muscle index

ⁱTATI: Total adipose tissue index

Arribas et al. Nutrition Journal (2025) 24:140 Page 7 of 13

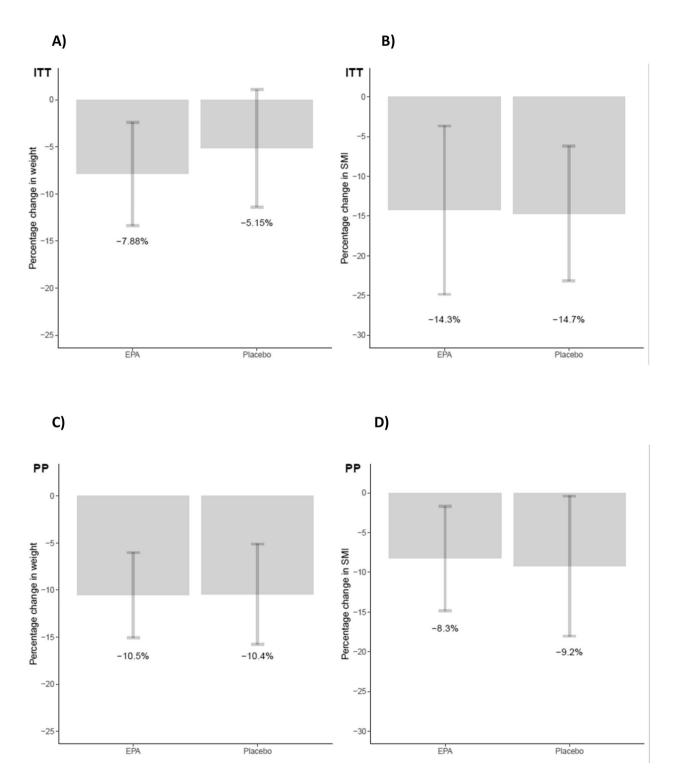


Fig. 3 Mean percentage change in body weight and skeletal muscle index (SMI) from baseline (T0) and end of treatment (T2) in the EPA and placebo groups according intention-to-treat (ITT) and per-protocol (PP) analysis. Bars represent mean percentage change from baseline (T0) to T2; error bars indicate 95% confidence intervals. Estimates are derived from an analysis of covariance model (ANCOVA) adjusted for age, sex, malnutrition, and baseline values

Arribas et al. Nutrition Journal (2025) 24:140 Page 8 of 13

Table 2 Comparisons of the coprimary end points for the intention-to-treat (ITT) and the per-protocol sample (PP)

	Intention-to-treat (ITT)		Per- protocol (PP)		
Variable	Contrast	Estimate (95% CI)	<i>P</i> -value*	Estimate (95% CI)	<i>P</i> -value*
% Weight change	EPA vs Placebo	-2.73 (-10.6; 5.17)	0.4747	-0.10 (-6.79; 6.59)	0.9752
Weight (kg)	EPA: T2 vs T0	-4.25 (-7.04; -1.45)	0.0373	-3.81 (-5.70; -1.91)	0.0009
	Placebo: T0 vs T2	-5.05 (-8.20; -1.90)	0.0256	-7.72 (-12.8; -6.40)	< 0.0001
BMI (kg/m ²)	EPA: T2 vs T0	-1.55 (-2.54;0.56)	0.0286	-1.35 (-1.96; -0.74)	0.0002
	Placebo: T2 vs T0	-1.75 (-2.92;0.58)	0.0438	-2.61 (-3.59; -1.63)	< 0.0001
% SMI change	EPA vs Placebo	-0.42 (-13.3; 12.5)	0.9477	-0.95 (-10.4; 8.44)	0.8365
SMI (cm ² /m ²)	EPA: T2 vs T0	-7.00 (-11.3; -2.66)	0.0235	-4.99 (-7.52; -2.46)	0.0015
	Placebo: T2 vs T0	-7.67 (-11.9; -3.47)	0.0067	-5.68 (-9.73; -1.62)	0.0660

This table represents estimated changes in clinical outcomes (weight, BMI, body composition) between baseline (T0), post-induction chemotherapy (T1) and end of treatment (T2) in the EPA and placebo groups

EPA vs Placebo: estimate of the mean difference between arms of the outcome percent change. Estimates are derived from an analysis of covariance model (ANCOVA) adjusted for age, sex, malnutrition. Each ANCOVA model further comprised the baseline variable of the respective outcome at T0

EPA (or Placebo): T2 vs T0: estimate of the mean difference between time points derived from a linear mixed model adjusted for age, sex and malnutrition. Each linear mixed model further comprised the baseline variable of the respective outcome at T0

*p-values of EPA and placebo: T2 vs T0 comparisons are obtained using Tukey's method for multiple post-hoc comparisons between time and group

For the ITT analysis imputations were performed based on age, sex, malnutrition, stage, body mass index and respective outcome at T0

in the EPA group and -1.75 kg/m² (95% CI: -2.92 to -0.58; p = 0.0438) in the placebo group (Table 2).

Regarding SMI, the mean percentage loss was -14.3% (95% CI: -24.9% to -3.66%) in the EPA group and -14.7% (95% CI: -23.2% to -6.19%) in the placebo group (Fig. 3B), with no significant difference between groups (difference: 0.42%, 95% CI: -12.5% to 13.3%; p=0.9477) (Table 2). Mean absolute reductions in SMI were also similar: -7.00 cm²/m² (95% CI: -11.3 to -2.66) in the EPA group and -7.67 cm²/m² (95% CI: -11.9 to -3.47) in the placebo group.

Other body composition compartments (VATI, SATI, TATI) showed no statistically significant differences between groups (see Supporting Information Table S1). Nevertheless, a trend toward smaller decreases in the EPA group was observed across several measures (see Supporting Information Figure S1).

PP analysis

The per-protocol analysis included 33 patients who completed the intervention at T2: 16 in the EPA group and 17 in the placebo group.

The mean percentage of weight loss was -10.5% (95% CI: -15.1% to -6.02%) in the EPA group and -10.4% (95% CI: -15.8% to 5.11%) in the placebo group (Fig. 3C), with no statistically significant difference between groups (-0.10%, 95% CI: -6.79 to 6.59; p = 0.9752) (Table 2).

When modeling weight dynamics over time, both groups showed significant weight loss. In the EPA group, the estimated mean change was -3.81 kg (95% CI: -5.70 to -1.91; p = 0.0009), compared to -7.72 kg (95% CI: -12.8 to -6.40; p < 0.001) in the placebo group. A similar trend was observed for BMI as shown in Fig. 4, with reductions of -1.35 kg/m² (95% CI: -1.96 to -0.74;

p = 0.0002) in the EPA group and -2.61 kg/m^2 (95% CI: -3.59 to -1.63; p < 0.0001) in the placebo group (Table 2).

SMI decreased in both groups, with a mean percentage loss of -8.3% (95% CI: -14.9% to -1.67%) in the EPA group and -9.2% (95% CI: -18.0% to -0.39%) in the placebo group (Fig. 3D). The difference was not statistically significant (mean difference: 0.95%, 95% CI: -10.4% to 8.44%; p=0.8365). Absolute changes in SMI were also similar: -4.99 cm²/m² (95% CI: -7.52 to -2.46) in the EPA group vs -5.68 cm²/m² (95% CI: -9.73 to -1.62) in the placebo group.

No statistically significant differences were observed in the percentage change of adipose compartments (VATI, SATI, TATI) between groups (Supporting Information Table S1). Nevertheless, a trend toward smaller decreases in the EPA group was again observed across all compartments (Supporting Information Figure S1).

EPA compliance

At baseline, plasma EPA levels (expressed as % of total fatty acids) were similar between groups: 0.7% in the EPA group vs 0.5% in the placebo group. At T1 and T2, plasma EPA levels increased in the EPA group but remained stable in the placebo group.

At T1, 10 patients (62.5%) in the EPA group showed increases in plasma EPA levels consistent with incorporation of the supplement, whereas no increased were observed in the placebo group. By the end of treatment (T2), approximately half of the patients in the EPA group maintained elevated EPA levels (Table 3).

A total of 17 participants (31.5%) discontinued the intervention due to intolerance. Reported side effects included nausea, vomiting, heartburn, gastric fullness, and reflux, and were reported in both groups: 9 in the EPA group and 8 in the placebo group.

Arribas et al. Nutrition Journal (2025) 24:140 Page 9 of 13

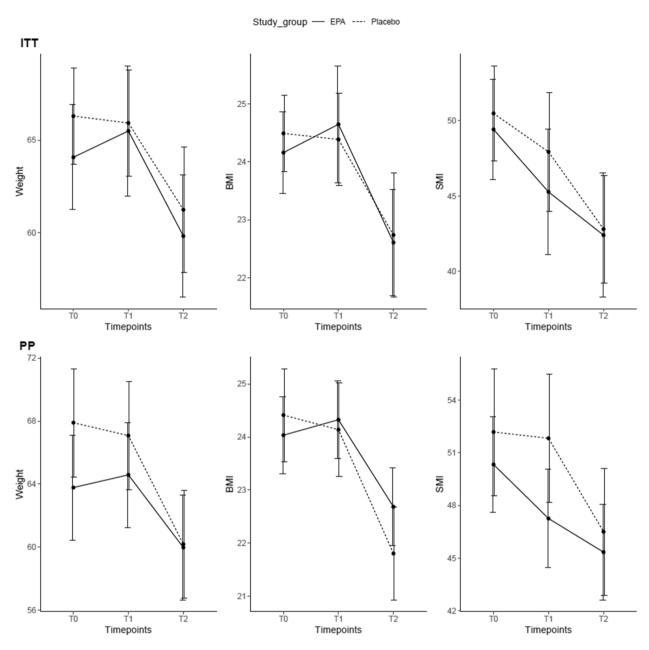


Fig. 4 Dynamic changes in body weight, body mass index (BMI) and skeletal muscle index (SMI) at baseline (T0), after induction chemotherapy (T1) and after the end of the oncological treatment (T2) according the intention-to-treat (ITT) and per-protocol (PP) analyses. Dots represent marginal means at each timepoint; error bars indicate 95% confidence intervals. Estimated are derived from a linear mixed models adjusted for age, sex, malnutrition status, and baseline values

Changes in nutritional, functional, inflammatory parameters over time

The evolution of nutritional, functional, and inflammatory parameters after induction chemotherapy (T1) and at the end of treatment (T2) is presented Table 3.

Nutritional status At baseline, 59.2% of patients in the EPA group were malnourished compared to 33.3% in the placebo group. By T1, malnutrition prevalence decreased in the EPA group and increased in the placebo group,

resulting in similar rates (50.0% vs. 47.6%). This pattern remained consistent at T2. Dietary intake, including energy and protein consumption, showed no significant intra- or between-group differences at any time point.

Nutritional support All participants received nutritional support in accordance with ESPEN guidelines. No significant differences were observed between groups regarding the need for different types of nutritional support at either T1 or T2. At baseline, two patients in the EPA group and

Arribas et al. Nutrition Journal (2025) 24:140 Page 10 of 13

Table 3 Nutritional, functional, inflammatory and compliance parameters at T1 and T2

	After induction chemotherapy (T1)		End of radical treat	ment (T2)
	EPA (n = 18)	Placebo (n=21)	EPA (n = 16)	Placebo (<i>n</i> = 17)
Malnutrition ^a n (%)	9 (50.0)	10 (47.6)	14 (87.5)	14 (82.4)
Energy intake, kcal/d	2401.1 (604.3)	2319.9 (494.6)	2222.8 (642.3)	2210.4 (770.6)
Protein intake, g/d	101.2 (33.1)	86.1 (31.6)	96.0 (36.6)	96.8 (37.5)
Enteral nutrition, n (%)	2 (7.4)	1 (3.7)	2 (11.8)	6 (35.3)
Hand-grip strength, kg	27.8 (8.5)	33.0 (9.6)	25.5 (8.2)	31.4 (12.2)
GPS ^b n (%)				
0	10 (58.8)	10 (50.0)	11 (68.8)	9 (56.2)
1	5 (29.4)	10 (50.0)	4 (25.0)	6 (37.5)
2	2 (11.8)	0 (0.0)	1 (6.2)	1 (6.2)
IL-6 ^c pg/mL	16.7 (23.8)	10.6 (7.9)	13.8 (15.2)	19.2 (24.5)
NLR ^d	2.5 (1.6)	2.6 (2.2)	8.4 (14.6)	3.5 (1.6)
EPA (% of total plasma fatty acids)	3.1 (2.3)	0.7 (0.5)	3.5 (3.1)	0.7 (0.4)

^{*}Values are presented as mean (standard deviation, SD) unless otherwise indicated. EPA measured in plasma and expressed as percentage of total identified fatty acids, based on gas chromatography analysis

one in the placebo group required enteral nutrition, and these same individuals remained on support at T1. By T2, six patients in the placebo group and two in the EPA group required enteral feeding.

Functionality Hand-grip strength was preserved in both groups throughout the duration of the treatment, with no significant variations observed between the groups or over the course of the treatment.

Inflammatory biomarkers There were no differences between groups regarding serum levels of proinflammatory biomarkers such as IL6, GPS and NLR.

Toxicity

No differences were observed in the incidence or severity of toxicities between the EPA and placebo groups.

At baseline, nine patients reported dysphagia to liquids (five in the EPA group, four in the placebo group), including two cases classified as grade 3. Dysphagia to solids (grade 3–4) affected 20 patients, and 19 reported odynophagia across all severity grades.

After induction chemotherapy, dysphagia persisted in two patients for liquids and in four for solids, with no cases exceeding grade 2 severity. Odynophagia remained in six patients, with one case classified as grade 3. Other nutrition impact symptoms emerged, including xerostomia (11 patients) and taste alterations (22 patients), all of mild to moderate severity (grade 1–2). Nausea and vomiting, likely related to EPA/placebo supplementation, were reported similarly in both groups.

By the end of treatment, dysphagia to liquids was reported in five patients, and dysphagia to solids in seven, prompting enteral nutrition in two patients from the EPA group and six from the placebo group. Xerostomia became the most prevalent symptom (29 patients), followed by dysgeusia (27 patients) and fatigue (26 patients). Anorexia progressively worsened throughout treatment, with the most pronounced impact observed after treatment completion.

Discussion

This study did not demonstrate a clear benefit of EPA supplementation in attenuating weight loss, BMI decline, or muscle depletion in patients with locally advanced SCCHN undergoing curative-intent treatment. Both intention-to-treat and per-protocol analyses failed to show statistically significant effects. However, among those who completed the protocol, a modest trend toward reduced weight and muscle loss in the EPA group was observed, despite most not achieving optimal biochemical compliance.

No significant differences were found in nutritional status, need for nutritional support, functionality, inflammatory biomarkers, or treatment-related toxicity. It is important to note that the study was not powered to detect differences in these secondary outcomes.

EPA has been explored for its potential role in counteracting muscle protein degradation through inhibition of proteolytic pathways such as the ubiquitin–proteasome system [48]. It was selected as the sole supplement in this trial based on previous evidence suggesting it is the main active component responsible for the muscle-preserving effects observed in fish oil formulations [49].

Previous meta-analyses have shown a positive association between omega-3 dose and body weight gain in cancer patients, but little effect on BMI or body composition [50]. In our study, EPA compliance was assessed

 $^{^{}m a}$ Malnutrition defined as Patients Generated Subjective Global Assessment (PG-SGA) B+C

^bGPS: Glasgow Prognostic Score

clL-6: interleukin 6

^dNLR: neutrophil-to-lymphocyte ratio

Arribas et al. Nutrition Journal (2025) 24:140 Page 11 of 13

biochemically, and plasma EPA levels confirmed partial adherence. Although prior studies suggested that ≥ 2 g/day of oral EPA supplementation may help stabilize weight [51], we found no correlation between plasma EPA levels and clinical outcomes, even in patients with higher biochemical incorporation.

To explore whether a dose—response relationship existed in our cohort, we conducted a series of post hoc correlation analyses between plasma EPA levels and clinical outcomes. However, no significant associations were found, even among patients with higher biochemical compliance.

Our findings contrast with a previous single-blind RCT in SCCHN patients that reported benefits in weight and lean mass using an EPA-enriched high-protein supplement [15]. However, in that study, differences in the supplement formulations may have confounded the effects attributed to EPA. In contrast, both groups in our trial received standardized nutritional support, isolating the role of EPA. Consistent with other trials [22], we did not observe significant changes in inflammatory biomarkers such as CRP or IL-6.

The potential of EPA to enhance chemotherapy sensitivity remains under investigation [52, 53]. In our cohort, disease control and toxicity profiles were similar between groups. Importantly, adverse effects did not seem to impair adherence or influence clinical outcomes.

Compared to previous trials, our study offers a homogeneous cohort of locally advanced SCCHN patients, all receiving a uniform treatment protocol. We also included a longer follow-up, capturing outcomes not only during treatment but also during the recovery phase, where post-treatment inflammation is known to persist. Most prior trials assessed EPA over 4 to 12 weeks, while our study spanned 6.5 months from baseline.

Limitations include a small sample size and high dropout rate. Although we enrolled sufficient participants for baseline assessments, over one-third discontinued the intervention, largely due to intolerance. The liquid formulation, chosen to accommodate swallowing difficulties, may have hindered compliance due to taste or gastrointestinal side effects. Use of alternative formulations (e.g., capsules) might improve adherence in future studies. Moreover, lack of randomization stratification may have introduced imbalances between groups.

In conclusion, this study did not find sufficient evidence that EPA supplementation significantly attenuates weight loss, BMI decline, or muscle loss in patients with locally advanced SCCHN undergoing curative-intent therapy. However, due to the limited sample size and adherence challenges, these findings should be interpreted cautiously. Further research with larger, stratified cohorts and optimized delivery methods is warranted to better determine the clinical impact of EPA.

Supplementary Information

The online version contains supplementary material available at https://doi.or q/10.1186/s12937-025-01203-8.

Supplementary Material 1.

Supplementary Material 2.

Acknowledgements

Ferrer Spain is acknowledged for supplementing EPA. We would like to express our gratitude to the patients and caregivers who

generously participated in the study. Their commitment and calegivers who generously participated in the study. Their commitment and collaboration were essential to its success. We also want to thank to the members of the Clinical Nutrition Unit and the Head and Neck Unit for their invaluable assistance in participant recruitment and the smooth execution of the study. Additionally, we are deeply grateful to the neuroradiology department at IDI-Bellvitge University Hospital for their support in coordinating the scheduling of L3 CT scans to coincide with routine cervical CT scans.

Trial registration

Clinical Trial registration: NCT02715596 (registered on March 22, 2016 at ClinicalTrials.gov).

Authors' contributions

LA, LH, RM contributed to the conceptualization of the study and methodology. LA, LH, ARGT and AE participated in the data curation. Formal analysis was carried out by AE. Funding acquisition was by MA and LA. LA, LH, IP, EV, AL, MT contributed to the investigation and resources. LA and AE prepared the original draft. RM and MT supervised the study. All authors contributed to the article and approved the submitted version.

Funding

This work was supported by Ferrer Spain. The sponsor had no role in the design of the study, data collection, analysis, interpretation of the results or writing of the manuscript.

Data availability

No datasets were generated or analysed during the current study.

Declarations

Ethics approval and consent to participate

The study was conducted in accordance with the principles of the Declaration of Helsinki and Good Clinical Practice, and was approved by the Ethics Committee for Clinical Research at Hospital Universitari de Bellvitge (PR261/14). All participants provided written informed consent prior to inclusion in the study.

Competing interests

The authors declare no competing interests.

Author detail:

¹Clinical Nutrition Unit, Catalan Institute of Oncology (ICO), L'Hospitalet de Llobregat, Gran Via de L'Hospitalet, 199-203, Barcelona 08908, Spain ²Bellvitge Biomedical Research Institute (IDIBELL), L'Hospitalet de Llobregat, Barcelona, Spain

³Medical Oncology Department, Catalan Institute of Oncology (ICO)-Badalona, B-ARGO Group, Barcelona, Spain

⁴Translational Program in Cancer Research (CARE), Germans Trias I Pujol Research Institute (IGTP), Badalona, Spain

⁵Medical Oncology Department, Catalan Institute of Oncology (ICO), ONCOBELL, L'Hospitalet de Llobregat, Barcelona, Spain

⁶Radiation Oncology Department, Catalan Institute of Oncology (ICO), L'Hospitalet de Llobregat, Barcelona, Spain

Received: 11 January 2025 / Accepted: 6 August 2025 Published online: 26 September 2025 Arribas et al. Nutrition Journal (2025) 24:140 Page 12 of 13

References

- Shachar SS, Williams GR, Muss HB, Nishijima TF. Prognostic value of sarcopenia in adults with solid tumours: a meta-analysis and systematic review. Eur J Cancer. 2016;57:58–67. https://doi.org/10.1016/j.ejca.2015.12.030.
- Martin L, Senesse P, Gioulbasanis I, Antoun S, Bozzetti F, Deans C, et al. Diagnostic criteria for the classification of cancer-associated weight loss. J Clin Oncol. 2015;33:90–9. https://doi.org/10.1200/JCO.2014.56.1894.
- Bril SI, Al-Mamgani A, Chargi N, Remeijer P, Devriese LA, de Boer JP, et al. The
 association of pretreatment low skeletal muscle mass with chemotherapy
 dose-limiting toxicity in patients with head and neck cancer undergoing primary chemoradiotherapy with high-dose cisplatin. Head Neck. 2022;44:189–
 200. https://doi.org/10.1002/hed.26919.
- Sealy MJ, Dechaphunkul T, van der Schans CP, Krijnen WP, Roodenburg JLN, Walker J, et al. Low muscle mass is associated with early termination of chemotherapy related to toxicity in patients with head and neck cancer. Clin Nutr. 2019. https://doi.org/10.1016/j.clnu.2019.02.029.
- van Rijn-Dekker MI, van den Bosch L, van den Hoek JGM, Bijl HP, van Aken ESM, van der Hoorn A, et al. Impact of sarcopenia on survival and late toxicity in head and neck cancer patients treated with radiotherapy. Radiother Oncol. 2020;147:103–10. https://doi.org/10.1016/j.radonc.2020.03.014.
- Takenaka Y, Oya R, Takemoto N, Inohara H. Predictive impact of sarcopenia in solid cancers treated with immune checkpoint inhibitors: a meta-analysis. J Cachexia Sarcopenia Muscle. 2021. https://doi.org/10.1002/JCSM.12755.
- Farhangfar A, Makarewicz M, Ghosh S, Jha N, Scrimger R, Gramlich L, et al. Nutrition impact symptoms in a population cohort of head and neck cancer patients: Multivariate regression analysis of symptoms on oral intake, weight loss and survival. Oral Oncol. 2014;50:877–83. https://doi.org/10.1016/j.oralon cology.2014.06.009.
- Arribas L, Hurtós L, Taberna M, Peiró I, Vilajosana E, Lozano A, et al. Nutritional changes in patients with locally advanced head and neck cancer during treatment. Oral Oncol. 2017;71:67–74. https://doi.org/10.1016/j.oraloncology. 2017.06.003.
- Silver HJ, Dietrich MS, Murphy BA. Changes in body mass, energy balance, physical function, and inflammatory state in patients with locally advanced head and neck cancer treated with concurrent chemoradiation after lowdose induction chemotherapy. Head Neck. 2007;29:893–900. https://doi.org/ 10.1002/hed.20607.
- Arribas L, Sabaté-Llobera A, Taberna M, Pallarés N, Narro Marin A, Virgili N, et al. Adequacy of nutritional support using computed tomography (CT) in patients with head and neck cancer (HNC) during chemo-radiotherapy (CRT). Eur J Clin Nutr. 2021. https://doi.org/10.1038/s41430-021-00863-z.
- Choi Y-C, Chan P-C, Cheung K-WA, Huang J-J, Wong K-LA, Doescher J, et al. Impact of weight loss on treatment interruption and unplanned hospital admission in head and neck cancer patients undergoing curative (chemo)radiotherapy in Hong Kong. Support Care Cancer. 2023;31:487. https://doi.or g/10.1007/s00520-023-07952-8.
- Della Valle S, Colatruglio S, La Vela V, Tagliabue E, Mariani L, Gavazzi C. Nutritional intervention in head and neck cancer patients during chemoradiotherapy. Nutrition. 2018;51–52:95–7. https://doi.org/10.1016/j.nut.2017.1 2.012.
- Kramer B, Wenzel A, Boerger M, Lippert B, Feist K, Petrasch R, et al. Long-term quality of life and nutritional status of patients with head and neck cancer. Nutr Cancer 2018:1–14. https://doi.org/10.1080/01635581.2018.1506492.
- Crowder SL, Douglas KG, Yanina Pepino M, Sarma KP, Arthur AE. Nutrition impact symptoms and associated outcomes in post-chemoradiotherapy head and neck cancer survivors: a systematic review. J Cancer Surviv. 2018;12:479–94. https://doi.org/10.1007/s11764-018-0687-7.
- Solís-Martínez O, Plasa-Carvalho V, Phillips-Sixtos G, Trujillo-Cabrera Y, Hernández-Cuellar A, Queipo-García GE, et al. Effect of eicosapentaenoic acid on body composition and inflammation markers in patients with head and neck squamous cell cancer from a public hospital in Mexico. Nutr Cancer. 2018;70:663–70. https://doi.org/10.1080/01635581.2018.1460678.
- Yeh K-Y, Wang H-M, Chang JW-C, Huang J-S, Lai C-H, Lan Y-J, et al. Omega-3 fatty acid-, micronutrient-, and probiotic-enriched nutrition helps body weight stabilization in head and neck cancer cachexia. Oral Surg Oral Med Oral Pathol Oral Radiol. 2013;116:41–8. https://doi.org/10.1016/j.oooo.2013.0 1.015.
- de Castro GS, Andrade MF, Pinto FCS, Faiad JZ, Seelaender M. Omega-3 fatty acid supplementation and its impact on systemic inflammation and body weight in patients with cancer cachexia—a systematic review and metaanalysis. Front Nutr. 2022;8:797513. https://doi.org/10.3389/fnut.2021.797513.

- Newell M, Mazurak V, Postovit LM, Field CJ. N-3 Long-chain polyunsaturated fatty acids, eicosapentaenoic and docosahexaenoic acid, and the role of supplementation during cancer treatment: a scoping review of current clinical evidence. Cancers (Basel). 2021;13:1206. https://doi.org/10.3390/cancers1 3061206.
- Jantharapattana K, Orapipatpong O. Efficacy of EPA-enriched supplement compared with standard formula on body weight changes in malnourished patients with head and neck cancer undergone surgery: a randomized study. Head Neck. 2020;42:188–97. https://doi.org/10.1002/hed.25987.
- Weed HG, Ferguson ML, Gaff RL, Hustead DS, Nelson JL, Voss AC. Lean body mass gain in patients with head and neck squamous cell cancer treated perioperatively with a protein- and energy-dense nutritional supplement containing eicosapentaenoic acid. Head Neck. 2011;33:1027–33. https://doi.org/10.1002/hed.21580.
- 21. Hanai N, Terada H, Hirakawa H, Suzuki H, Nishikawa D, Beppu S, et al. Prospective randomized investigation implementing immunonutritional therapy using a nutritional supplement with a high blend ratio of ω -3 fatty acids during the perioperative period for head and neck carcinomas. Jpn J Clin Oncol. 2018;48:356–61. https://doi.org/10.1093/jjco/hyy008.
- Carvalho TC, Cruz BCS, Viana MS, Martucci RB, Saraiva DCA, Reis PF. Effect of nutritional supplementation enriched with eicosapentaenoic acid on inflammatory profile of patients with oral cavity cancer in antineoplastic pretreatment: a controlled and randomized clinical trial. Nutr Cancer. 2017;69:428–35. https://doi.org/10.1080/01635581.2017.1274406.
- Schmidt N, Møller G, Bæksgaard L, Østerlind K, Stark KD, Lauritzen L, et al. Fish
 oil supplementation in cancer patients. Capsules or nutritional drink supplements? A controlled study of compliance. Clin Nutr ESPEN. 2020;35:63–8. htt
 ps://doi.org/10.1016/j.clnesp.2019.12.004.
- Kazemi-Bajestani SMR, Becher H, Butts C, Basappa NS, Smylie M, Joy AA, et al. Rapid atrophy of cardiac left ventricular mass in patients with non-small cell carcinoma of the lung. J Cachexia Sarcopenia Muscle 2019:jcsm.12451. https://doi.org/10.1002/jcsm.12451.
- ICO-ICSPraxis para el tratamiento medico y con irradiacion del cancer de orofaringe, cavidad oral, hipofaringe, laringe, nasofaringe y carcinoma escamoso cervical de origen desconocido. 2022. Available online at: https://ico.gencat.c at/web/.content/minisite/ico/professionals/documents/arxius/ICO-ICS-Praxi s-CabezaCuello.pdf. Accessed 28 Feb 2025.
- Vermorken JB, Remenar E, van Herpen C, Gorlia T, Mesia R, Degardin M, et al. Cisplatin, fluorouracil, and docetaxel in unresectable head and neck cancer. N Engl J Med. 2007;357:1695–704. https://doi.org/10.1056/NEJMoa071028.
- Pignon J-P, le Maître A, Maillard E, Bourhis J, MACH-NC Collaborative Group. Meta-analysis of chemotherapy in head and neck cancer (MACH-NC): an update on 93 randomised trials and 17, 346 patients. Radiother Oncol. 2009;92:4–14. https://doi.org/10.1016/j.radonc.2009.04.014.
- Bonner JA, Harari PM, Giralt J, Azarnia N, Shin DM, Cohen RB, et al. Radiotherapy plus cetuximab for squamous-cell carcinoma of the head and neck. N Engl J Med. 2006;354:567–78. https://doi.org/10.1056/NEJMoa053422.
- Hitt R, Mesía R, Lozano A, et al. Randomized Phase 3 Noninferiority Trial of Radiotherapy and Cisplatin vs Radiotherapy and Cetuximab After Docetaxel-Cisplatin-Fluorouracil Induction Chemotherapy in Patients With Locally Advanced Unresectable Head and Neck Cancer. Oral Oncol. 2022;134:106087. https://doi.org/10.1016/j.oraloncology.2022.106087.
- Read JA, Beale PJ, Volker DH, Smith N, Childs A, Clarke SJ. Nutrition intervention using an eicosapentaenoic acid (EPA)-containing supplement in patients with advanced colorectal cancer. Effects on nutritional and inflammatory status: a phase II trial. Support Care Cancer. 2007;15:301–7. https://doi.org/10.1007/s00520-006-0153-3.
- Fearon KCH, Barber MD, Moses AG, Ahmedzai SH, Taylor GS, Tisdale MJ, et al. Double-blind, placebo-controlled, randomized study of eicosapentaenoic acid diester in patients with cancer cachexia. J Clin Oncol. 2006;24:3401–7. ht tps://doi.org/10.1200/JCO.2005.04.5724.
- Mourtzakis M, Prado CMM, Lieffers JR, Reiman T, McCargar LJ, Baracos VE. A
 practical and precise approach to quantification of body composition in cancer patients using computed tomography images acquired during routine
 care. Appl Physiol Nutr Metab. 2008;33:997–1006. https://doi.org/10.1139/H0
 8-075.
- Prado CMM, Lieffers JR, McCargar LJ, Reiman T, Sawyer MB, Martin L, et al. Prevalence and clinical implications of sarcopenic obesity in patients with solid tumours of the respiratory and gastrointestinal tracts: a populationbased study. Lancet Oncol. 2008;9:629–35. https://doi.org/10.1016/S1470-20 45(08)70153-0.

Arribas et al. Nutrition Journal (2025) 24:140 Page 13 of 13

- Shen W, Punyanitya M, Wang Z, Gallagher D, St-Onge M-P, Albu J, et al. Total body skeletal muscle and adipose tissue volumes: estimation from a single abdominal cross-sectional image. J Appl Physiol. 2004;97:2333–8. https://doi. org/10.1152/japplphysiol.00744.2004.
- 35. Lee S, Janssen I, Ross R. Interindividual variation in abdominal subcutaneous and visceral adipose tissue: influence of measurement site. J Appl Physiol. 2004;97:948–54. https://doi.org/10.1152/japplphysiol.01200.2003.
- Thoresen L, Frykholm G, Lydersen S, Ulveland H, Baracos V, Prado CMM, et al. Nutritional status, cachexia and survival in patients with advanced colorectal carcinoma. Different assessment criteria for nutritional status provide unequal results. Clin Nutr. 2013;32:65–72. https://doi.org/10.1016/j.clnu.2012. 05.009.
- Ottery FD. Patient-generated subjective global assessment. In: The American Dietetic Association, editors. Clinical Guidelines to Oncology Nutrition. McCallum PD, editor. Chicago: The American Dietetic Association; 2000. p. 11–23.
- Fuchs-Tarlovsky V, Velasco Gimeno C, Arias-Soberón MD, Silva-Sánchez C, Álvarez-Altamirano K, Vedenne-Gutierrez F, et al. Translation, cultural adaptation, and assessment of the linguistic and content validity of the PG-SGA to the Spanish linguistic setting by cancer patients and healthcare professionals. Nutrition. 2024;128: 112567. https://doi.org/10.1016/j.nut.2024.112567.
- Luan C, Kuo L, Wang Y, Liao C, Kang C, Lee Y, et al. Utility of modified Glasgow prognostic score for head and neck squamous cell carcinoma: systematic review and meta-analysis. Head Neck. 2023;45:1856–67. https://doi.org/10.10 02/hed 27397
- Gui L, Cheng M, Zheng M, Ning C, Huo Q. Effects of omega-3 fatty acid supplementation on nutritional status and inflammatory response in patients with stage II-III NSCLC undergoing postoperative chemotherapy: a doubleblind randomized controlled trial. Front Nutr. 2023;10:1266584. https://doi.or g/10.3389/fnut.2023.1266584.
- 41. Mathiowetz V, Weber K, Volland G, Kashman N. Reliability and validity of grip and pinch strength evaluations. J Hand Surg Am. 1984;9:222–6.
- U.S. Department of Health and Human Services, National Institutes of Health, National Cancer Institute. Common Terminology Criteria for Adverse Events (CTCAE) Version 4.0. (v4.03: June 14, 2010). 2009.
- McGrath K. RefBasedMI: Reference-based imputation for longitudinal clinical trials with protocol deviation. R package version 0.2.0. 2024. https://cran.r-project.org/package=RefBasedMI. Accessed 12 Nov 2024.
- 44. Carpenter JR, Roger JH, Kenward MG. Analysis of longitudinal trials with protocol deviation: a framework for relevant, accessible assumptions, and

- inference via multiple imputation. J Biopharm Stat. 2013;23:1352–71. https://doi.org/10.1080/10543406.2013.834911.
- Lenth RV. emmeans: estimated marginal means, aka least-squares means. R package version 1.10.4. 2024. https://cran.r-project.org/package=emmeans. Accessed 12 Nov 2024.
- Hernán MA, Hernández-Díaz S, Robins JM. Randomized trials analyzed as observational studies. Ann Intern Med. 2013. https://doi.org/10.7326/0003-48 19-159-8-201310150-00709.
- 47. Hernán MA, Robins J. Causal inference: what if. Boca Raton: FL, USA; 2020.
- Khal J, Tisdale MJ. Downregulation of muscle protein degradation in sepsis by eicosapentaenoic acid (EPA). Biochem Biophys Res Commun. 2008;375:238– 40. https://doi.org/10.1016/j.bbrc.2008.08.004.
- Yamazaki H, Nishimura M, Uehara M, Kuribara-Souta A, Yamamoto M, Yoshikawa N, et al. Eicosapentaenoic acid changes muscle transcriptome and intervenes in aging-related fiber type transition in male mice. Am J Physiol Metab. 2021;320:E346–58. https://doi.org/10.1152/ajpendo.00184.2020.
- Ghoreishy SM, Zeraattalab-Motlagh S, Amiri Khosroshahi R, Hemmati A, Noormohammadi M, Mohammadi H. Dose-dependent impacts of omega-3 fatty acids supplementation on anthropometric variables in patients with cancer: results from a systematic review and meta-analysis of randomized clinical trials. Clin Nutr Res. 2024;13:186. https://doi.org/10.7762/cnr.2024.13.3.186.
- 51. Barber MD, Fearon KCH, Tisdale MJ, McMillan DC, Ross JA. Effect of a fish oil-enriched nutritional supplement on metabolic mediators in patients with pancreatic cancer cachexia. Nutr Cancer. 2001;40:118–24. https://doi.org/10.1 207/S15327914NC402_7.
- Zhang Y, Shen G, Meng T, Lv Z, Li X, Li J, et al. Eicosapentaenoic acid enhances the sensitivity of osteosarcoma to cisplatin by inducing ferroptosis through the DNA-PKcs/AKT/NRF2 pathway and reducing PD-L1 expression to attenuate immune evasion. Int Immunopharmacol. 2023;125: 111181. https://doi.or g/10.1016/j.intimp.2023.111181.
- Roeland EJ, Bohlke K, Baracos VE, Bruera E, Del Fabbro E, Dixon S, et al. Management of cancer cachexia: ASCO Guideline. J Clin Oncol. 2020;38:2438–53. https://doi.org/10.1200/JCO.20.00611.

Publisher's Note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.